

Thrombocytosis and megakaryocyte changes associated with PRCA

Tracking no: ADV-2023-012309R1

Joelle Abdallah (University of Texas Southwestern Medical School, United States) Robert Williams (University of Texas Southwestern Medical School, United States) Hussein Awada (Department of Translational Hematology and Oncology Research, Taussig Cancer Institute, Cleveland Clinic, United States) Ganesh Raman (University of Texas Southwestern Medical School, United States) Yusuf Ozcan (University of Texas Dallas, United States) Mark Orland (Department of Internal Medicine, Cleveland Clinic, United States) Mutlu Mete (Texas A&M University-Commerce, United States) Weina Chen (UT Southwestern Medical Center at Dallas, United States) Carmelo Gurnari (Taussig Cancer Institute, Cleveland Clinic, United States) Jaroslaw Maciejewski (Cleveland Clinic, United States) Taha Bat (University of Texas Southwestern, United States)

Abstract:

Conflict of interest: No COI declared

COI notes:

Preprint server: No;

Author contributions and disclosures: J.A., G.R.W and T.B. generated and conceived the study design. J.A., G.R.W, G.R., M.M., Y.O., W.C., and T.B. contributed to the table and manuscript; C.G., H.A., and J.P.M. reviewed the clinical data, took part in patients' selection and helped with writing the manuscript; J.A., G.R.W, G.R., M.M., Y.O., H.A., M.O., W.C., C.G., J.P.W., and T.B. reviewed clinical data and contribute to writing of this manuscript. All authors participated in data interpretation and critical review of the final paper and submission. All authors have read and agreed to the published version of the manuscript.

Non-author contributions and disclosures: No;

Agreement to Share Publication-Related Data and Data Sharing Statement: For original data, please contact taha.bat@utsouthwestern.edu

Clinical trial registration information (if any):

1 **Title: Thrombocytosis and megakaryocyte changes associated with PRCA**

2 **Running Title:** PRCA-associated thrombocytosis

3 Joelle Abdallah ^{1*}, Robert George Williams ^{1*}, Hussein Awada ², Ganesh Raman ¹, Yusuf Ozcan ³, Mark
4 Orland ², Mutlu Mete ⁴, Weina Chen ⁵, Carmelo Gurnari ^{2,6}, Jaroslaw P. Maciejewski ², Taha Bat ¹

5 1 Division of Hematology-Oncology, UT Southwestern Medical Center, Dallas, TX, USA

6 2 Translational Hematology and Oncology Research, Cleveland Clinic, Cleveland, OH 44195, USA

7 3 University of Texas Dallas, Dallas TX, USA

8 4 Department of Computer Science, Texas A&M University - Commerce, Commerce, TX, USA

9 5 Department of Pathology, UT Southwestern Medical Center, Dallas, TX, USA

10 6 Department of Biomedicine and Prevention, University of Rome Tor Vergata, 00133 Rome, Italy

11 *J.A. and R.G.W contributed equally to this manuscript

12 **Correspondence:**

13 Taha Bat, MD

14 Division of Hematology-Oncology, UT Southwestern Medical Center, Dallas, TX

15 5323 Harry Hines Blvd. Dallas, TX 75390-9255

16 Office: 214-648-4943|Fax: 214-648-4105

17 **Data Sharing statement**

18 All data are presented in the paper. Requests for additional information should be sent to the
19 corresponding author.

20

21 Orcid: 0000-0002-9879-3402

22 Text word count: 1195

23 Figure count: 1

24 Table count: 2

25 Reference count: 25

26 Primary category: Red cells, iron, and erythropoiesis

27 Secondary category: Platelets and thrombopoiesis

28

29

30 *J.A. and R.G.W contributed equally to this manuscript

31 Acquired pure red cell aplasia (PRCA) is a rare hematological disorder that results from failure of
32 erythropoiesis [1, 2] and can be distinguished from other bone marrow failure disorders by
33 reticulocytopenia and normal granulopoiesis and megakaryopoiesis[3, 4]. PRCA is often idiopathic and

34 likely due to cytotoxic T-cell mediated destruction of early erythroid precursors. Indeed, it can be
35 associated with conditions such as T-cell Large Granular Lymphocytic (T-LGL) Leukemia, B-cell
36 dyscrasia, thymoma, immunodeficiency, and infections[5-8].

37 Thrombocytosis refers to the abnormal elevation in platelet count of $>450,000/\mu\text{L}$. This condition can be
38 related to a primary process, usually associated with a myeloproliferative neoplasm known as essential
39 thrombocythemia, but more commonly presents as an epiphenomenon of other causes that include iron
40 deficiency, chronic inflammatory conditions, and asplenia[9, 10]. Thrombocytosis has not been associated
41 with PRCA. Herein, we report a cohort of PRCA patients with thrombocytosis that have not been defined
42 in the literature.

43 A retrospective analysis was conducted on patient records diagnosed with PRCA at the University of
44 Texas Southwestern and Cleveland Clinic Foundation between 2000 and 2022. The primary objective was
45 to investigate the presence of thrombocytosis and/or megakaryocyte changes in these PRCA cases.
46 Clinical, laboratory, and molecular data were abstracted, adhering to the guidelines established by the
47 Declaration of Helsinki and the respective participating institutions.

48 Among a total of 90 patients, comprehensive analysis of 27 cases were identified as having acquired
49 PRCA with thrombocytosis and/or megakaryocyte changes noted on bone marrow biopsies (Table 1)
50 examined. To assess treatment efficacy, hemoglobin and platelet count before and after resolution of
51 PRCA were compared. A Shapiro-Wilk test was used to assess normality. For normally distributed data, a
52 paired t-test was performed. For non-normally distributed data, a Wilcoxon matched-pairs signed rank
53 test was performed. For all relevant comparisons, a p-value < 0.05 was used to set statistical significance.

54
55
56 In a cohort of 90 PRCA patients, we identified 27 individuals with concurrent thrombocytosis and/or
57 megakaryocyte changes (Table 1).

58 Of the 27 patients, the mean hemoglobin count at diagnosis was 7.6 ± 0.7 g/dL, and the mean platelet
59 count at diagnosis was $389,000 \pm 80,400/\mu\text{L}$. 51.9% were female. 20 demonstrated megakaryocyte
60 hyperplasia, 4 demonstrated hyper-lobation, and 1 displayed megakaryocyte dysplasia. Three patients
61 tested positive for parvovirus on polymerase chain reaction, one patient had Chronic Lymphocytic
62 Leukemia, and ten patients had LGL leukemia.

63 To delve deeper into the potential mechanisms linking PRCA and thrombocytosis, we then examined
64 clinical variables in patients exhibiting thrombocytosis ($n=7$) following clinical response of their PRCA.
65 This was defined as the achievement of a hemoglobin level > 9 g/dL with red blood cell transfusion
66 independency. We excluded four patients due to unresolved hemoglobin levels or insufficient chart
67 information. Bone marrow biopsies at the time of resolution were not available for any of the examined
68 patients, as this procedure is generally not clinically indicated at this timepoint.

69 Among the subset of 7 patients with PRCA resolution, the mean hemoglobin count at diagnosis was $8.5 \pm$
70 1.4 g/dL, and the mean platelet count was $576,300 \pm 71,800$ platelets/ μL . Post-resolution of PRCA, the
71 mean hemoglobin count increased to 11.3 ± 1.3 g/dL, and the mean platelet count decreased to $431,600 \pm$
72 $78,600$ platelets/ μL . Statistical analysis revealed significant differences in mean hemoglobin and mean
73 platelet count before and after resolution, with p-values of 0.0225 and 0.0120, respectively. Interestingly,
74 two of seven patients still exhibited thrombocytosis even after resolution of their PRCA.

75 Next, the mean EPO levels of those patients with thrombocytosis were examined. Of the 7 patients who
76 had EPO levels drawn at the time of diagnosis, all had levels well over the upper limit of normal (26
77 mU/mL), 2476.5 +/- 2784.7 (range 72.4-10815) mU/mL.

78 In this study, we provide a detailed characterization of a cohort of patients with PRCA exhibiting notable
79 alterations in megakaryocytes. Our cohort is heterogenous, and the etiology of each patient's PRCA is
80 different. Therefore, there may not be one mechanism by which PRCA is linked to thrombocytosis.
81 However, we theorize that there are two potential mechanisms that may explain the phenomenon in some
82 cases.

83 We first hypothesize that the scarcity of erythroid production may redirect hematopoietic precursors
84 towards the megakaryocytic lineage. The process of hematopoiesis, encompassing both erythroid and
85 megakaryocyte lineages arising from a bipotential MEP, is well-documented[11-15]. We suggest that the
86 interruption of erythroid differentiation at a critical stage may funnel differentiation towards the
87 megakaryocytic lineage (Figure 1, I). Our findings lend support to this hypothesis, as the initial
88 thrombocytosis observed prior to treatment resolves upon PRCA resolution with treatment.

89 A similar phenomenon has been reported in the context of iron-deficient anemia leading to secondary
90 thrombocytosis, albeit the precise underlying mechanism remains elusive[16]. One proposed mechanism
91 posits that once the hematopoietic growth factors and cytokines required for erythrocyte development
92 become available, differentiation of the MEP cell veers away from the megakaryocyte lineage and reverts
93 towards the erythroid lineage. This skewing may be attributed to heightened MKL1 expression[14],
94 transcription factors determining the megakaryocyte lineage [18], or an increase in thrombopoietin, stem
95 cell factor, stromal-derived factor 1, or cytokines known to exert thrombopoietic effects[18].
96 Additionally, one study found that low iron in the bone marrow environment can bias MEP differentiation
97 towards the megakaryocyte lineage via a reduction in ERK signaling[19]. It is plausible that one or more
98 of these mechanisms may account for the PRCA-related thrombocytosis we have observed, although
99 further research is warranted for a comprehensive understanding.

100 In our cohort, many patients for whom EPO levels were measured at the time of PRCA diagnosis
101 exhibited levels well above the normal range. Therefore, we hypothesize that EPO may play a role in the
102 development of thrombocytosis (Figure 1, II). EPO signaling has been shown to have a synergistic effect
103 with TPO on thrombopoiesis[20]. A study involving TPO-knockout mice demonstrated that EPO exerts a
104 direct and TPO-independent influence on late-stage thrombopoiesis, resulting in increased production of
105 large platelets [21]. Furthermore, several human studies in healthy volunteers, uremic patients, and
106 chronic liver disease patients have reported significant short-term increases in platelet count following
107 EPO injections[22-25]. Thus, it is conceivable that, in certain PRCA patients, synergistic signaling
108 between EPO and TPO may be occurring upstream at the level of the bipotential MEP cell, leading to
109 increased megakaryocyte production and thrombocytosis.

110 The implications of thrombocytosis and megakaryocyte alterations in the context of PRCA remain
111 enigmatic, necessitating comprehensive datasets to unravel their role in the pathophysiology of the
112 disease and their potential as prognostic markers for treatment response. It is important to acknowledge
113 the limitations of this study, including a modest sample size secondary to the rarity of the disorder, and
114 the absence of post-treatment bone marrow biopsy results, which would have provided visual
115 confirmation of resolved megakaryocyte abnormalities.

116 Our findings in aggregate suggest that perhaps the definition of PRCA be expanded to include those with
117 megakaryocyte changes or thrombocytosis. Further research is warranted to establish a definitive
118 correlation and assess whether the course and response of PRCA to standard immunosuppressive

119 treatment differ in cases with and without thrombocytosis. Future investigations should also encompass
120 an examination of megakaryocyte hyperplasia and other abnormalities, such as hyper- and hypo-lobation,
121 associated with PRCA.

This study was approved by the institutional review boards of both the University of Texas
Southwestern Medical Center and Cleveland Clinic Foundation. Clinical, laboratory, and
molecular data were meticulously abstracted, adhering to the guidelines established by the
Declaration of Helsinki and the respective participating institutions.

122

123 **Author Contributions**

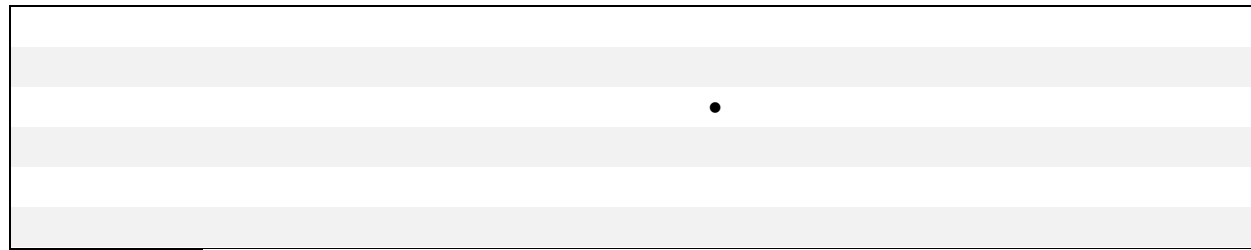
124 J.A., G.R.W and T.B. generated and conceived the study design. J.A., G.R.W, G.R., M.M., Y.O., W.C.,
125 and T.B. contributed to the table and manuscript; C.G., H.A., and J.P.M. reviewed the clinical data, took
126 part in patients' selection and helped with writing the manuscript; J.A., G.R.W, G.R., M.M., Y.O., H.A.,
127 M.O., W.C., C.G., J.P.W., and T.B. reviewed clinical data and contribute to writing of this manuscript.
128 All authors participated in data interpretation and critical review of the final paper and submission. All
129 authors have read and agreed to the published version of the manuscript.

130 **Disclosure of conflicts of interest**

131 The authors declare no conflicts of interest.

132 **References**

- 133 1. Gurnari, C. and J.P. Maciejewski, *How I manage acquired pure red cell aplasia in adults*. Blood,
134 2021. **137**(15): p. 2001-2009.
- 135 2. Sawada, K., N. Fujishima, and M. Hirokawa, *Acquired pure red cell aplasia: updated review of*
136 *treatment*. Br J Haematol, 2008. **142**(4): p. 505-14.
- 137 3. Means, R.T., Jr., *Pure red cell aplasia*. Blood, 2016. **128**(21): p. 2504-2509.
- 138 4. Ulirsch, J.C., et al., *The Genetic Landscape of Diamond-Blackfan Anemia*. Am J Hum Genet, 2018.
139 **103**(6): p. 930-947.
- 140 5. Xavier, R.D., et al., *Thymoma associated with pure red cell aplasia: a case report and literature*
141 *review*. Indian J Thorac Cardiovasc Surg, 2020. **36**(4): p. 404-408.
- 142 6. Frickhofen, N., et al., *Parvovirus B19 as a cause of acquired chronic pure red cell aplasia*. Br J
143 Haematol, 1994. **87**(4): p. 818-24.
- 144 7. Teague, D., et al., *Hepatitis C Infection Associated with Acquired Pure Red Cell Aplasia*. Trop Med
145 Infect Dis, 2022. **8**(1).
- 146 8. Lee, N.C.J., et al., *SARS-CoV-2 infection associated with aplastic anemia and pure red cell aplasia*.
147 Blood Adv, 2022. **6**(13): p. 3840-3843.
- 148 9. Rokkam, V.R. and R. Kotagiri, *Secondary Thrombocytosis*, in *StatPearls*. 2022: Treasure Island
149 (FL).
- 150 10. Evstatiev, R., et al., *Iron deficiency alters megakaryopoiesis and platelet phenotype independent*
151 *of thrombopoietin*. Am J Hematol, 2014. **89**(5): p. 524-9.
- 152 11. Hattangadi, S.M., et al., *From stem cell to red cell: regulation of erythropoiesis at multiple levels*
153 *by multiple proteins, RNAs, and chromatin modifications*. Blood, 2011. **118**(24): p. 6258-68.
- 154 12. Lodish, H., J. Flygare, and S. Chou, *From stem cell to erythroblast: regulation of red cell*
155 *production at multiple levels by multiple hormones*. IUBMB Life, 2010. **62**(7): p. 492-6.
- 156 13. Socolovsky, M., H.F. Lodish, and G.Q. Daley, *Control of hematopoietic differentiation: lack of*
157 *specificity in signaling by cytokine receptors*. Proc Natl Acad Sci U S A, 1998. **95**(12): p. 6573-5.



	<i>Megakaryocyte changes only</i>	<i>Thrombocytosis only</i>	<i>Both</i>	<i>Megakaryocyte changes and/or thrombocytosis</i>	<i>Neither (PRCA in the absence of thrombocytosis or megakaryocyte changes)</i>	Total
Number of patients	16	4	7	27	63	90
Mean age at diagnosis	57.6 +/- 8.3 (33.8-82)	68 +/- 16.1 (45.3-84.3)	53.5 +/- 11.2 (28-73.9)	58.1 +/- 6.2 (28-84.3)	56.7 +/- 5.1 (0-85)	57.1 +/- 4 (0-85)
Patients with BM Megakaryocyte changes						
• Dysplasia	1	0	0	1	0	1
• Hyperlobation	3	0	1	4	0	4
• Hyperplasia	13	0	7	20	0	20
• Total with any changes	16	0	7	23	0	23
Mean hemoglobin at diagnosis, g/dL	7.6 +/- 0.9 (3.5-9.6)	6.4 +/- 1.3 (4.5-7.4)	8.3 +/- 1.6 (5.1-12.4)	7.6 +/- 0.7 (3.5-12.4)	7.7 +/- 0.6 (2.6-11.7)	7.6 +/- 0.5 (2.6-12.4)
Mean platelet count, 100,000 platelets/ μ L	232.7 +/- 70.3 (9-440)	550.8 +/- 96.6 (476-695)	564.7 +/- 59.2 (472-693)	389 +/- 80.4 (9-695)	254.2 +/- 26.7 (45-432)	299.1 +/- 35.2 (9-695)
Mean ferritin at diagnosis, μ g/L	2309.4 +/- 1750.2 (648.4-6579)	2254.9 +/- 2350.7 (434.1-5505)	705 +/- 310.1 (315-1569)	1635.9 +/- 861.5 (315-6579)	2021 +/- 583.6 (50.1-7597)	1899.8 +/- 481 (50.1-7597)
Mean WBC at diagnosis, 1,000 WBC/ μ L	8.1 +/- 2.7 (3.9-18.8)	8.3 +/- 0.7 (7.8-9.4)	8 +/- 1.8 (3.3-10.3)	8.1 +/- 1.5 (3.3-18.8)	5.6 +/- 0.7 (2-12.6)	6.4 +/- 0.7 (2-18.8)
Female (% of total)	6 (37.5%)	2 (50%)	6 (85.7%)	14 (51.9%)	29 (46.0%)	43 (47.8%)
Mean bone marrow cellularity, %	55.3 +/- 10.9 (25-90)	53.8 +/- 26.9 (40-95)	73.6 +/- 8.2 (50-80)	60 +/- 8.2 (25-95)	39.6 +/- 5.4 (5-90)	1973.2 +/- 745.8 (46.6-11560)
Next generation sequencing findings	•STAT3 p.S614R c.1840A>C 8 •IDH1, SETBP1	•SF3B1 p.K666N 18.21% •BCOR p.1252_1253del 9%	•c.490G>T p.G164C 41.7%, c.6460G>A p.D2154N 41.4%, c.3974A>G p.K1325R 40.6%, c.588_589insACCCGC p.P196_P197insTR 13.0% •NF-kappaB2, JAK2, TYK 2 •c.490G>T p.G164C 43.2%, c.1292C>T p.P431L 41.7%, c.6460G>A p.D2154N 41.4%, c.3974A>G p.K1325R 40.6%, c.588_589insACCCGC p.P196_P197insTR 13.0%	•c.490G>T p.G164C 43.2%, c.1292C>T p.P431L 41.7%, c.6460G>A p.D2154N 41.4%, c.3974A>G p.K1325R 40.6%, c.588_589insACCCGC p.P196_P197insTR 13.0%	•SPTB, SPTA1, EPB42 •TET2 p.N275Iifs* 4.3% •RPS19, HFE C282Y, H63D •PB, ASXL1, U2AF1 •ASXL1, JAK2, STAT3, U2AF1 VUS •STAT3 p.D661Y VAF 12.7% •ASXL1 p.K912Q c.2734A>C VAF 50.3% and PTPN11 p.K131R c.392A>G 51.1% VAF •N6471 mutation in STAT3 gene	
Mean absolute reticulocyte count at diagnosis, reticulocytes/ μ L	2.924 +/- 3.162 (0.01-10)	0.013 +/- 0.004 (0.011-0.015)	3.685 +/- 3.322 (0.007-8.9)	2.84 +/- 1.987 (0.007-10)	1.45 +/- 0.884 (0-7.1)	1.894 +/- 0.881 (0-10)
Mean reticulocyte percent at diagnosis, %	0.684 +/- 0.647 (0.3-2)	0.543 +/- 0.283 (0.3-0.8)	0.263 +/- 0.084 (0.04-0.4)	0.459 +/- 0.231 (0.04-2)	0.5 +/- 0.191 (0.1-2.1)	0.487 +/- 0.149 (0.04-2.1)
Mean erythropoietin level, mU/mL	748.6 +/- 532.1 (46.6-1646)	2012 (n=1)	2553.9 +/- 3290 (72.4-10815)	1679 +/- 1542.6 (46.6-10815)	2105 +/- 845.3 (91-11560)	1973.2 +/- 745.8 (46.6-11560)

Parvovirus B19 Chronic Lymphocytic Leukemia	0	2	1	3	3	6
Large granular lymphocyte leukemia	0	1	0	1	1	2
	4	2	4	10	15	25

189

190 Table 2: Subset of 7 patients who had resolution of transfusion dependency

VARIABLE	MEAN BEFORE RESOLUTION ± CI	MEAN AFTER RESOLUTION ± CI	P
Platelet count, 100,000 platelets/μL	576.3 ± 71.8	431.6 ± 78.6	0.0120
Hemoglobin, g/dl	8.5 ± 1.4	11.3 ± 1.3	0.0225

191

192 **Figure legends**

193 Figure 1: **Proposed mechanism for PRCA-related thrombocytosis.** Abbreviations: MPP- multipotent
 194 progenitor; CFU- colony-forming unit; BFU- burst-forming unit; GEMM- granulocyte-erythrocyte-
 195 monocyte-megakaryocyte. GM- granulocyte-monocyte; Eo- eosinophil; E- erythrocyte; MEG-
 196 megakaryocyte; MEP- megakaryocyte-erythrocyte progenitor. EPO- Erythropoietin. TPO-
 197 Thrombopoietin. **(I)** Proposed mechanism I: Decreased erythropoiesis shunts differentiation toward
 198 megakaryocyte lineage. **(II)** Proposed mechanism II: Increased EPO due to PRCA leads to synergistic
 199 signaling between EPO and TPO at the bipotential MEP, stimulates TPO receptors and resulting in
 200 increaseds megakaryocyte formation. Created with BioRender.com.

