

Matched control analysis suggests R-CHOP followed by (R)-ICE may improve outcome in non-GCB DLBCL compared to R-CHOP

Tracking no: ADV-2023-011408R1

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Abstract:

Rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone (R-CHOP) is considered the standard-of-care for patients with advanced-stage diffuse large B-cell lymphoma (DLBCL), despite findings that non-germinal center B-cell-like (non-GCB) patients have significantly worse outcome with this regimen. We evaluated the prognostic significance of baseline risk factors, including cell of origin (COO) classified by the Hans algorithm, within an alternative chemoimmunotherapy program. At Memorial Sloan Kettering Cancer Center (MSK), 151 patients with DLBCL received sequential R-CHOP induction and (R)-ICE (rituximab, ifosfamide, carboplatin, and etoposide) consolidation. Outcome analysis based on COO was validated with a propensity score matched cohort treated with R-CHOP from the Mayo Clinic component of the Molecular Epidemiology Resource (MER). Among the GCB (n=69) and non-GCB (n=69) patients at MSK, event-free survival (EFS) of non-GCB was superior to that of GCB (HR 0.53, 95% CI 0.29-0.98). Overall survival (OS) demonstrated an association in the same direction but was not statistically significant (HR 0.68, 95% CI 0.33-1.42). Propensity score matched patients from MSK (n=108) demonstrated a small attenuation in the HRs for EFS (HR 0.57, 95% CI 0.27-1.18) and OS (HR 0.76, 95% CI 0.33-1.79) and were no longer statistically significant. In contrast, the matched MER cohort (n=108) demonstrated an EFS association (HR 1.17, 95% CI 0.70-1.95) and OS association (HR 1.13, 95% CI 0.64-2.00) in the opposite direction, but were also not statistically significant. R-CHOP induction and (R)-ICE consolidation may overcome the negative prognostic impact of the non-GCB phenotype, per the Hans algorithm, and can be preferentially selected for this population.

Conflict of interest: COI declared - see note

COI notes: M.J. Maurer reports research funding from Roche/Genentech, BMS, and GenMab, and serves on the advisory board of GenMab, and Adaptive Biotechnologies. J.T.F. is a full-time employee of Histowiz. M.J. Matasar reports compensation from Epizyme, Genentech, Roche, Kite, Bayer, Seagen, Celgene, ADC Therapeutics, IMV Therapeutics, and AstraZeneca. A.M. receives research support from Seattle Genetics, Merck, Bristol-Myers Squibb, and Incyte and receives honorarium from Kyowa Hakko Kirin Pharma, Miragen Therapeutics, Takeda Pharmaceuticals, ADC Therapeutics, Seattle Genetics, Cell Medica, Bristol-Myers Squibb, and Erytech Pharma. D.J.S. receives compensation from InPractice Elsevier and Seattle Genetics and is on speakers bureau for Medical Crossfire. A.N. received honoraria from Janssen, Pharmacylics, and Prime Oncology; consults for Medscape; serves on advisory board for Janssen; serves on the speakers bureau for Prime Oncology; and receives research funding for Rafael Pharma and Pharmacylics. S.M.H. received research funding from ADCT therapeutics, Aileron, Forty-Seven, Verastem, Kyowa Hakko Kirin, Millennium Pharmaceuticals Inc, Celgene, Trillium, and Daiichii Sankyo and consults for Astex, Affimed, Merck Sharp and Dome, Kyowa Hakko Kirin Pharma, Corvus Pharmaceuticals Inc., Celgene, Portola Pharmaceuticals, Takeda Millennium, Innate Pharma, Verastem, Miragen Therapeutics Inc, Seattle Genetics, and ADCT. P.A.H. receives research support from Portola, Novartis/GSK, Molecular Templates, and Janssen Pharmaceuticals and served as consultant for Karyopharm, Juno, Portola, Celgene, and AstraZeneca. T.M.H. reports research support from Genentech, and Sorrento, serves on the data monitoring committee of Seagen, Tess Therapeutics, and Eli Lilly & Co., and the scientific advisory board of Eli Lilly & Co., Morphosys, Incyte, Biegene, and Loxo Oncology. G.A.S. reports advisory boards/consulting fees from AbbVie, Atbtherapeutics, Bayer, BeiGene, Bristol Myers Squibb/Celgene, Debiopharm, Epizyme, F. Hoffmann-La Roche Ltd./Genentech, Inc., Genmab, Incyte, Ipsen, Janssen, Kite/Gilead, Loxo/Lilly, Molecular Partners, MorphoSys, Nordic Nanovector, Novartis, Regeneron, and Takeda, and is a shareholder of Owkin. C.H.M. has held an employment/leadership position/advisory role for Celgene, Genentech, Merck & Co., Seattle Genetics Inc., participated in an advisory board for Molecular Templates, and has received research funding from Pharmacylics, Genentech, Merck & Co., Inc., and Seattle Genetics Inc. A.D.Z. received research grants from AbbVie, Adaptive Biotechnologies, Bristol Myers Squibb, BeiGene, Genentech/Roche, and MEI Pharma, consulting fees from Amgen, AstraZeneca, BeiGene, Genentech/Roche, Janssen, JUNO/Celgene/Bristol Myers Squibb, Kite/Gilead, MEI Pharma, Pfizer, Pharmacylics, and Sandoz/Novartis, and serves on the scientific advisory board of Adaptive Biotechnologies, Lymphoma Research Foundation. The remaining authors declare no competing financial interests.

Preprint server: No;

Author contributions and disclosures: Contribution: K.S.B. and A.D.Z. designed the study; M.J. Matasar, A.J.M., D.J.S., A.N., M.L.P., S.M.H., P.A.H., C.S.P., J.R.C., T.M.H., G.A.S., G.S.N., C.H.M., and A.D.Z. provided study materials and patients; K.S.B., A.N.S., M.J. Maurer, J.R.C., T.M.H., G.S.N., and A.D.Z. coordinated and collected data; K.S.B., A.N.S., M.J. Maurer, J.R.C., G.S.N., and A.D.Z. analyzed and interpreted the data; K.S.B. and A.D.Z wrote the manuscript; all authors contributed to the final approval of the manuscript.

Non-author contributions and disclosures: No;

Agreement to Share Publication-Related Data and Data Sharing Statement: Data-sharing requests may be e-mailed to the corresponding author, Andrew D. Zelenetz, at zeleneta@mskcc.org.

Clinical trial registration information (if any): NCT00039195 and NCT00712582. Registered at ClinicalTrials.gov

Matched control analysis suggests R-CHOP followed by (R)-ICE may improve outcome in non-GCB DLBCL compared to R-CHOP

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Research support for the study:

MSK funding: CORE grant P30 CA008748 and Lymphoma SPORE grant 5 P50 CA192937

MER Funding: P50 CA97274 and U01 CA195568.

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Running Title: R-CHOP > (R)-ICE improves outcome in non-GCB DLBCL

Text Word Count: 3379 words (limit: 4000 words)

Abstract Word Count: 250 words (limit: 250 words)

Number of Figures and Tables: 2 Tables and 2 Figures (limit: 7 total)

Number of References: 50 references (limit 100 references)

Key Points:

- Intensified non-cross-resistant sequential R-CHOP followed by (R)-ICE may improve outcome in non-GCB DLBCL.
- The immunohistochemistry-based Hans algorithm may be used to stratify patients with DLBCL into prognostic subgroups.

Abstract

Rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone (R-CHOP) is considered the standard-of-care for patients with advanced-stage diffuse large B-cell lymphoma (DLBCL), despite findings that non-germinal center B-cell-like (non-GCB) patients have significantly worse outcome with this regimen. We evaluated the prognostic significance of baseline risk factors, including cell of origin (COO) classified by the Hans algorithm, within an alternative chemoimmunotherapy program. At Memorial Sloan Kettering Cancer Center (MSK), 151 patients with DLBCL received sequential R-CHOP induction and (R)-ICE (rituximab, ifosfamide, carboplatin, and etoposide) consolidation. Outcome analysis based on COO was validated with a propensity score matched cohort treated with R-CHOP from the Mayo Clinic component of the Molecular Epidemiology Resource (MER). Among the GCB (n=69) and non-GCB (n=69) patients at MSK, event-free survival (EFS) of non-GCB was superior to that of GCB (HR 0.53, 95% CI 0.29-0.98). Overall survival (OS) demonstrated an association in the same direction but was not statistically significant (HR 0.68, 95% CI 0.33-1.42). Propensity score matched patients from MSK (n=108) demonstrated a small attenuation in the HRs for EFS (HR 0.57, 95% CI 0.27-1.18) and OS (HR 0.76, 95% CI 0.33-1.79) and were no longer statistically significant. In contrast, the matched MER cohort (n=108) demonstrated an EFS association (HR 1.17, 95% CI 0.70-1.95) and OS association (HR 1.13, 95% CI 0.64-2.00) in the opposite direction, but were also not statistically significant. R-CHOP induction and (R)-ICE consolidation may overcome the negative prognostic impact of the non-GCB phenotype, per the Hans algorithm, and can be preferentially selected for this population. NCT00039195 and NCT00712582.

Registered at ClinicalTrials.gov

Introduction

Diffuse large B-cell lymphoma (DLBCL) is a heterogeneous group of B-cell lymphomas. It is the most common type of non-Hodgkin lymphoma, accounting for 30-40% of new diagnoses.^{1,2} Advanced-stage DLBCL is highly variable in its clinical behavior, and treatment with rituximab plus cyclophosphamide, doxorubicin, vincristine and prednisone (R-CHOP) is considered the standard for first-line therapy. However, some patients experience disease recurrence or refractory disease soon after R-CHOP, and numerous chemoimmunotherapy regimens have attempted to improve event-free survival (EFS) and overall survival (OS) in patients with these aggressive lymphomas.³⁻⁶ Strategies to improve outcome include risk-adapted therapies based on baseline prognostic factors that stratify patients into favorable versus less favorable groups.

DLBCL is a diagnostic category that includes morphologically similar tumors based on histology. Gene expression profiling (GEP) through microarray analysis has revealed three molecularly distinct subtypes: germinal center B cell-like (GCB), activated B-cell-like (ABC), and un-classifiable.^{7,8} These three DLBCL subtypes involve different oncogenic events and varying consequent prognoses, independent of the International Prognostic Index (IPI) score.^{9,10} However, the expensive technology and limited availability in clinical laboratories make its use impractical for many patients with DLBCL needing therapy. Thus, several surrogates using immunohistochemistry (IHC) algorithms have been developed to approximate GEP. The Hans model, which uses combined immunostaining of CD10, BCL6, and MUM1¹¹ to translate the ABC subtype into a similarly behaving group referred to as non-germinal center B-cell (non-GCB)

subtype, is the most widely used means of determining cell of origin (COO) in the real world. It has been validated in several studies as predictive of less favorable outcome in non-GCB patients compared to GCB patients when treated with R-CHOP,^{12,13} just as with the GEP classification gold standard. The prognostic difference of COO determined by IHC has not been universally reproducible,¹⁴⁻¹⁷ it remains a simple and more accessible method of classifying DLBCL into prognostically significant subgroups, showing a reasonably high concordance with GEP (86%).¹²

R-CHOP has been considered the standard-of-care frontline treatment for patients with advanced-stage DLBCL regardless of COO in the US, as reflected in the National Comprehensive Cancer Network guidelines,¹⁸ despite findings that non-GCB patients have significantly worse outcome with this regimen than GCB patients.^{9,12,13,19-22} Recent results of the POLARIX trial suggest that substitution of polatuzumab vedotin for vincristine may improve the outcome of patients with ABC DLBCL.²³ Findings from the LNH03-2B trial by the Groupe d'Etudes des Lymphomes de l'Adulte (GELA) demonstrated improved EFS and OS among non-GCB patients treated with R-ACVBP, an induction / consolidation immunochemotherapy program (see Supplemental Table 1), compared with R-CHOP, but no impact among GCB patients.²⁴

We previously conducted two highly-related phase II trials of sequential R-CHOP followed by (R)-ICE demonstrating excellent long-term outcome in first-line treatment of DLBCL.^{25,26} However, this treatment was associated with increased toxicity compared to R-CHOP, so we undertook subsequent analysis to determine if there was a sub-group

which had particular benefit to this regimen. R-CHOP followed by (R)-ICE is strikingly similar to the R-ACVBP induction followed by non-cross-resistant consolidation regimen in the LNH03-2B trial, leading to the hypothesis that R-CHOP followed by (R)-ICE may have differential outcome based on COO. To examine this question, we undertook a propensity score matched analysis comparing the MSK patients treated with R-CHOP > (R)-ICE with patients treated with R-CHOP from the Mayo Clinic component of the University of Iowa / Mayo Clinic Lymphoma Specialized Program of Research Excellence (SPORE) Molecular Epidemiology Resource (MER).²⁷

Methods

Key Eligibility Criteria

Two risk-adapted phase II studies treating patients with advanced-stage large cell lymphomas were approved by the MSK Institutional Review Board. Patients were required to have 1 to 3 adverse risk factors according to the age-adjusted IPI (aaIPI)²⁸. All patients were suitable to undergo stem cell rescue. In the first study, Protocol 01-142²⁵, eligible patients had a histologic diagnosis of CD20+ DLBCL or primary mediastinal B-cell lymphoma (PMBL). In the second study, MSK Protocol 08-026²⁶, eligible patients were diagnosed with CD20+ DLBCL, PMBL, or follicular lymphoma grade 3B (FL3B). For both studies, patients were not excluded if the bone marrow demonstrated involvement by small-cleaved cell lymphoma, and all patients had measurable disease by positron emission tomography with [¹⁸F]fluorodeoxyglucose (FDG-PET) scans, normal baseline cardiac function, serum creatinine ≤ 1.5 mg/dL (or creatinine clearance > 60 mL/min), absolute neutrophil count > 1000/μL, and platelets >

50,000/ μ L. Patients had to be hepatitis B surface antigen-negative and Hepatitis C negative. Exclusion criteria included known pregnancy or breast-feeding, human immunodeficiency virus infection, and central nervous system involvement.

Pathology Review and Cell of Origin Assessment

The department of hematopathology at MSK confirmed histologic diagnoses of DLBCL, PMBL, and FL3B, and classified DLBCL COO subtype using the IHC-based algorithm developed by Hans et al.¹¹ Paraffin-embedded tumor cells were stained with antibodies to CD10, BCL6 and MUM1, and cases were considered positive for an antigen if $\geq 30\%$ of the tumor cells were stained with that antigen. DLBCL cases were classified into 2 subtypes: GCB and non-GCB. GCB subtype was defined as any one of the following: CD10+ alone; both CD10+ and BCL6+; CD10-, BCL6+, and MUM1-. Non-GCB subtype was defined as any one of the following: both CD10- and BCL6-; CD10-, BCL6+, and MUM1+; negative for all three antigens.

Treatment

Treatment has been previously described in 01-142²⁵ and 08-026²⁶ (Supplemental Figure 1). In 01-142 (NCT00039195), initial therapy consisted of 4 cycles of R-CHOP-14 induction followed by FDG-PET. Patients who were FDG-PET negative received 3 cycles of ifosfamide, carboplatin, etoposide (ICE) consolidative chemotherapy, and those who were FDG-PET positive received consolidative chemotherapy of 2 cycles of ICE, 1 cycle of rituximab plus ICE (R-ICE), followed by carmustine, etoposide, cytarabine, and melphalan and autologous stem-cell rescue (HDT-ASCR). In 08-026

(NCT00712582), after induction therapy with 3 cycles of R-R-CHOP and a 4th cycle of CHOP alone, all patients underwent FDG-PET. Patients who had negative FDG-PET results and a baseline Ki-67 proliferation index of less than 80% received consolidative chemotherapy with 3 cycles of ICE, whereas those who were FDG-PET negative and baseline Ki-67 proliferation index of 80% or more received augmented R-ICE for 2 cycles. The cutoff point for biomarker Ki-67 was determined using the method of Mazumdar and Glassman.²⁹ If the interim FDG-PET results were positive, patients received consolidative chemotherapy with 2 cycles of augmented R-ICE followed by HDT-ASCR. In these 2 studies, patients with positive interim FDG-PET results had a repeat biopsy of the FDG-positive site to verify imaging findings. Only patients with a confirmed positive biopsy went on to receive HDT-ASCR, whereas those with negative biopsy were treated the same as those with FDG-PET negative imaging. Radiation was not used in any of the patients.

Comparison cohort

An external comparison cohort was assembled from patients enrolled at Mayo Clinic in the University of Iowa / Mayo Clinic Lymphoma SPORE MER.²⁷ Patients who received frontline R-CHOP in the MER were matched to the MSK cohort based on the following covariates: age, COO by the Hans algorithm (GCB vs non-GCB only), aalPI, year of diagnosis, sex, and diagnosis to treatment interval (DTI). A 1:1 nearest neighbor matching (NNM) with a 0.2 caliper cutoff³⁰ of MER to MSK patients was attempted on the 138 MSK patients with non-GCB and GCB subtypes. Only a subset of 108 MSK patients were matched within the specified caliper distance.

Statistical Analysis

Patient demographics and baseline characteristics were summarized and reported using descriptive statistics. EFS was defined as time from diagnosis until first of disease progression, relapse, initiation of unplanned lymphoma therapy due to lack of efficacy, or death from any cause. Patients alive without an event were censored at their last follow-up. OS was defined as time from disease diagnosis until death from any cause. Patients alive were censored at their last follow-up. EFS and OS rates were estimated using a Kaplan-Meier estimator. The prognostic impact of baseline risk factors on survival were assessed using univariable Cox proportional hazard models. The median follow-up was estimated using the reverse Kaplan-Meier method. Multivariate survival analyses were not performed due to limited power. A two-sided *P*-value of <0.05 was considered statistically significant. Statistical analyses were done using SAS 9.4 (SAS Institute Inc. Cary, NC, USA) and R (version 4.1.2) for Windows.

Results

From March 26, 2002 through November 3, 2006, 98 patients were enrolled onto MSK Protocol 01-142. From July 1, 2008 through May 28, 2013, 99 patients were enrolled onto MSK Protocol 08-026. For the purposes of this analysis, 44 patients with PMBL and 2 patients with FL3B were excluded. The patients with DLBCL from Protocols 01-142 and 08-026 had similar pre-treatment characteristics (Supplemental Table 2) and similar outcome after a median follow-up of 8.0 years (95% CI: 6.9 – 8.9; Supplemental Figures 2 and 3). This justified combining the two cohorts for analysis. A total of 151

patients with DLBCL were evaluable and their baseline clinical and demographic characteristics are summarized in Table 1. COO subtype were as follows: 69 GCB, 69 non-GCB, and 13 unclassified. Only 7 patients (4.6%) had a positive interim biopsy and received HDT-ASCR. Of those 7 patients, 3 were GCB, 2 were non-GCB, and 2 were unclassified. As a sensitivity analysis, we examined EFS and OS excluding all patients who underwent HDT-ASCR. This demonstrated very similar EFS and OS compared to the entire group, suggesting that transplantation had minimal impact on the overall outcome (Supplemental Figure 4).

Follow-up and Outcome

Median EFS and OS have not been reached (Figures 1A and 1C). The median follow-up for 151 patients is 8.0 years (95% CI 6.9 – 8.9). At median follow-up of 8.0 years, 47 and 31 patients in the MSK cohort had an event and died, respectively (4 of the deaths were unrelated to lymphoma and 3 were of unknown causes). The 2-year EFS was 85% (95% CI: 79 – 91) and 5-year OS was 86% (95% CI: 81 – 92). Notably, 6 of the 47 events had DLBCL at diagnosis but at progression either had marginal zone lymphoma or follicular lymphoma. Due to the lack of adequate tissue, molecular studies were not performed and clonal relationship to the original tumor remains uncertain. At MSK, patients who remained without evidence of disease at 5 years were referred back to their primary care physician, which truncated the length of follow-up available.

Impact of Baseline Prognostic Factors on Outcome

Associations between baseline clinical characteristics and outcome were calculated by univariate analyses. Among the 151 patients with DLBCL, there was no difference in EFS or OS when stratified by proliferation index Ki-67, aalPI score, and tumor bulk ≥ 10 cm (Supplemental Figure 5).

Outcome by COO showed superior EFS in the non-GCB subtype (2-year EFS 88%, 95% CI 81-96) compared to the GCB subtype (2-year EFS 78%, 95% CI 69-89), hazard ratio (HR) 0.53, 95% CI 0.29-0.98; $p = 0.04$ (Figure 1B). OS stratified by COO demonstrated an association in the same direction but was not statistically significant: 5-year OS of 87% (95% CI 79-95) in non-GCB vs 82% (95% CI 74-92) in GCB, HR 0.68, 95% CI 0.33-1.41; $p = 0.30$ (Figure 1D).

To validate this observation, we used the MER as an external comparison cohort of patients treated with standard of care R-CHOP. A 1:1 propensity score matching via the NNM approach with a 0.2 caliper cutoff³⁰ was applied to the 138 MSK patients with non-GCB and GCB subtypes; ultimately 108 patients were matched within the specified caliper cutoff. Other than aalPI, there were no clinical, follow-up, or outcome differences between the 108 MSK patients with available matches and the 30 unmatched patients (Supplemental Table 3, Supplemental Figure 6). As a consequence of the propensity score matching, there were fewer patients with high and high-intermediate risk disease in the matched MSK cohort. The clinical characteristics of the propensity score matched cohort from MER and MSK are shown in Table 2. The MER cohort consisted of 108 patients with DLBCL diagnosed between 2002 through 2014 who underwent first-line

treatment with R-CHOP. Despite propensity score matching, some minor differences existed between the populations. Compared to the MSK cohort, the MER cohort had a slightly higher proportion of low-intermediate risk aalPI (27% in MER vs 23% in MSK) and a lower proportion of high-intermediate risk aalPI (61% in MER vs 64% in MSK). Most prominently, the median DTI was shorter in the MER cohort (15 days in MER vs 21 days in MSK); however, quartile 3 representing the longest delay in treatment initiation was similar (29 days in MER vs 28 days in MSK). At median follow-up of 11 years, 59 and 48 patients in the MER cohort had an event and died, respectively.

Among the 108 matched MSK patients, the 2-year EFS was 88% (95% CI 80-98) for non-GCB and 80% (95% CI 71-92) for GCB (HR 0.57, 95% CI 0.27-1.18; $p = 0.13$; Figure 2B), and the 5-year OS was 86% (95% CI 77-96) for non-GCB and 86% (95% CI 77-95) for GCB (HR 0.76, 95% CI 0.33-1.79; $p = 0.54$; Figure 2D). Though the non-GCB subtype also showed favorable EFS (and was only slightly attenuated) in the matched subset, the association did not remain statistically significant. Compared to the MSK cohort, the MER cohort demonstrated an EFS association (HR 1.17, 95% CI 0.70-1.95; $p = 0.56$) and OS association (HR 1.13, 95% CI 0.64-2.00; $p = 0.67$) in the opposite direction with respect to COO, but these associations were not statistically significant.

Discussion

In the full MSK cohort treated with R-CHOP > (R)-ICE, the EFS of the non-GCB subtype was superior to that of the GCB subtype, and OS by COO showed an association in the same direction but was not statistically significant. Favorable EFS in the non-GCB

subtype was also seen in the 108 MSK patients with available matches from the MER, though the association was not significant in this subset. Matched patients from the MER cohort treated with R-CHOP alone demonstrated an EFS association in the opposite direction (i.e., >1), although HRs were weak and not statistically significant (Figure 2B). Although some minor differences in the baseline characteristics existed between the propensity score matched MSK and MER patients, we do not believe they account for the reversed association between EFS and COO observed in the two cohorts. We hypothesize that the R-CHOP induction (R)-ICE consolidation chemoimmunotherapy regimens may overcome the negative prognostic impact of the non-GCB phenotype, possibly by selectively targeting oncogenic events activated in non-GCB tumors.

The COO as determined by IHC has historically had a weak association with outcomes in the MER cohort (unpublished data not shown). However, the use of digital gene expression-based algorithms via Nanostring and/or RNA-Seq in a subset of N = 475 patients in the MER has shown significant associations with outcomes (ABC vs GCB EFS HR = 1.41, 95% CI: 1.08-1.83). The lack of significance observed in the MER cohort analyzed in this study may be due to IHC based approach for COO calling and/or small sample size. Analysis in the full MER cohort should be done to make conclusions regarding COO in the MER cohort which is outside the scope of the present study.

The R-CHOP > (R)-ICE combination therapy in 01-142 and 08-026 have similarities to the induction and non-cross-resistant consolidation phase used in the LNH03-2B trial by

GELA.³¹ With only 7 of the 151 MSK patients (4.6%) receiving HDT-ASCR (2 out of 7 were non-GCB), we do not believe transplantation accounts for the favorable EFS among patients with non-GCB tumors in the MSK cohort compared with the propensity score matched MER cohort. The LNH03-2B trial was restricted to patients 18-59 years of age with only a single IPI risk factor. The younger age is similar to the age distribution in the MSK cohort: median age of the MSK and LNH03-2B cohorts were 54 and 48 years, respectively. A dose intensity comparison of the 3 regimens is shown in Supplemental Table 3. The induction phase in all 3 regimens has 4 cycles with shortened intervals, where doxorubicin and cyclophosphamide are given at higher doses. The consolidation phase in all 3 regimens consist of sequential treatment with new chemotherapy drugs not included during induction. Therefore, we might suggest that the non-cross-resistant consolidation phase targeted oncogenic pathways specific to non-GCB tumor cells, like the activation of the antiapoptotic nuclear factor-kappa B (NF- κ B) which can inhibit chemotherapy.³²⁻³⁴ Secondary analyses of the GELA LNH03-2B trial showed more favorable outcome in non-GCB patients treated with an induction regimen of R-ACVBP and consolidated with methotrexate, rituximab, ifosfamide, etoposide, and cytarabine²⁴. Molina et al. suggest that this may be associated with a suppression of NF- κ B activity by methotrexate, which sensitized the non-GCB cells to the remaining chemotherapy drugs in the consolidation phase. Since our regimens did not include methotrexate, the current data do not support the conclusion that this drug is the basis for superior outcome in non-GCB tumors.

Some reports have shown enhanced activity of chemotherapy with bortezomib in non-GCB but not in GCB DLBCL,^{35,36} which also support targeting the NF- κ B pathway as an effective treatment approach for the genetically distinct non-GCB subtype. Although the primary analysis of the REMoDL-B trial at a median follow-up of 30 months found no benefit of bortezomib on outcome in the ABC subgroup determined via GEP,³⁷ the updated 5-year survival results demonstrated improved PFS and OS with the addition of bortezomib to R-CHOP in ABC patients.³⁸ Other large-scale multicenter phase III studies have also attempted to improve outcome in untreated non-GCB tumors by adding targeted agents to standard R-CHOP: the PHOENIX trial with ibrutinib³⁹ and the ROBUST trial with lenalidomide.⁴⁰ Although ABC DLBCL tumors showed promising response to both ibrutinib and lenalidomide in preclinical and phase I/II studies,⁴¹⁻⁴³ including the ECOG-ACRIN E1412 phase II study where the addition of lenalidomide to R-CHOP demonstrated a potential clinical benefit in newly diagnosed DLBCL regardless of COO (both GCB and ABC),⁴⁴ results of the PHOENIX and ROBUST phase III trials did not demonstrate a definitive benefit in the ABC subgroup. Further investigation of treatment regimens for the various DLBCL molecular groups is needed.

The addition of etoposide to the R-CHOP regimen (R-CHOEP) has been shown to improve outcome in young patients with high-risk DLBCL.⁴⁵ However, it is uncertain whether baseline biological markers like COO are prognostic of this effect. Some studies demonstrated no significant difference in outcome between GCB vs non-GCB (as determined by the Hans algorithm) after treatment with R-CHOEP.^{46,47} Frontzek et al.⁴⁸ also found no OS advantage with R-CHOEP and R-MegaCHOEP (high dose

chemotherapy plus rituximab followed by ASCR) between GCB and ABC, as determined by Lymph2CX.⁴⁹ Gang et al. found that COO (by Hans) predicted that GCB DLBCL had superior outcome after treatment with R-CHOEP.⁵⁰ In contrast, sequential R-CHOP > (R)-ICE therapy improved the outcome for non-GCB DLBCL. Therefore, we do not think the impact of sequential therapy is simply attributable to the addition of etoposide.

The recent results of the POLARIX trial suggest that substitution of polatuzumab vedotin for vincristine (pola-R-CHP) may improve PFS in the ABC phenotype as determined by the Nanostring Lymph2Cx assay.²³ The magnitude of PFS benefit Tilly et al. observed with pola-R-CHP in the ABC phenotype compared to GCB (HR 0.4, 95% CI 0.2-0.6) may be slightly greater than the magnitude of EFS benefit we saw with R-CHOP > (R)-ICE in the non-GCB phenotype compared to GCB (HR 0.57, 95% 0.27 – 1.18). However, the high cost of polatuzumab vedotin limits its global availability and the R-CHOP > (R)-ICE regimen may represent a cost-effective alternative for patients with non-GCB DLBCL in certain resource-constrained regions of the world.

Although the Hans model is not a perfect surrogate for COO classification by GEP,¹² our results support its use as a means of stratifying patients with DLBCL into prognostic subgroups. R-CHOP is less effective against non-GCB tumors compared with GCB tumors. Similar to the findings in the GELA trial LNH03-2B, we observed improved EFS in non-GCB patients when treated with R-CHOP induction followed by non-cross-resistant (R)-ICE consolidation. This offers a cost-effective treatment approach for this

population with a poor outcome. A strength of this analysis is that the improvement in outcome was seen using the Hans algorithm based on IHC rather than the more precise GEP, which is not readily available in clinical laboratories. In conclusion, our results suggest that an intensified non-cross-resistant sequential chemoimmunotherapy regimen, such as R-CHOP > (R)-ICE combination therapy, could be preferentially selected for patients with non-GCB tumors as classified by the widely applied Hans algorithm.

Acknowledgements

This research was supported in part by CORE grant P30 CA008748, Lymphoma SPORE grant 5 P50 CA192937, and P50 CA97274 and U01 CA195568.

Authorship

Contribution: K.S.B. and A.D.Z. designed the study; M.J. Matasar, A.J.M., D.J.S., A.N., M.L.P., S.M.H., P.A.H., C.S.P., J.R.C., T.M.H., G.A.S., G.S.N., C.H.M., and A.D.Z. provided study materials and patients; K.S.B., A.N.S., M.J. Maurer, J.R.C., T.M.H., G.S.N., and A.D.Z. coordinated and collected data; K.S.B., A.N.S., M.J. Maurer, J.R.C., G.S.N., and A.D.Z. analyzed and interpreted the data; K.S.B. and A.D.Z. wrote the manuscript; all authors contributed to the final approval of the manuscript.

Conflict-of-interest disclosure: M.J. Maurer reports research funding from Roche/Genentech, BMS, and GenMab, and serves on the advisory board of GenMab, and Adaptive Biotechnologies. J.T.F. is a full-time employee of Histowiz. M.J. Matasar reports compensation from Epizyme, Genentech, Roche, Kite, Bayer, Seagen, Celgene, ADC Therapeutics, IMV Therapeutics, and AstraZeneca. A.M. receives research support from Seattle Genetics, Merck,

Bristol-Myers Squibb, and Incyte and receives honorarium from Kyowa Hakko Kirin Pharma, Miragen Therapeutics, Takeda Pharmaceuticals, ADC Therapeutics, Seattle Genetics, Cell Medica, Bristol-Myers Squibb, and Erytech Pharma. D.J.S. receives compensation from InPractice Elsevier and Seattle Genetics and is on speakers bureau for Medical Crossfire. A.N. received honoraria from Janssen, Pharmacyclics, and Prime Oncology; consults for Medscape; serves on advisory board for Janssen; serves on the speakers bureau for Prime Oncology; and receives research funding for Rafael Pharma and Pharmacyclics. S.M.H. received research funding from ADCT therapeutics, Aileron, Forty-Seven, Verastem, Kyowa Hakko Kirin, Millennium Pharmaceuticals Inc, Celgene, Trillium, and Daiichii Sankyo and consults for Astex, Affimed, Merck Sharp and Dome, Kyowa Hakko Kirin Pharma, Corvus Pharmaceuticals Inc., Celgene, Portola Pharmaceuticals, Takeda Millennium, Innate Pharma, Verastem, Miragen Therapeutics Inc, Seattle Genetics, and ADCT. P.A.H. receives research support from Portola, Novartis/GSK, Molecular Templates, and Janssen Pharmaceuticals and served as consultant for Karyopharm, Juno, Portola, Celgene, and AstraZeneca. T.M.H. reports research support from Genentech, and Sorrento, serves on the data monitoring committee of Seagen, Tess Therapeutics, and Eli Lilly & Co., and the scientific advisory board of Eli Lilly & Co., Morphosys, Incyte, Biegene, and Loxo Oncology. G.A.S. reports advisory boards/consulting fees from AbbVie, Atbtherapeutics, Bayer, BeiGene, Bristol Myers Squibb/Celgene, Debiopharm, Epizyme, F. Hoffmann–La Roche Ltd./Genentech, Inc., Genmab, Incyte, Ipsen, Janssen, Kite/Gilead, Loxo/Lilly, Molecular Partners, MorphoSys, Nordic Nanovector, Novartis, Regeneron, and Takeda, and is a shareholder of Owkin. C.H.M. has held an employment/leadership position/advisory role for Celgene, Genentech, Merck & Co., Seattle Genetics Inc., participated in an advisory board for Molecular Templates, and has received research funding from Pharmacyclics, Genentech, Merck & Co., Inc., and Seattle Genetics Inc. A.D.Z. received research grants from AbbVie, Adaptive Biotechnologies, Bristol Myers Squibb, BeiGene, Genentech/Roche, and MEI Pharma, consulting fees from Amgen, AstraZeneca,

BeiGene, Genentech/Roche, Janssen, JUNO/Celgene/Bristol Myers Squibb, Kite/Gilead, MEI Pharma, Pfizer, Pharmacyclics, and Sandoz/Novartis, and serves on the scientific advisory board of Adaptive Biotechnologies, Lymphoma Research Foundation. The remaining authors declare no competing financial interests.

References

1. A clinical evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. The Non-Hodgkin's Lymphoma Classification Project. *Blood*. Jun 1 1997;89(11):3909-18.
2. Coiffier B. Diffuse large cell lymphoma. *Curr Opin Oncol*. Sep 2001;13(5):325-34. doi:10.1097/00001622-200109000-00003
3. Pfreundschuh M, Trumper L, Osterborg A, et al. CHOP-like chemotherapy plus rituximab versus CHOP-like chemotherapy alone in young patients with good-prognosis diffuse large-B-cell lymphoma: a randomised controlled trial by the MabThera International Trial (MInT) Group. *Lancet Oncol*. May 2006;7(5):379-91. doi:10.1016/S1470-2045(06)70664-7
4. Wilson WH, Dunleavy K, Pittaluga S, et al. Phase II study of dose-adjusted EPOCH and rituximab in untreated diffuse large B-cell lymphoma with analysis of germinal center and post-germinal center biomarkers. *J Clin Oncol*. Jun 1 2008;26(16):2717-24. doi:10.1200/JCO.2007.13.1391
5. Leonard JP, Martin P, Barrientos J, Elstrom R. Targeted treatment and new agents in diffuse large B-cell lymphoma. *Semin Hematol*. Jul 2008;45(3 Suppl 2):S11-6. doi:10.1053/j.seminhematol.2008.07.004
6. Gianni AM, Bregni M, Siena S, et al. High-dose chemotherapy and autologous bone marrow transplantation compared with MACOP-B in aggressive B-cell lymphoma. *N Engl J Med*. May 1 1997;336(18):1290-7. doi:10.1056/NEJM199705013361804
7. Alizadeh AA, Eisen MB, Davis RE, et al. Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. *Nature*. Feb 3 2000;403(6769):503-11. doi:10.1038/35000501
8. Rosenwald A, Wright G, Leroy K, et al. Molecular diagnosis of primary mediastinal B cell lymphoma identifies a clinically favorable subgroup of diffuse large B cell lymphoma related to Hodgkin lymphoma. *J Exp Med*. Sep 15 2003;198(6):851-62. doi:10.1084/jem.20031074
9. Rosenwald A, Wright G, Chan WC, et al. The use of molecular profiling to predict survival after chemotherapy for diffuse large-B-cell lymphoma. *N Engl J Med*. Jun 20 2002;346(25):1937-47. doi:10.1056/NEJMoa012914
10. Lossos IS, Czerwinski DK, Alizadeh AA, Wechser MA, Tibshirani R, Botstein D, Levy R. Prediction of survival in diffuse large-B-cell lymphoma based on the expression of six genes. *N Engl J Med*. Apr 29 2004;350(18):1828-37. doi:10.1056/NEJMoa032520
11. Hans CP, Weisenburger DD, Greiner TC, et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. *Blood*. Jan 1 2004;103(1):275-82. doi:10.1182/blood-2003-05-1545
12. Meyer PN, Fu K, Greiner TC, et al. Immunohistochemical methods for predicting cell of origin and survival in patients with diffuse large B-cell lymphoma treated with rituximab. *J Clin Oncol*. Jan 10 2011;29(2):200-7. doi:10.1200/JCO.2010.30.0368
13. Fu K, Weisenburger DD, Choi WW, et al. Addition of rituximab to standard chemotherapy improves the survival of both the germinal center B-cell-like and non-germinal center B-cell-like subtypes of diffuse large B-cell lymphoma. *J Clin Oncol*. Oct 1 2008;26(28):4587-94. doi:10.1200/JCO.2007.15.9277
14. Gutierrez-Garcia G, Cardesa-Salzman T, Climent F, et al. Gene-expression profiling and not immunophenotypic algorithms predicts prognosis in patients with diffuse large B-cell lymphoma treated with immunochemotherapy. *Blood*. May 5 2011;117(18):4836-43. doi:10.1182/blood-2010-12-322362
15. Ott G, Ziepert M, Klapper W, et al. Immunoblastic morphology but not the immunohistochemical GCB/nonGCB classifier predicts outcome in diffuse large B-cell lymphoma in the RICOVER-60 trial of the DSHNHL. *Blood*. Dec 2 2010;116(23):4916-25. doi:10.1182/blood-2010-03-276766

16. Copie-Bergman C, Gaulard P, Leroy K, et al. Immuno-fluorescence in situ hybridization index predicts survival in patients with diffuse large B-cell lymphoma treated with R-CHOP: a GELA study. *J Clin Oncol*. Nov 20 2009;27(33):5573-9. doi:10.1200/JCO.2009.22.7058
17. Salles G, de Jong D, Xie W, et al. Prognostic significance of immunohistochemical biomarkers in diffuse large B-cell lymphoma: a study from the Lunenburg Lymphoma Biomarker Consortium. *Blood*. Jun 30 2011;117(26):7070-8. doi:10.1182/blood-2011-04-345256
18. National Comprehensive Cancer Network. B-Cell Lymphomas (Version 5. 2021). Accessed August 2, 2023, <https://jnccn.org/view/journals/jnccn/19/11/article-p1218.xml?ArticleBodyColorStyles=full%20html>
19. Lenz G, Wright G, Dave SS, et al. Stromal gene signatures in large-B-cell lymphomas. *N Engl J Med*. Nov 27 2008;359(22):2313-23. doi:10.1056/NEJMoa0802885
20. Malumbres R, Chen J, Tibshirani R, et al. Paraffin-based 6-gene model predicts outcome in diffuse large B-cell lymphoma patients treated with R-CHOP. *Blood*. Jun 15 2008;111(12):5509-14. doi:10.1182/blood-2008-02-136374
21. Natkunam Y, Farinha P, Hsi ED, et al. LMO2 protein expression predicts survival in patients with diffuse large B-cell lymphoma treated with anthracycline-based chemotherapy with and without rituximab. *J Clin Oncol*. Jan 20 2008;26(3):447-54. doi:10.1200/JCO.2007.13.0690
22. Rimsza LM, Leblanc ML, Unger JM, et al. Gene expression predicts overall survival in paraffin-embedded tissues of diffuse large B-cell lymphoma treated with R-CHOP. *Blood*. Oct 15 2008;112(8):3425-33. doi:10.1182/blood-2008-02-137372
23. Tilly H, Morschhauser F, Sehn LH, et al. Polatuzumab Vedotin in Previously Untreated Diffuse Large B-Cell Lymphoma. *N Engl J Med*. Jan 27 2022;386(4):351-363. doi:10.1056/NEJMoa2115304
24. Molina TJ, Canioni D, Copie-Bergman C, et al. Young patients with non-germinal center B-cell-like diffuse large B-cell lymphoma benefit from intensified chemotherapy with ACVBP plus rituximab compared with CHOP plus rituximab: analysis of data from the Groupe d'Etudes des Lymphomes de l'Adulte/lymphoma study association phase III trial LNH 03-2B. *J Clin Oncol*. Dec 10 2014;32(35):3996-4003. doi:10.1200/JCO.2013.54.9493
25. Moskowitz CH, Schoder H, Teruya-Feldstein J, et al. Risk-adapted dose-dense immunochemotherapy determined by interim FDG-PET in Advanced-stage diffuse large B-Cell lymphoma. *J Clin Oncol*. Apr 10 2010;28(11):1896-903. doi:10.1200/JCO.2009.26.5942
26. Schoder H, Zelenetz AD, Hamlin P, et al. Prospective Study of 3'-Deoxy-3'-18F-Fluorothymidine PET for Early Interim Response Assessment in Advanced-Stage B-Cell Lymphoma. *J Nucl Med*. May 2016;57(5):728-34. doi:10.2967/jnumed.115.166769
27. Cerhan JR, Link BK, Habermann TM, et al. Cohort Profile: The Lymphoma Specialized Program of Research Excellence (SPORE) Molecular Epidemiology Resource (MER) Cohort Study. *Int J Epidemiol*. Dec 1 2017;46(6):1753-1754i. doi:10.1093/ije/dyx119
28. International Non-Hodgkin's Lymphoma Prognostic Factors P. A predictive model for aggressive non-Hodgkin's lymphoma. *N Engl J Med*. Sep 30 1993;329(14):987-94. doi:10.1056/NEJM199309303291402
29. Mazumdar M, Glassman JR. Categorizing a prognostic variable: review of methods, code for easy implementation and applications to decision-making about cancer treatments. *Stat Med*. Jan 15 2000;19(1):113-32. doi:10.1002/(sici)1097-0258(20000115)19:1<113::aid-sim245>3.0.co;2-o
30. Austin PC. An Introduction to Propensity Score Methods for Reducing the Effects of Confounding in Observational Studies. *Multivariate Behav Res*. May 2011;46(3):399-424. doi:10.1080/00273171.2011.568786
31. Recher C, Coiffier B, Haioun C, et al. Intensified chemotherapy with ACVBP plus rituximab versus standard CHOP plus rituximab for the treatment of diffuse large B-cell lymphoma (LNH03-2B): an open-

- label randomised phase 3 trial. *Lancet*. Nov 26 2011;378(9806):1858-67. doi:10.1016/S0140-6736(11)61040-4
32. Young RM, Staudt LM. Targeting pathological B cell receptor signalling in lymphoid malignancies. *Nat Rev Drug Discov*. Mar 2013;12(3):229-43. doi:10.1038/nrd3937
33. Orlowski RZ, Baldwin AS, Jr. NF-kappaB as a therapeutic target in cancer. *Trends Mol Med*. Aug 2002;8(8):385-9. doi:10.1016/s1471-4914(02)02375-4
34. Orlowski RZ, Kuhn DJ. Proteasome inhibitors in cancer therapy: lessons from the first decade. *Clin Cancer Res*. Mar 15 2008;14(6):1649-57. doi:10.1158/1078-0432.CCR-07-2218
35. Dunleavy K, Pittaluga S, Czuczman MS, et al. Differential efficacy of bortezomib plus chemotherapy within molecular subtypes of diffuse large B-cell lymphoma. *Blood*. Jun 11 2009;113(24):6069-76. doi:10.1182/blood-2009-01-199679
36. Ruan J, Martin P, Furman RR, et al. Bortezomib plus CHOP-rituximab for previously untreated diffuse large B-cell lymphoma and mantle cell lymphoma. *J Clin Oncol*. Feb 20 2011;29(6):690-7. doi:10.1200/JCO.2010.31.1142
37. Davies A, Cummin TE, Barrans S, et al. Gene-expression profiling of bortezomib added to standard chemoimmunotherapy for diffuse large B-cell lymphoma (REMoDL-B): an open-label, randomised, phase 3 trial. *Lancet Oncol*. May 2019;20(5):649-662. doi:10.1016/S1470-2045(18)30935-5
38. Davies AJ, Barrans S, Stanton L, et al. Differential Efficacy From the Addition of Bortezomib to R-CHOP in Diffuse Large B-Cell Lymphoma According to the Molecular Subgroup in the REMoDL-B Study With a 5-Year Follow-Up. *J Clin Oncol*. Mar 27 2023;JCO2300033. doi:10.1200/JCO.23.00033
39. Younes A, Sehn LH, Johnson P, et al. Randomized Phase III Trial of Ibrutinib and Rituximab Plus Cyclophosphamide, Doxorubicin, Vincristine, and Prednisone in Non-Germinal Center B-Cell Diffuse Large B-Cell Lymphoma. *J Clin Oncol*. May 20 2019;37(15):1285-1295. doi:10.1200/JCO.18.02403
40. Nowakowski GS, Chiappella A, Gascoyne RD, et al. ROBUST: A Phase III Study of Lenalidomide Plus R-CHOP Versus Placebo Plus R-CHOP in Previously Untreated Patients With ABC-Type Diffuse Large B-Cell Lymphoma. *J Clin Oncol*. Apr 20 2021;39(12):1317-1328. doi:10.1200/JCO.20.01366
41. Wilson WH, Young RM, Schmitz R, et al. Targeting B cell receptor signaling with ibrutinib in diffuse large B cell lymphoma. *Nat Med*. Aug 2015;21(8):922-6. doi:10.1038/nm.3884
42. Zhang LH, Kosek J, Wang M, Heise C, Schafer PH, Chopra R. Lenalidomide efficacy in activated B-cell-like subtype diffuse large B-cell lymphoma is dependent upon IRF4 and cereblon expression. *British journal of haematology*. Feb 2013;160(4):487-502. doi:10.1111/bjh.12172
43. Hernandez-Ilizaliturri FJ, Deeb G, Zinzani PL, et al. Higher response to lenalidomide in relapsed/refractory diffuse large B-cell lymphoma in nongerminal center B-cell-like than in germinal center B-cell-like phenotype. *Cancer-Am Cancer Soc*. Nov 15 2011;117(22):5058-66. doi:10.1002/cncr.26135
44. Nowakowski GS, Hong F, Scott DW, et al. Addition of Lenalidomide to R-CHOP Improves Outcomes in Newly Diagnosed Diffuse Large B-Cell Lymphoma in a Randomized Phase II US Intergroup Study ECOG-ACRIN E1412. *J Clin Oncol*. Apr 20 2021;39(12):1329-1338. doi:10.1200/JCO.20.01375
45. Gang AO, Strom C, Pedersen M, et al. R-CHOEP-14 improves overall survival in young high-risk patients with diffuse large B-cell lymphoma compared with R-CHOP-14. A population-based investigation from the Danish Lymphoma Group. *Ann Oncol*. Jan 2012;23(1):147-153. doi:10.1093/annonc/mdr058
46. Leppa S, Jorgensen J, Tierens A, et al. Patients with high-risk DLBCL benefit from dose-dense immunochemotherapy combined with early systemic CNS prophylaxis. *Blood Adv*. May 12 2020;4(9):1906-1915. doi:10.1182/bloodadvances.2020001518
47. Basic-Kinda S, Radman I, Dujmovic D, et al. R-CHOEP14 in younger high-risk patients with large B cell lymphoma: an effective front-line regimen with cardiac toxicity: a real-life, single-center experience. *Ann Hematol*. Jun 2021;100(6):1517-1524. doi:10.1007/s00277-020-04353-3

48. Frontzek F, Ziepert M, Nickelsen M, et al. Rituximab plus high-dose chemotherapy (MegaCHOEP) or conventional chemotherapy (CHOEP-14) in young, high-risk patients with aggressive B-cell lymphoma: 10-year follow-up of a randomised, open-label, phase 3 trial. *Lancet Haematol.* Apr 2021;8(4):e267-e277. doi:10.1016/S2352-3026(21)00022-3
49. Scott DW, Wright GW, Williams PM, et al. Determining cell-of-origin subtypes of diffuse large B-cell lymphoma using gene expression in formalin-fixed paraffin-embedded tissue. *Blood.* Feb 20 2014;123(8):1214-7. doi:10.1182/blood-2013-11-536433
50. Gang AO, Pedersen MO, Knudsen H, et al. Cell of origin predicts outcome to treatment with etoposide-containing chemotherapy in young patients with high-risk diffuse large B-cell lymphoma. *Leukemia & lymphoma.* Jul 2015;56(7):2039-46. doi:10.3109/10428194.2014.982645

Characteristic	Full Cohort (n = 151)
Age at diagnosis (in years)	
Median (range)	54 (21 – 71)
Sex	
Male	83 of 151 (55%)
Female	68 of 151 (45%)
LDH > ULN	120 of 151 (79%)
KPS ≤ 70%	52 of 151 (34%)
Stage II	10 of 151 (7%)
III-IV	141 of 151 (93%)
Extranodal sites > 1	81 of 151 (54%)
Bone marrow biopsy +	40 of 151 (26%)
aalPI score	
LIR	27 of 151 (18%)
HIR	86 of 151 (57%)
HR	38 of 151 (25%)
Bulk > 10 cm	32 of 151 (21%)
COO	
GCB	69 of 151 (46%)
Non-GCB	69 of 151 (46%)
Unclassified	13 of 151 (9%)

Table 1. Baseline Characteristics of MSK Cohort. Abbreviations: MSK, Memorial Sloan Kettering; GCB, germinal center B-cell-like; LDH, lactate dehydrogenase; ULN, upper limit of normal; KPS, Karnofsky Performance Status; aalPI, age-adjusted International Prognostic Index; LIR, low-intermediate risk; HIR, high-intermediate risk; HR, high risk; COO, cell of origin.

Characteristic	Full MSK cohort with GCB and non-GCB subtype (n = 138)	MSK subset with available matches (n = 108)	MER propensity score matched cohort (n = 108)
Age at diagnosis (years)			
Median (range)	54 (21 – 71)	55 (22 – 71)	54 (20 – 71)
Q1, Q3	45, 62	45, 62	45, 64
Sex			
Male	76 of 138 (55%)	62 of 108 (57%)	59 of 108 (55%)
Female	62 of 138 (45%)	46 of 108 (43%)	49 of 108 (45%)
aalPI score			
aalPI			
LIR	25 of 138 (18%)	25 of 108 (23%)	29 of 108 (27%)
HIR	80 of 138 (58%)	69 of 108 (64%)	66 of 108 (61%)
HR	33 of 138 (24%)	14 of 108 (13%)	13 of 108 (12%)
COO			
GCB	69 of 138 (50%)	56 of 108 (52%)	55 of 108 (51%)
Non-GCB	69 of 138 (50%)	52 of 108 (48%)	53 of 108 (49%)
DTI (days)			
Median (range)	21 (1 – 77)	21 (1 – 54)	15 (2 – 109)
Q1, Q3	14, 28	14, 28	8, 29
Year of Diagnosis	2002 - 2013	2002 - 2013	2002 - 2014

Table 2. Patient Characteristics of 1:1 NNM with a 0.2 caliper cutoff of MER to MSK patients. Abbreviations: NNM, nearest neighbor matching; MSK, Memorial Sloan Kettering; MER, Molecular Epidemiology Resource; Q, quartile; aalPI, age-adjusted

International Prognostic Index; LIR, low-intermediate risk; HIR, high-intermediate risk; HR, high risk; COO, cell of origin; GCB, germinal center B-cell-like; DTI, diagnosis to treatment interval.

Figure Legends:

Figure 1. Survival endpoints for full MSK cohort treated with R-CHOP induction > (R)-ICE consolidation (n=151): (A) EFS; (B) EFS by COO; (C) OS; (D) OS by COO. MSK, Memorial Sloan Kettering; R-CHOP, rituximab plus cyclophosphamide, doxorubicin, vincristine and prednisone; R-ICE, rituximab plus ifosfamide, carboplatin and etoposide; EFS, event free survival; COO, cell of origin; OS, overall survival; GCB, germinal center B-cell-like; CI, confidence interval; ref, reference.

Figure 2. Comparing GCB and non-GCB patients from MSK cohort with available matches treated with R-CHOP induction > (R)-ICE consolidation (n=108) versus matched MER cohort using NNM approach with a 0.2 caliper cutoff treated with R-CHOP (n=108): (A) EFS; (B) EFS by COO; (C) OS; (D) OS by COO; *1 MER patient had missing EFS data; GCB, germinal center B-cell-like; MSK, Memorial Sloan Kettering; MER, Molecular Epidemiology Resource; NNM, nearest neighbor matching; EFS, event free survival; OS, overall survival; R-CHOP, rituximab plus cyclophosphamide, doxorubicin, vincristine and prednisone; R-ICE, rituximab plus ifosfamide, carboplatin and etoposide; COO, cell of origin.

Figure 1

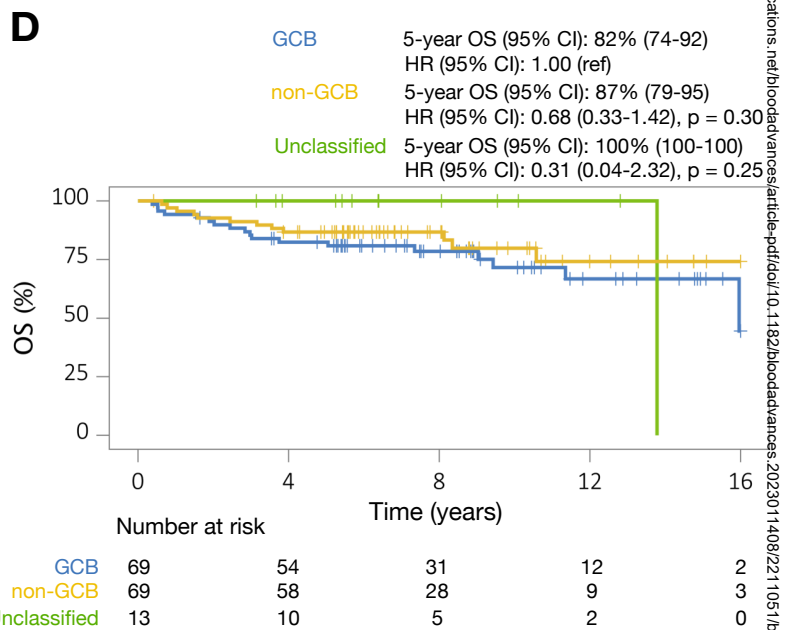
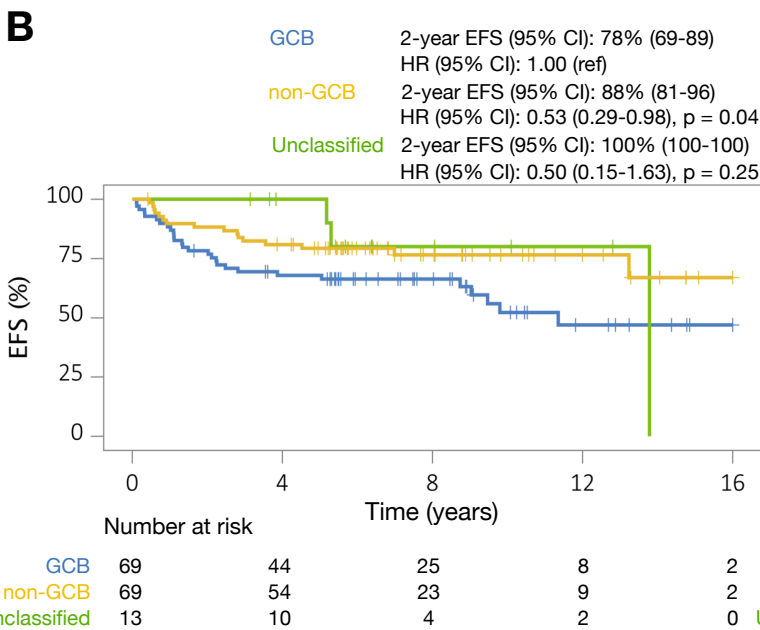
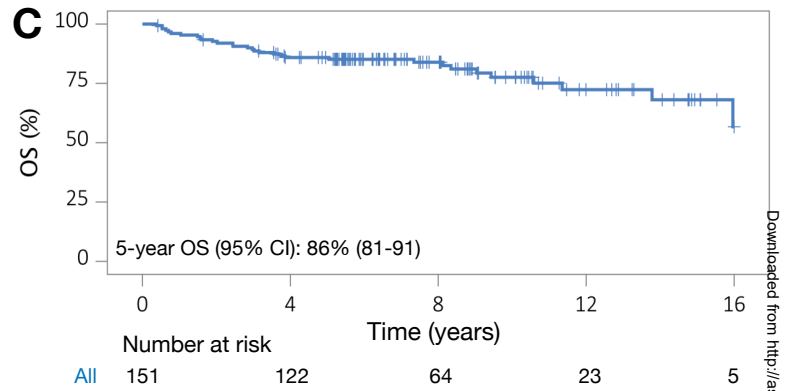
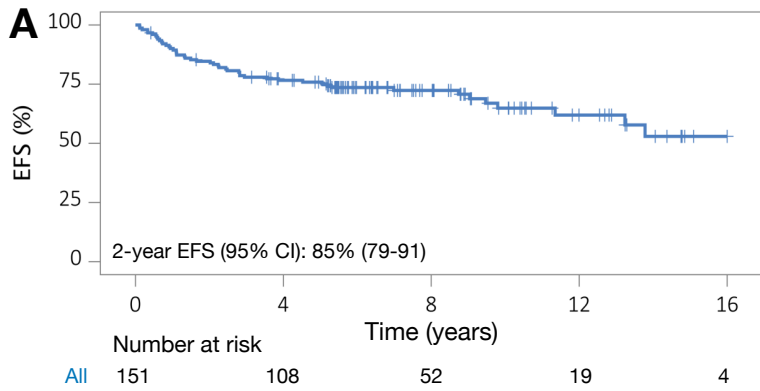


Figure 2

