

## TO THE EDITOR:

# Inhibition of casein kinase 1 $\delta$ disrupts translation initiation and exerts potent antilymphoma activity

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Global protein synthesis rate plays a critical role in the governance of cell growth, metabolism, and tumor cell proliferation. Stabilization of the eukaryotic translation initiation factor 4F (eIF4F) complex, comprised of the eIF4E/eIF4A/eIF4G subunits, is a hallmark of resistance to anti-BRAF and anti-MEK therapies in several melanoma and colon cancer models.<sup>1-3</sup> This occurs through persistent hyper-phosphorylation of the negative regulator 4E-BP1, which leads to release of eIF4E from 4E-BP1 and hyperactivation of eIF4F-mediated translation initiation. Here, we identify casein kinase 1 $\delta$  (CK1 $\delta$ ) as a novel regulator of mRNA translation and promising target for cancer drug development.

Both CK1 $\delta$  and CK1 $\epsilon$  are components of pre-40S ribosome subunits and are collectively required for the maturation of 40S ribosomes during translation.<sup>4</sup> CK1 $\epsilon$  is required for translation initiation by upregulating 4E-BP1 phosphorylation in some models of breast cancer.<sup>5</sup> However, in lymphoma, knockdown of CK1 $\epsilon$  does not significantly impact 4E-BP1 phosphorylation.<sup>6</sup> Furthermore, PF4800567, a selective CK1 $\epsilon$  inhibitor, failed to repress 4E-BP1 phosphorylation in lymphoma cells.<sup>6</sup> These observations question whether CK1 $\epsilon$  is broadly involved in regulating translation in cancer models, particularly lymphoma. We present evidence that CK1 $\delta$  is an activator of translation initiation in models of lymphoma. CK1 $\delta$  acts by stimulating 4E-BP1 phosphorylation and is associated with the m<sup>7</sup>G cap. Chemical inhibition of CK1 $\delta$  strongly inhibits 4E-BP1 phosphorylation, eIF4F complex assembly, and translation initiation and demonstrates potent and specific antilymphoma activity in vitro and in vivo.

We found that PF670462, a dual CK1 $\epsilon$ /CK1 $\delta$  inhibitor, reduces phosphorylation of 4E-BP1 in the mantle cell lymphoma line Z-138 (supplemental Figure 1A). This result, combined with our previous work targeting CK1 $\epsilon$  with a dual inhibitor of PI3K $\delta$ , umbralisib, led us to hypothesize that CK1 $\delta$  may play an important role in regulating translation initiation in lymphoma via phosphorylation of 4E-BP1. SR-3029 is the most potent selective CK1 $\delta$  inhibitor that is widely available<sup>7</sup> and was used in this study to interrogate these relationships. Consistent with our hypothesis, we found that SR-3029 potently inhibited phosphorylation of 4E-BP1 in Z-138 cells and other lymphoma cell lines representing mantle cell lymphoma (MCL; Figure 1A; supplemental Figure 1B) and diffuse large B-cell lymphoma (DLBCL; supplemental Figure 1C).

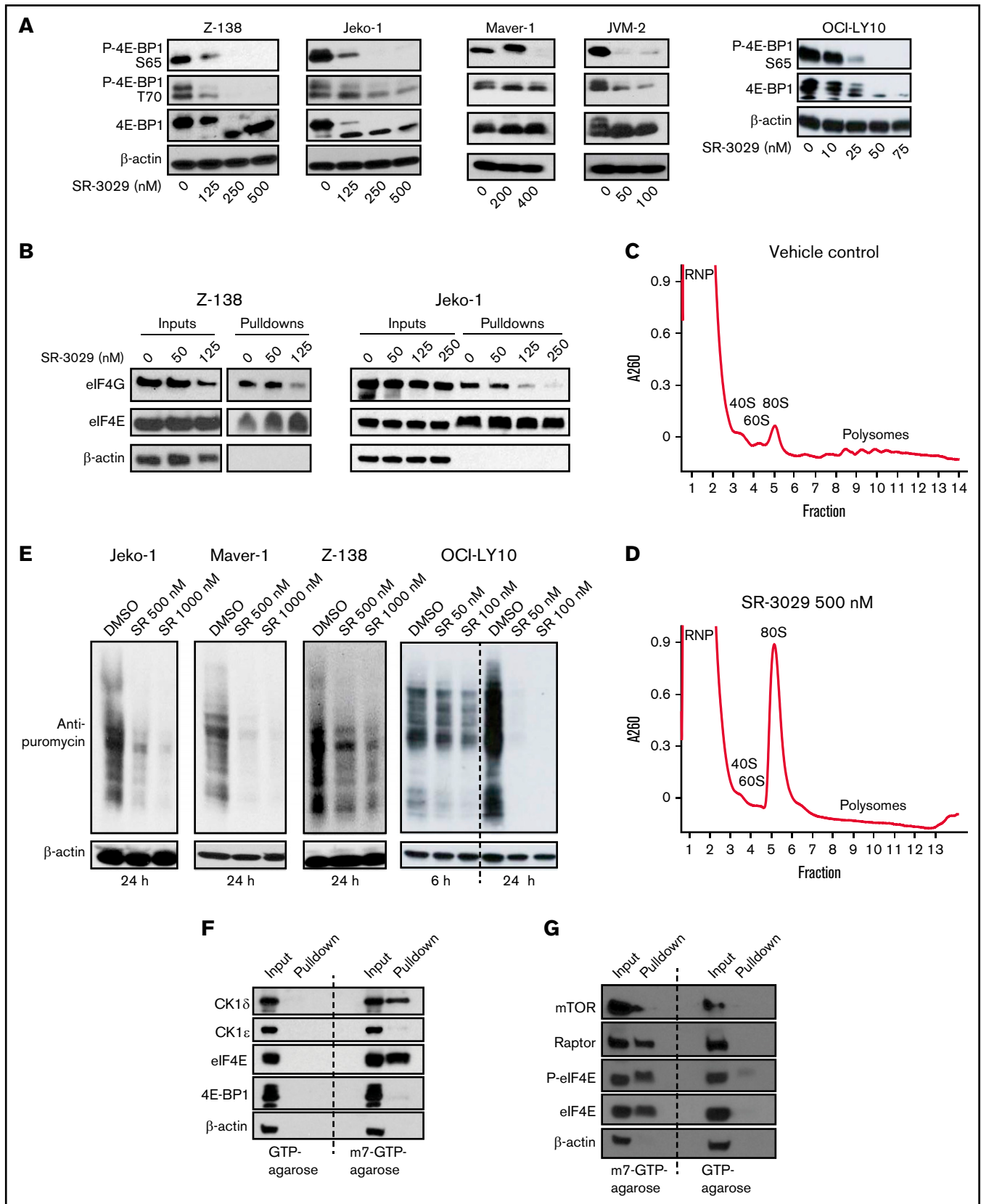
Given the critical role of 4E-BP1 phosphorylation in releasing eIF4E for assembly of the eIF4F complex<sup>8</sup> (comprising eIF4G, eIF4E, and eIF4A), the above results led us to question whether SR-3029 may inhibit translation initiation. We first investigated whether SR-3029 affects cap recruitment of eIF4F using a m<sup>7</sup>GTP cap binding assay.<sup>9</sup> In this assay, eIF4E binds directly to the mRNA m<sup>7</sup>G cap<sup>10</sup> and serves as a loading control, whereas the level of eIF4G in the pulldown fraction indicates the efficiency of eIF4F assembly. We observed that SR-3029 substantially reduced the amount of eIF4G pulled down by the m<sup>7</sup>GTP-agarose beads in the MCL cell lines Z-138 and Jeko-1, without significantly reducing the level of cap-bound eIF4E (Figure 1B). To assess the effects of SR-3029 on global protein synthesis, we

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The full-text version of this article contains a data supplement.

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**Figure 1.**

conducted polysome profiling analysis. In Z-138 lymphoma cells treated with dimethyl sulfoxide (DMSO), the 40S, 60S, and 80S ribosome peaks were easily identified, and polysome levels were elevated (Figure 1C). SR-3029 treatment resulted in a substantial increase in the 80S monosome peak at the expense of polysomes indicating that CK1 $\delta$  inhibition, either directly or indirectly, repressed global translation (Figure 1D).

To further characterize how SR-3029 influences global protein synthesis we performed a surface sensing of translation (SUnSET) assay, in which puromycin was used to terminate translation elongation. The levels of terminated polypeptides can be determined with an anti-puromycin antibody<sup>11</sup> and indicate global protein synthesis levels. SR-3029 substantially inhibited incorporation of puromycin in multiple MCL cell lines (Jeko-1, Maver-1, and Z-138) and 1 DLBCL cell line (OCI-LY10; Figure 1E). Collectively, these results suggest that the CK1 $\delta$  inhibitor SR-3029 represses protein synthesis in lymphoma cells via inhibiting 4E-BP1 phosphorylation and eIF4F assembly.

The above results implicating CK1 $\delta$  in translation initiation led us to suggest that CK1 $\delta$  may be associated with the mRNA cap, which we assessed using a m<sup>7</sup>GTP cap-binding assay. We observed that CK1 $\delta$  was pulled down by m<sup>7</sup>GTP agarose beads in Z-138 lymphoma cells in a manner similar to the cap-binding protein eIF4E (Figure 1F). We reasoned that binding of CK1 $\delta$  and eIF4E to m<sup>7</sup>GTP beads was specific based on the following criteria: (1) control GTP beads failed to pull down these proteins and (2) CK1 $\epsilon$  was not pulled down by m<sup>7</sup>GTP beads (Figure 1F). The m<sup>7</sup>GTP beads (but not GTP beads) also pulled down mTOR, Raptor, eIF4E, and phosphorylated eIF4E (Figure 1G), which are components of the translation preinitiation complex (PIC).<sup>12</sup> These results suggest that CK1 $\delta$  is associated with components of the PIC and may regulate translation initiation via this association.

Given the importance of translation in blood cancer cell proliferation and the role of CK1 $\delta$  in upregulating translation, we reasoned that the CK1 $\delta$  inhibitor SR-3029 may effectively block tumor growth. We found that SR-3029 potently inhibited the viability of 4 human MCL cell lines using the Cell Titer Glo assay (Figure 2A), with IC<sub>50</sub> values at 48 hours in the range of 22.3 to 202.3 nM (supplemental Table 1). SR-3029 also inhibited viability in human cell lines of DLBCL, the most common aggressive lymphoma (Figure 2B). A notable exception was the DLBCL cell line VAL, which was resistant to SR-3029 and lacks 4E-BP1.<sup>13</sup>

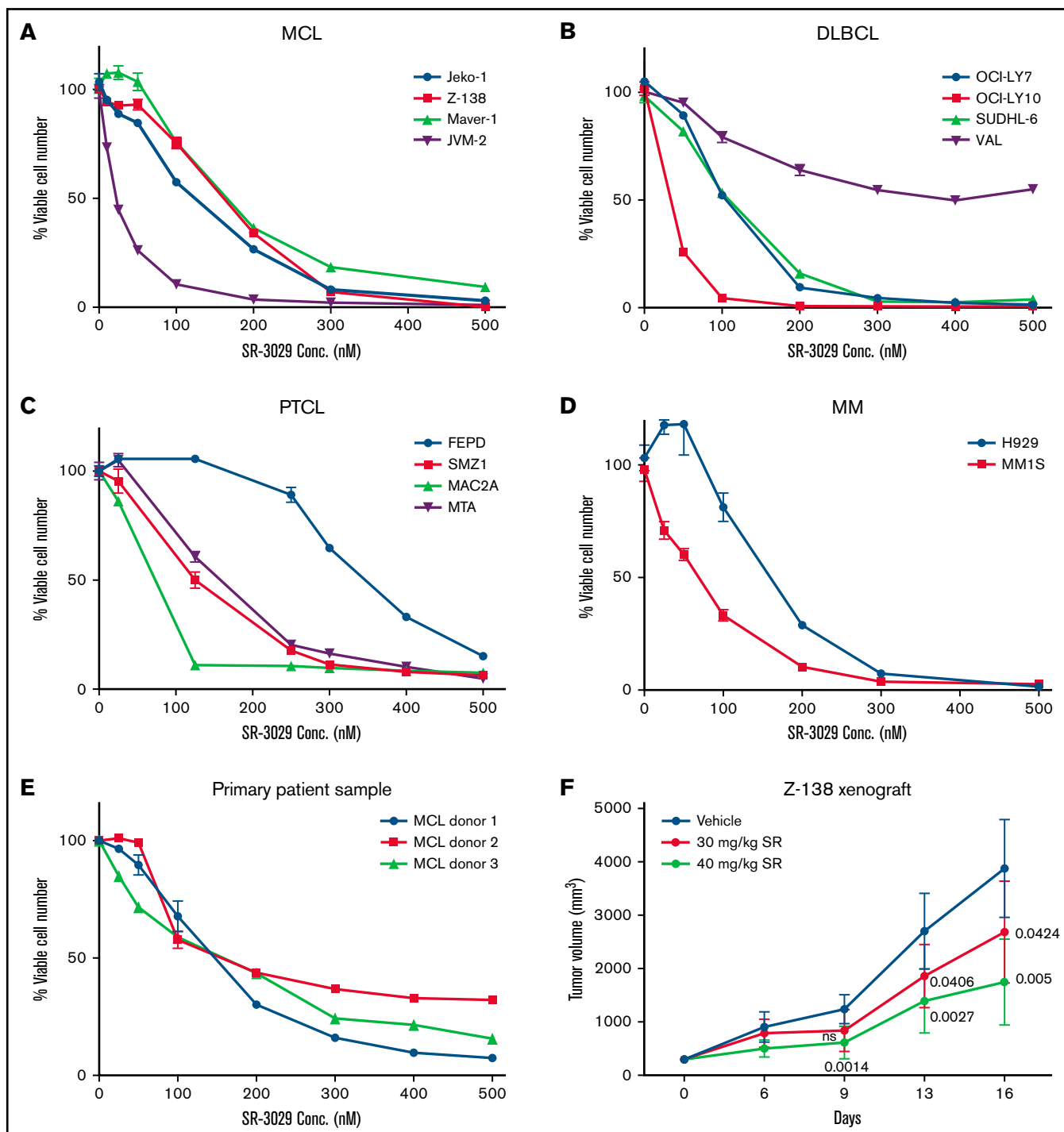
Presumably, these cells may be able to bypass inhibitors that act via 4E-BP1. Although peripheral T-cell lymphoma (PTCL) has a poorer prognosis compared with B-cell lymphomas because of the lack of effective treatment, PTCL cell lines were also sensitive to SR-3029 (Figure 2C). Multiple myeloma (MM) cell lines were highly sensitive to SR-3029 (Figure 2D). Moreover, primary lymphoma cells from patients with MCL were sensitive to SR-3029 (Figure 2E). Importantly, SR-3029 (30-40 mg/kg daily dosing by oral gavage) exhibited a dose-dependent tumor growth inhibition in a mouse xenograft model of MCL, established using Z-138 cells injected subcutaneously in the flank of SCID beige mice (Figure 2F). No significant body weight loss was observed in any treatment group (supplemental Figure 2A) indicating that SR-3029 is well tolerated and that its effects are relatively specific to lymphoma cells in this context.

Because SR-3029 has been reported to kill MDA-MB-231 human breast cancer cells through inhibiting nuclear localization of  $\beta$ -catenin,<sup>14</sup> we set out to determine whether SR-3029 had this effect in lymphoma. SR-3029 treatment did not reduce nuclear  $\beta$ -catenin in the MCL cell line Z-138 and DLBCL cell line OCI-LY10 (supplemental Figure 2B-C), despite that SR-3029 potently inhibited phosphorylation of 4E-BP1. Taken together, these results suggest that the broad antilymphoma activity of SR-3029 is mediated primarily by downregulation of translation.

In this study, we demonstrate that CK1 $\delta$  is a positive regulator of translation initiation. This conclusion is supported by the observations that CK1 $\delta$  inhibition disrupts 4E-BP1 phosphorylation, eIF4F assembly, and global translation. Furthermore, CK1 $\delta$  exhibits m<sup>7</sup>GTP cap-binding activity in a manner similar to eIF4E. A future challenge will be to determine the precise mechanism by which CK1 $\delta$  stimulates translation initiation in lymphoma cells, which could be direct or indirect, and will require an understanding of the substrate(s) of CK1 $\delta$ .

We present evidence that blocking translation in blood cancer cells via inhibition of CK1 $\delta$  may be an innovative strategy to treat broad subtypes of B- and T-cell lymphomas and multiple myeloma. Although SR-3029 may exhibit off-target (CK1 $\delta$ ) effects at high concentrations, at the low concentrations and in the lymphoma models we tested, SR-3029 appears to specifically inhibit CK1 $\delta$ -mediated translation. In the lymphoma models studied, the effects of CK1 $\delta$  inhibition do not invoke the mechanism of repressing  $\beta$ -catenin nuclear localization. In breast cancer and possibly some other cancer models, the primary role of CK1 $\delta$

**Figure 1 (continued) CK1 $\delta$  inhibitor SR-3029 downregulates translation initiation.** (A) CK1 $\delta$  inhibition disrupts 4E-BP1 phosphorylation. Five lymphoma cell lines, Z-138, Jeko-1, Maver-1, and JVM-2 (representing mantle cell lymphoma), and OCI-LY10 (representing diffuse large B-cell lymphoma), were treated with SR-3029 or vehicle control for 24 hours. Protein levels were determined by immunoblot. P-, phosphorylated protein. (B) CK1 $\delta$  inhibition disrupts eIF4F assembly. The lymphoma cell lines Z-138 and Jeko-1 were treated with SR-3029 or DMSO control at the indicated concentrations for 24 hours and then processed for the m<sup>7</sup>GTP cap binding assay using m<sup>7</sup>GTP agarose beads to pull down cap binding proteins, which were analyzed by immunoblot. The level of eIF4G reflects eIF4F assembly, whereas the level of eIF4E is constant and serves as the loading control. (C-D) CK1 $\delta$  inhibition disrupts global translation. Z-138 cells were treated with DMSO vehicle control (C) or SR-3029 500 nM (D) for 6 hours. Cell lysates were fractionated on 10% to 50% sucrose density gradients by centrifugation. The x axis shows fractions collected, and the y axis shows absorbance at OD<sub>260</sub>. Shown are the ribonucleoprotein particle fraction (RNP), ribosomal subunits (40S, 60S, and 80S), and polysomes. (E) SUnSET translation assay. Bulk translation was determined in 3 MCL cell lines and 1 DLBCL cell line treated with SR-3029 at the indicated concentrations or DMSO control for 24 hours (or 6 and 24 hours in the case of OCI-LY10). Puromycin was then added at the final concentration of 1  $\mu$ g/mL for 30 minutes. Cell lysates were analyzed by immunoblot using anti-puromycin or anti- $\beta$ -actin antibodies. (F-G) CK1 $\delta$  associates with the mRNA cap structure in lymphoma. m<sup>7</sup>GTP cap binding assays were performed in untreated Z-138 lymphoma cells. (F) Protein levels of CK1 $\delta$ , CK1 $\epsilon$ , eIF4E, 4E-BP1, and  $\beta$ -actin were determined in the input and pulldown fractions by immunoblotting. (G) Protein levels of mTOR, Raptor, eIF4E, P-eIF4E, and  $\beta$ -actin were determined in the input and pulldown fractions by immunoblotting.



**Figure 2. CK1 $\delta$  inhibitor SR-3029 potently inhibits tumor growth in broad subtypes of blood cancers.** (A-E) SR-3029 potently inhibits viability of lymphomas. Lymphoma cell lines or primary lymphoma cells were treated with SR-3029 or DMSO control at the concentrations indicated on the *x* axis for 48 hours. Cell viability was quantitated using the Cell-Titer Glo (Promega) assay and is plotted on the *y* axis. Shown is the average of 3 experiments from the 48-hour treatment, presented as mean  $\pm$  standard error of the mean. (F) A mouse xenograft model of human lymphoma was established using the Z-138 cell line in SCID beige mice. Tumor volume (*y* axis) is plotted against the time of treatment (*x* axis) for 2 treatment cohorts and the vehicle control. *P* values of the treatment cohorts vs control are indicated as determined by repeated-measure analysis of variance.

may be to mediate Wnt/ $\beta$ -catenin signaling as previously reported.<sup>14</sup> At the therapeutic effective dose ranges, SR-3029 appears to be well tolerated in mice based on our results and

those of previously reported.<sup>14</sup> Other observations supporting a favorable therapeutic window include: normal peripheral blood mononuclear cells from healthy donors were relatively resistant to

SR-3029 in contrast to malignant cell lines (supplemental Figure 2D), and CK1 $\delta$  protein level was more abundant in lymphoma cell lines compared with PBMCs from the healthy donors (supplemental Figure 2E). Although preliminary in nature, these results suggest that the selective CK1 $\delta$  inhibitor SR-3029 could be developed into a safe and effective treatment for patients with lymphomas that are dependent on dysregulated protein translation. Inhibition of CK1 $\delta$  may offer a new mechanistic approach for the treatment of hematologic malignancies.

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## References

1. Boussemaert L, Malka-Mahieu H, Girault I, et al. eIF4F is a nexus of resistance to anti-BRAF and anti-MEK cancer therapies. *Nature*. 2014;513(7516):105-109.
2. Pelletier J, Graff J, Ruggero D, Sonenberg N. Targeting the eIF4F translation initiation complex: a critical nexus for cancer development. *Cancer Res*. 2015;75(2):250-263.
3. Malka-Mahieu H, Newman M, Désaubry L, Robert C, Vagner S. Molecular pathways: the eIF4F translation initiation complex—new opportunities for cancer treatment. *Clin Cancer Res*. 2017;23(1):21-25.
4. Zemp I, Wandrey F, Rao S, et al. CK1 $\delta$  and CK1 $\epsilon$  are components of human 40S subunit precursors required for cytoplasmic 40S maturation. *J Cell Sci*. 2014;127(Pt 6):1242-1253.
5. Shin S, Wolgamott L, Roux PP, Yoon S-O. Casein kinase 1 $\epsilon$  promotes cell proliferation by regulating mRNA translation. *Cancer Res*. 2014;74(1):201-211.
6. Deng C, Lipstein MR, Scotto L, et al. Silencing c-Myc translation as a therapeutic strategy through targeting PI3K $\delta$  and CK1 $\epsilon$  in hematological malignancies. *Blood*. 2017;129(1):88-99.
7. Bibian M, Rahaim RJ, Choi JY, et al. Development of highly selective casein kinase 1 $\delta/1\epsilon$  (CK1 $\delta/\epsilon$ ) inhibitors with potent antiproliferative properties. *Bioorg Med Chem Lett*. 2013;23(15):4374-4380.
8. Gingras A-C, Gygi SP, Raught B, et al. Regulation of 4E-BP1 phosphorylation: a novel two-step mechanism. *Genes Dev*. 1999;13(11):1422-1437.
9. Choo AY, Yoon S-O, Kim SG, Roux PP, Blenis J. Rapamycin differentially inhibits S6Ks and 4E-BP1 to mediate cell-type-specific repression of mRNA translation. *Proc Natl Acad Sci USA*. 2008;105(45):17414-17419.
10. Marcotrigiano J, Gingras A-C, Sonenberg N, Burley SK. Cocystal structure of the messenger RNA 5' cap-binding protein (eIF4E) bound to 7-methyl-GDP. *Cell*. 1997;89(6):951-961.
11. Schmidt EK, Clavarino G, Ceppi M, Pierre P. SUNSET, a nonradioactive method to monitor protein synthesis. *Nat Methods*. 2009;6(4):275-277.
12. Holz MK, Ballif BA, Gygi SP, Blenis J. mTOR and S6K1 mediate assembly of the translation preinitiation complex through dynamic protein interchange and ordered phosphorylation events. *Cell*. 2005;123(4):569-580.
13. Mallya S, Fitch BA, Lee JS, So L, Janes MR, Fruman DA. Resistance to mTOR kinase inhibitors in lymphoma cells lacking 4EBP1. *PLoS One*. 2014;9(2):e88865.
14. Rosenberg LH, Lafitte M, Quereda V, et al. Therapeutic targeting of casein kinase 1 $\delta$  in breast cancer. *Sci Transl Med*. 2015;7(318):318ra202.