

# Acute erythroid leukemia is enriched in *NUP98* fusions: a report from the Children's Oncology Group

Karen M. Chisholm,<sup>1,2</sup> Amy E. Heerema-McKenney,<sup>3</sup> John K. Choi,<sup>4</sup> Jenny Smith,<sup>5</sup> Rhonda E. Ries,<sup>5</sup> Betsy A. Hirsch,<sup>6</sup> Susana C. Raimondi,<sup>4</sup> Todd A. Alonzo,<sup>7</sup> Yi-Cheng Wang,<sup>8</sup> Richard Aplenc,<sup>9</sup> Lillian Sung,<sup>10</sup> Alan S. Gamis,<sup>11</sup> Soheil Meshinchi,<sup>5</sup> and Samir B. Kahwash<sup>12</sup>

<sup>1</sup>Department of Laboratories, Seattle Children's Hospital, Seattle, WA; <sup>2</sup>Department of Laboratory Medicine and Pathology, University of Washington Medical Center, Seattle, WA; <sup>3</sup>Pathology and Laboratory Medicine Institute, Cleveland Clinic, Cleveland, OH; <sup>4</sup>Department of Pathology, University of Alabama at Birmingham, Birmingham, AL; <sup>5</sup>Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, WA; <sup>6</sup>Division of Laboratory Medicine, University of Minnesota Medical Center, Fairview, Minneapolis, MN; <sup>7</sup>Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, CA; <sup>8</sup>Children's Oncology Group, Monrovia, CA; <sup>9</sup>Children's Hospital of Philadelphia, Philadelphia, PA; <sup>10</sup>Division of Hematology/Oncology, Department of Pediatrics, The Hospital for Sick Children, Toronto, ON, Canada; <sup>11</sup>Children's Mercy Hospitals & Clinics, Kansas City, MO; and <sup>12</sup>Department of Pathology and Laboratory Medicine, Nationwide Children's Hospital, Columbus, OH

## Key Points

- Pediatric PEL is rare and associated with poor overall survival.
- *NUP98* fusions are increased in acute erythroid leukemias, especially those with PELs, per the 2008 World Health Organization classification.

Acute erythroid leukemia (AEL) is a rare subtype of acute myeloid leukemia (AML) primarily affecting older adults and was previously classified into erythroid/myeloid and pure erythroid subtypes. In this pediatric AEL study, we evaluated morphologic, immunophenotypic, cytogenetic, molecular, and clinical data of 24 (1.2%) cases from all cases undergoing central pathology review in Children's Oncology Group trials AAML0531 and AAML1031. Of 24 cases, 5 had a pure erythroid phenotype, and 19 had an erythroid/myeloid phenotype. *NUP98* fusions were highly enriched in patients with AEL, occurring in 7 of 22 cases for which molecular data were available (31.8% vs 6.7% in other AML subtypes). Of 5 cases of pure erythroid leukemias (PELs), 3 had *NUP98* fusions, and 4 had complex karyotypes. Erythroid/myeloid leukemias were reclassified by using the 2017 World Health Organization hematopathology classification as: myelodysplastic syndrome (MDS) with excess blasts-1 (n = 3), MDS with excess blasts-2 (n = 7), AML (nonerythroid, n = 5), and unknown MDS/AML (n = 4); the 5 cases of nonerythroid AML included 1 with an *NUP98-NSD1* fusion, 2 with myelodysplasia-related changes, and 1 with a complex karyotype. Three cases of MDS with excess blasts-2 also had *NUP98* rearrangements. *WT1* mutations were present in 5 of 14 cases, all erythroid/myeloid leukemia. Outcomes assessment revealed statistically poorer overall survival (5-year, 20% ± 36% vs 66% ± 23%; *P* = .004) and event-free survival (5-year, 20% ± 36% vs 46% ± 23%; *P* = .019) for those with PEL than those with erythroid/myeloid leukemia. Our study supports that AEL is a morphologically and genetically heterogeneous entity that is enriched in *NUP98* fusions, with the pure erythroid subtype associated with particularly adverse outcomes.

## Introduction

Acute myeloid leukemia (AML) has an incidence of 8.8 per 1 million children aged 0 to 19 years,<sup>1</sup> with acute erythroid leukemia (AEL) being the rarest AML subtype. Historically, AEL has been classified into 2 subtypes: (1) pure erythroid leukemia (PEL), defined as a bone marrow with >80% immature (undifferentiated or pro-normoblastic) erythroid cells and lacking 20% myeloblasts (also previously classified as French-American-British [FAB] M6b); and (2) erythroid/myeloid leukemia, defined as

Submitted 18 June 2020; accepted 19 October 2020; published online 7 December 2020. DOI 10.1182/bloodadvances.2020002712.

De-identified individual participant data for the results reported in this paper are available in supplemental Table 1. Data-sharing requests may also be sent to the

corresponding author (Karen M. Chisholm; e-mail: karen.chisholm@seattlechildrens.org).

The full-text version of this article contains a data supplement.

© 2020 by The American Society of Hematology

a bone marrow with >50% erythroid cells and myeloblasts accounting for >20% of all nonerythroid cells (also previously classified as FAB M6a).<sup>2-4</sup> This classification is confusing, and some authors have questioned whether cases of acute erythroid/myeloid leukemia with <20% myeloblasts of all cells should be classified as myelodysplastic syndrome (MDS) with excess blasts.<sup>5-9</sup> The most recently revised 2017 World Health Organization (WHO) classification eliminated the erythroid/myeloid leukemia subtype; currently, only the PEL subtype remains, defined as >80% immature erythroid precursors, of which  $\geq 30\%$  are proerythroblasts.<sup>10</sup>

Even before the erythroid/myeloid subtype was eliminated, AEL (AML-M6) accounted for <5% of cases of AML and was the most uncommon AML FAB type in both children and adults (3%-5%).<sup>11,12</sup> In the Children's Cancer Group (CCG) AML clinical trial CCG-2891, cases of AML-M6 represented 2.2% of all de novo MDS and AML.<sup>11</sup> A slight male predominance has been noted.<sup>11,13</sup> AML-M6 patients also have a worse prognosis, with lower overall survival (OS) and event-free survival (EFS) than FAB M0-M5 patients.<sup>11</sup> The reason for this poor prognosis remains to be identified and potentially includes the actual pathologic diagnosis, karyotype, and/or the molecular genotype.

In the current study, we report the morphologic, immunohistochemical, cytogenetic, and molecular features of non-Down syndrome AEL in patients enrolled in the Children's Oncology Group (COG) trials AAML0531 and AAML1031 with the goal of determining if any of these factors affect prognosis.

## Methods

### Patients

AAML0531 was a COG phase 3 trial that administered intensive chemotherapy to children, adolescents, and young adults with newly diagnosed AML. Participants were randomized to receive or not receive gemtuzumab ozogamicin (Mylotarg; Pfizer Inc, New York, NY) during the first cycle (for all patients) and the fourth cycle (for those who did not undergo hematopoietic stem cell transplantation).<sup>14</sup> Details of the treatment regimen have been previously reported. The trial included 1022 eligible patients (age range, 1 month to 29.99 years) with de novo AML enrolled between August 2006 and June 2010 from participating COG institutions ( $n = 181$ ). Eleven of the included patients from this trial analyzed herein were also included in the analysis of acute erythroleukemia by Iacobucci et al.<sup>15</sup> AAML1031 was also a COG phase 3 randomized trial comparing standard chemotherapy with or without bortezomib in individuals with de novo AML and without Down syndrome. In addition, sorafenib was given to consenting participants with a high *FLT3* internal tandem duplication (ITD) allelic ratio ( $>0.4$ ).<sup>16</sup> This trial increased risk stratification based on minimal residual disease, *FLT3* ITD allelic ratio, *NPM1* mutations, and *CEBP $\alpha$*  mutations to identify patients who would benefit from hematopoietic stem cell transplantation. The trial included 1231 eligible patients with de novo AML (age range, 1 month-29.99 years) enrolled between June 2011 and July 2017 from participating COG institutions ( $n = 193$ ). Because there were no statistically significant differences in OS or EFS between these studies, we combined all of the patients with AEL.

These studies were approved by the National Cancer Institute's central Institutional Review Board, and the institutional review boards of all participating institutions. All patients or their parents

gave written informed consent or assent, according to institutional regulations. Patients' age at diagnosis and their sex were recorded.

### Morphologic assessment

All cases with an institutional diagnosis of AEL were identified from the AAML0531 and AAML1031 databases. Blood smears, bone marrow aspirate, bone marrow core biopsies, flow cytometry reports, and immunohistochemistry reports underwent central pathology review to confirm diagnosis. Slides were re-reviewed for the presence of multilineage dysplasia. Central pathology review was performed on 864 (84.5%) of all cases in AAML0531 and 1070 (86.9%) of all cases in AAML1031. Patients were excluded if material was not available or insufficient for central pathology review. Because AAML0531 and AAML1031 were conducted when the 2001 WHO and 2008 WHO classifications of hematopoietic tumors, respectively, were used,<sup>4,17</sup> AEL included PEL and erythroid/myeloid leukemia subtypes. Cases included were morphologically AEL even if multilineage dysplasia and/or an 11q23 genetic abnormality were present.

### AML reclassification

Included cases were reclassified by using the 2017 WHO definitions of AML, not otherwise specified (NOS), AML with recurrent genetic abnormalities, and AML with myelodysplasia-related changes (AML-MRC).<sup>10,18,19</sup> Accordingly, as the erythroid/myeloid subtype of AEL has been eliminated, these cases were reclassified as AML or MDS with excess blasts 1 or 2 (EB-1 or EB-2).<sup>20</sup>

### Cytogenetic and molecular assessments

Results of cytogenetic analyses for both studies were centrally reviewed and recorded by using the International System of Human Cytogenetic Nomenclature. The number and types of cytogenetic abnormalities were noted, including the presence or absence of *inv(16)/t(16;16)*, *t(8;21)*, and *11q23/KMT2A* abnormalities. Screening for *FLT3* ITD and other mutations in *FLT3*, *NPM1*, *CEBP $\alpha$* , and *WT1* were performed as previously described.<sup>21-24</sup> Fusion transcripts were detected by RNA-sequencing (RNA-seq) with fusion detection algorithms STAR-Fusion v1.1.0 and TransABySS v1.4.10. RNA-seq was completed for total RNA from primary patient samples purified by using the AllPrep DNA/RNA/miRNA Universal Kit (#80224; Qiagen, Valencia, CA), using the QIAcube system. The messenger RNA libraries were prepared for 75 bp strand-specific, paired-end sequencing by using the ribodepletion 2.0 protocol by the British Columbia Genome Sciences Centre (Vancouver, BC, Canada). STAR-Fusion was run by using default parameters with the premade GRCh37 resource library with Gencode v19 annotations ([https://data.broadinstitute.org/Trinity/CTAT\\_RESOURCE\\_LIB/](https://data.broadinstitute.org/Trinity/CTAT_RESOURCE_LIB/)).<sup>25</sup> TransABySS parameters were set to retain fusions with breakpoint reads  $\geq 1$ , flanking pairs  $\geq 2$  counts, and spanning reads  $\geq 2$  counts.<sup>26</sup> Transcriptomic data are available through the dbGaP TARGET: Acute Myeloid Leukemia study (accession, phs000465.v19.p8).<sup>27</sup>

### Outcomes assessment

Data from AAML0531 and AAML1031 were current as of 31 March 2019. Variables included morphologic classifications, presence or absence of multilineage dysplasia, complex karyotypes, identified genetic mutations, and identified fusions. The Kaplan-Meier method<sup>28</sup> was used to estimate OS (defined as time from study entry to death) and EFS (time from study entry until failure to achieve complete

remission [CR] during induction, relapse, or death). Relapse risk (RR) was calculated by cumulative incidence methods and defined as time from the end of induction I for patients in CR to relapse or death, in which deaths without a relapse were considered competing events.<sup>29</sup> Patients who withdrew from therapy due to relapse, persistent central nervous system disease, or refractory disease with  $\geq 20\%$  bone marrow blasts by the end of induction I were categorized as induction I failures.

## Statistical analyses

Statistical analyses were performed on the following variables: ages at diagnosis, morphologic classifications, presence or absence of multilineage dysplasia, certain karyotypic abnormalities [specifically *inv(16)/t(16;16)*, *t(8;21)*, *11q23/KMT2A* abnormalities, and complex karyotypes], identified genetic mutations, and identified fusions. Given the small size of some of these subgroups and because the comparisons are ad hoc analyses, the study was more exploratory in nature. The  $\chi^2$  test was used to test the significance of observed differences in proportions, and Fisher's exact test was used when data were sparse. The Student *t* test was used to compare means and distributions of 2 groups. The significance of predictor variables was tested with the log-rank statistic for OS and EFS and with Gray's statistic for RR. All estimates are reported with 2 times the Greenwood standard errors. Children lost to follow-up were censored at their date of last known contact. Testing the interaction of the comparison variable with the log of the survival time indicated that the proportional hazards assumption was not violated for any of the comparisons. A value of  $P < .05$  was considered statistically significant.

## Results

### Leukemia morphologic and immunophenotypic classifications

For the 2 COG trials, 24 (1.2%) of 1934 cases that underwent central pathology review were confirmed as AEL (PEL or acute erythroid/myeloid leukemia). According to morphology and immunophenotype, there were 5 cases of PEL and 19 of erythroid/myeloid leukemia. As depicted in Table 1, at least one erythroid antigen (CD71, glycophorin A, CD36, PAS, and E-cadherin) was present in each case. Myeloid markers such as myeloperoxidase, CD13, CD33, CD34, CD117, and HLA-DR were more often expressed in the erythroid/myeloid subtype than in the pure erythroid subtype. Similar to other AML, aberrantly expressed antigens included CD4 and CD7. Rarely, megakaryocytic antigens such as CD41, CD42b, and CD61 were also positive. Of the 24 cases, 8 had multilineage dysplasia, all of which were previously classified as the erythroid/myeloid subtype. In total, the 24 cases were reclassified morphologically as follows: PEL ( $n = 5$ ), MDS-EB-1 ( $n = 3$ ), MDS-EB-2 ( $n = 7$ ), AML (nonerythroid leukemia) ( $n = 3$ ), AML-MRC (by multilineage dysplasia) ( $n = 2$ ), and MDS/AML unknown due to an unknown percentage of total myeloid blasts ( $n = 4$ ).<sup>10,20</sup>

### Patient characteristics

The 24 cases of morphologic AEL were split evenly between male and female subjects (male:female ratio, 13:11) (Table 1). The median age of patients was 10.2 years (range, 0.5-21.0 years). However, patients with PEL were significantly younger (median, 2.3 years; range, 0.5-2.6 years) than those with erythroid/myeloid leukemia (median, 13.5 years; range, 1.0-21.0;  $P < .01$ ).

## Cytogenetic assessment

For all 24 cases, cytogenetics was confirmed by central cytogenetics review. Compared with all other centrally reviewed cases of AML, these cases of AEL lacked the classic cytogenetic findings of *inv(16)/t(16;16)*, *t(8;21)*, and *11q23/KMT2A* abnormalities, the last finding being statistically significant ( $P = .012$ ) (Figure 1). Figure 2 displays the spectrum of cytogenetic findings identified in cases of AEL. Overall, 9 cases had normal karyotypes, 8 of which were erythroid/myeloid leukemias; 4 of these cases showed multilineage dysplasia. Seven cases had complex karyotypes, defined as  $\geq 3$  chromosomal abnormalities, all occurring in patients aged 1.0 to 2.6 years. Only 1 other patient was aged  $< 6$  years (a 6-month-old child with PEL and a normal karyotype); the remaining 4 cases of PEL had complex karyotypes. Of the total 7 cases of complex karyotypes, only 1 MDS-EB-2 had multilineage dysplasia. Deletions of 13q were prevalent in those with complex karyotypes (5 of 7 complex karyotype cases). One case of MDS-EB-2 having multilineage dysplasia did not have a complex karyotype but had an MDS-defining cytogenetic abnormality of *t(3;21)(q26;q22)*. There were no cases of monosomy 5/del(5q), monosomy 7/del(7q), or abnormalities of 12p. The most common aneuploidies were trisomy 6 (13%), trisomy 8 (21%), and trisomy 21 (13%), which were present in cases of PEL, MDS-EB-1, MDS-EB-2, and AML (nonerythroid).

### Final 2017 WHO classifications

Considering morphology and karyotype, the 5 cases of PEL would be recategorized by the 2017 WHO classification as AML-MRC ( $n = 4$ ) and AML, NOS PEL ( $n = 1$ ). The 19 cases of erythroid/myeloid subtype would be classified as MDS-EB-1 ( $n = 3$ ), MDS-EB-2 ( $n = 7$ ), AML-MRC ( $n = 2$  due to multilineage dysplasia and  $n = 1$  due to complex karyotype), AML, NOS, nonerythroid subtype ( $n = 2$ ), and unknown MDS/AML ( $n = 4$ , due to unknown percentage of total myeloid blasts).

### Genetic mutations and fusions

Figure 2 lists the prognostic markers used for cases of leukemia. *FLT3* ITD was identified in 4 (17%) cases, all of which were erythroid/myeloid leukemia. Allelic ratios ranged from 0.01 to 0.09 and were not considered high. All 4 were aged  $> 6$  years (range, 6.4-16.4 years), and 3 had normal karyotypes. No *FLT3* or *CEBP $\alpha$*  point mutations were identified in those cases tested. There was only 1 case with an identified *NPM1* mutation (ie, an erythroid/myeloid leukemia). However, 36% (5 of 14) of tested cases had identified *WT1* mutations, all of which were erythroid/myeloid leukemias; in fact, most of these 5 erythroid/myeloid leukemias had more than one *WT1* mutation. These mutations are listed in the dbGaP TARGET: Acute Myeloid Leukemia study (accession, phs000465.v19.p8).<sup>27</sup> Finally, in the 11 cases with *TP53* mutation testing, only one had a single mutation (p.P47S), a case of MDS-EB-1.

In addition to karyotype analysis, RNA-seq analysis was performed for 22 patients with AEL for expression and fusion profiling. In these sequenced cases, 9 fusions were identified, of which 7 involved *NUP98*, for a total percentage of 32% with *NUP98* fusions (Figures 1 and 2). The 7 cases with *NUP98* rearrangements had fusion partners of *KDM5A* ( $n = 4$ ), *NSD1* ( $n = 2$ ), and *SET* ( $n = 1$ ). Notably, 3 (60%) of 5 cases of PEL had *NUP98* fusions. Of the 4 cases with *NUP98-KDM5A* translocations, 2 had PEL and 2 had

**Table 1. Comparison of pathologic findings between morphologic subtypes of AEL**

Morphologic classification	PEL	Erythroid/ myeloid subtype: MDS-EB-1	Erythroid/ myeloid subtype: MDS-EB-2	Erythroid/myeloid subtype: AML (not pure erythroid)	Erythroid/ myeloid subtype: AML with multilineage dysplasia	Erythroid/myeloid subtype: unknown MDS/AML
No. of cases	5	3	7	3	2	4
Multilineage dysplasia, n/N (%)	0/5 (0)	2/3 (67)	4/7 (57)	0/3 (0)	2/2 (100)	0/4 (0)
<b>Age, y</b>						
Median	2.3	13.5	16.0	15.2	6.4	14.6
Range	0.5-2.6	11.4-16.4	1.3-21.0	1.0-16.2	6.4-6.4	9.00-19.0
Male:female ratio	2:3	2:1	3:4	2:1	1:1	1:3
<b>Flow cytometry/immunohistochemistry, n/N (%)</b>						
CD36	1/1 (100)	NA	1/2 (50)	1/1 (100)	NA	1/1 (100)
CD71	2/2 (100)	2/2 (100)	2/2 (100)	2/2 (100)	1/1 (100)	2/2 (100)
Glycophorin A	3/3 (100)	1/1 (100)	2/2 (100)	NA	0/1 (0)	2/2 (100)
PAS	2/2 (100)	NA	NA	1/1 (100)	1/1 (100)	NA
E-cadherin	1/1 (100)	NA	NA	NA	NA	NA
CD4	2/3 (67)	1/2 (50)	1/2 (50)	2/2 (100)	1/1 (100)	NA
CD7	0/3 (0)	1/3 (33)	3/4 (75)	1/1 (100)	0/1 (0)	2/2 (100)
CD13	2/5 (40)	3/3 (100)	5/5 (100)	2/2 (100)	1/1 (100)	3/3 (100)
CD33	1/4 (25)	3/3 (100)	5/5 (100)	2/2 (100)	1/1 (100)	4/4 (100)
CD34	0/4 (0)	3/3 (100)	3/5 (80)	NA	1/1 (100)	2/3 (67)
CD38	1/3 (33)	1/1 (100)	1/1 (100)	2/2 (100)	1/1 (100)	4/4 (100)
CD41	1/2 (50)	0/1 (0)	1/1 (100)	1/1 (100)	0/1 (0)	NA
CD42b	1/1 (100)	NA	NA	NA	0/1 (0)	NA
CD61	2/2 (100)	0/2 (0)	2/3 (66)	1/1 (100)	0/1 (0)	NA
CD45	4/5 (80)	1/1 (100)	2/2 (100)	NA	1/1 (100)	1/1 (100)
CD64	0/3 (0)	0/1 (0)	1/3 (33)	2/2 (100)	0/1 (0)	1/1 (100)
CD117	1/5 (20)	2/2 (100)	5/6 (83)	2/2 (100)	1/1 (100)	4/4 (100)
HLA-DR	3/5 (60)	3/3 (100)	2/5 (40)	1/2 (50)	1/1 (100)	4/4 (100)
MPO	0/5 (0)	3/3 (100)	2/2 (100)	2/3 (66)	1/1 (100)	1/3 (33)

MPO, myeloperoxidase; NA, not available.

MDS-EB-2 (erythroid/myeloid subtype). Of these *NUP98-KDM5A* cases, 3 had complex karyotypes and 1 had a normal metaphase karyotype (age range, 1.4–6.0 years). The 2 cases of *NUP98-NSD1* translocations (ages 6.4 and 16.0 years) had erythroid/myeloid leukemia. Another alternative *NUP98* fusion was identified with *SET* in a case of PEL. Two additional fusions were identified: a variant t(3;5)(q25.3;q35.1) (*NPM1-MLF1*) identified in a case of erythroid/myeloid leukemia and a t(3;21)(q26;q22) (*RUNX1-MECOM*) identified in a case of MDS-EB-2. There were no rearrangements of *KMT2A*.

## Outcomes

Table 2 details the 5-year OS and 5-year EFS of all risk factors in this group of 24 AEL patients. Comparison of the entire AEL cohort vs that of all other FAB subtypes who underwent central pathology review (n = 1910) revealed no statistically significant differences in OS, EFS, or RR in those who achieved CR. Only 4 of the 19 patients with known transplant status underwent stem cell transplant, all with erythroid/myeloid leukemia; the rate of transplantation in the cases of AEL did not differ between trials, nor did outcomes between those who did and did not undergo

transplant. Including all 24 cases, when cases of pure erythroid and erythroid/myeloid subtypes of AEL (Figure 3A-B) were separated, there was no significant difference in OS or EFS for those with erythroid/myeloid leukemias or other FAB subtypes. However, patients with PEL had significantly lower OS than those with erythroid/myeloid leukemia (5-year, 20% ± 36% vs 66% ± 23%; *P* = .004) as well as EFS (5-year, 20% ± 36% vs 46% ± 23%; *P* = .019). In addition, the RR in those who achieved CR was higher in patients with PEL than those with erythroid/myeloid leukemia (5-year, 67% ± 0% vs 55% ± 32%; *P* = .035). None of the cases of PEL underwent transplant in this study.

The presence of any fusion protein did not affect 5-year OS or EFS, nor did the presence of a *NUP98* fusion (Figure 3C-D). The PEL with *NUP98-SET* fusion and the 2 erythroid/myeloid leukemias with *NUP98-NSD1* fusions had a 5-year OS of 100% ± 0%. However, the 4 cases with *NUP98-KDM5A* fusions had much worse 5-year OS and EFS (both 25% ± 43%), although these results were not statistically significant compared with all cases without *NUP98-KDM5A* fusions (5-year OS and EFS, 59% ± 24% and 38% ± 23%, respectively). Of the 4 cases with *NUP98-KDM5A*



**Table 2. Outcome measures in pediatric AEL**

Variable	n	5-y OS, % ± 2SE%	5-y EFS, % ± 2SE%
<b>Morphology</b>			
M0-M5, M7 FAB	1910	65 ± 2*	50 ± 2*
M6 FAB	24	56 ± 21	41 ± 20
PEL	5	20 ± 36†	20 ± 36†
Erythroid/myeloid leukemia	19	66 ± 23*	46 ± 23*
MDS (new WHO classification)	10	56 ± 33	48 ± 33
AML (new WHO classification)	5	80 ± 36	40 ± 44
<b>Multilineage dysplasia</b>			
Absent	16	48 ± 26	38 ± 24
Present	8	73 ± 33	50 ± 35
<b>Karyotype</b>			
Not complex	17	55 ± 26	34 ± 24
Complex	7	57 ± 37	57 ± 37
<b>Mutations</b>			
<i>WT1</i> wild type	9	53 ± 35	56 ± 33
<i>WT1</i> mutation	5	60 ± 44	20 ± 36
<i>TP53</i> wild type	10	57 ± 23	40 ± 31
<i>TP53</i> mutation	1	0 ± 0	0 ± 0‡
<b>Fusion status</b>			
No fusions	13	45 ± 28	38 ± 27
Fusion positive	9	65 ± 33	30 ± 33
<i>NUP98</i> fusion positive	7	54 ± 40	43 ± 37
<i>NUP98-KDM5A</i> positive	4	25 ± 43	25 ± 43
<i>NUP98-NSD1</i> positive	2	100 ± 0	NA
<i>NUP98-SET</i> positive	1	100 ± 0	100 ± 0
<i>NPM1-MLF1</i> positive	1	NA	0 ± 0
<i>RUNX1-MECOM</i> positive	1	NA	0 ± 0
<b>Morphology + fusion</b>			
PEL without <i>NUP98</i> fusion	2	0 ± 0	0 ± 0
PEL with <i>NUP98</i> fusion	3	33 ± 54	33 ± 54
PEL with <i>NUP98-KDM5A</i> fusion	2	0 ± 0	0 ± 0
MDS with <i>NUP98</i> fusion	3	50 ± 71	67 ± 54
MDS with <i>NUP98-KDM5A</i> fusion	2	50 ± 71	50 ± 71
AML (nonerythroid) with <i>NUP98-NSD1</i> fusion	1	100 ± 0	0 ± 0

SE, standard error.

\*Significantly better compared with PEL.

†Significantly worse compared with M0-M5, M7 FAB group and erythroid/myeloid leukemia cohort.

‡Significantly worse compared with those without *TP53* mutations.

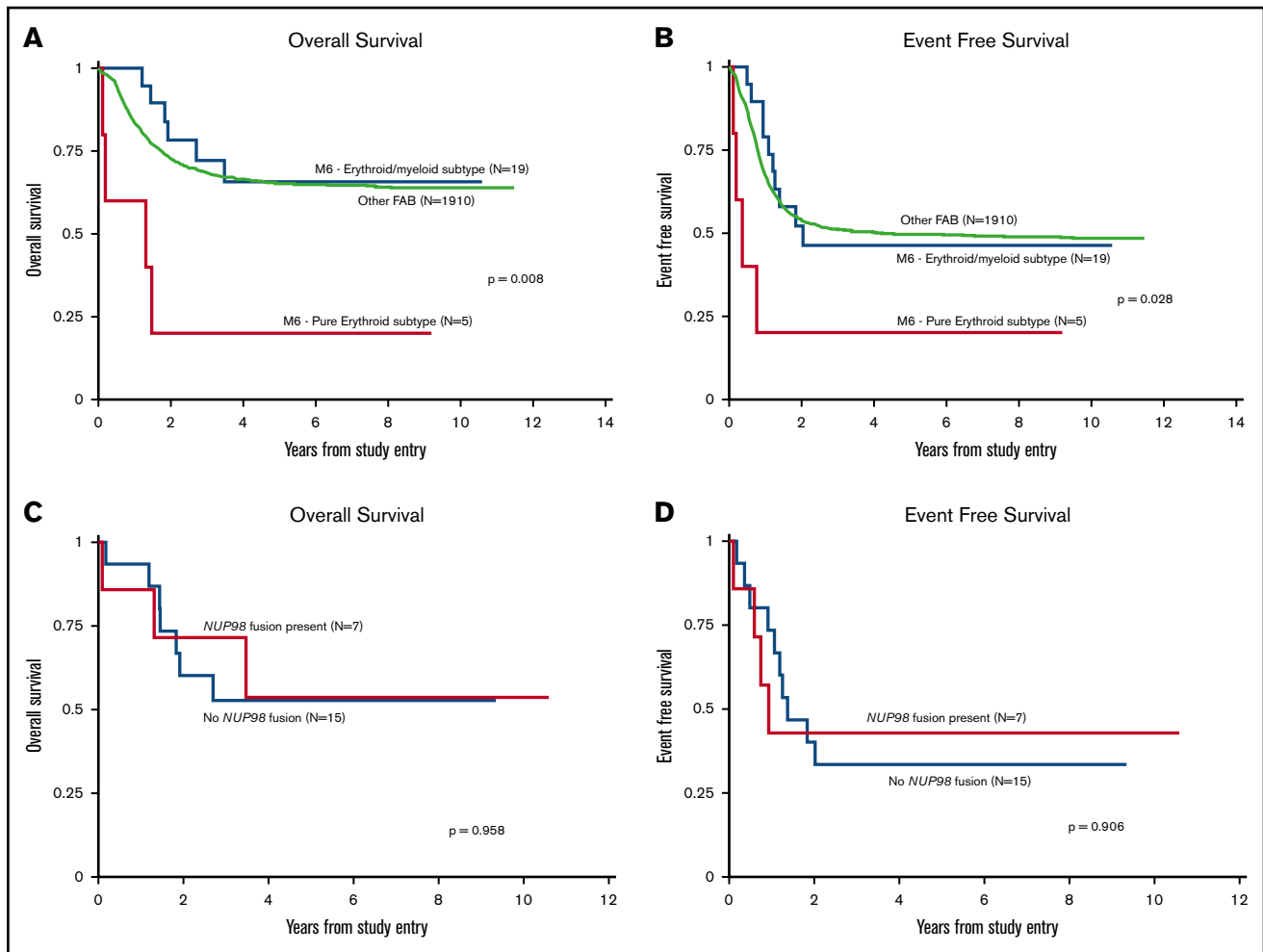
rearrangements). These 19 cases did not have significantly worse outcomes compared with the rest of the AML subtypes in these 2 COG trials.

AEL is a rare subtype of AML in children. In these 2 COG trials, only 24 (1.2%) of all central pathology review cases of AML had the AEL immunophenotype. Because this disease usually affects older individuals, studies on childhood AEL are rare. Despite the small number of patients in the current study, it is one of the largest on

children and young adults with AEL, and one of the few that uses the current WHO standards of classification. The previous COG trial (CCG-2891) reported 19 cases of AML FAB M6 (2.2% of the study).<sup>11</sup> Of note, that COG study included both erythroid/myeloid and pure erythroid subtypes of M6 but required that myeloblasts constitute >30% of all nonerythroid cells, and it therefore may have underestimated the number of cases of AEL.<sup>17</sup> Median ages of cases were similar for the previous CCG-2891 trial and the current study (both, 10.2 years).<sup>11</sup>

The CCG-2891 pediatric MDS/AML trial reported a 5-year OS and EFS of 31.6% ± 21% and 21.1% ± 19%, respectively, in cases of AEL.<sup>11</sup> However, that cohort included both pure erythroid and erythroid/myeloid subtypes. The current study shows that patients with PEL have markedly poorer outcomes than those with erythroid/myeloid leukemia. Although previous studies report reduced survival for patients with PEL in adult and mixed cohorts,<sup>15,30</sup> to the best of our knowledge, our study is the first to show this finding in a mainly pediatric cohort. Although this small cohort of PEL cases had a high rate of complex karyotypes (80%), the presence of a complex karyotype did not affect outcomes, suggesting that other factors drive the poor prognosis associated with those with PEL.

Of the 22 cases analyzed by using RNA-seq, 9 (41%) had identified fusions, 7 (32%) of which involved *NUP98*. This finding is substantially higher than the rate of 3.8% (22 of 574) reported in a pediatric AML cohort,<sup>31</sup> the rate of 6.6% (82 of 1231) in all pediatric AML cases in AAML1031, and the rate of 20% (7 of 35) in pediatric AEL reported by Iacobucci et al<sup>15</sup> (although 4 of the *NUP98* fusion patients included in the study by Iacobucci et al are also present in our study). Specifically, the rate of *NUP98* fusions in patients with morphologic PEL was 60% in the current study (none of whose fusions were identified in the study by Iacobucci et al). The supplemental data from Iacobucci et al identified 2 of 4 pediatric PEL with *NUP98* fusions, also supporting this increased rate of *NUP98* fusions in this specific type of pediatric leukemia. Presence of *NUP98* fusions has been associated with poor OS in patients with AEL, with a 5-year OS of 35.7% reported in a small pediatric series (n = 7) by Iacobucci et al. In both the study by Iacobucci et al and our series, the most common *NUP98* fusion partner was *KDM5A*, although *NUP98* is known to have multiple fusion partners. Again, we note that there was some overlap of cases between the current study and that by Iacobucci et al, including 4 shared cases that had *NUP98* fusions (two *NUP98-NSD1* and two *NUP98-KDM5A*). The specific *NUP98-KDM5A* cryptic fusion is typically found in acute megakaryoblastic leukemias<sup>32-35</sup> but rarely identified in FAB M0, M2, M5, and M6 leukemias.<sup>31,33,36</sup> Interestingly, 2 of the *NUP98-KDM5A* cases reported in the current cohort expressed megakaryocytic antigens such as CD41, CD42b, and CD61 in addition to erythroid antigens glycophorin A and CD71. The incidence of *NUP98-KDM5A* for all pediatric leukemias is 1.4%,<sup>33</sup> but it is enriched in acute megakaryoblastic leukemias with rates of 9% to 11.5%.<sup>33,34,36,37</sup> The median age of patients with acute megakaryoblastic leukemia and this translocation is 1.8 to 1.9 years (range, 0.8-8.5 years)<sup>33,34</sup> similar to the median and age range of the 4 cases in our cohort. In the study by Hara et al,<sup>36</sup> 3 of the 4 cases of *NUP98-KDM5A* fusion were associated with complex karyotypes and 1 had a normal karyotype, which is the same as the ratio reported in our series. The prognosis of patients with acute megakaryoblastic leukemia and *NUP98-KDM5A* fusions is reportedly poor, with one cohort by de Rooij et al<sup>34</sup> having



**Figure 3. Kaplan-Meier curves.** OS (A) and EFS (B) according to morphologic subtype. All other centrally reviewed leukemias were compared with the AEL of the pure erythroid subtype and the AEL of the erythroid/myeloid subtype. As shown, cases with PEL subtype had a significantly worse OS and EFS than those of the erythroid/myeloid subtype. OS (C) and EFS (D) of AEL cases according to the presence or absence of *NUP98* fusion.

a 4-year OS of  $36\% \pm 13\%$ , another cohort of de Rooij et al<sup>37</sup> reporting a 5-year OS of  $35\% \pm 16\%$ , and a third cohort of de Rooij et al<sup>33</sup> reporting a 5-year OS of  $22\% \pm 14\%$ .<sup>34,36</sup> Our series show a similar 5-year OS of  $25\% \pm 43\%$  for cases of AEL with *NUP98-KDM5A* fusions. In general, this particular fusion may portend as a poor prognosis for AEL as it did in these prior studies of acute megakaryoblastic leukemia.

Given that there is immunophenotypic overlap between some of our *NUP98-KDM5A* fusion cases and acute megakaryoblastic leukemia, and that this fusion is associated with such a poor prognosis, the diagnosis of AML with *NUP98-KDM5A* may warrant being classified as a recurrent genetic abnormality. Although the 4 cases of leukemia with *NUP98-KDM5A* did not have a statistically significant difference in outcome in our series compared with those without this specific fusion, this finding is most likely hampered by the small number of cases in our series and will need to be validated in a larger cohort. If confirmed, this fusion seems to be more important than the morphologic and immunophenotypic findings.

Alternatively, as megakaryocytic antigens were identified in 3 additional cases (including the PEL with *NUP98-SET* fusion),

there may just be more immunophenotypic overlap between AEL and acute megakaryoblastic leukemia. Of note, the rates of expression of these antigens may be underrepresented due to the manner of data collection for COG trials and/or the lack of these antigens on initial leukemia panels, as 14 cases did not have megakaryocytic antigens reported. Rare cases of AML with mixed megakaryoblastic and erythroblastic origin have been reported,<sup>38,39</sup> and shared messenger RNAs have been found to be expressed in these different leukemia subtypes.<sup>40</sup>

Although this study is one of the largest cohorts of children and young adults with AEL, it is still hampered by small numbers, especially when analyzing those with PEL. Given the small size of these groups, this study is more exploratory, and the comparisons being made are ad hoc analyses. In addition, we combined patients from 2 different clinical trials; the combination of cases from these trials was feasible, as they did not have statistically significant differences in OS or EFS.<sup>16</sup> However, we cannot determine if there were similar enrollment biases in the AAML0531 compared with those in AAML1031 that were previously reported.<sup>41</sup> Although our subgroups of PEL and erythroid/myeloid leukemias had outcomes

similar to those of prior studies, our findings of increased *NUP98* fusions, especially in those with PEL, will need to be validated in a larger study. In addition, we were unable to compare our cohort of newly classified MDS cases to a similar MDS cohort, as neither COG trial included cases with <20% blasts unless they were classified as AEL or with a specific recurrent genetic abnormality. Similarly, we did not perform case-control analyses using AEL with other AML from these trials.

In summary, although we could not determine the exact etiology underlying the poor prognosis of children with PEL, our study did find that 2 of 5 cases of PEL had *NUP98-KDM5A* fusions, and both were associated with poor outcomes. The poor outcomes were not related to presence of a complex karyotype, any *NUP98* fusion, or multilineage dysplasia. However, our study did not include a large number of PEL cases, and we therefore could not perform a comprehensive statistical analysis. We recommend that future studies analyze larger cohorts that include data on transcriptomic and epigenetic changes to identify the underlying causes for poor outcomes in these patients.

## Acknowledgments

The authors thank Vani Shanker from the Department of Scientific Editing (St. Jude Children's Research Hospital, Memphis, TN), for her thorough reading and editing of the manuscript. They also thank the patients and families for participating in these COG trials.

This research was supported by COG Chairs grant U10CA098543, NCTN Network Group Operations Center grant U10CA180886,

Statistics and Data Center grant U10CA098413, and NCTN Statistics and Data Center grant U10CA180899 from the National Institutes of Health, National Cancer Institute; St. Baldrick's Foundation; Seattle Children's Hospital Mark Alan Bomgardner Endowment (K.M.C.); and Canada Research Chair in Pediatric Oncology Supportive Care (L.S.).

## Authorship

Contribution: A.E.H.-M., J.K.C., and S.B.K. performed central pathology reviews; K.M.C., A.E.H.-M., J.K.C., and S.B.K. reviewed cases for multilineage dysplasia; B.A.H. and S.C.R. performed central cytogenetic reviews; J.S. and R.E.R. screened molecular and fusion results; T.A.A. and Y.-C.W. performed statistics; K.M.C., T.A.A., and Y.-C.W. analyzed the data; A.S.G., R.A., S.M., and L.S. were COG study chairs and vice chairs; K.M.C. wrote the manuscript; and all authors edited and approved the final version of the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: K.M.C., 0000-0002-1898-1252; J.K.C., 0000-0002-2861-0180; R.A., 0000-0001-7482-5644; A.S.G., 0000-0003-1513-2893; S.B.K., 0000-0001-8944-2128.

Correspondence: Karen M. Chisholm, Seattle Children's Hospital, Department of Laboratories, 4800 Sand Point Way NE, OC.8.720, Seattle, WA 98105; e-mail: karen.chisholm@seattlechildrens.org.

## References

1. Howlader N, Noone AM, Krapcho M, et al SEER Cancer Statistics Review, 1975-2017, National Cancer Institute, Bethesda, MD. [https://seer.cancer.gov/csr/1975\\_2017/](https://seer.cancer.gov/csr/1975_2017/) (based on November 2019 SEER data submission). Accessed 1 August 2020.
2. Bennett JM, Catovsky D, Daniel MT, et al. Proposals for the classification of the acute leukaemias. French-American-British (FAB) co-operative group. *Br J Haematol*. 1976;33(4):451-458.
3. Bennett JM, Catovsky D, Daniel MT, et al. Proposed revised criteria for the classification of acute myeloid leukemia. A report of the French-American-British Cooperative Group. *Ann Intern Med*. 1985;103(4):620-625.
4. Brunning RD, Matutes E, Flandrin G, et al. Acute myeloid leukemia, not otherwise categorised. In: Jaffe ES, Harris NL, Stein H, Vardiman JW, eds. *World Health Organization Classification of Tumours: Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues*, Lyon, France: International Agency for Research on Cancer; 2001:91-105.
5. Hasserjian RP, Zuo Z, Garcia C, et al. Acute erythroid leukemia: a reassessment using criteria refined in the 2008 WHO classification. *Blood*. 2010; 115(10):1985-1992.
6. Honda Y, Manabe A, Tsuchida M, et al; From the MDS Committee, the Japanese Society of Pediatric Hematology. Clinicopathological characteristics of erythroblast-rich RAEB and AML M6a in children. *Int J Hematol*. 2008;88(5):524-529.
7. Liu CJ, Hong YC, Yang CF, et al. Clinicopathologic features and outcome of acute erythroid leukemia based on 2008 revised World Health Organization classification. *Leuk Lymphoma*. 2012;53(2):289-294.
8. Selby DM, Valdez R, Schnitzer B, Ross CW, Finn WG. Diagnostic criteria for acute erythroleukemia. *Blood*. 2003;101(7):2895-2896.
9. Wang SA, Tang G, Fadare O, et al. Erythroid-predominant myelodysplastic syndromes: enumeration of blasts from nonerythroid rather than total marrow cells provides superior risk stratification. *Mod Pathol*. 2008;21(11):1394-1402.
10. Arber DA, Brunning RD, Orazi A, et al. Acute myeloid leukaemia, NOS. In: Swerdlow SH, Campo E, Harris NL, eds., et al. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*, Lyon, France: International Agency for Research on Cancer; 2017:156-168.
11. Barnard DR, Alonzo TA, Gerbing RB, Lange B, Woods WG; Children's Oncology Group. Comparison of childhood myelodysplastic syndrome, AML FAB M6 or M7, CCG 2891: report from the Children's Oncology Group. *Pediatr Blood Cancer*. 2007;49(1):17-22.
12. Bennett JM, Begg CB. Eastern Cooperative Oncology Group study of the cytochemistry of adult acute myeloid leukemia by correlation of subtypes with response and survival. *Cancer Res*. 1981;41(11 pt 2):4833-4837.
13. Malkin D, Freedman MH. Childhood erythroleukemia: review of clinical and biological features. *Am J Pediatr Hematol Oncol*. 1989;11(3): 348-359.



14. Gamis AS, Alonzo TA, Meshinchi S, et al. Gemtuzumab ozogamicin in children and adolescents with de novo acute myeloid leukemia improves event-free survival by reducing relapse risk: results from the randomized phase III Children's Oncology Group trial AAML0531. *J Clin Oncol.* 2014;32(27):3021-3032.
15. Iacobucci I, Wen J, Meggendorfer M, et al. Genomic subtyping and therapeutic targeting of acute erythroleukemia. *Nat Genet.* 2019;51(4):694-704.
16. Aplenc R, Meshinchi S, Sung L, et al. Bortezomib with standard chemotherapy for children with acute myeloid leukemia does not improve treatment outcomes: a report from the Children's Oncology Group. *Haematologica.* 2020;105(7):1879-1886.
17. Arber DA, Brunning RD, Orazi A, et al. Acute myeloid leukemia, not otherwise specified. In: Swerdlow SH, Campo E, Harris NL, eds., et al. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*, Lyon, France: International Agency for Research on Cancer; 2008:134-136..
18. Arber DA, Brunning RD, Le Beau MM, et al. Acute myeloid leukaemia with recurrent genetic abnormalities. In: Swerdlow SH, Campo E, Harris NL, eds., et al. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*, Lyon, France: International Agency for Research on Cancer; 2017:130-149..
19. Arber DA, Brunning RD, Orazi A, et al. Acute myeloid leukaemia with myelodysplasia-related changes. In: Swerdlow SH, Campo E, Harris NL, eds., et al. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*, Lyon, France: International Agency for Research on Cancer; 2017:150-152..
20. Hasserjian RP, Orazi A, Brunning RD, et al. Myelodysplastic syndromes: overview. In: Swerdlow SH, Campo E, Harris NL, eds., et al. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*, Revised 4th ed.. Lyon, France: International Agency for Research on Cancer; 2017:98-106..
21. Brown P, McIntyre E, Rau R, et al. The incidence and clinical significance of nucleophosmin mutations in childhood AML. *Blood.* 2007;110(3):979-985.
22. Ho PA, Alonzo TA, Gerbing RB, et al. Prevalence and prognostic implications of CEBPA mutations in pediatric acute myeloid leukemia (AML): a report from the Children's Oncology Group. *Blood.* 2009;113(26):6558-6566.
23. Ho PA, Zeng R, Alonzo TA, et al. Prevalence and prognostic implications of WT1 mutations in pediatric acute myeloid leukemia (AML): a report from the Children's Oncology Group. *Blood.* 2010;116(5):702-710.
24. Meshinchi S, Woods WG, Stirewalt DL, et al. Prevalence and prognostic significance of Flt3 internal tandem duplication in pediatric acute myeloid leukemia. *Blood.* 2001;97(1):89-94.
25. Haas BJ, Dobin A, Li B, Stransky N, Pochet N, Regev A. Accuracy assessment of fusion transcript detection via read-mapping and de novo fusion transcript assembly-based methods. *Genome Biol.* 2019;20(1):213.
26. Robertson G, Schein J, Chiu R, et al. De novo assembly and analysis of RNA-seq data. *Nat Methods.* 2010;7(11):909-912.
27. Bolouri H, Farrar JE, Triche T Jr., et al. The molecular landscape of pediatric acute myeloid leukemia reveals recurrent structural alterations and age-specific mutational interactions [published corrections appear in *Nat Med.* 2018;24(4):526 and *Nat Med.* 2019;25(3):530]. *Nat Med.* 2018;24(1):103-112.
28. Kaplan EL, Meier P. Nonparametric estimation from incomplete observations. *J Am Stat Assoc.* 1958;53(282):457-481.
29. Kalbfleisch JD, Prentice RL. *The Statistical Analysis of Failure Time Data.* 2nd ed.. Hoboken, NJ: John Wiley & Sons, Inc.; 2002.
30. Liu W, Hasserjian RP, Hu Y, et al. Pure erythroid leukemia: a reassessment of the entity using the 2008 World Health Organization classification. *Mod Pathol.* 2011;24(3):375-383.
31. Struski S, Lagarde S, Bories P, et al. NUP98 is rearranged in 3.8% of pediatric AML forming a clinical and molecular homogenous group with a poor prognosis. *Leukemia.* 2017;31(3):565-572.
32. Bisio V, Zampini M, Tregnago C, et al. NUP98-fusion transcripts characterize different biological entities within acute myeloid leukemia: a report from the AIEOP-AML group. *Leukemia.* 2017;31(4):974-977.
33. de Rooij JD, Hollink IH, Arentsen-Peters ST, et al. NUP98/JARID1A is a novel recurrent abnormality in pediatric acute megakaryoblastic leukemia with a distinct HOX gene expression pattern. *Leukemia.* 2013;27(12):2280-2288.
34. de Rooij JD, Masetti R, van den Heuvel-Eibrink MM, et al. Recurrent abnormalities can be used for risk group stratification in pediatric AMKL: a retrospective intergroup study. *Blood.* 2016;127(26):3424-3430.
35. van Zutven LJ, Onen E, Velthuisen SC, et al. Identification of NUP98 abnormalities in acute leukemia: JARID1A (12p13) as a new partner gene. *Genes Chromosomes Cancer.* 2006;45(5):437-446.
36. Hara Y, Shiba N, Ohki K, et al. Prognostic impact of specific molecular profiles in pediatric acute megakaryoblastic leukemia in non-Down syndrome. *Genes Chromosomes Cancer.* 2017;56(5):394-404.
37. de Rooij JD, Branstetter C, Ma J, et al. Pediatric non-Down syndrome acute megakaryoblastic leukemia is characterized by distinct genomic subsets with varying outcomes. *Nat Genet.* 2017;49(3):451-456.
38. Daniëls L, Guerti K, Vermeulen K, et al. Acute myeloid leukaemia of mixed megakaryocytic and erythroid origin: a case report and review of the literature. *Acta Clin Belg.* 2007;62(5):308-314.
39. Wang SA, Hasserjian RP. Acute erythroleukemias, acute megakaryoblastic leukemias, and reactive mimics: a guide to a number of perplexing entities. *Am J Clin Pathol.* 2015;144(1):44-60.
40. Linari S, Vannucchi AM, Ciolli S, et al. Coexpression of erythroid and megakaryocytic genes in acute erythroblastic (FAB M6) and megakaryoblastic (FAB M7) leukaemias. *Br J Haematol.* 1998;102(5):1335-1337.
41. Winestone LE, Getz KD, Rao P, et al. Disparities in pediatric acute myeloid leukemia (AML) clinical trial enrollment. *Leuk Lymphoma.* 2019;60(9):2190-2198.