TO THE EDITOR:

CBL-MZ is not a single biological entity: evidence from genomic analysis and prolonged clinical follow-up

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> The term "clonal B cell lymphocytosis of marginal zone origin" (CBL-MZ)^{1,2} has recently been suggested for asymptomatic individuals whose routine blood count shows a persistent modest lymphocytosis that is usually accompanied by bone marrow involvement. This immunophenotype is suggestive of marginal zone/postgerminal center derivation, but no other features of a chronic B cell lymphoproliferative disorder are found, other than a low-level paraprotein in some cases. Cases with a clonal lymphocyte count $<5 \times 10^9$ /L would fall within the revised World Health Organization (WHO) category of non-chronic lymphocytic leukemia-type monoclonal B-cell lymphocytosis. In 3 previous series of CBL-MZ consisting of 102, 53, and 16 cases with median follow-ups (FUs) of 60, 34, and 44 months, respectively,^{1,3,4} the overall incidence of progression was 15.7%, with the majority (11.1%) developing splenomegaly that frequently did not require treatment. However, it remains unclear whether CBL-MZ is the precursor to 1 or several well-defined WHO entities and what factors predict disease progression. To address this, we performed a genomic analysis of a well-characterized cohort of CBL-MZ cases with long FU and provide evidence to show that CBL-MZ is not a single biological entity.

> This study includes data from 37 patients with CBL-MZ diagnosed and managed at the Royal Bournemouth Hospital. Clinical, routine laboratory, morphological, immunophenotypic, immunogenetic, and cytogenetic data have been reported on 36 cases using previously described methods.¹ Informed patient consent was obtained according to the Declaration of Helsinki, and the ethical aspect of the study was approved by the Somerset Research and Ethics Committee.

DNA from blood-derived tumor cells (n = 37) at diagnosis was analyzed with a bespoke HaloPlex Target Enrichment System (Agilent Technologies) that enriched 2.39 Mb of genomic DNA for the coding regions of 768 genes, as previously described.⁵ From this panel, the following candidate genes were selected for Sanger validation (primers and conditions are listed in supplemental Table 1) based on the high prevalence of somatic mutations in similar mature B-cell malignancies: *KLF2* and *NOTCH2* (splenic marginal zone lymphoma [SMZL]),⁵⁻⁹ *CCND3* and *BCOR* (splenic diffuse red pulp lymphoma [SDRPL]),^{10,11} *MAP2K1* (hairy cell variant [HCL-v]),¹² *MYD88* (lymphoplasmacytic lymphoma [LPL]),¹³ *BRAF V600E* (hairy cell leukemia and nodal marginal zone lymphoma [MZL]),^{14,15} and *TNFAIP3* and *TP53* (not disease specific). DNA from buccal cells (n = 22) was used to confirm the somatic origin of 14 of 15 variants identified in 7 of these genes.

The study included 20 men and 17 women (1.2:1 ratio). The median age at presentation was 73.2 years (range 47.8-95.5 years). Key clinical and laboratory data and additional demographic and cytogenetic data are provided in Table 1 and supplemental Table 2, respectively. Lymphocyte morphology was heterogeneous in all cases, with a variable percentage of villous and lymphoplasmacytoid cells. No case had the typical morphological features of HCL-v or SDRPL. The immunophenotype was uniform, with expression of moderate Smlg, CD19, and CD49d and lack of CD10, CD38, and CD5, with the exception of 5 cases with weak CD5 positivity.

With a median FU of 9.6 years (range 2.5-22.4), 28 of 37 cases (75.7%) remained stable of whom 11 have died after a median FU of 8.8 years (range 2.5-14.3), and 17 remained stable after a median FU of 9.7 years (range 2.9-14.6 years). Nine of 37 (24.3%) cases showed evidence of progressive disease, and 3 died. The median time to progression was 69 months (range 47-175). Seven patients

	Candidate gene mutations	None	TP53	MYD88	None	KLF2, NOTCH2	TNFAIP3	None	MYD88	MYD88	None	None	None	CCND3	MYD88	TP53, CCND3	None	None	TP53, CCND3	MYD88	None	None	None	None	None	None	None	None	MAP2K1	None	None		None
	Key cytogenetics abnormalities	del(3q)	*CK inc i(17)(q10)	None	del(14q)	del(14q)	ð	dup(3q)	Х	t(2;7)	None	+12	+12	None	+12	i(17) (q10)	None	None	del(7q), +12	None	None	None	+3,+12, i(17)(q10)	None	None	None	del(7q)	del(14q)	None	del(7q)	i(17) (q10)	None	
	% IGVH identity	89.7	92.36	95.49	96.41	98.13	94.14	96.63	93.47	96.6	87.37	96.63	89.8	96.37	93.4	95.51	100	97.54	96.88	95.49	100	92.74	98.32, 91.93	95.92	92.65	89.51	100	91.7	93.06	94.76	93.85	96.41	
	IGHV gene usage	V5-51	V3-64*01/*02	V3-30*03/*18/V3-30-5*01	V4-34*01/*02/*07	V1-2*04	V4-34*01/*02	V5-51*01	V4-34*01/*02	V3-73*02	V3-66*01/*04	V6-1*01	V4-34*01/*02/*12	V4-4*02	V3-23*01/V3-23D*01	V4-59*01	V3-66*02	V4-59*01	V3-7*02	V3-7*01	V1-2*04	V4-38-2*01	V6-1*01(9 bp ins), V4-59*01	V4-34*01/*02	V4-34*01/*02	V1-69*01/*11/*12/V1-69D*01	V4-34*01	V4-30-2*01	V3-9*01	V3-21*01/*02	V1-3*01	V4-39*01	
	Presentation paraprotein, g/L	No	No	Mk	No	No	No	No	GK 1.0	ML 1.7	No	No	No	No	GK 9.8	No	No	No	No	GK 5.2	MK 0.5	No	No	No	No	No	ML trace	No	No	GK trace	No	No	
Overall	survival, mo	177.48	89.49	134	164.44	51.09	118.51	45.9	35.94	269	171.86	95.57	115.68	136.51	29.73	39.39	53.85	126.78	39.66	107.89	19.88	105	96.16	90.94	88.41	40.77	82	149.13	75.43	160.72	68.34	105.13	
Time to	progression, mo	147	65	66	119	47	69	57	150	175	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	na	
	Diagnosis at progression	SLLU	SMZL	H TZWS	SMZL	SMZL	H MZH	H SMZL H	LPL H	SLLU	na	па	na	па	na	na	na	па	na	na	na	na	na	na	na	na	na	na	na	na	na	na	
	Progressive disease	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	
	Case	-	7	ო	4	ß	9	7	80	o	10	÷	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	

Table 1. Patient immunogenetics, cytogenetics, and mutational data

Case	Progressive disease	Diagnosis at progression	Time to progression, mo	Overall survival, mo	Presentation paraprotein, g/L	IGHV gene usage	% IGVH identity	Key cytogenetics abnormalities	Candidate gene mutations
34	No	na	na	146.92	No	V4-39*01	94.07	+12	None
35	No	na	na	130.83	No	V5-51*01	97.98	None	None
36	No	na	na	5192	No	V3-23*01/V3-23D*01	96.18	i(17)(q10)	None
37	No	па	na	153.53	GL trace	V1-69-2*01	96.63	None	None
CK, coi	mplex karyotype based c	in >3 G-banding aberrati	ons; GL, G-lambda; I	 Histological c 	diagnosis; MK, M-kappa;	na, not applicable; SLLU, splenic	B-cell lymphoma/leuke	∍mia unclassifiable.	

(cases 1-5, 7, and 9) developed splenomegaly that was accompanied by progressive lymphocytosis in 4 cases. In 2 patients, splenic histopathology confirmed a diagnosis of SMZL. In 3 patients, lymphocyte morphology and marrow histology at progression, combined with immunogenetic and karyotypic features, were also consistent with SMZL. Two cases, classified as splenic B cell lymphoma/leukemia unclassifiable, were too frail for further investigation; 1 (case 9) had a t(2;7)(p11;q21.2) translocation at diagnosis, which has been associated with MZLs,16,17 and 1 (case 1) exhibited progressive lymphocytosis with large circulating lymphoid cells. Case 8 developed heavy marrow infiltration with small nonvillous lymphocytes in conjunction with a low-level IgGK paraprotein and cytogenetic analysis showing del(6q) and iso(18q); LPL was considered a likely diagnosis. Case 6 underwent biopsy of orbital and abdominal wall masses, both of which showed histological and immunophenotypic features of MZL.

Fifteen genomic mutations, involving all candidate genes screened, with the exception of BCOR and BRAF V600E, were identified in 12 cases (Table 1). The most frequent was MYD88 in 5 (13.5%) cases, involving L265P (n = 3) or S219C (n = 2) and indicating that screening CBL-MZ cases only for the L265P mutation is likely to miss cases with alternate MYD88 mutations. Three patients had histologically proven SMZL, 1 had a t(2;7)(p11;q21.2) translocation, and 1 had LPL. None of these cases had a mutation of CXCR4 (data not shown). Three cases (8.1%) had mutations of TP53, all accompanied by TP53 loss, and 3 had PEST domain CCND3 mutations, although none had the typical features of SDRPL, and all had stable disease. The sole case with a MAP2K1 mutation (E203K; deleterious and damaging by Mutationtaster and Polyphen2, respectively) was stable with a FU of 66 months. Extended immunophenotypic analysis showed expression of SmlgG, FMC7, CD22, and CD11c (weak) and lack of CD103 and CD25, consistent with HCL-v, splenic B cell lymphoma/leukemia unclassifiable, or SMZL. NOTCH2 and KLF2 mutations were present in a single case that used IGHV1-2*04 and progressed to SMZL. The patient with orbital lymphoma had the recently noted association of a TNFAIP3 mutation and IGHV4-34 usage in this subset of MZL.¹⁸ Three patients had repeat genomic analysis at evolution, and no new mutations were found.

Neither cytogenetic nor immunogenetic data measured at presentation correlated with the natural history of CBL-MZ. In contrast, 5 of 9 patients with progressive disease had ≥ 1 mutation (3 MYD88 mutations; single cases with TP53, NOTCH2, KLF2, TNFAIP3 mutations), of which TP53 and NOTCH2 mutations have been associated with disease progression in MZLs, compared with 6 of 28 patients with stable disease (P = .034). Five of 6 patients with stable disease who had mutations (3 CCND3 mutations, 2 mutations in both MYD88 and TP53) died of unrelated causes. Their median FU was considerably shorter (39 months) than that of the other stable cases, raising the question of whether their CBL-MZ would have progressed with extended FU.

In summary, our clinical outcome data indicate that CBL-MZ usually pursues a stable course, but the higher rate of progression in this study compared with previous studies probably reflects the longer FU and reinforces the need for long-term clinical management and patient education on when to seek medical advice. CBL-MZ can evolve into several well-defined WHO disorders, especially those of marginal zone origin. The genomic data are consistent with this observation because, although the genomic abnormalities in CBL-MZ

Table 1. (continued)

overlap with those found in any of the well-defined entities into which it could evolve, the incidence of mutations is lower and does not mirror any specific disease. However, important caveats are the relatively small number of cases in the current study and the lack of concordance among genomic studies in other rare disorders, such as HCL-v¹⁹and SDRPL.^{11,20,21} Further larger studies, ideally including immunogenetic, whole genomic sequencing, and epigenetic data, will be required to confirm the relationship between CBL-MZ and established WHO disorders and to identify additional drivers of progressive disease. In the interim, CBL-MZ remains a useful term to define a group of asymptomatic patients with well-defined clinical, morphological, and immunophenotypic features requiring long-term FU.

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