Presence of N regions in the clonotypic DJ rearrangements of the immunoglobulin heavy-chain genes indicates an exquisitely short latency in t(4;11)-positive infant acute lymphoblastic leukemia

Karin Fasching, Simon Panzer, Oskar A. Haas, Arndt Borkhardt, Rolf Marschalek, Frank Griesinger, and E. Renate Panzer-Grümayer

Childhood acute lymphoblastic leukemia (ALL) is frequently initiated in utero at a time of developmentally regulated insertion of N regions into the DJ_H rearrangements of immunoglobulin heavy-chain (Ig_H) genes. Here it is shown that N regions are present in the clonotypic DJ_H rearrangements in 11 of 12 infant ALLs

with t(4;11). These data are compared with the 122 previously published DJ_H sequences and were found to have a pattern similar to that of ALL in children older than 3 years at diagnosis but were unlike that in children younger than 3 years who predominantly lack N regions. These findings, therefore, indicate that

t(4;11)-positive infant ALL is initiated later in fetal development than most B-cell precursor ALL from children younger than 3 years and that they have a shorter latency period already in utero. (Blood. 2001;98:2272-2274)

© 2001 by The American Society of Hematology

Introduction

Childhood acute lymphoblastic leukemia (ALL) is a heterogeneous group of leukemias with a predominance of the B-cell precursor (BCP) phenotype. A minority of these leukemias is associated with a translocation involving the mixed-lineage leukemia (*MLL*) gene on chromosome 11q23 that is fused, in 50% of patients, to the *AF4* gene on chromosome 4q21.¹ The chromosomal translocation t(4;11)(q21;q23) occurs mostly in infant ALL and confers a dismal prognosis in this age group, whereas in older children and adults the prognosis does not differ from t(4;11)-negative cases.^{2,3} Differences in the chromosomal breakpoints of the *MLL* gene between infants and children or adults with t(4;11) ALL suggest different mechanisms for the development of these instances of ALL⁴ and, thus, their different biologic functions.

Greaves⁵ proposed a 2-step model for the development of ALL with an initiating event in utero, followed by a second mutation leading to overt leukemia. Indeed, leukemia-specific chromosomal translocations and clonotypic antigen-receptor gene rearrangements at birth recently confirmed the initiation of childhood ALL in utero.⁶⁻⁹ Thus, depending on the time of clinical manifestation, the latency period varies among the different types of leukemia. In contrast to leukemias with long latency periods for which a chromosomal translocation and additional postnatal mutations are required,⁹ the extremely short latency periods in infant ALL (with *MLL* rearrangements) suggest only limited further mutagenic requirements.¹ It seems likely that not only postnatal but also prenatal development of the disease is rapid.

Assuming that the gene fusion resulting from t(4;11) in infant ALL is indeed an initiating event, its origin must be restricted to a period between the beginning of B lymphopoiesis in the fetal liver—ie, the 6th gestational week¹⁰—and possibly a few months

Submitted March 2, 2001; accepted May 29, 2001.

Supported in part by the Österreichische Kinderkrebshilfe and by a grant from the Deutsche Krebshilfe (Bo 10-124.3).

before the diagnosis of leukemia.¹¹ This translocation is assumed to occur in a cell that has already started to rearrange its Ig_H genes. The time period of these rearrangement processes during fetal development can be determined by the presence or absence of N regions between the DJ_H joinings. The addition of N nucleotides requires terminal deoxynucleotidyl transferase (TdT) that is not initially present in fetal lymphopoiesis but that has been observed by the end of the first trimester of gestation.¹²⁻¹⁴

We therefore used the leukemia clone-specific junctional regions of DJ_H rearrangements to determine the time point of a first mutation in utero in t(4;11) ALL in infancy. We show the presence of N regions between the DJ_H joinings in 11 of 12 infant ALLs with t(4;11) indicating their initiation at a later time during fetal development than most other leukemias that become apparent during the first 3 years of life.

Study design

The occurrence of t(4;11) ALL was analyzed in 13 infants. Inclusion criteria were patient age younger than 1 year at diagnosis and the presence of at least one clonotypic Ig_H rearrangement (Figure 1). All leukemias had a pro B phenotype with frequent coexpression of myeloid markers. The local institutional ethical committee approved the study, and informed consent was obtained from the parents.

DNA was extracted from cells by standard procedures.¹⁵ Amplification of the ret-oncogen confirmed the integrity of DNA in all samples. Clonal Ig_H rearrangements were determined by V_{H} -family–specific and J_{H} consensus primers for the amplification of $V(D)J_{H}$ rearrangements and of primer sets for the amplification of all incomplete DJ_{H} rearrangements, as described previously.^{16,17} Amplified products were sequenced directly, and involved gene segments were identified by BLAST sequence similarity

Reprints: E. Renate Panzer-Grümayer, Children's Cancer Research Institute, St Anna Kinderspital, Kinderspitalgasse 6, A-1090 Vienna, Austria; e-mail: panzer@ccri.univie.ac.at.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 U.S.C. section 1734.

© 2001 by The American Society of Hematology

From the Children's Cancer Research Institute, St Anna Kinderspital, and Clinic for Blood Group Serology, University of Vienna, Austria; the Department of Pediatric Hematology/Oncology, Justus Liebig Universität, Giessen, Institute of Pharmaceutical Biology, University of Frankfurt; and the Department of Hematology/Oncology, University of Göttingen, Germany

Figure 1. Nucleotide sequence of Ig_H rearrangements in t(4;11) infant ALL. Trimming of the rearranged segments is indicated by the numbers of nucleotides. N nucleotides between DD segments are shown in italics; shaded areas indicate sequence homology between 2 rearrangements.

Pat.no	DH		N		JH	VDJH gene segments
1	ctaactgggga	-0	J1-pseudogene	-0	J1	DH7-27/J1
2	aggatattgtactaatggtgtatgct	-5	gttcc	-2	J5	DH2-8/J5
3	gtattacgatattttgactggttattataa	-1	ag	-2	J5	DH3-9/J5
4a	aggatattgtagtggtggtagctg	-7	1	-4	J6	DH2-15/J6
4b	gcttcccttattgtagtggtggtagetg	-7	1	-4	J 6	DH3-9/DH2-15/J6
5	aggatattgtactaatggtgtatgc	-6	cccgatttcccccctaagccctttc	-4	J4	DH2-8/J4
6	gtattacgatattttgactggttat	-6	gt	-0	J4	DH3-9/J4
7	gtattacgatattttgactggttatt	-5	cctt	-5	J6	DH3-9/J6
8a	ggtattacgatttttggagtggttattat	-3	gggt	-8	J6	DH3-3/J6
8b	atattgtagtagtaccagctgctata	-2	gttggatgg	-4	J5	VH3/DH2-2/J5
9	gtattacgatattttgactggttattataac	-0	cctcg	-2	J5	VH4/DH3-9/J5
10a	gatattgtagtagtaccagctgctat	-0	tgagggggt	-5	J6	VH4/DH2-2/J6
10b	atattgtagtagtaccagetgetat	-0	tgagggggt	-5	J6	VH6/DH2-2/J6
11	tagcagtggctggtac	-0	ggagg	-6	J4	VH3/DH6-19/J4
12	tagcagctcgtc	-1	gcccccg	-7	J4	VH3/DH6-6/J4
13			gcgggc	-4	J5	VH3/J5

searches (http://www.ncbi.nlm.nih.gov/BLAST/) and by comparison with published sequences of all known human immunoglobulin genes (http:// www.ebi.acc.uk), allowing the assignment of nucleotides to either the V, D, and J regions or to the P and N regions by exclusion.

Results and discussion

We identified 16 Ig_H rearrangements in 13 infants with t(4;11) ALL (Figure 1A). Ten leukemias had 1 rearrangement, and 3 leukemias (patients 4, 8, 10) had 2 rearrangements. We considered only 1 of the 2 rearrangements (in patients 4 and 10) because they had identical DJ_H regions. We excluded the DJ_H rearrangement in patient 1 because the sequence between D_H and J_H was homologous to the J1 pseudogene. Thus, 13 unique rearrangements were analyzed for the inclusion of N regions. We observed in patient 4 a D-D fusion, but only the D_H gene segment most proximal to the J_H segment was included in the analysis. As depicted in Figure 1, only 1 of the 13 unique sequences lacked N regions at the DJ_{H} junction. The results from this study are compared with those from 122 previously published cases (Table 1). It appears that the lack of N regions in t(4;11) infant ALL is less common than in children with ALL who are younger than 3 years at diagnosis but that they are about as common as in children older than 3 years.^{15,18,19} Figure 1 illustrates the use of D_H and J_H families, similar to that reported previously.16,18,19

The data from this study indicate that most DJ_H regions from infant ALL with t(4;11) contain N nucleotides that developed at a time of TdT activity; hence, they were more mature than those without N regions. Interestingly, other leukemias, diagnosed in children before the age of 3, do not have N regions in their DJ_H junctions^{15,18,19} and thus have a longer latency period than the

Table 1. Occurrence of N regions between the DJ_H junction of children with BCP ALL and their ages at diagnosis

Age at diagnosis (y)	No. of ALL/study	N + n	N – n	Reference
Younger than 1 year	12	11	1	This study
Younger than 3 years	16	2	16	Wassermann et al ¹⁸
	8	5	5	Steenbergen et al ¹⁹
	14	4	12	Schneider et al ¹⁵
Older than 3 years	46	41	10	Wassermann et al ¹⁸
	39	35	12	Steenbergen et al ¹⁹

Leukemias are considered N+ if one DJ_H junction has N nucleotides inserted. They are considered N- if one DJ_H junction lacks N nucleotides. Leukemias are included in both groups if one rearrangement contains N nucleotides and the other does not.

t(4;11) infant leukemias. There are 2 groups of BCP ALL that have N regions in their clonotypic DJ_H junctions, namely t(4;11) infant ALL with a manifestation mostly in the first year of life and other leukemias with a clinical manifestation after the 3rd year of life.^{18,19} Both rearrange their DJ_H segments at a similar time during gestational development. It is obvious, however, that t(4;11) ALL has a remarkably shorter latency than the others.

We propose a model for the relation between the time of initiation of the leukemia, characterized by the clonotypic DJ_H rearrangements, and the age of the children at clinical manifestation of BCPALL (Figure 2). Ig_H rearrangements that lack N regions occur during a narrow time window-the first weeks of B lymphopoiesis in fetal liver that is TdT negative. Transformed cells with such rearrangements most likely acquire additional mutations, leading to leukemias during the first 3 years of life. Ig_H rearrangements with the addition of N nucleotides in the DJ_H junction occur later in gestation, when TdT has already been activated. These leukemias become clinically apparent during the first year of life if a t(4;11) chromosomal translocation started leukemogenesis or, in its absence, after the 3rd year of life. Alternatively, the t(4;11) translocation arises in a TdT-negative primitive cell without Ig_H rearrangements. This target cell may represent a B-plus myeloid lymphoid stem cell, as described by Cumano et al²⁰ in mouse fetal liver, which would be unique in specific stages of in utero hematopoiesis. The N region-positive DJ_H rearrangement may be a later addition during progression to leukemia. Then, unrelated rearrangements are expected, such as in t(9;22) B lymphoid blast crisis of chronic myeloid leukemia.16 However, in our series, no leukemia had multiple unrelated Ig_H rearrangements, but 2 leukemias had related rearrangements. In addition, the target cell for the t(4;11) translocation may be a rare progenitor with TdT expression at earlier stages of fetal lymphopoiesis than common B precursor cells. No such cells have been identified thus far in humans.



Figure 2. Time frame. Development of N- and N+ BCP leukemias in childhood.

It is assumed that a chromosomal translocation is an initiating event in leukemogenesis,^{9,21} which can be induced by apoptotic stimuli that lead to the generation of gene fusions in B precursor cells, thus rescuing a cell programmed to die.²² This assumption is supported by the findings that most Ig_H rearrangements in ALL are either incomplete or not potentially productive,¹⁶ underlining the immaturity of these cells, which would not survive without a transformation. It is further hypothesized that additional mutations are required for the development of leukemia. However, our

findings support the hypothesis that the t(4;11) is either sufficient for leukemogenesis or provokes efficiently further changes that lead eventually to leukemia in infancy.^{1,11}

Acknowledgment

This article is dedicated to Helmut Gadner for his 60th birthday.

References

- Dimartino JF, Cleary ML. MLL rearrangements in haematological malignancies: lessons from clinical and biological studies. Br J Haematol. 1999; 106:614-626.
- Dordelmann M, Reiter A, Borkhardt A, et al. Prednisone response is the strongest predictor of treatment outcome in infant acute lymphoblastic leukemia. Blood. 1999;94:1209-1217.
- Biondi A, Cimino G, Pieters R, Pui CH. Biological and therapeutic aspects of infant leukemia. Blood. 2000;96:24-32.
- Reichel M, Gillert E, Angermüller A, et al. Biased distribution of chromosomal breakpoints involving the MLL gene in infants versus children and adults with t(4;11) ALL. Oncogene. 2001;20:2900-2907.
- 5. Greaves M. A natural history of pediatric acute leukemias. Blood. 1993;82:1043-1051.
- Gale KB, Ford AM, Repp R, et al. Backtracking leukemia to birth: identification of clonotypic gene fusion sequences in neonatal blood spots. Proc Natl Acad Sci U S A. 1997;94:13950-13954.
- Fasching K, Panzer S, Haas OA, Marschalek R, Gadner H, Panzer-Grümayer ER. Presence of clone-specific antigen receptor gene rearrangements at birth indicates an in utero origin of diverse types of early childhood acute lymphoblastic leukemia. Blood. 2000;95:2722-2724.
- Yagi T, Hibi S, Tabata Y, et al. Detection of clonotypic IgH and TCR rearrangements in the neonatal blood spots of infants and children with B-cell precursor acute lymphoblastic leukemia. Blood. 2000;96:264-268.
- 9. Wiemels JL, Cazzaniga G, Daniotti M, et al. Pre-

natal origin of acute lymphoblastic leukemias in children. Lancet. 1999;354:1499-1503.

- Gathings WE, Lawton AR, Cooper MD, et al. Immunofluorescent studies of the development of pre-B cells, B lymphocytes and immunoglobulin isotype diversity in humans. Eur J Immunol. 1977;7:804-810.
- Greaves M. Molecular genetics, natural history and the demise of childhood leukemia. Eur J Cancer. 1999;35:173-185.
- Bodger MP, Janossy G, Bollum FJ, Burford GD, Hoffbrand AV. The ontogeny of terminal deoxynucleotidyl transferase positive cells in the human fetus. Blood. 1983;61:1125-1131.
- Raaphorst FM, Timmers E, Kenter MJ, Van Tol MJ, Vossen JM, Schuurman RK. Restricted utilization of germ-line VH3 genes and short diverse third complementarity-determining regions (CDR3) in human fetal B lymphocyte immunoglobulin heavy chain rearrangements. Eur J Immunol. 1992;22:247-251.
- Cuisinier AM, Gauthier L, Boubli L, Fougereau M, Tonnelle C. Mechanisms that generate human immunoglobulin diversity operate from the 8th week of gestation in fetal liver. Eur J Immunol. 1993;23:110-118.
- Schneider M, Panzer S, Stolz F, Fischer S, Gadner H, Panzer-Grümayer ER. Crosslineage TCR delta rearrangements occur shortly after the DJ joinings of the IgH genes in childhood precursor B ALL and display age-specific characteristics. Br J Haematol. 1997;99:115-121.
- Height SE, Swansbury GJ, Matutes E, Treleaven JG, Catovsky D, Dyer MJS. Analysis of clonal re-

arrangements of the Ig heavy chain locus in acute leukemia. Blood. 1996;87:5242-5250.

- Szczepanski T, Pongers-Willemse MJ, Langerak AW, et al. Ig heavy chain gene rearrangements in T-cell acute lymphoblastic leukemia exhibit predominant DH6–19 and DH7–27 gene usage, can result in complete V-D-J rearrangements, and are rare in Tcell receptor ab lineage. Blood. 1999;93:4079-4085.
- Wassermann R, Galili N, Yoshinori Y, Reichard BA, Shane S, Rovera G. Predominance of fetal type DJH joining in young children with B precursor lymphoblastic leukemia as evidence for an in utero transforming event. J Exp Med. 1992;176: 1577-1581.
- Steenbergen EJ, Verhagen OJHM, Van Leuwen EF, et al. B precursor acute lymphoblastic leukemia third complementarity-determining regions predominantly represent an unbiased recombination repertoire: leukemic transformation frequently occurs in fetal life. Eur J Immunol. 1994; 24:900-908.
- Cumano A, Paige CJ, Iscove N, Brady G. Bipotential precursor B cells and macrophages in murine fetal liver. Nature. 1992;356:612-615.
- Wiemels JL, Ford AM, Van Wering ER, Postma A, Greaves M. Protracted and variable latency of acute lymphoblastic leukemia after *TEL-AML1* gene fusion in utero. Blood. 1999;94:1057-1062.
- Stanulla M, Wang J, Chervinsky DS, Thandla S, Aplan PD. DNA cleavage within the MLL breakpoint cluster region is a specific event which occurs as part of higher-order chromatin fragmentation during the initial stages of apoptosis. Mol Cell Biol. 1997;17:4070-4079.