

Transcriptional activation of urokinase by the Krüppel-like factor Zf9/COPEB activates latent TGF- β 1 in vascular endothelial cells

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Understanding the regulation of genes controlling fibrinolysis and matrix homeostasis is essential for elucidating the basis of tissue repair. A recently described novel Krüppel-like factor, Zf9, is up-regulated in acute liver injury in activated hepatic stellate cells. Because Zf9 can be induced widely, its activity was examined in vascular endothelium, a key cell in vascular injury. Zf9 is induced as an immediate-early response gene in bovine aortic endothelial cells (BAECs) following treatment with serum or phorbol ester. Zf9 transcriptionally activates urokinase plasminogen activator (uPA).

Recombinant Zf9-GST binds to wild-type but not mutated 'GC-box' motifs within the human uPA promoter (−63 to −32), with greatest affinity to the middle of 3 contiguous GC boxes. Transient transfection of Zf9 drives transactivation of a full-length uPA promoter- and GC box-construct, but not a uPA promoter-construct devoid of GC boxes. Transactivation of uPA by Zf9 is also supported in *Drosophila* S2 cells. Most importantly, transiently transfected Zf9 up-regulates endogenous uPA messenger RNA and activity in BAECs, resulting in increased bioactive transforming growth factor-

beta (TGF- β) via enhancement of proteolytic activation of the latent molecule. Furthermore, concomitant expression of Zf9 and uPA proteins was observed in arterial endothelial cells after balloon injury in rats, suggesting a potential role of Zf9 in uPA expression not only in vitro but also in vivo. These findings suggest a role of Zf9 in the injury response by enhancing uPA synthesis and subsequent activation of latent TGF- β . (Blood. 2000;95:1309-1316)

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Introduction

Coordinated gene expression is a key component of the response to tissue injury. Classes of induced genes include extracellular matrix proteins,¹⁻⁴ growth factors/cytokines such as transforming growth factor- β (TGF- β),¹⁻⁶ and proteases such as urokinase plasminogen activator (uPA).¹⁻⁹ uPA was originally discovered as a fibrinolytic factor responsible for conversion of plasminogen to plasmin in dissolving vascular thrombosis.^{10,11} However, it also has a key role in tissue remodeling,^{7,9,12} metastasis,^{13,14} apoptosis,^{14,15} and immunity¹⁴ either directly or through activation of other enzymes including matrix proteases^{9,13,16} and cytokines.^{5,17} uPA has been implicated in the activation of latent TGF- β . By activating plasminogen to plasmin, uPA can stimulate proteolytic activation of latent TGF- β in several tissues and cell types especially in pathogenic situations.^{15,17-25} TGF- β in turn mediates key components of the injury response. It down-regulates the inflammation, modulates growth inhibition and differentiation, and enhances extracellular matrix production in a variety of cell types.¹⁻⁴ Vascular endothelial cells are a potent source of uPA, and given their critical position in tissue remodeling, are a key regulator of the injury response.⁶ Thus, control of uPA/plasmin and thereby TGF- β levels in endothelial cells is an important event in the response to tissue injury.

Circulating levels of plasminogen are fairly constant, so cellular plasmin levels are controlled primarily by plasminogen activator (PA) activity. Cellular PA activity is, in turn, controlled not only by regulation of uPA synthesis but also by levels of its receptor (uPAR).^{7,11} and inhibitor (PAI-1).¹⁰ uPAR is essential for localizing the PA activity on the cell surface leading to acceleration of plasminogen activation,^{7,11} whereas PAI-1 rapidly inhibits PA activity.¹⁰ Thus, the balance between uPA, uPAR, and PAI-1 levels is critical for regulating the level of net fibrinolytic activity.^{10,11} Regulation of uPA gene expression is therefore a key control point of cellular PA activity. It is known that uPA expression by vascular cells is up-regulated during the process of tissue repair in vivo.^{7,8,12} However, the molecular basis for injury-induced uPA gene expression is not completely understood. Several transcription factors have been reported to participate in this response. These include LFB3/HNF²⁶ CRE-binding protein,⁷ Ets-1,²⁷ PEA3,²⁸⁻³⁰ UEF1-4,²⁸⁻³⁰ AP-1,²⁸⁻³¹ and NF- κ B³² demonstrated in the cultured cells and Egr-1 in injured rat aorta.³³ Sp1, the initial Krüppel-like zinc finger protein,^{34,35} is also shown to be another important transcriptional activator of uPA in vitro through its interaction with 3 contiguous GC boxes immediately upstream of the TATA box in the uPA promoter.^{36,37}

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Zf9/COPEB/GBF (soon to be renamed KLF6) is a novel zinc finger transcription factor recently cloned from liver, placenta, and leukocyte complementary DNA (cDNA) libraries.³⁸⁻⁴¹ The molecule belongs to the family of Krüppel-like transcription factors,⁴²⁻⁵² all of which contain 3 contiguous C₂H₂ zinc fingers at the carboxyl-terminal domain, and recognize either GC box motifs or CACCC motifs in responsive promoters.³⁹⁻⁵² A proximal role for Zf9 in response to tissue injury has been suggested by its rapid induction in activated hepatic stellate cells, the key fibrogenic cell type in liver injury and repair.^{38,39} Moreover, Zf9 transactivates key genes comprising the injury response, including collagen $\alpha 1(I)$, TGF- $\beta 1$, and types I and II TGF- β receptors in hepatic stellate cells.^{39,53}

Given its widespread expression and interaction with GC box motifs,³⁹⁻⁴¹ we have explored a potential role for Zf9 in uPA gene expression by endothelial cells. We have identified Zf9 messenger RNA (mRNA) and protein in cultured endothelial cells, which are induced by serum or phorbol esters. Zf9 stimulates uPA transcription and endogenous uPA activity in these cells, leading to proteolytic activation of latent TGF- $\beta 1$. Furthermore, concomitant expression of Zf9 and uPA proteins was induced in arterial endothelial cells following carotid balloon injury in rats. The data suggest that Zf9 may have an important role in regulation of PA/plasmin activity and TGF- $\beta 1$ activity in vascular endothelium.

Materials and methods

Materials

Phorbol myristate acetate (PMA) was purchased from Sigma (Chemical Co, St Louis, MO). Rabbit polyclonal antibodies against rat Zf9 and rat uPA were obtained from Santa Cruz Biotechnology (Santa Cruz, CA) and American Diagnostica (Greenwich, CT), respectively. Recombinant human TGF- $\beta 1$ and its antibody, which neutralizes all subtypes of TGF- β s, were from R&D System (Minneapolis, MN) and Genzyme Diagnostics (Cambridge, MA), respectively.

Plasmids and cells

Zf9 mammalian (*Zf9-pCIneo*) and *Drosophila* (*Zf9-pAC*) expression plasmids were constructed as previously described.³⁹ The uPA-luciferase expressing vector, pGL2-2350, containing the human uPA promoter (−2345 to 32), its deletion mutant, pGL2-GC, containing the first 65 bp and GC box and TATA box region (−2345 to −2280 and −91 to +32, respectively), and another deletion mutant, pGL2- Δ GC, which contains uPA promoter devoid of the GC box region (−71 to −28), were as described.^{36,37} Bovine aortic endothelial cells (BAECs) were isolated and grown in α minimal essential medium (α MEM; GIBCO BRL, Life Technologies, Rockville, MD) containing 10% fetal calf serum (FCS). *Drosophila* S2 cells were generous gifts from Dr H. Tanaka, National Institute of Sericultural and Entomological Science, Tsukuba, Japan and maintained in Shields and Sang M3 insect medium (Sigma) containing 10% FCS.

Northern blotting

Isolation of total RNA from BAECs and its Northern blot analyses were performed as described previously.⁵⁴ RNA was separated through 1% agarose-formaldehyde gel electrophoresis and transferred onto the Biotrans nylon membranes (Pall Biosupport, New York, NY). Membranes were hybridized with ³²P-labeled cDNA for either human Zf9,³⁹ bovine uPA,³⁷ bovine uPAR,⁵⁴ or human PAI-1,⁵⁴ and rehybridized with chicken glyceraldehyde-3-phosphate dehydrogenase (GAPDH) as an internal control.⁵⁴ Signals were quantitated using a Fujix BAS 2000 Bioimaging analyzer (Fuji Photo-Film, Tokyo, Japan).

Western blotting

Western blotting was performed as described previously³⁷ using rabbit anti-Zf9 polyclonal antibody (final 1:10,000 dilution), and goat antirabbit IgG antibodies conjugated with peroxidase (Jackson ImmunoResearch Laboratories, West Grove, PA). The signals were detected with an Amersham-Pharmacia (Buckinghamshire, UK) ECL system. The blot was reprobed with antirabbit GAPDH (Chemicon Int., Temecula, CA).

Gel shift assay

Rat *z9* cDNA was cloned into pGEX-3T (Pharmacia Biotech, Sweden), and transformed in *Escherichia coli* BL21. Following induction by 2 mM isopropyl β -D-thiogalactoside for 4 hours, the cells were disrupted by sonication, and Zf9-glutathione S-transferase (GST) fusion protein was purified using glutathione-Sepharose (Amersham-Pharmacia) from cell homogenates. Purified GST protein (Santa Cruz) was used as a control. Oligonucleotides corresponding to −63 to −32 of the human uPA promoter (wild-type uPA GC box),³⁷ which contains 3 GC boxes, as well as mutant oligos, in which mutations were made in 1 or all of 3 GC boxes, were synthesized. Sequences of these oligonucleotides are indicated in Figure 2A. Oligonucleotides were double-stranded and labeled with γ -³²P adenosine triphosphate (DuPont, Wilmington, DE) by polynucleotide kinase (Boehringer Mannheim, Mannheim, Germany). For gel shift assays, 20 ng of Zf9-GST fusion protein was preincubated for 15 minutes at 4°C with or without unlabeled oligonucleotide, then 3.5 ng labeled oligonucleotide (10,000 μ Ci/mol) was added in the presence of 1 μ g of dI-dC (Pharmacia) in 20 μ L binding buffer (20 mM Hepes, pH 7.4, containing 1 mM MgCl₂, 10 μ M ZnSO₄, 20 mM KCl, and 8% glycerol). The reaction mixture was incubated for 15 minutes at 4°C and separated on a 6% polyacrylamide gel. The gel was dried and exposed on films for a Fujix BAS 2000 Bioimaging analyzer (Fuji Photo-Film).

Transient transfection

The BAEC cultures were plated on 35-mm dishes at 80% confluency. *Drosophila* S2 cells were grown in 35-mm dishes (5×10^6 cells/dish). Transient transfection was performed using Lipofectamine Plus reagents (GIBCO, BRL, Life Technologies Inc, Rockville, MD) in 1 mL serum-free medium containing either *Zf9-pCIneo* or *Zf9-pAC*, with or without 1 μ g pGL2-2350, along with pRL-CMV (*Renilla* luciferase, 100 ng/dish, Promega, Madison, WI) as an internal standard to normalize transfection efficiency. pCIneo or pAC was used as empty vector to adjust total amount of DNA transfected always to 2 μ g per each sample. After a 4-hour incubation, 1 mL of medium containing 20% FCS was added to the cultures, and then further incubated for 48 hours. Thereafter, either luciferase activity or endogenous PA activity was determined in cell lysates, or total RNA was isolated and used for Northern blotting.

Luciferase assay

Cells were harvested by scraping (BAECs) or by centrifugation (S2 cells) and lysed into 120 μ L of the commercial lysis buffer (Promega). Luciferase activity was determined in 10 μ L of lysate using a commercial kit (Promega) and luminometer (Turner Designs Instrument, Sunnyvale, CA). Transfection efficiency was normalized by *Renilla* luciferase activity measured in the same cell lysate at the same time.

Assay of cellular PA activity and membrane plasmin activity

Cellular PA activity was measured using the chromogenic substrate S2403 as described previously and expressed as urokinase unit per milligram of protein in the sample.³⁷ Protein concentration was measured by bicinchoninic acid (BCA) (Pierce, Rockford, IL) assay using bovine serum albumin (BSA) as the standard. Plasmin bound to the cell surface was recovered with tranexamic acid and assayed as described previously.⁵⁴ This plasmin is originally derived from plasminogen present in the serum.

Zf9 and uPA expression in vivo

The distal half of the left common carotid artery of a 10-week-old Sprague-Dawley rat was denuded of endothelium by 3 passages of a 2-F

balloon catheter inserted through the left external carotid artery as reported previously.⁵⁵ Pentobarbital (intraperitoneal, 30 mg/kg) and 1% xylocaine (local) were used for anesthesia. Animals were killed with an overdose of pentobarbital, and the artery was perfused fixed with 3% paraformaldehyde and excised. Paraffin-embedded serial sections were stained using Vectastain ABC elite kit (Vector Laboratories, Burlingame, CA) according to the manufacturer's instructions with antibodies against rat Zf9 (final 1:500 dilution), rat uPA (final 1:50 dilution), or nonimmune rabbit serum (final 1:50 dilution). Color development was made with diaminobenzidine tetrahydrochloride and nuclear counterstain with hematoxylin. Procedures were examined and authorized by the institutional committee.

TGF- β assays

Measurement of TGF- β was performed using either a bioassay (cellular PA assay with BAECs) or enzyme-linked immunosorbent assays (ELISAs) specific for either TGF- β 1 or TGF- β 2 (Promega). BAEC cultures were transfected with Zf9 expression vector and incubated for 36 hours. The cultures were then rinsed with phosphate-buffered saline (PBS) and further incubated with 1 mL of α MEM containing 0.1% BSA for an additional 12 hours to generate conditioned medium. Cellular PA assays for TGF- β were performed as described previously.⁵⁶ Briefly, following an 8-hour incubation of test BAECs with each sample conditioned medium in the presence or absence of 20 μ g/mL either anti-TGF- β antibody or nonimmune antibody, cell lysate was prepared and PA activity levels of each cell lysate were determined. The concentration of TGF- β in the conditioned medium was calculated from the decrease in PA levels by comparison with a standard curve made with recombinant human TGF- β 1. The specificity of the assays was verified by controls using anti-TGF- β antibody. The amount of total (active plus latent) TGF- β was determined following conversion of all latent TGF- β to active TGF- β by acidification of the sample (pH 3, 1 hour at room temperature), followed by neutralization. The lowest limit of this bioassay is about 2 pg/mL. For ELISAs, conditioned medium was concentrated 10-fold on Centrprep concentrator (Amicon Inc, Beverly, MA) and assayed for either active TGF- β 1 or TGF- β 2 according to the manufacturer's directions, because both ELISAs detect a minimum of 25 pg/mL TGF- β 1 and 32 pg/mL TGF- β 2, respectively. Final concentrations were calculated accounting for measured losses. The concentration of total TGF- β s was determined in unconcentrated medium immediately after acid activation of all latent TGF- β s.

Statistics

Significance was determined by the 2-tailed *t* test.

Results

Expression of Zf9 by BAECs

We first determined whether BAECs express Zf9. As seen in Figure 1A, BAECs maintained in serum-free medium barely expressed a Zf9 mRNA transcript of 4.5 kb (lane 1). Incubation of the cells with 10% FCS-containing medium induced Zf9 mRNA expression in a time-dependent manner after 2 hours (lanes 3-6). This induction was potentiated by inclusion of 10 ng/mL PMA (lanes 7-11). Similar induction of Zf9 was also observed in protein levels using Western blot (Figure 1B). PMA alone induced Zf9 mRNA and protein expression in the absence of serum (data not shown). When serum was removed after a 5-hour exposure in the absence of PMA, Zf9 expression disappeared within 3 to 4 hours (Figure 1C and D, lanes 1-5). On the other hand, the down-regulation of Zf9 by serum withdrawal in the presence of PMA is only partial (Figure 1C and D, lanes 6-10), because PMA can stimulate Zf9 production. These results indicate that Zf9 is a serum- and PMA-inducible factor expressed by BAECs. Furthermore, PMA may slow the degradation of Zf9. None of the following serum factors induced Zf9 when added individually to serum-free medium: platelet-derived growth factor, TGF- β , tumor necrosis factor- α , and interleukin-1 β , or retinoic acid.

Next, the effect of the protein synthesis inhibitor, cycloheximide, on Zf9 mRNA expression was assessed to determine if induction by serum comprised an immediate-early response. Addition of 6 μ g/mL cycloheximide did not abrogate the induction of Zf9 mRNA by serum, but rather resulted in prominent superinduction of the transcript (Figure 1E, lanes 3 and 4), confirming the immediate-early nature of Zf9 induction in BAECs.

Binding of recombinant Zf9 to the uPA GC box

To determine if Zf9 interacts with the uPA promoter sequences, we examined binding of Zf9-GST to native and mutated GC boxes within the uPA promoter by gel shift assay (Figure 2B). Zf9-GST fusion protein strongly bound to the wild-type uPA GC box (lane 2), whereas none was detected for GST alone (lane 4). Two shifted

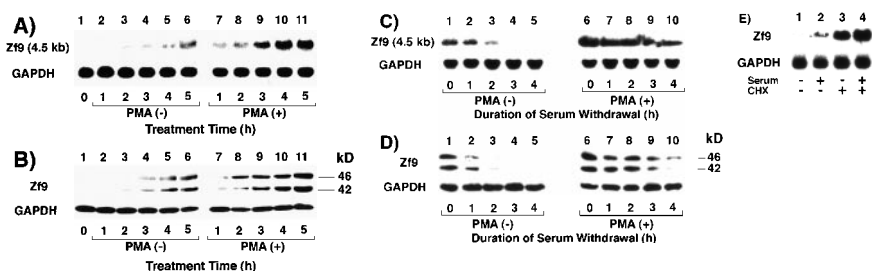


Figure 1. Expression of Zf9 by BAECs. (A) and (B) Time course in induction by serum and PMA. After a 5-hour preincubation with serum-free medium confluent BAEC cultures in either 15-cm dishes or 6-cm dishes were incubated with 10% FCS-containing medium for the indicated times, in the absence and presence of 10 ng/mL PMA. Cell lysate was prepared from each 15-cm dish, total RNA was isolated, and 30 μ g of each RNA was used for detection of Zf9 mRNA in Northern blotting (panel A). Cells on each 6-cm dish were scraped into 60 μ L of sodium dodecyl sulfate (SDS) sample buffer, and rapidly passed 20 times through a 17-gauge needle to shear nucleic acids. A portion (30 μ L) of each cell lysate was immediately subjected to SDS-polyacrylamide gel electrophoresis on 13% resolving gel without reducing conditions, and changes in Zf9 protein levels were analyzed by Western blotting as described in "Materials and Methods" (panel B). Lane 1, control; lanes 2-6, without PMA; lanes 7-11, with PMA. Each experiment was repeated 5 times with similar results and representative results are shown. (C) and (D) Disappearance of Zf9 expression by BAECs after depletion of serum. Following a 5-hour incubation with 10% serum \pm 10 ng/mL PMA, BAECs were then incubated with serum-free medium \pm 10 ng/mL PMA, and harvested at increasing intervals thereafter. The Zf9 mRNA levels (panel C) and Zf9 protein levels (panel D) were analyzed by Northern blotting and Western blotting, respectively. Lanes 1-5, without PMA; lanes 6-10, with PMA. Each experiment was repeated 3 times with similar results and representative results are shown. (E) Superinduction of Zf9 mRNA in the presence of cycloheximide. After a preincubation in serum-free medium for 5 hours, confluent BAEC cultures were incubated for 3 hours with either fresh serum-free medium or 10% serum-containing medium in the absence or presence of 6 μ g/mL cycloheximide (CHX). Cell lysates were prepared and the changes of Zf9 mRNA levels were assessed by Northern blotting. Lane 1, no stimulus; lane 2, serum alone; lane 3, cycloheximide alone; lane 4, serum plus cycloheximide. The experiment was repeated 4 times with similar results and a representative result is shown.

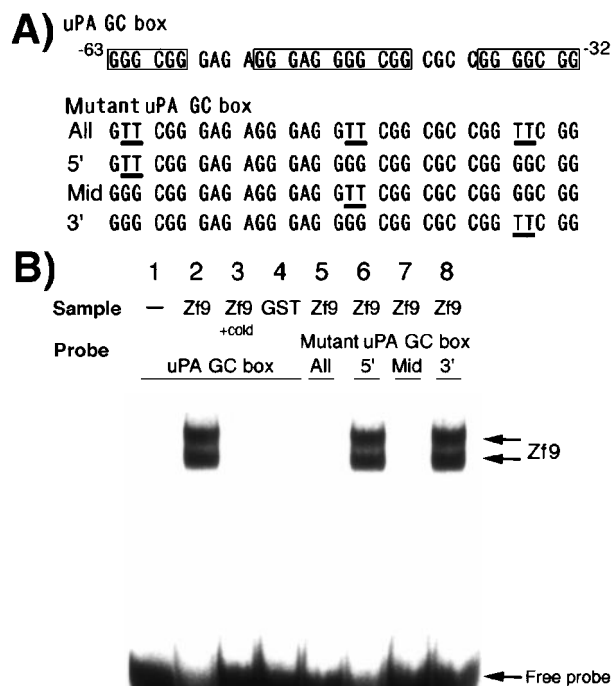


Figure 2. Binding of Zf9-GST to GC boxes in the uPA promoter. Binding of Zf9-GST fusion protein to the 32 P-labeled uPA GC box oligonucleotide was determined by gel shift assay. Four different mutant oligonucleotides were tested, which contained mutations in one or all of the GC boxes as shown. Sequences of these oligonucleotides are indicated in panel A. Protein-DNA complexes were separated through a 6% polyacrylamide gel and visualized on an image analyzer (panel B). Lane 1, wild uPA GC box alone; lane 2, wild uPA GC box + Zf9-GST; lane 3, wild uPA GC box + Zf9-GST in the presence of a 20-fold excess of unlabeled (cold) oligonucleotide; lane 4, wild uPA GC box + GST; lanes 5-8, mutant uPA GC boxes + Zf9-GST. Lane 5, oligonucleotide with mutations in all 3 GC boxes (abbreviated as "All"); lane 6, mutant oligonucleotide in which 5' site GC box was mutated (abbreviated as "5'"); lane 7, mutant oligonucleotide in which the middle GC box was mutated (abbreviated as "Mid"); lane 8, mutant oligonucleotide in which 3' site GC box was mutated (abbreviated as "3'"). The experiment was repeated 3 times with similar results in all experiments; representative results are shown.

bands were detected. In experiments not shown, the higher molecular weight band corresponds to intact Zf9 based on Western blotting of the nondenaturing gel following transfer to nylon, and a lower molecular weight band represents a degradation product containing only the DNA binding domain.³⁹ Interaction of both intact and degraded Zf9 with labeled oligonucleotide was completely inhibited by a 20-fold excess of unlabeled oligonucleotide (lane 3). Addition of anti-Zf9 antibody abolished rather than supershifted the interaction between recombinant Zf9 and the oligonucleotide (data not shown), similar to what we reported previously using a consensus GC box oligonucleotide as a probe.³⁹

We determined which GC boxes among 3 uPA promoter GC boxes are critical for Zf9 binding. Intact Zf9 did not bind to the oligonucleotides containing mutations in the middle GC box (lanes 5 and 7), but did bind to the mutants preserving the middle GC box (lanes 6 and 8), indicating that this middle GC box is the most critical for interaction between Zf9 and the uPA promoter.

Enhancement of PA activity by Zf9 via transactivation of uPA gene

Transactivation of uPA by Zf9 was explored using transient transfection. Cotransfection of a uPA reporter (pGL2-2350) and the Zf9 expression vector (*Zf9-pCIneo*) in BAECs resulted in a

concentration-dependent enhancement of luciferase activity (Figure 3A). Relative luciferase activity increased maximally to 6-fold compared to the control cells transfected only with pCIneo empty vector. To confirm that transactivation of uPA by Zf9 was not dependent on coexpression of Sp1, the same analysis was performed in *Drosophila* S2 cells, which are devoid of endogenous Zf9 as well as Sp1-like activity.^{35,39} A similar effect was obtained and interestingly, the relative activity was greater in this cell type, probably due to low basal activity (Figure 3B). In Figure 3C we compared the effect of Zf9 expression on transactivation of wild-type uPA promoter (*pUK-Luc*) or its mutants, either containing only GC and TATA boxes (*pUK GC-Luc*) or deficient in GC boxes (*pUK ΔGC-Luc*). The basal activities of *pUK GC-Luc* (sample 3) and *pUK ΔGC-Luc* (sample 5) were, respectively, 360% and 13% of that of the wild-type *pUK-Luc* (sample 1), suggesting that the GC box region is necessary for full transactivation of the uPA gene as reported previously.³⁶ Zf9 transfection enhanced transactivation of both wild-type *pUK-Luc* and *pUK GC-Luc* to the

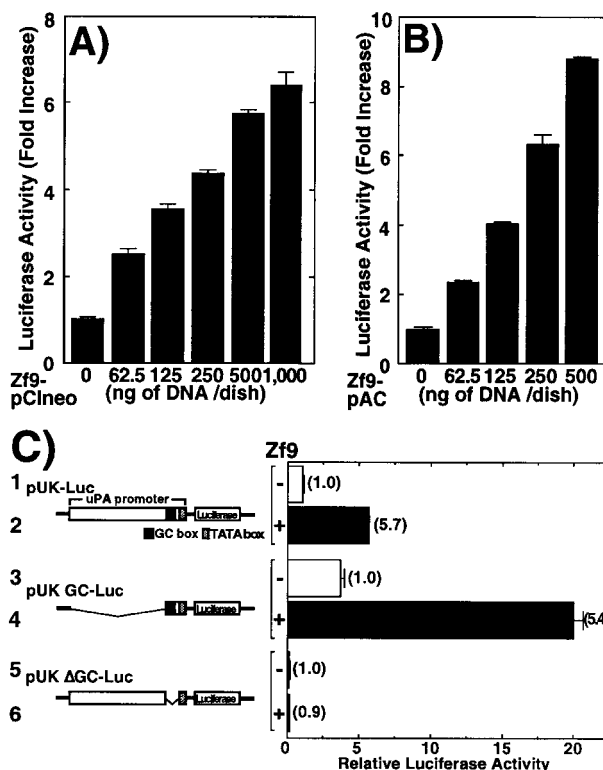


Figure 3. Transactivation of the uPA promoter by Zf9 in BAECs and *Drosophila* S2 cells. (A) and (B) BAEC and *Drosophila* S2 cell cultures grown on 35-mm dishes were cotransfected with a combination of the indicated amounts of Zf9 expression vector (*Zf9-pCIneo* or *Zf9-pAC*) and 1 μ g of pGL2-2350, luciferase reporter gene fused with the uPA promoter, as described in "Materials and Methods." After a 48-hour incubation, cell lysates were prepared, luciferase activity in each lysate was determined and expressed as fold increase. Panel A, BAECs; panel B, *Drosophila* S2 cells. Each value represents the average \pm SD from triplicate determinations. Each experiment was repeated 3 times with similar results and representative results are shown. (C) Transactivation by Zf9 of the uPA promoter via GC box regions. BAEC cultures were cotransfected with a combination of 500 ng each of either pCIneo or *Zf9-pCIneo* plus 1 μ g each of either pGL2-2350 (pUK-Luc), pGL2-GC (pUK GC-Luc), or pGL2-ΔGC (pUK ΔGC-Luc). Cell lysates were prepared, and luciferase activity of each lysate was determined. Data are expressed as relative luciferase activity compared to the activity of pUK-Luc cotransfected with pCIneo alone. The numbers in parentheses to the right of each bar indicate fold-induction calculated for each reporter. Samples 1 and 2, pUK-Luc; samples 3 and 4, pUK GC-Luc; samples 5 and 6, pUK ΔGC-Luc. Odd numbers, pCIneo; even numbers, *Zf9-pCIneo*. Each value represents the average \pm SD from triplicate determinations. Experiment was repeated 3 times with similar results and representative results are shown.

same extent (5.4~5.7-fold), but not of *pUK ΔGC-Luc*, indicating that transactivating activity of Zf9 requires GC boxes within uPA promoter.

We next examined the effect of transiently transfected Zf9 on endogenous uPA expression and activity in BAEC cultures. uPA mRNA expression in BAEC cultures was up-regulated 200% as determined by densitometry normalized to GAPDH expression following transfection of *z9* cDNA despite the typically low (< 20%) efficiency of transient transfection (Figure 4A, upper bands). Because both uPAR and PAI-1 promoters also contain GC boxes,^{57,58} we examined the effect of Zf9 on these mRNAs. In contrast to uPA, mRNA expression of these genes was not affected by Zf9 (Figure 4A, second and third bands). The selective increase in uPA mRNA but not its inhibitor or receptor suggested that Zf9 might enhance net PA activity. Indeed, transient transfection with Zf9 increased endogenous cellular PA activity to 200% (Figure 4B). Furthermore, membrane-associated plasmin levels were increased from 9.8 ng/10⁶ cells to 27.5 ng/10⁶ cells after transfection with Zf9 for 48 hours. These results suggest that Zf9 promotes uPA transcription through GC box region and enhances surface plasmin levels of BAEC cultures.

Coordinated expression of Zf9 and uPA proteins by vascular endothelial cells of the carotid artery and aorta after balloon injury

Vascular endothelial cells express a small amount or no uPA in normal arteries, but uPA expression dramatically increases after vascular injury.^{12,55} To study the role of Zf9 in response to vascular injury in vivo, we injured the distal half of the left carotid artery of rats with a balloon catheter and immunostained carotid artery with Zf9 and uPA antibodies at various time points after the injury. At 3 hours after the injury, neither Zf9 nor uPA immunoreactivity was detected in the vascular endothelial cells or in the smooth muscle cells (Figure 5, panels A and B, respectively). Within 2 days of

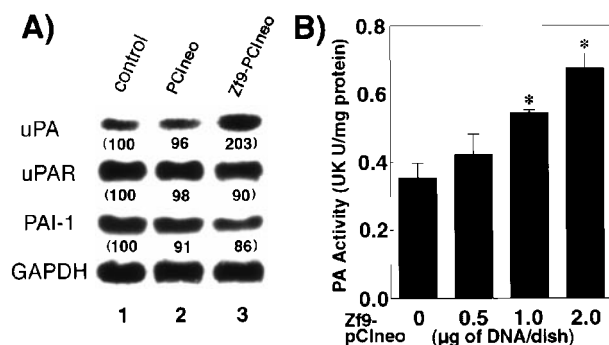


Figure 4. Effect of Zf9 on endogenous PA activity in BAEC cultures. (A) Effect on mRNA expression of uPA, uPAR, and PAI-1. BAEC cultures grown on 10-cm dishes were transfected with 8 μg of *Zf9-pCIneo*. As a control, cells were transfected with the same amount of pCIneo. Forty-eight hours after transfection, cell lysates were prepared, total RNA was extracted, and mRNA expression of uPA, uPAR, PAI-1, and GAPDH in each sample was assessed by Northern blotting. Radioactivity of each band was detected on an image analyzer, and relative intensity of each band was quantitated, normalized against the intensity of GAPDH, and expressed as arbitrary units in parentheses under each band. Each experiment was repeated 3 times with similar results in all, and representative results are shown. (B) Effect on BAEC PA activity levels. BAEC cultures grown on 35-mm dishes were transfected with the indicated amounts of *Zf9-pCIneo*. Forty-eight hours after transfection, cells were harvested in lysis buffer, and PA levels in each lysate were determined using chromogenic substrate, S-2403, and expressed as urokinase (UK) unit (U)/mg of protein in the sample. Each value represents the average ± SD from triplicate determinations. Points marked by an asterisk differ significantly ($P < 0.05$) from control cells transfected with 2 μg of pCIneo alone. Experiment was repeated 4 times with similar results in all, and representative results are shown.

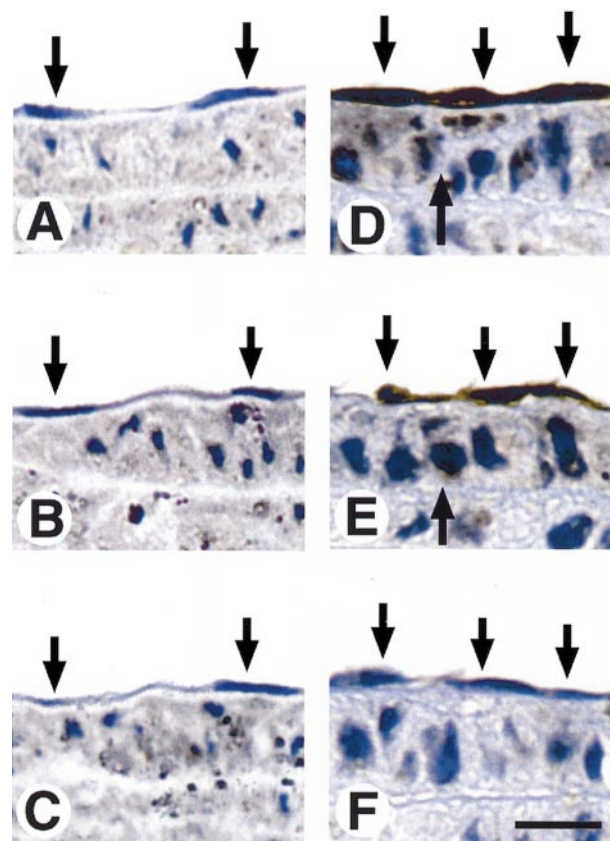


Figure 5. Concomitant expression of Zf9 and uPA in arterial endothelial cells after carotid balloon injury in rats. The distal half of the left carotid artery of rats was injured with a balloon catheter. At 3 hours and 2 days after injury, the carotid artery was perfusion fixed with 3% paraformaldehyde, excised, fixed again with 3% paraformaldehyde, and paraffin embedded. Serial sections were stained with anti-Zf9 antibody (panels A and D), anti-uPA antibody (panels B and E) or nonimmune antibody (panels C and F) as described in the Materials and Methods. Panels A through F show the typical staining pattern of the adjacent proximal half of carotid artery at 3 hours (panels A-C) and 2 days (panels D-F) after injury. Down arrows indicate the endothelial cells. Up arrows indicate the smooth muscle cells. Scale bar, 50 μm.

injury, however, both Zf9 and uPA immunoreactivities were detected in the vascular endothelial cells (down arrows in Figure 5, panels D and E, respectively), whereas neither was detected in the endothelial cells of uninjured control rats. Weak immunoreactivity was also detected in the medial smooth muscle cells of the injured carotid artery (up arrows). Interestingly, Zf9 was induced not only in endothelial cells near the wound edge, but also in the adjacent proximal half of carotid artery, as well as in the endothelial cells of the aorta in the same animal. Nonimmune serum did not give any positive staining (Figure 5, panels C and F).

Induction of TGF-β by Zf9 via enhancement of PA activity

Finally, we examined whether Zf9 has biologic activity in this system via enhancement of PA/plasmin activity. We speculated that up-regulation of PA/plasmin by Zf9 might result in the activation of latent TGF-β as has been demonstrated in other systems.^{25,59,60} This hypothesis was tested by 2 different assays. First, as determined by bioassays, we observed that transient transfection of Zf9 cDNA induced the generation of active TGF-β in BAEC cultures during a subsequent 12-hour incubation with serum-free medium (Figure 6A, sample 3). This increase was eliminated by inclusion of a plasmin inhibitor, aprotinin, during the preparation of the condi-

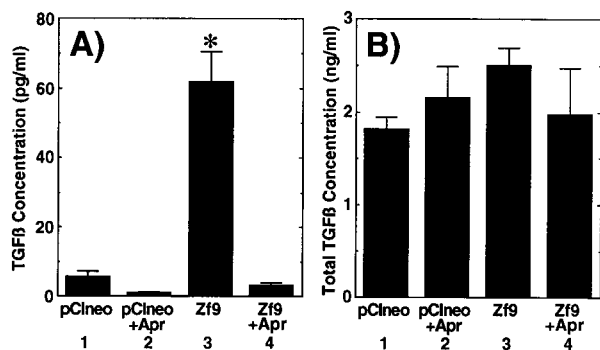


Figure 6. Activation of latent TGF- β following transfection of Zf9. BAEC cultures grown on 35-mm dishes were transfected with 1 μ g of pCneo or Zf9-pCneo as described above. After a 36-hour incubation, medium was changed to serum-free α MEM containing 0.1% BSA, and cells were further incubated for 12 hours in the absence and presence of 50 μ g/mL aprotinin (Apr). The concentration of active (panel A) and total (panel B) TGF- β in each conditioned medium was determined by the bioassays as described in "Materials and Methods." Each value represents the average \pm SD from triplicate determinations. Each experiment was repeated 3 times with similar results in all, and representative results are shown. Sample 1, pCneo; sample 2, pCneo with inclusion of aprotinin; sample 3, Zf9-pCneo; sample 4, Zf9-pCneo with inclusion of aprotinin. Points marked by an asterisk differ significantly ($P < 0.05$) from the control (sample 1).

tioned medium (Figure 6A, sample 4), suggesting that TGF- β formation resulted from proteolytic activation of latent TGF- β .^{17-25,59,60} On the other hand, total TGF- β levels only moderately (35%) increased (Figure 6B, sample 3), and there were no statistically significant differences between each sample. A similar result was also obtained with TGF- β 1 and TGF- β 2 ELISAs, although the basal levels of active TGF- β s (sum of TGF- β 1 and TGF- β 2 concentrations) were higher than those determined with the bioassay (Table 1). Transient transfection of Zf9 did not alter endogenous TGF- β 1 and TGF- β 2 mRNA levels in BAEC cultures (data not shown). These data indicate that Zf9 can induce TGF- β activity in BAECs and that the increased TGF- β in this system derived mainly from conversion of the latent to the active form of the cytokine, due at least in part to increased fibrinolytic activity.

Discussion

We have demonstrated a potential role of Zf9 in regulating vascular injury through its marked stimulatory effects on uPA gene expression and TGF- β activity. Cultured BAECs express Zf9 in response to serum as well as PMA (see Figure 1A), which is a model agonist of a hitherto known early response gene, *Egr-1*³³; we have not yet identified the responsible serum factor. The kinetic analysis of Zf9 mRNA and protein (see Figures 1A-D) suggests that both are labile, and therefore withdrawal of appropriate stimulus culminates in their rapid disappearance. This observation, combined with superinduction by cycloheximide (see Figure 1E), supports the role of Zf9 as an immediate-early gene, as previously observed in hepatic stellate cells in vivo.³⁸ As shown in Figure 1B and D, Zf9 protein is expressed by BAECs as a doublet of 46 kD and 42 kD, consistent with our previous finding in hepatic stellate cells.³⁹ Furthermore, transient transfection of human or rat Zf9 into BAECs also yielded the same doublet in Western blot (data not shown). As we have suggested previously, the 14-kD or 10-kD size differences from 32 kD, which is the predicted size of the unmodified polypeptide, may be explained by posttranslational modifications, but this has not yet been tested directly.³⁹

We have also demonstrated that both the interaction between recombinant Zf9 and the uPA promoter (see Figure 2) and transactivation of the uPA gene by Zf9 (see Figure 3) are dependent on GC boxes within the promoter in vitro. Incubation of Zf9-GST with mutated promoter sequences establishes that the middle GC box is the most essential for Zf9 binding. The substantial increase in endogenous uPA activity after transient transfection of Zf9 (see Figure 4), despite a typical transfection efficiency of only 10% to 20%, underscores the potency of this factor in up-regulating uPA expression. Importantly, vascular injury caused by a balloon catheter induces the expression of Zf9 by endothelial cells in vivo (see Figure 5). Concomitant induction of uPA in this tissue underscores the notion that Zf9 may play an important role in the uPA gene expression induced by vascular injury. This finding supports our in vitro observations and agrees with findings of previous investigators characterizing uPA induction after vascular injury in vivo.¹² We acknowledge, however, that the current data do not establish a direct role of Zf9 in uPA expression in vivo.

Zf9 belongs to an enlarging family of zinc finger Krüppel-like transcription factors.⁴²⁻⁵² The physiologic roles and distribution of these factors are distinct.⁵² Sp1 is widely expressed in various types of cells at a fairly constant level and is involved in basal transactivation of many genes including uPA.^{36,37} Zf9 is also widely expressed but is an early response gene in both hepatic stellate^{38,39} and vascular endothelial cells (current study) during the wound healing response. In this context, Zf9 resembles Egr-1, a non-Krüppel-like zinc finger transcription factor that has been identified also as an early gene induced by serum, PMA, and tissue injury, and can substitute for Sp1 in promoting transcription of genes including uPA.³³ Therefore, it is of great interest to know how Sp1, Egr-1, Zf9, and other Krüppel-like factors interact in regulating uPA expression. In preliminary studies we have documented a synergism and physical interaction between Zf9 and Sp1.⁶¹ This contrasts with the interplay of Sp1 and Egr-1.³³

We previously demonstrated that Zf9 transactivates promoters of collagen α 1(I), TGF- β 1, and its signaling receptors in hepatic stellate cells.^{39,53} Activation of latent TGF- β via uPA induction provides an additional mechanism through which TGF- β 1 activity is augmented by Zf9 (see Figure 6 and Table 1). This has also been observed in other systems in which uPA is up-regulated, for example, in response to basic fibroblast growth factor or retinoic acid.^{59,60} Activation of latent TGF- β has been implicated as an important regulator of vascular smooth muscle cell differentiation, endothelial cell quiescence, and extracellular matrix production during vessel morphogenesis. It seems that latent TGF- β is

Table 1. Summary of the results of TGF- β 1 and TGF- β 2 ELISAs

Vector	Aprotinin (50 μ g/mL)	Active TGF- β (pg/mL)		Total TGF- β (pg/mL)	
		TGF- β 1	TGF- β 2	TGF- β 1	TGF- β 2
pCneo	—	20.2 \pm 3.3	18.1 \pm 3.0	850 \pm 64	1035 \pm 65
pCneo	+	15.2 \pm 2.2	14.7 \pm 2.0	963 \pm 47	851 \pm 126
Zf9-pCneo	—	65.3 \pm 7.2*	48.6 \pm 6.5*	1,029 \pm 47*	1,118 \pm 51
Zf9-pCneo	+	37.4 \pm 1.1†	29.8 \pm 1.0†	1,041 \pm 79	1,035 \pm 53

The concentration of active and total TGF- β 1 as well as TGF- β 2 in each conditioned medium prepared in Figure 6 was determined by subtype-specific ELISAs as described in "Materials and Methods." Each value represents the average \pm SD from triplicate determinations. Each experiment was repeated 3 times with similar results in all, and representative results are shown.

*P values obtained by a comparison between pCneo and Zf9-pCneo are less than 0.05 except for total TGF- β 2 concentration.

†P values obtained by a comparison between Zf9-pCneo and Zf9-pCneo + aprotinin are less than 0.05 for both active TGF- β 1 and TGF- β 2 concentrations.

activated through different mechanisms under physiologic and pathologic situations. It is intriguing to determine if Zf9 might also stimulate the expression of other TGF- β activators, such as thrombospondin and integrin, $\alpha v\beta 6$.^{62,63} We speculate that Zf9 may promote different GC box-containing gene promoters in a tissue and cell context-dependent manner. For example, the current (see Figure 3B) and previous^{39,53} data suggest that in S2 cells Zf9 transactivates the SV40 promoter as well as promoters of TGF- $\beta 1$ and uPA, but not promoters of collagen $\alpha 1(I)$, types I and II TGF- β receptors. Although the percentage of active TGF- β in our system was low (2.4% in Figure 6 and 4.3%-6.3% in Table 1), 40 to 60 pg/mL TGF- β is enough to exert its biologic effects on BAECs as we and others have reported.^{23,56,59} We cannot explain the reason for the differences in the concentration of active TGF- β determined by the biologic assays (see Figure 6) and ELISAs (see Table 1). We speculate that artificial activation might occur during concentration of the conditioned medium prior to ELISA, resulting in high basal levels. It would be difficult to evaluate if serum-induced Zf9 also generates active TGF- β in BAEC cultures because 10% serum used in the current study already contains 99.5 ± 4.7 pg/mL active TGF- $\beta 1$ and 11.2 ± 3.8 pg/mL active TGF- $\beta 2$ as measured by ELISA. This preexisting TGF- β might counteract the ability of Zf9 to enhance uPA expression, inferring that the system

is self-regulated, similar to what has been reported previously in other systems.⁶⁴

In summary, we have shown that both in vitro and in vivo Zf9 is expressed by vascular endothelial cells in response to certain stimuli and can promote transactivation of uPA, which leads to activation of latent TGF- β , suggesting that this system may have an important role in vascular injury response. The present data and previous findings^{25,38,39} strongly suggest that rapid, early induction of Zf9 may promote fibrogenesis by potentiating the proteolytic activation of latent TGF- β in hepatic stellate cells. It will be important to determine if Zf9 regulates uPA activity and TGF- $\beta 1$ responses in other in vivo models of tissue injury including atherosclerosis and restenosis,^{6,20} cutaneous wound healing,² pulmonary intestinal pneumonia and fibrosis,^{22,63} and hepatic fibrosis.^{1,39}

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References

- Olaso E, Friedman SL. Molecular regulation of hepatic fibrogenesis. *J Hepatol*. 1998;29:836.
- Martin P. Wound healing—aiming for perfect skin regeneration. *Science*. 1997;276:75.
- Alcolado R, Arthur MJ, Iredale JP. Pathogenesis of liver fibrosis. *Clin Sci*. 1997;92:103.
- Border WA, Noble NA. Transforming growth factor β in tissue fibrosis. *N Engl J Med*. 1994;331:1286.
- Michalopoulos GK, DeFrances MC. Liver regeneration. *Science*. 1997;276:60.
- Ross R. Atherosclerosis—an inflammatory disease. *N Engl J Med*. 1999;340:115.
- Tkachuk V, Stepanova V, Little PJ, Bobik A. Regulation and role of urokinase plasminogen activator in vascular remodelling. *Clin Exp Pharmacol Physiol*. 1996;23:759.
- Reidy MA, Irvin C, Lindner V. Migration of arterial wall cells: expression of plasminogen activators and inhibitors in injured rat arteries. *Circ Res*. 1996;78:405.
- Carmeliet P, Moons L, Lijnen R, et al. Urokinase-generated plasmin activates matrix metalloproteinases during aneurysm formation. *Nat Genet*. 1997;17:439.
- Bachmann F. The plasminogen-plasmin enzyme system. In: Colman RW, Hirsh J, Marder VJ, Salzman EW, eds. *Hemostasis and Thrombosis: Basic Principles and Clinical Practice*, 3rd ed. Philadelphia, PA: JB Lippincott; 1994:1592.
- Ellis V, Danø K. u-Plasminogen activator. In: Barrett AJ, Rawlings ND, Woessner JF, eds. *Handbook of Proteolytic Enzymes*. London, UK: Academic Press; 1998:177.
- Carmeliet P, Moons L, Herbert J-M, et al. Urokinase but not tissue plasminogen activator mediates arterial neointima formation in mice. *Circ Res*. 1997;81:829.
- Mignatti P, Rifkin DB. Biology and biochemistry of proteinases in tumor invasion. *Physiol Rev*. 1993;73:161.
- Werb Z. ECM and cell surface proteolysis: regulating cellular ecology. *Cell*. 1997;91:439.
- Herbert JM, Carmeliet P. Involvement of u-PA in the anti-apoptotic activity of TGF β for vascular smooth muscle cells. *FEBS Lett*. 1997;413:401.
- Mazzieri R, Masiero L, Zanetta L, et al. Control of type IV collagenase activity by components of the urokinase-plasmin system: a regulatory mechanism with cell-bound reactants. *EMBO J*. 1997;16:2319.
- Flaumenhaft R, Kojima S, Abe M, Rifkin DB. Activation of latent transforming growth factor β . *Adv Pharmacol*. 1993;24:51.
- Brauer PR, Yee JA. Cranial neural crest cells synthesize and secrete a latent form of transforming growth factor β that can be activated by neural crest cell proteolysis. *Dev Biol*. 1993;155:281.
- Yee JA, Yan L, Dominguez JC, Allan EH, Martin TJ. Plasminogen-dependent activation of latent transforming growth factor beta (TGF β) by growing cultures of osteoblast-like cells. *J Cell Physiol*. 1993;157:528.
- Grainger DJ, Kemp PR, Liu AC, Lawn RM, Metcalfe JC. Activation of transforming growth factor- β is inhibited in transgenic apolipoprotein(a) mice. *Nature*. 1994;370:460.
- Nunes I, Shapiro RL, Rifkin DB. Characterization of latent TGF- β activation by murine peritoneal macrophages. *J Immunol*. 1995;155:1450.
- Khalil N, Corne S, Whitman C, Yacyszyn H. Plasmin regulates the activation of cell-associated latent TGF- $\beta 1$ secreted by rat alveolar macrophages after in vivo bleomycin injury. *Am J Respir Cell Mol Biol*. 1996;15:252.
- Nunes I, Gleizes PE, Metz CN, Rifkin DB. Latent transforming growth factor- β binding protein domains involved in activation and transglutaminase-dependent cross-linking of latent transforming growth factor- β . *J Cell Biol*. 1997;136:1151.
- Cai J, Zheng T, Lotz M, Zhang Y, Masood R, Gill P. Glucocorticoids induce Kaposi's sarcoma cell proliferation through the regulation of transforming growth factor- β . *Blood*. 1997;89:1491.
- Okuno M, Moriwaki H, Imai S, et al. Retinoids exacerbate rat liver fibrosis by inducing the activation of latent TGF- β in liver stellate cells. *Hepatology*. 1997;26:913.
- Soubt MK, Marksitzer R, Menoud PA, Nagamine Y. Role of tissue-specific transcription factor LFB3 in a cyclic AMP-responsive enhancer of the urokinase-type plasminogen activator gene in LLC-PK1 cells. *Mol Cell Biol*. 1998;18:4698.
- Iwasaka C, Tanaka K, Abe M, Sato Y. Ets-1 regulates angiogenesis by inducing the expression of urokinase-type plasminogen activator and matrix metalloproteinase-1 and the migration of vascular endothelial cells. *J Cell Physiol*. 1996;169:522.
- De Cesare D, Palazzolo M, Blasi F. Functional characterization of COM, a DNA region required for cooperation between AP-1 sites in urokinase gene transcription. *Oncogene*. 1996;13:2551.
- Berthelsen J, Vandekerckhove J, Blasi F. Purification and characterization of UEF3, a novel factor involved in the regulation of the urokinase and other AP-1 controlled promoters. *J Biol Chem*. 1996;271:3822.
- De Cesare D, Palazzolo M, Berthelsen J, Blasi F. Characterization of UEF-4, a DNA-binding protein required for transcriptional synergism between two AP-1 sites in the human urokinase enhancer. *J Biol Chem*. 1997;272:23,921.
- Miralles F, Parra M, Caelles C, Nagamine Y, Féliz J, Muñoz-Cánoves P. UV irradiation induces the murine urokinase-type plasminogen activator gene via the c-Jun N-terminal kinase signaling pathway: requirement of an AP1 enhancer element. *Mol Cell Biol*. 1998;18:4537.
- Reuning U, Wilhelm O, Nishiguchi T, et al. Inhibition of NF- κ B-Rel A expression by antisense oligodeoxynucleotides suppresses synthesis of urokinase-type plasminogen activator (uPA) but not its inhibitor PAI-1. *Nucleic Acids Res*. 1995;23:3887.
- Khachigian LM, Lindner V, Williams AJ, Collins T. Egr-1-induced endothelial gene expression: a common theme in vascular injury. *Science*. 1996;271:1427.
- Kadonaga JT, Courey AJ, Ladika J, Tjian R. Distinct regions of Sp1 modulate DNA binding and transcriptional activation. *Science*. 1988;242:1566.
- Courey AJ, Tjian R. Analysis of Sp1 in vivo reveals multiple transcriptional domains, including a novel glutamine-rich activation motif. *Cell*. 1988;55:887.

36. Verde P, Boast S, Franzè A, Robbiati F, Blasi F. An upstream enhancer and a negative element in the 5' flanking region of the human urokinase plasminogen activator gene. *Nucleic Acids Res.* 1988;16:10,699.
37. Suzuki Y, Shimada J, Shudo K, Matsumura M, Crippa MP, Kojima S. Physical interaction between retinoic acid receptor and Sp1: mechanism for induction of urokinase by retinoic acid. *Blood.* 1999;93:4264.
38. Lalazar A, Wong L, Yamasaki G, Friedman SL. Early genes induced in hepatic stellate cells during wound healing. *Gene.* 1997;195:235.
39. Ratzliff V, Lalazar A, Wong L, et al. Zf9, a Krüppel-like transcription factor up-regulated in vivo during early hepatic fibrosis. *Proc Natl Acad Sci U S A.* 1998;95:9500.
40. Koritschoner NP, Bocco JL, Panzetta-Dutari GM, Dumur CI, Flury A, Patrino LC. A novel human zinc finger protein that interacts with the core promoter element of a TATA box-less gene. *J Biol Chem.* 1997;272:9573.
41. Suzuki T, Yamamoto T, Kurabayashi M, Nagai R, Yazaki Y, Horikoshi M. Isolation and initial characterization of GBF, a novel DNA-binding zinc finger protein that binds to the GC-rich binding sites of the HIV-1 promoter. *J Biochem.* 1998;124:389.
42. Sogawa K, Imataka H, Yamasaki Y, Kusume H, Abe H, Fujii-Kuriyama Y. cDNA cloning and transcriptional properties of a novel GC box-binding protein, BTB2. *Nucleic Acids Res.* 1993;21:1527.
43. Miller IJ, Bieker JJ. A novel, erythroid cell-specific murine transcription factor that binds to the CACCC element and is related to the *Krüppel* family of nuclear proteins. *Mol Cell Biol.* 1993;13:2776.
44. Anderson KP, Kern CB, Crable SC, Lingrel JB. Isolation of a gene encoding a functional zinc finger protein homologous to erythroid Krüppel-like factor: identification of a new multigene family. *Mol Cell Biol.* 1995;15:5957.
45. Crossley M, Whitelaw E, Perkins A, Williams G, Fujiwara Y, Orkin SH. Isolation and characterization of the cDNA encoding BKL/TEF-2, a major CACCC-box-binding protein in erythroid cells and selected other cells. *Mol Cell Biol.* 1996;16:1695.
46. Shields JM, Christy RJ, Yang VW. Identification and characterization of a gene encoding a gut-enriched Krüppel-like factor expressed during growth arrest. *J Biol Chem.* 1996;271:20,009.
47. Garrett-Sinha LA, Eberspaecher H, Seldin MF, de Crombrughe B. A gene for a novel zinc-finger protein expressed in differentiated epithelial cells and transiently in certain mesenchymal cells. *J Biol Chem.* 1996;271:31,384.
48. Yet SF, McAnulty MM, Foltz SC, et al. Human EZF, a Krüppel-like zinc finger protein, is expressed in vascular endothelial cells and contains transcriptional activation and repression domains. *J Biol Chem.* 1998;273:1026.
49. Tachibana I, Imoto M, Adjei PN, et al. Overexpression of the TGF β -regulated zinc finger encoding gene, TIEG, induces apoptosis in pancreatic epithelial cells. *J Clin Invest.* 1997;99:2365.
50. Cook T, Gebelein B, Mesa K, Madek A, Urrutia R. Molecular cloning and characterization of *TIEG2* reveals a new subfamily of transforming growth factor- β -inducible Sp1-like zinc finger-encoding genes involved in the regulation of cell growth. *J Biol Chem.* 1998;273:25,929.
51. Matsumoto N, Laub F, Aldabe R, et al. Cloning the cDNA for a new human zinc finger protein defines a group of closely related Krüppel-like transcription factors. *J Biol Chem.* 1998;273:28,229.
52. Philipsen S, Suske G. A tale of three fingers: the family of mammalian Sp/KLF transcription factors. *Nucleic Acids Res.* 1999;27:2991.
53. Kim Y, Ratzliff V, Choi SG, et al. Transcriptional activation of transforming growth factor β 1 and its receptors by the Krüppel-like factor Zf9/core promoter-binding protein and Sp1: potential mechanisms for autocrine fibrogenesis in response to injury. *J Biol Chem.* 1998;273:33,750.
54. Kojima S, Muramatsu H, Amanuma H, Muramatsu T. Midkine enhances fibrinolytic activity of bovine endothelial cells. *J Biol Chem.* 1995;270:9590.
55. Clowes AW, Clowes MM, Au YPT, Reidy MA, Belin D. Smooth muscle cells express urokinase during mitogenesis and tissue-type plasminogen activator during migration in injured rat carotid artery. *Circ Res.* 1990;67:61.
56. Kojima S, VerNooy R, Moscatelli D, Amanuma H, Rifkin DB. Lipopolysaccharide inhibits activation of latent transforming growth factor- β in bovine endothelial cells. *J Cell Physiol.* 1995;163:210.
57. Soravia E, Grebe A, De Luca P, et al. A conserved TATA-less proximal promoter drives basal transcription from the urokinase-type plasminogen activator receptor gene. *Blood.* 1995;86:624.
58. Johnson MR, Bruzdinski CJ, Winograd SS, Gelehrter TD. Regulatory sequences and protein-binding sites involved in the expression of the rat plasminogen activator inhibitor-1 gene. *J Biol Chem.* 1992;267:12,202.
59. Kojima S, Rifkin DB. Mechanism of retinoid-induced activation of latent transforming growth factor- β in bovine endothelial cells. *J Cell Physiol.* 1993;155:323.
60. Flaumenhaft R, Abe M, Mignatti P, Rifkin DB. Basic fibroblast growth factor-induced activation of latent transforming growth factor β in endothelial cells: regulation of plasminogen activator activity. *J Cell Biol.* 1992;118:901.
61. Ratzliff V, Kim SJ, Kim YS, Dang Q, Wong L, Friedman SL. A key role for Zf9 in hepatic fibrosis via its transcriptional activation of TGF β 1 and types I and II TGF β receptor genes in rat stellate cells [abstract]. *Hepatology.* 1997;26:185A.
62. Crawford SE, Stellmach V, Murphy-Ullrich JE, et al. Thrombospondin-1 is a major activator of TGF- β 1 in vivo. *Cell.* 1998;93:1159.
63. Munger JS, Huang X, Kawakatsu H, et al. The integrin α v β 6 binds and activates latent TGF β 1: a mechanism for regulating pulmonary inflammation and fibrosis. *Cell.* 1999;96:319.
64. Sato Y, Tsuboi R, Lyons R, Moses H, Rifkin DB. Characterization of the activation of latent TGF- β by co-cultures of endothelial cells and pericytes or smooth muscle cells: a self-regulating system. *J Cell Biol.* 1990;111:757.