

## PHAGOCYTES

## Red Blood Cells Inhibit Apoptosis of Human Neutrophils

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Oxidative stress has been implicated in the triggering of apoptosis in neutrophils. Because red blood cells (RBCs) are well known to scavenge oxidants including  $H_2O_2$ , we tested the hypothesis that RBCs inhibit apoptosis of neutrophils by reducing intracellular oxidative stress. Apoptosis of neutrophils was evaluated by light microscopy and DNA gel electrophoresis. We found that coculture with RBCs protected against neutrophil apoptosis. Neither physical contact between RBCs and neutrophils nor the cellular integrity of RBCs was required to protect against neutrophil apoptosis.

Neutrophil apoptosis was promoted by exogenous  $H_2O_2$  but suppressed by catalase, indicating a role for  $H_2O_2$  as a mediator of apoptosis. The protective effect of RBCs against apoptosis was due to catalase and glutathione metabolism because blocking of these antioxidant systems in RBCs attenuated the protective effect of RBCs. These results suggest that neutrophils are protected against apoptosis in the circulation.

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**M**ATURE NEUTROPHILS are short-lived cells. They have a circulating half-life of less than 10 hours<sup>1,2</sup> and once they have migrated into tissue they die via apoptosis within several days without the requirement for any apparent inductive stimuli.<sup>3,4</sup> This rapid and spontaneous apoptosis in neutrophils is believed to provide a neutrophil clearance mechanism that would act to limit tissue injury and lead to resolution of inflammation.<sup>4</sup>

Recent studies have suggested that circulating neutrophils can undergo apoptosis in the vasculature.<sup>5,6</sup> However, if neutrophils undergo apoptosis rapidly in the vasculature, they would die before responding to inflammatory stimuli and infiltrating tissue. This suggests the need for a mechanism in which neutrophil apoptosis is inhibited in the circulation. Recently, platelets have been shown to inhibit neutrophil apoptosis,<sup>7</sup> supporting this view. However, little attention has been paid to interactions between neutrophils and red blood cells (RBCs), which are about 20-fold more abundant than platelets in the circulation.

Several lines of evidence indicate that reactive oxygen species (ROS) are involved in apoptosis of neutrophils. Neutrophil apoptosis is inhibited under hypoxic conditions, which dramatically decreases the generation of ROS.<sup>8</sup> Apoptosis is also inhibited in neutrophils obtained from patients with chronic granulomatous disease who carry a mutation that limits ROS generation.<sup>9</sup> Neutrophil apoptosis is promoted by exogenous  $H_2O_2$  but delayed by antioxidants including catalase,<sup>8</sup> indicating a role for  $H_2O_2$  as a mediator of neutrophil apoptosis.

In the circulation, RBCs are 1,000-fold more abundant than neutrophils. RBCs have large amounts of antioxidants such as

catalase, glutathione, and superoxide dismutase,<sup>10-15</sup> and RBC membranes are permeable to  $H_2O_2$  and  $O_2^-$ ,<sup>10</sup> suggesting that RBCs can scavenge ROS. RBCs have been shown to protect against  $H_2O_2$ -mediated damage to cells or tissues.<sup>11-14</sup> RBCs scavenge  $H_2O_2$  generated by neutrophils<sup>16</sup> and therefore block formation of hydroxy radical ( $\cdot OH$ ) and hypochlorite ( $HOCl$ ).<sup>15</sup> Because  $H_2O_2$  can freely cross the cell membrane and therefore readily diffuses out of cells if generated intracellularly,<sup>17</sup> the reduction of extracellular  $H_2O_2$  by antioxidants is expected to decrease intracellular  $H_2O_2$ .<sup>18</sup>

In this report, we show that RBCs inhibit apoptosis of neutrophils by reducing intracellular oxidant stress. Our data suggest that neutrophils may be protected against apoptosis in the circulation.

## MATERIALS AND METHODS

**Reagents.** All reagents for cell culture were obtained from GIBCO Life Technologies, Inc (Gaithersburg, MD). 3-amino-1,2,4-triazole, 1-chloro-2,4-dinitrobenzene, mercaptosuccinate, and 4,4'-diisothiocyanatostilbene-2,2'-disulfonic acid (DIDS) were purchased from Sigma-Aldrich Japan K.K. (Tokyo, Japan).

**Blood samples.** Blood samples were obtained from healthy volunteers by venipuncture of the forearm vein and were immediately anticoagulated with 10 mmol/L EDTA.

**Preparation and treatment of RBCs.** RBCs were isolated by centrifugation of the blood samples at 200g for 10 minutes, gentle removal of platelet-rich plasma and leukocytes, washing three times with phosphate-buffered saline (PBS), and resuspension in PBS at 50% hematocrit. For some experiments, RBC antioxidant systems were blocked as previously described. RBC catalase was irreversibly inhibited by incubating a cell suspension with 50 mmol/L aminotriazole (catalase inhibitor) and 0.4 mmol/L  $H_2O_2$  for 1 hour at 37°C.<sup>19</sup> RBC glutathione was blocked by treating a cell suspension with either 2 mmol/L chlorodinitrobenzene (glutathione depleting agent),<sup>20</sup> or 0.2 mmol/L mercaptosuccinate (glutathione peroxidase inhibitor)<sup>21</sup> for 1 hour at 37°C. If necessary, RBCs were treated simultaneously with aminotriazole, chlorodinitrobenzene, and mercaptosuccinate to inhibit both catalase and glutathione metabolism, which account for the majority of the  $H_2O_2$ -scavenging activity of RBCs. RBC transport of  $O_2^-$  was blocked by incubating a cell suspension with 0.1 mmol/L DIDS for 1 hour at 37°C.<sup>10</sup> Before use, RBCs treated as above were washed three times in PBS to remove excess reagents. In some experiments, RBCs were lysed by brief sonication with an ultrasonic sonicator.

**Preparation of neutrophils.** Human neutrophils were isolated by dextran sedimentation and centrifugation on a Histopaque gradient (without endotoxin; Sigma) as previously described.<sup>22</sup> Contaminating erythrocytes were removed by hypotonic water lysis. The isolated

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neutrophils were washed twice with PBS and resuspended in RPMI1640 containing 5% heat-inactivated fetal calf serum (FCS), 100 U/mL penicillin, and 100 µg/mL streptomycin. To avoid accidental neutrophil activation and clumping, neutrophil preparations were carefully washed by centrifugation at 200g, the supernatant was aspirated, cells were gently resuspended with a pipet, and sudden changes in temperature were avoided. The purity of neutrophil populations was greater than 95% on May-Grünwald-Giemsa stain, and neutrophil viability was greater than 98% as determined by trypan blue dye exclusion. Less than 5% of neutrophil preparations showed a polarized shape, a sensitive marker for neutrophil activation.<sup>23</sup>

**Coculture of neutrophils with RBCs.** Neutrophils ( $5 \times 10^4$ ) were mixed with autologous RBCs at ratios of 1:10, 1:100, and 1:1000, and cultured in 96-well round-bottom plates (Becton Dickinson, Lincoln Park, NJ) at 37°C in a humidified incubator containing 5% CO<sub>2</sub>. In some experiments, neutrophils and RBCs were separately cultured in a 24-well plate using the cell culture insert (pore size, 0.4 µm; Becton Dickinson Labware, Franklin Lakes, NJ). Neutrophils ( $2.5 \times 10^5$ ) were placed in the lower chamber separated by a membrane from the upper chamber in which RBCs were placed ( $2.5 \times 10^6$ ).

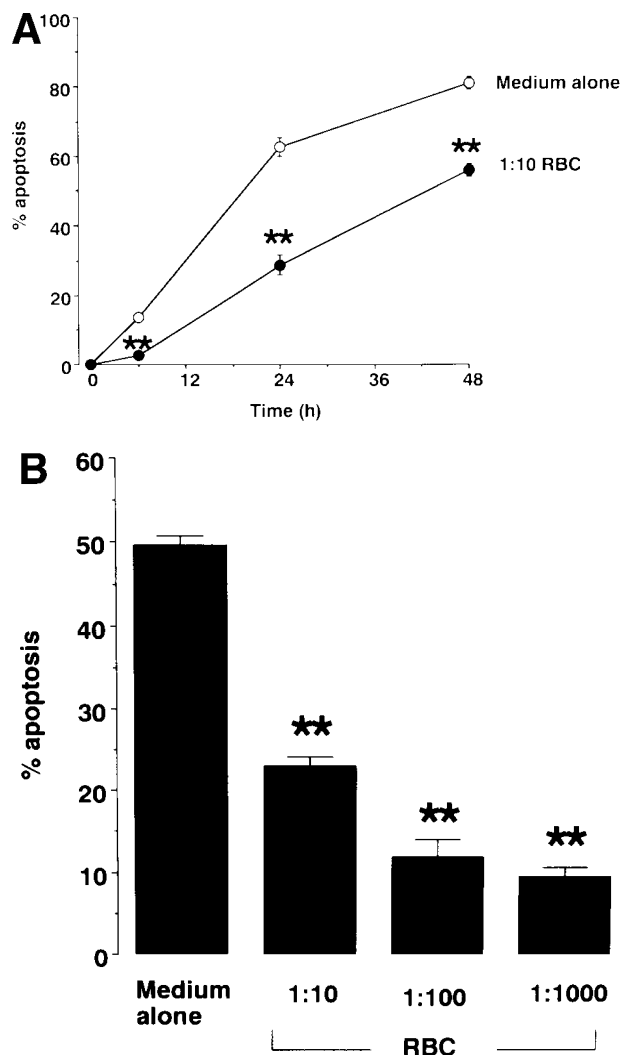
**Apoptosis assay.** Aliquots of neutrophils were cytospun on glass slides and air-dried. Slides were stained with May-Grünwald-Giemsa. Apoptosis was assessed based on nuclear pyknosis or chromatin condensation together with cytoplasmic vacuolation on oil-immersion microscopy.<sup>3</sup> Three hundred cells were scored in each experiment to determine the percentage of apoptotic cells.

**DNA fragmentation assay.** DNA fragmentation was evaluated using Apoptosis Ladder Detection Kit (Wako, Tokyo, Japan) according to the manufacturer's instructions. Briefly, neutrophils ( $1 \times 10^4$ ) were resuspended in lysis buffer containing RNase and proteases. Then, DNA was extracted in extraction buffer, precipitated by adding isopropanol, washed in 70% ethanol, air-dried, and resuspended in Tris-EDTA (TE) buffer. DNA was electrophoresed on a 1% agarose gel, stained with SYBR Green I (Molecular Probes, Inc, Eugene, OR), and visualized under ultraviolet light.

**Statistics.** Results are presented as mean  $\pm$  standard error of mean (SEM). Comparisons were made by Student's *t*-test or analysis of variance (ANOVA) with Scheffe's correction as appropriate. *P* < .05 was accepted as significant.

## RESULTS

**RBCs inhibit spontaneous apoptosis of neutrophils.** When cultured in RPMI 1640 containing 5% FCS, neutrophils constitutively underwent apoptosis, with 62.8% of cells apoptotic at 24 hours and 81.3% apoptotic by 48 hours as assessed by light microscopy (Fig 1A). The constitutive rate of apoptosis in neutrophils shown here is within the range reported previously.<sup>3,9</sup> When neutrophils were cultured with RBCs at a ratio of 1:10, neutrophil apoptosis was significantly inhibited at each time point evaluated (Fig 1A). Inhibition of apoptosis by RBCs was also dose-dependent (Fig 1B). When neutrophils were cultured with RBCs at a ratio of 1:1,000, a physiological ratio in the blood, apoptosis was decreased by about 80%. Figure 2 shows typical micrographs of neutrophils cultured for 24 hours with or without RBCs at a ratio of 1:100. Neutrophil-RBC contact was not required to inhibit apoptosis because apoptosis was inhibited even when neutrophils and RBCs were cultured separately using culture inserts. Percentage of apoptotic neutrophils after 24-hour culture with or without RBCs at a ratio of 1:10 was  $85.8\% \pm 1.0\%$  for no RBCs versus  $55.8\% \pm 1.7\%$  for contact culture and  $68.1\% \pm 2.6\%$  for separate culture (*P* < .01). The integrity of RBCs was not required because apoptosis was inhibited even when neutrophils were cultured with RBC



**Fig 1.** Effect of RBCs on neutrophil apoptosis. Neutrophils were cultured for varying times (A) or 16 hours (B) in RPMI 1640 containing 5% FCS in the presence or absence of 1:10 to 1:1000 RBCs. Apoptotic cells were identified by morphology on light microscopy and % apoptosis was obtained from 300 counts per experiment. Data represents mean  $\pm$  SEM of 6 experiments. \*\* *P* < .01 versus cells in the absence of RBCs.

lysates (data not shown). The protection of neutrophil apoptosis by RBCs was substantiated by the results of DNA gel electrophoresis, which showed that DNA ladder formation was inhibited when neutrophils were cultured with RBCs (Fig 3).

**Role of H<sub>2</sub>O<sub>2</sub> as a mediator of neutrophil apoptosis.** To determine the role of H<sub>2</sub>O<sub>2</sub> in neutrophil apoptosis, we cultured neutrophils in the presence or absence of either catalase or H<sub>2</sub>O<sub>2</sub>. As shown in Fig 4, application of catalase inhibited neutrophil apoptosis whereas H<sub>2</sub>O<sub>2</sub> promoted apoptosis. These results are consistent with a previous report<sup>9</sup> suggesting that H<sub>2</sub>O<sub>2</sub> is a mediator of neutrophil apoptosis.

**Catalase and glutathione are responsible for protection of neutrophil apoptosis by RBCs.** To determine the contribution of each of RBC antioxidant to protection against neutrophil apoptosis, RBC antioxidant systems were pharmacologically

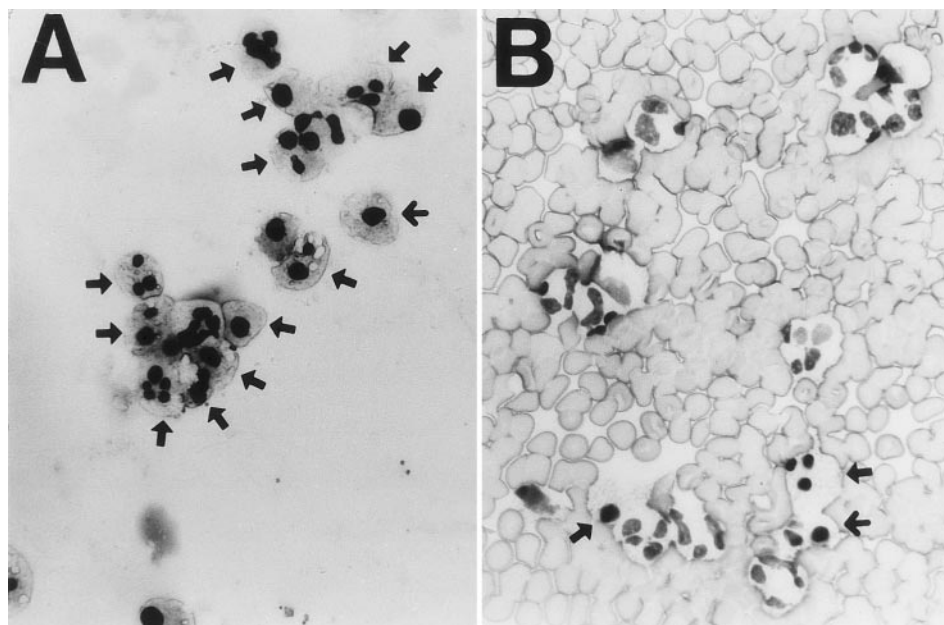


Fig 2. Light microscopy of neutrophils cultured for 24 hours without (A) or with (B) RBCs at a ratio of 1:100 ( $\times 400$ ). Arrows indicate apoptotic cells showing nuclear pyknosis or chromatin condensation together with cytoplasmic vacuolation. Note apoptosis is inhibited in the presence of RBCs.

blocked before analysis of the protective effect on apoptosis (Fig 5). Inhibition of RBC catalase by pretreatment with aminotriazole partially suppressed the protection against neutrophil apoptosis by RBCs. Likewise, inhibition of glutathione metabolism with chlorodinitrobenzene or mercaptosuccinate partially suppressed the protection against apoptosis. Simultaneous inhibition of catalase and glutathione metabolism, which account for the majority of the  $\text{H}_2\text{O}_2$ -scavenging activity of RBCs, completely abolished the protection. In contrast, pretreatment of RBCs with the anion channel blocker DIDS, which blocks transport of  $\text{O}_2^-$  across RBC membranes, had little effect on the protection against apoptosis by RBCs. These results suggest that the inactivation of  $\text{H}_2\text{O}_2$  by catalase and glutathione metabolism is the mechanism by which RBCs protect neutrophils against apoptosis.

#### DISCUSSION

There is increasing evidence to indicate that ROS play an important role in spontaneous- and stimulus-mediated apoptosis

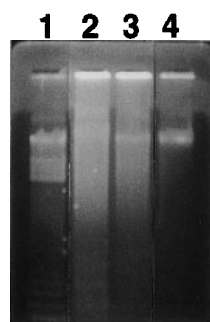


Fig 3. Effect of RBCs on nucleosomal DNA fragmentation in neutrophils. Neutrophils were cultured for 24 hours in the presence or absence of 1:1000 RBCs. Cells were harvested and DNA fragmentation was analyzed by agarose gel electrophoresis. Lane 1, molecular weight marker; lane 2, cells cultured without RBCs; lane 3, cells cultured with RBCs; lane 4, cells isolated freshly.

of neutrophils.<sup>8,9,24-27</sup> Our data showing that neutrophil apoptosis is promoted by exogenous  $\text{H}_2\text{O}_2$  but blocked by catalase confirms a previous study<sup>9</sup> indicating  $\text{H}_2\text{O}_2$  as a mediator of neutrophil apoptosis. Although cellular sources of  $\text{H}_2\text{O}_2$  are numerous and difficult to determine, the major source of endogenous  $\text{H}_2\text{O}_2$  is a leak from the mitochondrial electron transport chain. All aerobic cells constantly produce small amounts of  $\text{O}_2^-$  via mitochondrial respiratory enzymes.  $\text{O}_2^-$  is then dismutated to form  $\text{H}_2\text{O}_2$ . Many types of cells also generate

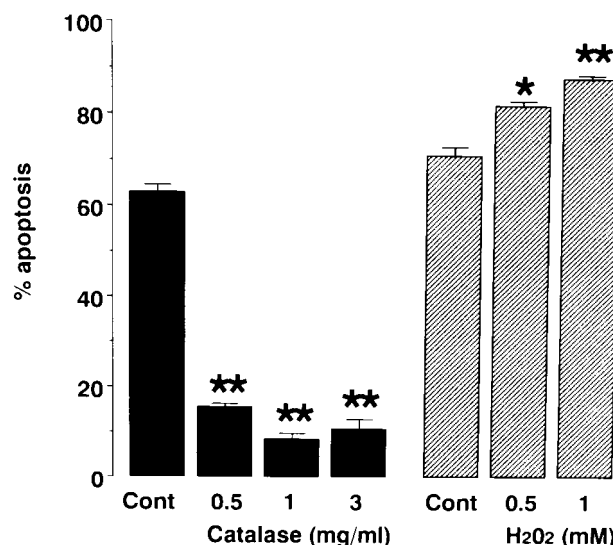


Fig 4. Effects of catalase and  $\text{H}_2\text{O}_2$  on neutrophil apoptosis. Neutrophils were cultured for 24 hours in the presence or absence of catalase and  $\text{H}_2\text{O}_2$ . Apoptosis was evaluated as described in Fig 1. To avoid inactivation of  $\text{H}_2\text{O}_2$  by serum, serum-free medium was used to assess the effect of  $\text{H}_2\text{O}_2$  on neutrophil apoptosis. This probably reflects increased apoptosis in  $\text{H}_2\text{O}_2$ -untreated control cells ( $70.8\% \pm 2.0\%$ ) compared with apoptosis in catalase-untreated control cells ( $62.8\% \pm 1.7\%$ ). Data represents mean  $\pm$  SEM of 4 experiments. \*  $P < .05$ , \*\*  $P < .01$  versus control cells in the absence of catalase or  $\text{H}_2\text{O}_2$ .

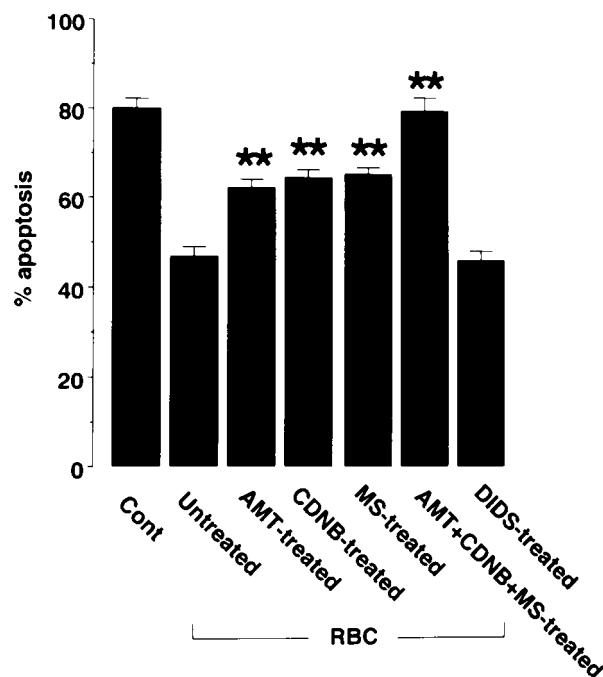


Fig 5. Effects of antioxidant inhibitors on the ability of RBCs to inhibit neutrophil apoptosis. RBCs were pretreated with the catalase inhibitor aminotriazole (AMT), the glutathione metabolism inhibitors chlorodinitrobenzene (CDNB) and mercaptosuccinate (MS), a combination of AMT, CDNB, and MS, or the  $O_2^-$  transport inhibitor DIDS. Excess reagents were removed by washing RBCs three times in PBS. Then, RBCs were mixed with neutrophils at a neutrophil/RBC ratio of 1:10. After 48 hours of culture, neutrophil apoptosis was evaluated as described in Fig 1. Data represents mean  $\pm$  SEM of five experiments. \*\* $P < .01$  versus untreated RBCs.

$O_2^-/H_2O_2$  via various enzyme systems including arachidonic acid metabolism, microsomal cytochrome systems, and DNA synthesis enzymes. Furthermore, phagocytes such as neutrophils are endowed with nicotinamide adenine dinucleotide phosphate (NADPH) oxidase that releases large amounts of  $O_2^-$  and  $H_2O_2$  on stimulation.<sup>28</sup> Thus, the exact source of  $H_2O_2$ -mediating apoptosis of neutrophils remains to be determined. Nevertheless, our data together with others<sup>9</sup> indicate that intracellular  $H_2O_2$ , at least in part, functions as a mediator of neutrophil apoptosis.

The major finding in our study is that RBCs inhibit apoptosis of neutrophils. A number of studies have suggested that RBCs can scavenge  $H_2O_2$  within their microenvironment. For example, RBCs can protect against  $H_2O_2$ -mediated damage to cells and tissues.<sup>12,13,15,16</sup> Because  $H_2O_2$  can freely cross the cell membrane<sup>17</sup> and there are 1000-fold more RBCs than neutrophils in the blood, the elimination of extracellular  $H_2O_2$  by RBCs is expected to decrease  $H_2O_2$  within neutrophils.

RBCs are endowed with high activities of catalase and glutathione peroxidase, which together probably account for the majority of  $H_2O_2$ -scavenging activity of RBCs.<sup>11,13,15,29</sup> Specific inhibition of these RBC antioxidants in the present study showed that both catalase and glutathione metabolism are responsible and sufficient for protection of apoptosis by RBCs. In contrast, inhibition of  $O_2^-$  transport across RBC membranes had no protective effect against apoptosis. These data imply that

inactivation of  $H_2O_2$  by catalase and glutathione within RBCs mediates the protective effect against neutrophil apoptosis.

The fact that RBCs can inhibit neutrophil apoptosis in vitro suggests that neutrophil apoptosis is inhibited in the circulation in vivo. Because blood plasma contains few antioxidant enzymes,<sup>30,31</sup> RBCs may constitute a major antioxidant system that suppresses neutrophil apoptosis in the blood. However, it should also be noted that although we found that RBCs inhibited neutrophil apoptosis in vitro, this effect was rather small at 6 hours of culture and became apparent after this point of time. Because neutrophils have a circulating half-life of less than 10 hours,<sup>1,2</sup> extrapolation of our in vitro data to in vivo situations requires caution.

Apoptosis is believed to provide a mechanism of neutrophil clearance that limits tissue injury and leads to resolution of inflammation.<sup>4</sup> Recently, apoptosis of neutrophils has also been shown to occur in the circulation.<sup>5,6</sup> However, it remains unclear whether apoptosis occurs as rapidly in the circulation as in tissue. If so, a considerable amount of circulating neutrophils would die before reaching their target tissue on inflammatory stimulation. Thus, it seems reasonable to speculate that there is a mechanism by which apoptosis of neutrophils is suppressed in the circulation. A recent study has shown that platelets also inhibit apoptosis of neutrophils although the mechanism is not well defined.<sup>7</sup> Taken together, our data suggest that neutrophils are protected against apoptosis in the circulation.

## REFERENCES

1. Cartwright GE, Athens JW, Wintrobe MM: The kinetics of granulopoiesis in normal man. *Blood* 24:780, 1964
2. Dancy JT, Deubelbeiss KA, Harker LA, Finch CA: Neutrophil kinetics in man. *J Clin Invest* 58:705, 1976
3. Savill JS, Wyllie AH, Henson JE, Walport MJ, Henson PM, Haslett C: Macrophage phagocytosis of aging neutrophils in inflammation. Programmed cell death in the neutrophil leads to its recognition by macrophages. *J Clin Invest* 83:865, 1989
4. Haslett C: Granulocyte apoptosis and inflammatory disease. *Br Med Bull* 53:669, 1997
5. Bincknell S, Van Eeden SF, Hayashi S, Hards J, English D, Hogg JC: A non-radioisotopic method for tracing neutrophils in vivo using 5'-bromo-2-deoxyuridine. *Am J Respir Cell Mol Biol* 10:16, 1994
6. Matsuba KT, Van Eeden SF, Bicknell SG, Walker BAM, Hayashi S, Hogg JC: Apoptosis in circulating PMN: Increased susceptibility in L-selectin-deficient PMN. *Am J Physiol* 272 (Heart Circ Physiol 41):H2852, 1997
7. Andonegui G, Trevani AS, López DH, Raiden S, Giordano M, Geffner JR: Inhibition of human neutrophil apoptosis by platelets. *J Immunol* 158:3372, 1997
8. Hannah S, Mecklenburgh K, Rahman I, Bellingan GJ, Greening A, Haslett C, Chilvers ER: Hypoxia prolongs neutrophil survival in vitro. *FEBS Lett* 372:233, 1995
9. Kasahara Y, Iwai K, Yachie A, Ohta K, Konno A, Seki H, Miyawaki T, Taniguchi N: Involvement of reactive oxygen intermediates in spontaneous and CD95(Fas/APO-1)-mediated apoptosis of neutrophils. *Blood* 89:1748, 1997
10. Lynch RE, Fridovich I: Permeation of the erythrocyte stroma by superoxide radical. *J Biol Chem* 253:4697, 1978
11. Van Asbeck BS, Hoidal J, Vercellotti GM, Schwartz BA, Moldow CF, Jacob HS: Protection against lethal hyperoxia by tracheal insufflation of erythrocytes: Role of red cell glutathione. *Science* 227:756, 1985
12. Toth KM, Clifford DP, Berger EM, White CW, Repine JE: Intact



human erythrocytes prevent hydrogen peroxide-mediated damage to isolated perfused rat lungs and cultured bovine pulmonary artery endothelial cells. *J Clin Invest* 74:292, 1984

13. Agar NS, Sadrzadeh SMH, Hallaway PE, Eaton JW: Erythrocyte catalase. A somatic oxidant defense? *J Clin Invest* 77:319, 1986
14. Brown JM, Grosso MA, Terada LS, Beehler CJ, Toth KM, Whitman GJ, Harken AH, Repine JE: Erythrocytes decrease myocardial hydrogen peroxide levels and reperfusion injury. *Am J Physiol* 256: H584, 1989
15. Test ST, Weiss SJ: Quantitative and temporal characterization of the extracellular H<sub>2</sub>O<sub>2</sub> pool generated by human neutrophils. *J Biol Chem* 259:399, 1984
16. Winterbourn CC, Stern A: Human red cells scavenge extracellular hydrogen peroxide and inhibit formation of hypochlorous acid and hydroxyl radical. *J Clin Invest* 80:1486, 1987
17. Chance B, Sies H, Boveris E: Hydroperoxide metabolism in mammalian organs. *Physiol Rev* 59:527, 1979
18. Okuda S, Nishiyama N, Saito H, Katsuki H: Hydrogen peroxide-mediated neuronal cell death induced by an endogenous neurotoxin, 3-hydroxykynurenine. *Proc Natl Acad Sci USA* 93:12553, 1996
19. Burke TM, Wolin MS: Hydrogen peroxide elicits pulmonary arterial relaxation and guanylate cyclase activation. *Am J Physiol* 252:H721, 1987
20. Awasthi YC, Garg HS, Dao DD, Patridge CA, Strivastva SK: Enzymatic conjugation of erythrocyte glutathione with 1-chloro-2,4-dinitrobenzene: The fate of glutathione conjugate in erythrocytes and the effect of glutathione depletion on hemoglobin. *Blood* 58:733, 1981
21. Michiels C, Remacle J: Use of the inhibition of enzymatic antioxidant system in order to evaluate their physiological importance. *Eur J Biochem* 177:435, 1988
22. English D, Anderson BR: Single-step separation of red blood cells, granulocytes, and mononuclear leukocytes on discontinuous density gradients of Ficoll-Hypaque. *J Immunol Methods* 5:249, 1974
23. Haslett C, Guthrie LA, Kopaniak MA, Johnson Jr RB, Henson PM: Modulation of multiple neutrophil functions by preparative methods or trace concentrations of bacterial lipopolysaccharide. *Am J Pathol* 119:101, 1985
24. Watson RWG, Redmond HP, Wang JH, Bouchier-Hayes D: Mechanisms involved in sodium arsenite-induced apoptosis of human neutrophils. *J Leukoc Biol* 60:625, 1997
25. Watson RWG, Rotstein OD, Jimenez M, Parodo J, Marshall JC: Augmented intracellular glutathione inhibits Fas-triggered apoptosis of activated human neutrophils. *Blood* 89:4175, 1997
26. Oishi K, Machida K: Inhibition of neutrophil apoptosis by antioxidants in culture medium. *Scand J Immunol* 45:21, 1997
27. Kettritz R, Falk RJ, Jennette JC, Gaido ML: Neutrophil superoxide release is required for spontaneous and FMLP-mediated but not for TNF  $\alpha$ -mediated apoptosis. *J Am Soc Nephrol* 8:1091, 1997
28. Babior BM, Kipnes RS, Curnutte JT: Biological defense mechanisms. The production by leukocytes of superoxide, a potential bactericidal agent. *J Clin Invest* 52:741, 1973
29. Cohen G, Hochstein P: Glutathione peroxidase: The primary agent for the elimination of hydrogen peroxide in erythrocytes. *Biochemistry* 2:1420, 1963
30. Marklund SL, Holme E, Hellner L: Superoxide dismutase in extracellular fluids. *Clin Chim Acta* 126:41, 1982
31. Halliwell B, Gutteridge JMC: Oxygen free radicals and iron in relation to biology and medicine. Some problems and concepts. *Arch Biochem Biophys* 246:501, 1986