

## REVIEW ARTICLE

**Role of Calcium in Glucocorticosteroid-Induced Apoptosis of Thymocytes and Lymphoma Cells: Resurrection of Old Theories by New Findings**

By Clark W. Distelhorst and George Dubyak

**M**ORE THAN 50 years ago, investigators observed that the thymus gland undergoes hypertrophy in association with adrenocortical insufficiency and atrophy in association with adrenocortical excess.<sup>1,2</sup> These observations led to the realization that adrenal corticosteroids (ACS) have a "lympholytic effect," which in turn stimulated clinical investigators to treat lymphomas with ACS for the first time in the late 1940s.<sup>3,4</sup> Today, we know that ACS do not directly lyse thymocytes and lymphoma cells, but rather induce apoptosis.<sup>5</sup> Apoptosis, or programmed cell death, is a genetically regulated process in which the cell is active in producing its own death, a type of cellular suicide for the sake of maintaining homeostasis in the cellular community.<sup>6-8</sup>

Although our understanding of apoptosis has advanced tremendously in recent years, the mechanism by which ACS induce apoptosis in thymocytes and lymphoma cells is not completely understood. In the 1970s, Sibley and Tompkins determined that the initial step in ACS-induced apoptosis is mediated through the ACS receptor and requires translocation of the receptor from the cytoplasm into the nucleus.<sup>9</sup> In the nucleus, the ACS receptor functions as a transcription factor, enhancing or repressing the expression of a selected repertoire of genes.<sup>10</sup> ACS may repress expression of genes necessary for cell survival by attenuating AP-1 (c-Fos/c-Jun) transcription factor activity,<sup>11</sup> or may induce the transcription of genes involved in carrying out the death program. The evidence in support of the latter concept is twofold: first, inhibitors of RNA and protein synthesis inhibit ACS-induced apoptosis<sup>12,13</sup>; and second, the transactivation domain of the ACS receptor is required for apoptosis induction by ACS.<sup>14</sup>

Progress in identifying ACS-inducible genes that mediate apoptosis had been slow. Now, in the past year, two ACS-inducible genes have been formally implicated in mediating apoptosis. One of the genes encodes a purinergic receptor, P<sub>2X</sub><sub>1</sub>, that functions as an ATP-gated calcium channel,<sup>15</sup> whereas the other gene encodes an inositol 1,4,5-trisphosphate receptor (IP<sub>3</sub>R) that functions as an IP<sub>3</sub>-gated calcium channel.<sup>16</sup> The fact that both of these genes encode calcium channels has reawakened interest in the role of calcium, a major intracellular second messenger molecule, as a mediator of apoptosis. In this review, we first summarize the evidence for involvement of these genes in apoptosis. Then, we attempt to integrate these novel findings with previous experimental evidence suggesting

a role for calcium in signaling glucocorticoid-induced apoptosis in both thymocytes and lymphoma cells.

P<sub>2X</sub><sub>1</sub> RECEPTORS AND APOPTOSIS

Recognition that purinergic receptors are involved in ACS-induced apoptosis evolved out of an effort by Owens et al<sup>17</sup> to identify differentially expressed mRNAs associated with ACS-induced apoptosis in thymocytes. One of the differentially expressed mRNAs identified by these investigators, termed RP-2, was induced early in the course of ACS-induced apoptosis. However, the identity of the RP-2 sequence was not elucidated until 5 years later, when it was discovered that RP-2 corresponds to a partial sequence of a gene encoding an ATP-gated cation channel, termed purinergic receptor or P<sub>2X</sub> receptor.<sup>18,19</sup> This discovery is particularly intriguing, because extracellular ATP has been reported to induce apoptosis in thymocytes by increasing the intracellular concentration of calcium.<sup>20,21</sup>

The P<sub>2X</sub> receptors constitute a family of at least seven members, distributed throughout central and peripheral neurons, smooth muscles, epithelia, developing skeletal neuromuscular junctions, lymphocytes, platelets, and macrophages. P<sub>2X</sub> receptors are nonselective cation channels with significant permeability to calcium. P<sub>2X</sub> receptors engage in intercellular communication by detecting the regulated (synaptic) or lytic (cell death) release of intracellular metabolites such as ATP. Unlike other well-known ion channels gated by extracellular ligands (eg, nicotinic, serotonin, 5-HT<sub>3</sub>), P<sub>2X</sub> receptors are characterized by only two transmembrane domains with intracellular amino- and carboxy-termini.<sup>22</sup> RP-2 corresponds to the subfamily member P<sub>2X</sub><sub>1</sub> (originally termed P<sub>2X</sub>R1), recently cloned from *vas deferens* and PC12 cells.<sup>18,19</sup> Curiously, P<sub>2X</sub> receptors are structurally similar to some of the *Caenorhabditis elegans* gene products implicated in neuronal cell selection and death.

*From the Departments of Medicine, Pharmacology, and Physiology and Biophysics, Case Western Reserve University, Cleveland, OH.*

*Submitted July 21, 1997; accepted September 22, 1997.*

*Address reprint requests to Clark W. Distelhorst, MD, Division of Hematology/Oncology, Department of Medicine, Case Western Reserve University, 10900 Euclid Ave, Cleveland, OH, 44106-4937.*

*© 1998 by The American Society of Hematology.*

*0006-4971/98/9103-0018\$3.00/0*

The link between P2X<sub>1</sub> receptor expression and ACS-induced apoptosis in rat thymocytes was recently established by Chvatchko et al.<sup>15</sup> These investigators found that P2X<sub>1</sub> receptors were upregulated in thymocytes during ACS-induced apoptosis. Furthermore, extracellular ATP enhanced ACS-induced apoptosis and antagonists of ATP substantially reduced ACS-induced apoptosis, suggesting that ACS-induced apoptosis is dependent on P2X<sub>1</sub> receptor activation by extracellular ATP.

#### IP<sub>3</sub>R AND APOPTOSIS

The other ACS-inducible gene implicated in mediating apoptosis of both thymocytes and lymphoma cells encodes the Type 3 IP<sub>3</sub>R.<sup>16</sup> The IP<sub>3</sub>R has been extensively characterized and plays a central role in calcium signaling.<sup>23,24</sup> Typically, IP<sub>3</sub> generated in response to G-protein-coupled receptors and receptor tyrosine kinases binds to the IP<sub>3</sub> receptor that spans the endoplasmic reticulum (ER) membrane.<sup>23,24</sup> The binding of IP<sub>3</sub> induces transient opening of the receptor, allowing calcium to flow from the ER lumen into the cytoplasm, thereby producing a transient elevation of cytosolic calcium that in turn activates signal transduction kinases. Although IP<sub>3</sub>R are most prominently located on the ER membrane, there is evidence for localization of the IP<sub>3</sub>R to the plasma membrane, enabling the IP<sub>3</sub>R to mediate entry of extracellular calcium into the cytoplasm<sup>16</sup> (and references therein). In cells undergoing apoptosis in response to ACS, immunocytochemical studies localized the IP<sub>3</sub>R (specifically subtype 3) to the plasma membrane.<sup>16</sup> Upregulation of IP<sub>3</sub>R in S49 lymphoma cells was associated with increased cytosolic calcium concentration, suggesting that increased expression of IP<sub>3</sub>R on the plasma membrane increased calcium entry.<sup>16</sup> Significantly, repression of ACS-induced IP<sub>3</sub>R expression by transfecting cells with an IP<sub>3</sub>R antisense plasmid not only decreased the induction of IP<sub>3</sub>R, but also inhibited the induction of apoptosis by ACS.<sup>16</sup>

#### RELATIONSHIP OF P2X<sub>1</sub> AND IP<sub>3</sub>R TO EARLIER EVIDENCE LINKING CALCIUM WITH ACS-INDUCED APOPTOSIS

The involvement of calcium in ACS-induced apoptosis was first suggested by Kaiser and Edelman,<sup>25</sup> who discovered that extracellular calcium is necessary for induction of apoptosis in thymocytes by ACS. This observation, which has been confirmed by others,<sup>12,13</sup> suggests that extracellular calcium uptake mediates ACS-induced apoptosis. The concept that calcium signals ACS-induced apoptosis is further supported by evidence that the calmodulin inhibitor, calmidazolium, interferes with ACS-induced thymocyte apoptosis.<sup>26,27</sup> However, in contrast to the situation in thymocytes, extracellular calcium is unnecessary for ACS-induced apoptosis of peripheral lymph node lymphocytes and lymphoma cells.<sup>28-33</sup>

These earlier findings are fully consistent with the patterns of expression of the P2X<sub>1</sub> receptor. P2X<sub>1</sub> expression was detected in thymocytes, but not in peripheral (lymph node) T lymphocytes.<sup>15</sup> Furthermore, there is a strong correlation between P2X<sub>1</sub> expression and the susceptibility of individual thymocyte subsets to ACS-induced apoptosis. In the thymus gland, immature CD4<sup>+</sup>CD8<sup>+</sup> thymocytes residing within the cortex are programmed to undergo apoptosis in response to ACS, whereas the more mature CD4<sup>+</sup>CD8<sup>-</sup> or CD4<sup>-</sup>CD8<sup>+</sup> thymocytes lo-

cated in the medulla and circulating T lymphocytes are less sensitive to ACS-induced apoptosis.<sup>34-37</sup> Significantly, P2X<sub>1</sub> expression was detected only in CD4<sup>+</sup>CD8<sup>+</sup> thymocytes, but not in peripheral (lymph node) T lymphocytes.<sup>15</sup>

The earlier findings suggesting a role of extracellular calcium in mediating ACS-induced apoptosis of thymocytes are also consistent with evidence that ACS treatment increases the expression of the Type 3 IP<sub>3</sub>R in cortical, but not medullary, thymocytes.<sup>16</sup> However, evidence that ACS induce the expression of Type 3 IP<sub>3</sub>R on the plasma membrane of S49 cells, a T-cell lymphoma line,<sup>16</sup> appears to be less consistent with earlier evidence indicating the extracellular calcium is not required for induction of apoptosis in lymphoma cells by ACS.<sup>29-32</sup> Moreover, it is possible that the localization of the IP<sub>3</sub>R to plasma membrane was a consequence of apoptotic bleb formation in ACS-treated lymphoma cells. The apoptotic blebs that form at the cell surface during apoptosis are membranous structures composed of ER membrane.<sup>38</sup> Thus, further work will be required to determine whether IP<sub>3</sub>R induced by ACS treatment are located primarily on the plasma membrane, or are located on the ER membrane and then relocated to the cell surface as apoptotic blebs form.

Although there is considerable evidence that extracellular calcium is not required for apoptosis induction by ACS in lymphoma cells, a role for calcium in mediating ACS-induced apoptosis of lymphoma cells has been supported by several findings. First, calmodulin gene expression is increased in T-cell lymphoma cells after treatment with ACS.<sup>27</sup> Second, calmidazolium interferes with apoptosis in ACS-treated lymphoma cells.<sup>27</sup> Third, stable expression of a cDNA encoding the high-affinity calcium-binding protein, calbindin, inhibited ACS-induced apoptosis in lymphoma cells.<sup>39</sup> Where, then, might the calcium come from that mediates ACS-induced apoptosis in these cells? We and others have detected a diminution of the ER calcium pool in ACS-treated lymphoma cells.<sup>33,40</sup> Thus, one theory is that calcium release from the ER, perhaps via IP<sub>3</sub>R located on the ER membrane, may be involved in signaling apoptosis in ACS-treated lymphoma cells.

#### HOW DOES CALCIUM SIGNAL APOPTOSIS?

Although the novel findings of the past year have provided molecular evidence of a role for calcium in ACS-induced apoptosis, the specific role that calcium plays in death induction and the signal transduction pathway initiated by cytosolic calcium elevation and how it leads to apoptosis are unknown. In the case of T-cell receptor (TCR)-mediated apoptosis, calcium in combination with calmodulin activates calcineurin, a cytosolic protein phosphatase that dephosphorylates and thereby activates the transcription factor NF-AT<sub>c</sub>, leading to increased transcription of calcium-regulated, immediate-early genes, including Nur77.<sup>41</sup> Calcineurin function, and hence TCR-mediated apoptosis, is inhibited by the potent immunosuppressants cyclosporin A and FK506.<sup>42</sup> However, these agents do not inhibit ACS-induced apoptosis.<sup>42</sup> Furthermore, ACS- and TCR-mediated apoptotic pathways are mutually antagonistic<sup>42</sup> and calcineurin activation protects T cells from ACS-induced apoptosis.<sup>43</sup> Also, Nur77 is not significantly induced by ACS.<sup>41</sup> One possible lead is a recently identified calcium-binding protein, ALG-2, that has been implicated as necessary for ACS-induced

apoptosis, but its precise role in the apoptotic process has not been defined.<sup>44</sup>

Another concept deserving further consideration is that while calcium mediates apoptosis of ACS-treated cells, it might not be necessary for ACS-induced cell death. This concept is based on a recent report by Iseki et al,<sup>45</sup> who found that high concentrations of the intracellular calcium chelator Quin-2/AM inhibited DNA fragmentation in ACS-treated thymocytes, but did not inhibit cell death. In their hands, calmodulin inhibitors blocked DNA fragmentation, but markedly enhanced cytolysis. One interpretation of these findings is that calcium might be required for DNA cleavage during apoptosis, but not for cell death. Thus, calcium might be involved in endonuclease activation, as part of the degradation phase of apoptosis, but might not be necessary for cell death.

However, the role of calcium in endonuclease activation varies among different types of lymphocytes. Whereas the endonuclease responsible for apoptotic DNA fragmentation in ACS-treated thymocytes requires calcium for maximal activity,<sup>5,13,31,46-49</sup> DNA is cleaved by a calcium-independent endonuclease in the CEM human lymphoblast cell line.<sup>50</sup> These observations correlate with the evidence that extracellular calcium uptake is required for ACS-induced thymocyte apoptosis, but not for ACS-induced apoptosis of lymphoma cell.<sup>12,13,25,28</sup>

Another potential role for calcium in ACS-induced apoptosis might be in protease activation. The calcium-dependent neutral protease, calpain, is activated in the course of apoptosis induction in ACS-treated thymocytes, and a calpain inhibitor appears to inhibit ACS-induced thymocyte cell death.<sup>51</sup> Moreover, destruction of the nuclear structural protein lamin, an event that appears to precede endonucleolytic DNA cleavage during ACS-induced apoptosis, is inhibited by a calpain inhibitor.<sup>52,53</sup> Currently, there is little mechanistic insight into how ACS-induced cytosolic calcium elevation might lead to activation of interleukin-1 $\beta$  converting enzyme-like proteases, which clearly play a prominent role in ACS-induced apoptosis, as well as other forms of apoptosis.<sup>54</sup>

#### SUMMARY

Since the initial observations by Kaiser and Edelman,<sup>25,28</sup> interest in the role of calcium in ACS-induced apoptosis has wavered, in part because of the fact that extracellular calcium is only necessary for induction of apoptosis in thymocytes, but not in peripheral lymphocytes or lymphoma cells. Now, as a result of molecular evidence implicating two separate ligand-gated calcium channels in ACS-induced apoptosis, interest in the role of calcium is sure to be renewed. The major challenge lies in determining the signal transduction pathway through which ACS-induced calcium fluxes mediate apoptosis.

#### NOTE ADDED IN PROOF

Recent results (Jayaraman T, Marks AR: *Mol Cell Biol* 17:3005, 1997) indicate that T cells deficient in IP<sub>3</sub>RI are resistant to ACS-induced apoptosis, providing additional evidence for a role of calcium in signaling ACS-induced apoptosis.

#### REFERENCES

1. Dougherty TF, White A: Effect of pituitary adrenotropic hormone on lymphoid tissue. *Proc Soc Exp Biol Med* 53:132, 1943

2. Heilman RR, Kendall EC: The influence of 11-dehydro-17-hydroxy-corticosterone (compound E) on the growth of a malignant tumor in the mouse. *Endocrinology* 34:416, 1944

3. Pearson OH, Eliel LP, Rawson RW, Dobriner K, Rhoads CP: ACTH- and cortisone-induced regression of lymphoid tumors in man: A preliminary report. *Cancer* 2:943, 1949

4. Pearson OH, Eliel LP: Use of pituitary adrenocorticotrophic hormone (ACTH) and cortisone in lymphomas and leukemias. *JAMA* 144:1349, 1950

5. Wyllie AH: Glucocorticoid-induced thymocyte apoptosis is associated with endogenous endonuclease activation. *Nature* 284:555, 1980

6. Raff MC: Social controls on cell survival and cell death. *Nature* 356:397, 1992

7. Thompson CB: Apoptosis in the pathogenesis and treatment of disease. *Science* 267:1456, 1995

8. Yang E, Korsmeyer SJ: Molecular thanatopsis: A discourse on the BCL2 family and cell death. *Blood* 88:386, 1996

9. Sibley CH, Tompkins GM: Isolation of lymphoma cell variants resistant to killing by glucocorticoids. *Cell* 2:213, 1974

10. Beato M: Gene regulation by steroid hormones. *Cell* 56:335, 1989

11. Helmborg A, Auphan N, Caelles C, Karin M: Glucocorticoid-induced apoptosis of human leukemic cells is caused by the repressive function of the glucocorticoid receptor. *EMBO J* 14:452, 1995

12. Wyllie AH, Morris RG, Smith AL, Dunlop D: Chromatin cleavage in apoptosis: Association with condensed chromatin morphology and dependence on macromolecular synthesis. *J Pathol* 142:67, 1984

13. Cohen JJ, Duke RC: Glucocorticoid activation of a calcium-dependent endonuclease in thymocyte nuclei leads to cell death. *J Immunol* 132:38, 1984

14. Dieken ES, Miesfeld RL: Transcriptional transactivation functions localized to the glucocorticoid receptor N terminus are necessary for steroid induction of lymphocyte apoptosis. *Mol Cell Biol* 12:589, 1992

15. Chvatchko Y, Valera S, Aubry J-P, Renno T, Buell G, Bonnefoy J-Y: The involvement of an ATP-gated ion channel, P2X<sub>1</sub>, in thymocyte apoptosis. *Immunity* 5:275, 1996

16. Khan AA, Soloski MJ, Sharp AH, Schilling G, Sabatini DM, Li S-H, Ross CA, Snyder SH: Lymphocyte apoptosis: Mediation by increased type 3 inositol 1,4,5,-trisphosphate receptor. *Science* 273:503, 1996

17. Owens GP, Hahn WE, Cohen JJ: Identification of mRNAs associated with programmed cell death in immature thymocytes. *Mol Cell Biol* 11:4177, 1991

18. Brake AJ, Wagenbach MJ, Julius D: New structural motif for ligand-gated ion channels defined by an ionotropic ATP receptor. *Nature* 371:519, 1994

19. Valera S, Hussy N, Evans RJ, Adami N, North RA, Surprenant A, Buell G: A new class of ligand-gated ion channel defined by P2X receptor for extracellular ATP. *Nature* 371:516, 1994

20. Pizzo P, Zanovello P, Bronte V, Di Virgilio F: Extracellular ATP causes lysis of mouse thymocytes and activates a plasma membrane ion channel. *Biochem J* 274:139, 1991

21. Zheng LM, Zychlinsky A, Liu C-C, Ojcius DM, Young JD-E: Extracellular ATP as a trigger for apoptosis or programmed cell death. *J Cell Biol* 112:279, 1991

22. North RA: Families of ion channels with two hydrophobic segments. *Curr Opin Cell Biol* 8:474, 1996

23. Berridge MJ: Inositol trisphosphate and calcium signalling. *Nature* 361:315, 1993

24. Clapham DE: Calcium signaling. *Cell* 80:259, 1995

25. Kaiser N, Edelman IS: Calcium dependence of glucocorticoid-induced lymphocytolysis. *Proc Natl Acad Sci USA* 74:632, 1977

26. McConkey DJ, Nicotera P, Hartzell P, Bellomo G, Wyllie AH,

Orrenius S: Glucocorticoids activate a suicide process in thymocytes through an elevation of cytosolic Ca<sup>2+</sup> concentration. *Arch Biochem Biophys* 269:365, 1989

27. Dowd DR, MacDonald PN, Komm BS, Haussler MR, Miesfeld R: Evidence for early induction of calmodulin gene expression in lymphocytes undergoing glucocorticoid-mediated apoptosis. *J Biol Chem* 266:18423, 1991
28. Kaiser N, Edelman IS: Further studies on the role of calcium in glucocorticoid-induced lymphocytolysis. *Endocrinology* 103:936, 1978
29. Nicholson ML, Young DA: Effect of glucocorticoid hormones in vitro on the structural integrity of nuclei in corticosteroid-sensitive and -resistant lines of lymphosarcoma P1798. *Cancer Res* 38:3673, 1978
30. Nicholson ML, Young DA: Independence of the lethal actions of glucocorticoids on lymphoid cells from possible hormone effects on calcium uptake. *J Supramol Struct* 10:165, 1979
31. Alnemri ES, Litwack G: Glucocorticoid-induced lymphocytolysis is not mediated by an induced endonuclease. *J Biol Chem* 264:4104, 1989
32. Bansal N, Houle AG, Melnykovich G: Dexamethasone-induced killing of neoplastic cells of lymphoid derivation: Lack of early calcium involvement. *J Cell Physiol* 143:105, 1990
33. Bian X, Hughes FM, Huang Y, Cidlowski JA, Putney JW: Roles of cytoplasmic Ca<sup>2+</sup> and intracellular Ca<sup>2+</sup> stores in induction and suppression of apoptosis in S49 cells. *Am J Physiol* 272 (Cell Physiol 41):C1241, 1997
34. Claman HS: Corticosteroids and lymphoid cells. *N Engl J Med* 287:388, 1971
35. Ranelletti FO, Piantelli M, Iacobelli S, Musiani P, Longo P, Lauriola L, Marchetti P: Glucocorticoid receptors and in vitro corticosteroid sensitivity of peanut-positive and peanut-negative human thymocyte subpopulations. *J Immunol* 127:849, 1981
36. Cederig R, Dialynas DP, Fitch FW, MacDonald HR: Precursors of T cell growth factor producing cells in the thymus. *J Exp Med* 158:1654, 1983
37. Cohen JJ: Programmed cell death in the immune system. *Adv Immunol* 50:55, 1991
38. Casciola-Rosen LA, Anhalt G, Rosen A: Autoantigens targeted in systemic lupus erythematosus are clustered in two populations of surface structures on apoptotic keratinocytes. *J Exp Med* 179:1317, 1994
39. Dowd DR, MacDonald PN, Komm BS, Haussler MR, Miesfeld RL: Stable expression of the calbindin-D28K complementary DNA interferes with the apoptotic pathway in lymphocytes. *Mol Endocrinol* 6:1843, 1992
40. Lam M, Dubyak G, Distelhorst CW: Effect of glucocorticosteroid treatment on intracellular calcium homeostasis in mouse lymphoma cells. *Mol Endocrinol* 7:686, 1993
41. Liu Z-G, Smith SW, McLaughlin KA, Schwartz LM, Osborne BA: Apoptotic signals delivered through the T-cell receptor of a T-cell hybrid require the immediate-early gene *nur77*. *Nature* 367:281, 1994
42. Zacharchuk CM, Mercep M, Chakraborti PK, Simons SS, Ashwell JD: Programmed T lymphocyte death: Cell activation- and steroid-induced pathways are mutually antagonistic. *J Immunol* 145:4037, 1990
43. Zhao Y, Tozawa Y, Iseki R, Mukai M, Iwata M: Calcineurin activation protects T cells from glucocorticoid-induced apoptosis. *J Immunol* 154:6346, 1995
44. Vito P, Lacana E, D'Adamio LD: Interfering with apoptosis: Ca<sup>2+</sup>-binding protein ALG-2 and alzheimer's disease gene ALG-3. *Science* 271:521, 1996
45. Iseki R, Kudo Y, Iwata M: Early mobilization of Ca<sup>2+</sup> is not required for glucocorticoid-induced apoptosis in thymocytes. *J Immunol* 151:5198, 1993
46. Vedeckis WV, Bradshaw HDJ: DNA fragmentation in S49 lymphoma cells killed with glucocorticoids and other agents. *Mol Cell Endocrinol* 30:215, 1983
47. Jones DP, McConkey DJ, Nicotera P, Orrenius S: Calcium-activated DNA fragmentation in rat liver nuclei. *J Biol Chem* 264:6398, 1989
48. Gaido ML, Cidlowski JA: Identification, purification, and characterization of a calcium-dependent endonuclease (NUC18) from apoptotic rat thymocytes. *J Biol Chem* 266:18580, 1991
49. Ellis RE, Yuan J, Horvitz HR: Mechanisms and functions of cell death. *Annu Rev Cell Biol* 7:663, 1991
50. Alnemri ES, Litwack G: Activation of internucleosomal DNA cleavage in human CEM lymphocytes by glucocorticoid and novobiocin. *J Biol Chem* 265:17323, 1990
51. Squier MKT, Miller ACK, Malkinson AM, Cohen JJ: Calpain activation in apoptosis. *J Cell Physiol* 159:229, 1994
52. Neamati N, Fernandez A, Wright S, Kiefer J, McConkey DJ: Degradation of lamin B1 precedes oligonucleosomal DNA fragmentation in apoptotic thymocytes and isolated thymocyte nuclei. *J Immunol* 154:3788, 1995
53. McConkey DJ: Calcium-dependent, interleukin 1 $\beta$ -converting enzyme inhibitor-insensitive degradation of lamin B1 and DNA fragmentation in isolated thymocyte nuclei. *J Biol Chem* 271:22398, 1996
54. Henkart PA: ICE family proteases: Mediators of all apoptotic cell death? *Cell* 4:195, 1996