

jaundice, in which oxidative hemolysis may play an important role, is more common among newborns with the *A/*C genotype than among those with the *A/*A genotype.

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REFERENCES

1. Bottini E, Lucarelli P, Agostino R, Palmarino R, Businco L, Antognoni G: Favism: Association with erythrocyte acid phosphatase phenotype. *Science* 171:409, 1971
2. Bottini E: Favism: Current problems and investigations. *J Med Genet* 10:154, 1973
3. Arese P, De Flora A: Pathophysiology of hemolysis in glucose-6-phosphate dehydrogenase deficiency. *Semin Hematol* 27:1, 1990
4. Low PS: Structure and function of the cytoplasmic domain of band 3: Center of erythrocyte membrane-peripheral protein interactions. *Biochim Biophys Acta* 864:145, 1986

5. Dissing J: Human "red cell" acid phosphatase (ACP1) genetic, catalytic and molecular properties. PhD Thesis: Kobenhavn University, Kobenhavn, Denmark, 1993
6. Stefani M, Caselli A, Bucciantini M, Pazzagli L, Dolfi F, Camici G, Manao G, Ramponi G: Dephosphorylation of tyrosine phosphorylated synthetic peptides by rat liver phosphotyrosine protein phosphatase isoenzymes. *FEBS Lett* 326:131, 1993
7. Boivin P, Galand C: The human red cell acid phosphatase is a phosphotyrosine protein phosphatase which dephosphorylates the membrane protein band 3. *Biochem Biophys Res Commun* 134:557, 1986
8. Harrison ML, Rathinavelu P, Arese P, Geahlen RL, Low PS: Role of band 3 tyrosine phosphorylation in the regulation of erythrocyte glycolysis. *J Biol Chem* 266:4106, 1991
9. Bottini E, Modiano G: Effect of oxidized glutathione on human red cell acid phosphatases. *Biochem Biophys Res Commun* 17:260, 1964
10. Bottini E, Modiano G: On the effect of oxidized glutathione and acetylphenylhydrazine on red cell phosphatase. *Blood* 27:281, 1966
11. Bottini E, Modiano G, Santolamazza C, Filippi G, Businco L: Studies of the in vitro effects of oxidized glutathione and acetylphenylhydrazine on acid phosphatases of human red blood cells. An experimental model for the investigation of hemolytic drug action at the molecular level. *Clin Chim Acta* 31:243, 1971
12. Bottini E, Lucarelli P, Bastianon V, Gloria F: Erythrocyte acid phosphatase polymorphism and haemolysis. *J Med Genet* 9:434, 1972

Autologous Stem Cell Transplantation: Exogenous Granulocyte Colony-Stimulating Factor or Granulocyte-Macrophage Colony-Stimulating Factor Modulate the Endogenous Cytokine Levels

To the Editor:

Autologous bone marrow (BM) and peripheral blood (PB) stem cell transplantation (SCT) is widely used in a variety of hematological malignancies and solid tumors.¹ After high-dose conditioning therapy,² the recovery of hematopoiesis is dependent on stem cell self-renewal and differentiation into lineage-committed progenitors, which undergo differentiation and maturation to morphologically recognizable precursors and terminal cells circulating in PB.

SCT is associated with absolute leukopenia before hematopoietic recovery.² Administration of recombinant granulocyte-macrophage colony-stimulating factor (GM-CSF) and G-CSF accelerates hematopoietic recovery.³ A number of investigators have detected elevated plasma G-CSF,^{4,9} GM-CSF,⁴ interleukin-3 (IL-3),¹⁰ M-CSF,^{7,11} IL-6,^{4,5,11} and IL-8 following myeloablation and marrow or PB progenitor cell transplantation. Time-course studies have demonstrated kinetic profiles that imply a relationship between cytokine levels and engraftment. In this context, we have previously shown that (1) G-CSF, IL-8, and IL-6 levels peak at the time of neutrophil nadir^{8,9}; (2) G-CSF, IL-8, and IL-6 peak levels directly correlate with maximum neutrophil counts. Hence, it was proposed that these three cytokines are coordinately secreted after SCT and mediate the production and activation of neutrophils.^{8,9} Finally, recent studies^{11,12} have provided evidence that accessory cells do not contribute to the production of endogenous cytokines released after SCT; in fact, patients transplanted with purified CD34⁺ cells release cytokines, including G-CSF, IL-6, IL-8, and IL-3, at levels similar to those observed in patients transplanted with total PBMCs.^{11,12}

In the present study, the PBSCT model has been analyzed to investigate (1) the influence of exogenous G-CSF/erythropoietin (Epo) and GM-CSF/Epo on the production of IL-3, GM-CSF, G-CSF, M-CSF, IL-8, and IL-6; (2) the relationship between IL-5 release and eosinophil response, and the effect of exogenous GM-

Table 1. Ovarian Cancer Patients: Main Clinical Features, Infused Cells, and PBMC Recovery

Patient No.	Age	Infused PBMCs		PBMC Recovery		
		MNC (× 10 ⁶ /kg)	CFU-GM (× 10 ⁴ /kg)	WBC >1 × 10 ⁹ /L days	PMNC >0.5 × 10 ⁹ /L days	Platelets >50 × 10 ⁹ /L days
1	35	11.7	40.3	10	10	11
2	36	7.3	59.2	11	12	10
3	38	13.0	67.2	9	10	10
4	35	9.2	19.4	10	10	12
5	47	8.9	19.2	11	11	11
6	44	5.8	46.1	11	11	11
7	43	7.8	130.1	7	7	11
8	57	6.1	36.0	8	8	10
9	47	7.9	51.9	8	8	9
10	54	6.9	18.6	9	9	11
11	36	7.7	22.3	10	10	14
12	46	8.4	35.5	10	10	10
13	54	5.4	12.5	8	8	13
14	44	4.5	44.0	9	9	10
15	44	7.0	46.2	8	7	12
16	56	1.7	33.2	9	9	11
17	40	6.7	45.1	9	8	10
18	55	5.0	N.V.	10	10	11
19	51	3.3	11.9	9	9	13
20	60	5.8	N.V.	10	9	12
21	57	5.1	N.V.	9	9	10

Patients 1 to 6 PBSCT	Patients 7 to 15 PBSCT + G-CSF/Epo	Patients 16 to 21 PBSCT + G-CSF/Epo
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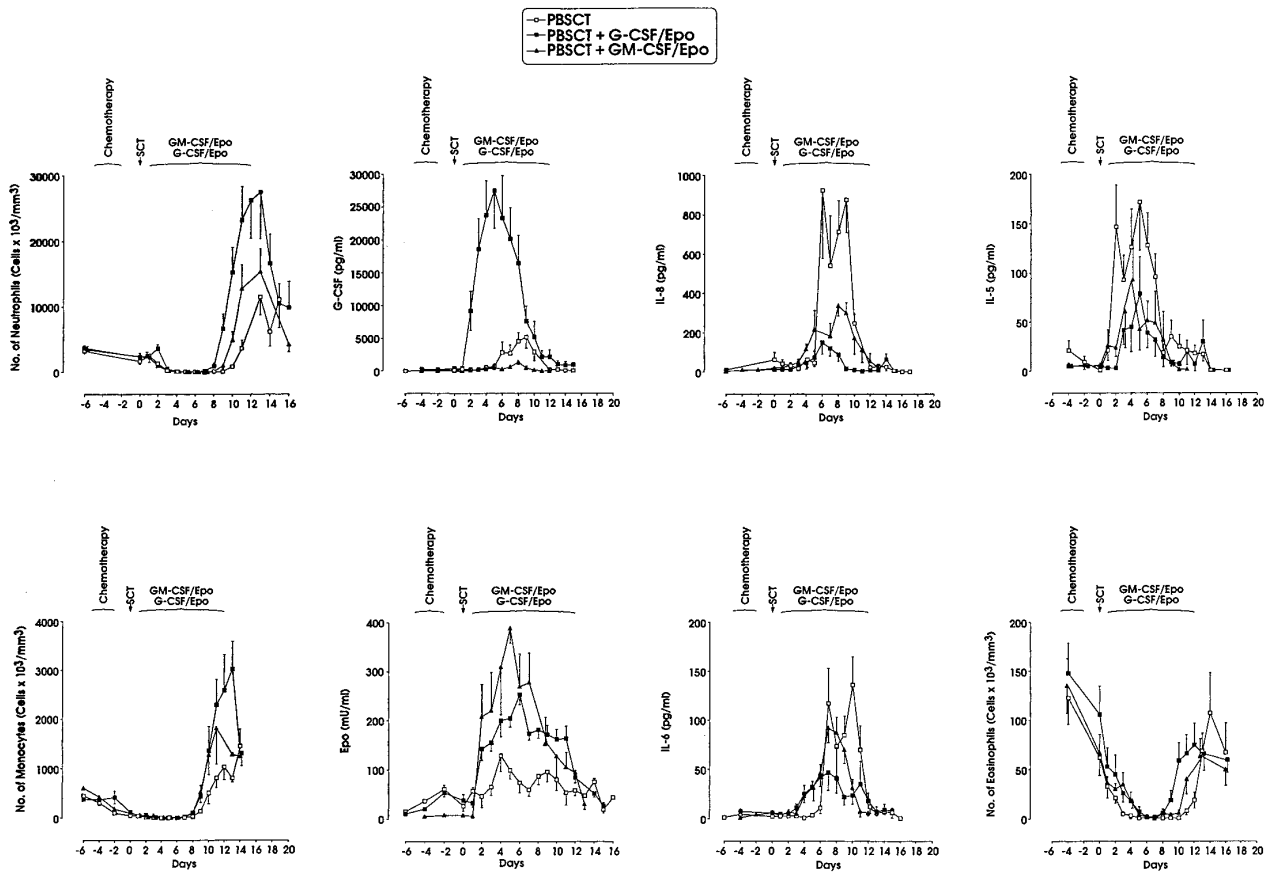


Fig 1. Neutrophil, monocyte, and eosinophil recovery, and G-CSF, Epo, IL-6, IL-8, and IL-5 plasma levels in 21 ovarian cancer patients (6 treated with no exogenous HGFs, 9 with G-CSF/Epo, and 6 with GM-CSF/Epo) undergoing SCT after high-dose chemotherapy. Chemotherapy days were -5 to -3 and SCT was performed on day 0. Mean \pm SEM values are presented.

CSF and G-CSF on these parameters; and (3) the relationship of lactoferrin (Lf) plasma levels with the initial G-CSF peak and the subsequent neutrophil rescue. Modifications of plasma cytokine levels may be due to changes in production, release, consumption, or clearance. This clinical approach allows to establish temporal correlations between hematopoietic growth factor (HGF) administration and fluctuations of endogenous serum cytokines. However, the underlying cause/effect mechanisms remain largely hypothetical. Furthermore, because exogenous GM-CSF or G-CSF was combined with Epo, the specific role of each cytokine cannot be determined. Despite these limitations intrinsic to the clinical setting, the results provide novel information on the interaction between exogenous HGFs and the SCT-related cytokine release network.

We have also monitored the plasma concentrations of HGFs (ie, IL-3, GM-CSF, G-CSF, Epo, IL-8, IL-6, and IL-5) and granule proteins (Lf and myeloperoxidase [Mpo]) in 21 ovarian cancer patients undergoing autologous PB SCT (Table 1).

G-CSF levels were markedly higher after exogenous G-CSF/Epo, as expected; furthermore, they were significantly lower following GM-CSF/Epo than after SCT alone; GM-CSF levels, moderately elevated after exogenous GM-CSF/Epo, were similar in the G-CSF/Epo and control groups. Finally, patients treated with either G-CSF/Epo or GM-CSF/Epo showed higher Epo plasma concentrations than the group receiving no exogenous HGFs (Fig 1 and data not shown).

When compared with SCT alone, (1) G-CSF/Epo treatment mod-

erately increased the IL-3 level peaking at day +10, and did not significantly modify M-CSF level (data not shown), whereas it almost completely suppressed IL-8 and moderately decreased IL-6 and IL-5 levels (Fig 1); (2) GM-CSF/Epo treatment increased M-CSF, associated with a moderate increase of IL-3 levels peaking at day +10, moderately decreased both IL-5 and IL-8 concentration and did not modify IL-6 (Fig 1 and data not shown).

These observations indicate that exogenous G-CSF treatment induces a marked decrease of IL-8 levels and moderately decreased IL-6 concentrations, whereas exogenous GM-CSF markedly reduces G-CSF and IL-8 concentrations and increases M-CSF levels. The increase of IL-3 and the decrease of IL-5 levels were similarly observed in patients treated with G-CSF/Epo or GM-CSF/Epo.

Altogether, these results indicate that exogenous HGFs modulate endogenous cytokine levels; consideration of these aspects may contribute to optimize HGF treatment protocols in the clinical SCT setting.

We also evaluated the relationship between these phenomena and Lf release: (1) in all three groups the Lf concentration is strictly and directly correlated with the neutrophil response with respect to time-response patterns and plasma levels. (2) In patients subjected to SCT alone, the G-CSF decrease after the G-CSF peak coincides with initiation of the Lf response; the inverse correlation between these two parameters is highly significant (data not shown). Altogether, these temporal correlations after SCT provide circumstantial evi-

dence that, after the G-CSF peak, the generated granulocytes release Lf, which could negatively feedback on the release of G-CSF, thus in line with the experimental model proposed by Broxmeyer.¹³

ACKNOWLEDGMENT

We thank M. Fontana and C. Mastropietro for typing the manuscript, D. Marinelli for editorial assistance, and M. Teragnoli for graphics. This study was supported in part by CNR (ACRO Project, No. 95.00525.39, No. 95.00545.39), Rome and AIRC, Milan, Italy.

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REFERENCES

1. Kessinger A, Armitage JO: The evolving role of autologous peripheral stem cell transplantation following high-dose therapy for malignancies. *Blood* 77:211, 1991
2. Brandt SJ, Peters WP, Atwater SK, Kurtzberg J, Borowitz MJ, Jones RB, Shpall EJ, Bast RC Jr, Gilbert CJ, Oette DH: Effect of recombinant human granulocyte-macrophage colony-stimulating factor on hematopoietic reconstitution after high-dose chemotherapy and autologous bone marrow transplantation. *N Engl J Med* 318:869, 1988
3. Nemunaitis J, Rabinowe SN, Singer JW, Bierman PJ, Vose JM, Freedman AS, Onetto N, Gillis S, Oette D, Gold M, Buckner CD, Hansen JA, Ritz J, Appelbaum FR, Armitage JO, Nadler LM: Recombinant granulocyte-macrophage colony-stimulating factor after autologous bone marrow transplantation for lymphoid cancer. *N Engl J Med* 324:1773, 1991
4. Sallerfors B, Olofsson T, Lenhoff S: Granulocyte-macrophage colony-stimulating factor (GM-CSF) and granulocyte-stimulating factor (G-CSF) in serum in bone marrow transplanted patients. *Bone Marrow Transplant* 8:191, 1991
5. Rabinowitz J, Petros WP, Stuart AR, Peters WP: Characterization of endogenous cytokine concentrations after high-dose chemotherapy with autologous bone marrow support. *Blood* 81:2452, 1993
6. Cairo MS, Suen Y, Sender L, Gillan ER, Ho W, Plunkett JM, Van de Ven C: Circulating granulocyte colony-stimulating factor (G-CSF) levels after allogeneic and autologous bone marrow transplantation: Endogenous G-CSF production correlates with myeloid engraftment. *Blood* 79:1869, 1992
7. Haas R, Gericke G, Witt B, Cayeux S, Hunstein W: Increased serum levels of granulocyte colony-stimulating factor after autologous bone marrow or blood stem cell transplantation. *Exp Hematol* 21:109, 1993
8. Baiocchi G, Scambia G, Benedetti P, Menichella G, Testa U, Pierelli L, Martucci R, Foddai ML, Bizzi B, Mancuso S, Peschle C: Autologous stem cell transplantation: Sequential production of hematopoietic cytokines underlying granulocyte recovery. *Cancer Res* 53:1297, 1993
9. Testa U, Martucci R, Rutella S, Scambia G, Sica S, Benedetti Panici P, Pierelli L, Menichella G, Leone G, Mancuso S, Peschle C: Autologous stem cell transplantation: Release of early- and late-acting growth factors relates with hemopoietic ablation and recovery. *Blood* 84:3532, 1994
10. Brugger W, Heimfeld S, Berenson RJ, Mertelsmann R, Kanz L: Reconstitution of hematopoiesis after high-dose chemotherapy by autologous cells generated ex vivo. *N Engl J Med* 333:283, 1995
11. Watts MJ, Jones HM, Sullivan AM, Langabeer SE, Jamieson E, Fielding A, Williams C, Berenson RJ, Goldstone AH, Linch DC: Accessory cells do not contribute to G-CSF or IL-6 production nor to rapid haematological recovery following peripheral blood stem cell transplantation. *Br J Haematol* 919:767, 1995
12. Sica S, Testa U, Rutella S, Martucci R, Chiusolo P, Etuk B, Di Mario A, Leone G, Peschle C: Endogenous cytokine production after immunoselected CD34⁺ peripheral blood progenitor cell transplant: Comparison with unfractionated progenitor transplant. *Br J Haematol* 1996 (in press)
13. Broxmeyer HE, Smithyman A, Eger RR: Identification of lactoferrin as granulocyte-derived inhibitor of colony-stimulating activity production. *J Exp Med* 148:1052, 1978