

Correlation of age of disease presentation with affected gene. Molteni et al describe deleterious germ line variants causing cytopenias and hypocellular bone marrows that vary across the age spectrum, with alleles in *GATA2* and Fanconi anemia genes common in younger individuals and *DDX41* in older individuals. Genes in which these alleles were identified are plotted as a function of age of presentation (on the x-axis) vs the diagnosis of the individual (top, ICUS/CCUS; bottom, myeloid malignancies; on the y-axis). Professional illustration by Somersault18:24.

ACMG recommend germ line genetic testing when the pretest probability of a positive finding is >5%.⁸⁻¹⁰ Therefore, we should be offering genetic counseling and testing to a larger and larger population of patients: patients with MDS aged <40 years; patients with MDS of any age going to related HCT; and, now based on this study, individuals with unexplained cytopenias and hypocellular marrows. These investigators had limited ability to predict those with germ line predisposition based on demographic data, clinical presentation, or family history, demonstrating that it is not obvious a priori who will have germ line risk. Thus, one wonders if/when we will have sufficient data to justify germ line genetic testing as standard of care for all patients undergoing evaluations for persistent cytopenias, HMs, and/or allogeneic HCT.

Conflict-of-interest disclosure: The author declares no competing financial interests. ■

REFERENCES

- Molteni E, Bono E, Galli A, et al. Prevalence and clinical expression of germ line predisposition to myeloid neoplasms in adults with marrow hypocellularity. *Blood*. 2023;142(7):643-657.
- Makishima H, Bowman TV, Godley LA. DDX41-associated susceptibility to myeloid neoplasms. *Blood*. 2023;141(13):1544-1552.
- Feurstein S, Churpek JE, Walsh T, et al. Germline variants drive myelodysplastic syndrome in young adults. *Leukemia*. 2021; 35(8):2439-2444.

- Feurstein S, Trottier AM, Estrada-Merly N, et al. Germ line predisposition variants occur in myelodysplastic syndrome patients of all ages. *Blood*. 2022;140(24):2533-2548.
- McReynolds LJ, Rafati M, Wang Y, et al. Genetic testing in severe aplastic anemia is required for optimal hematopoietic cell transplant outcomes. *Blood*. 2022;140(8): 909-921.
- Nie D, Zhang J, Wang F, et al. Fanconi anemia gene-associated germline predisposition in aplastic anemia and hematologic malignancies. *Front Med*. 2022;16(3):459-466.
- Sahoo SS, Kozyra EJ, Wlodarski MW. Germline predisposition in myeloid neoplasms: Unique genetic and clinical features of *GATA2* deficiency and *SAMD9/SAMD9L* syndromes. *Best Pract Res Clin Haematol*. 2020;33(3):1011-97.
- Daly MB, Pilarski R, Yurgelun MB, et al. NCCN Guidelines Insights: Genetic/Familial High-Risk Assessment: Breast, Ovarian, and Pancreatic, Version 1.2020. *J Natl Compr Canc Netw*. 2020;18(4):380-391.
- Miller DT, Lee K, Abul-Husn NS, et al. ACMG SF v3.1 list for reporting of secondary findings in clinical exome and genome sequencing: a policy statement of the American College of Medical Genetics and Genomics (ACMG). *Genet Med*. 2022;24(7):1407-1414.
- Haverfield EV, Esplin ED, Aguilar SJ, et al. Physician-directed genetic screening to evaluate personal risk for medically actionable disorders: a large multi-center cohort study. *BMC Med*. 2021;19(1):199.

<https://doi.org/10.1182/blood.2023021165>
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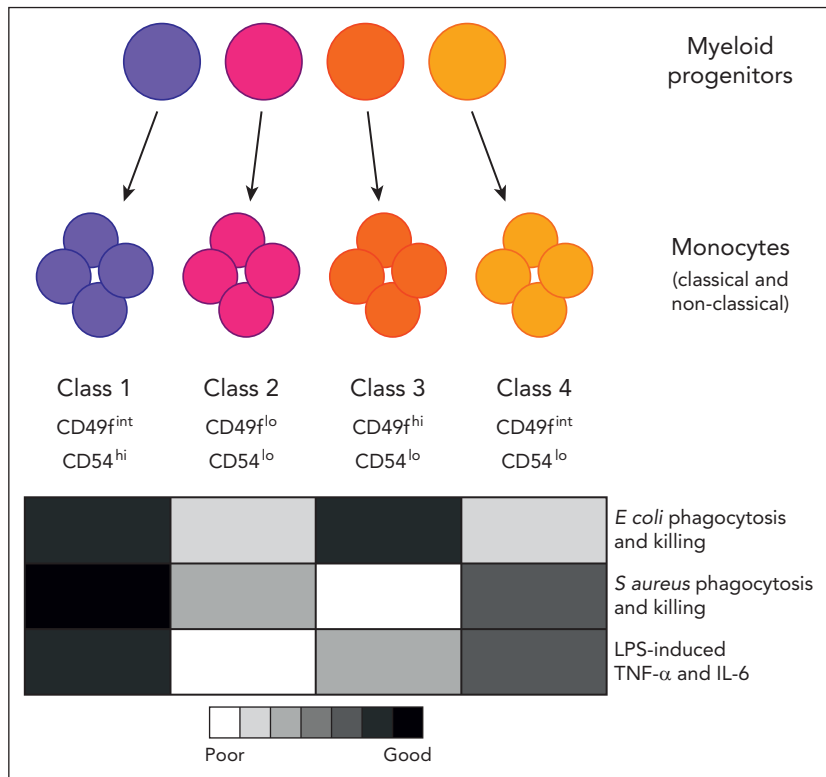
PHAGOCYTES, GRANULOCYTES, AND MYELOPOIESIS

Comment on *Rhee et al*, page 658

Progenitor diversity defines monocyte roles

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In this issue of *Blood*, Rhee et al¹ provide evidence that individual monocytes and their progeny may not be as plastic as previously thought. Instead, their data suggest that the apparent flexibility of monocytes to respond appropriately to diverse stimuli reflects selection of functionally diverse monocyte subtypes that arise from heterogeneous preprogrammed myeloid progenitors (see figure).



Four functionally specialized subtypes of monocytes arise from distinct myeloid progenitors. Epigenetic preprogramming of the monocytes is evident in the progenitors that produce them. IL-6, interleukin-6; TNF- α , tumor necrosis factor α .

Monocytes and the macrophages they produce have diverse roles and have long been described as functionally plastic, although it is unclear whether this reflects the adaptability of individual cells or a collective property of the whole population. Rhee et al have identified 4 classes of monocytes in mouse and human blood that have specialized functional properties and appear to be independently selected in response to specific challenges.

Using an ER-Hoxb8 fusion construct delivered into mouse bone marrow progenitors, the authors derived monocytes from clones of individual myeloid progenitor cells. Evaluation of the progeny of 25 clones revealed 4 monocyte subtypes with distinct gene expression signatures and functional properties. The monocyte classes differed in their relative capacity for chemotaxis, production of inflammatory mediators following lipopolysaccharide (LPS) stimulation, and phagocytosis and killing of *Escherichia coli* versus *Staphylococcus aureus*.

RNA sequencing analysis predicted differentially expressed surface markers that allowed the authors to identify the 4 monocyte classes in both mouse and human blood and to validate their functional differences in primary monocytes in vitro and in vivo. They also showed that upon infection with *E. coli* or *S. aureus*, the monocytes that were best equipped to kill the bacteria accumulated at the infection site.

In vivo differentiation experiments with barcoded progenitors provided additional evidence of the clonal origin of the monocyte subtypes, and in vitro and in vivo analyses demonstrated little or no interconversion between classes during homeostasis or under conditions of inflammatory stress. Instead, comparison of the progenitor clones and their monocyte progeny revealed that the transcriptional signatures of the monocyte subtypes were defined epigenetically in the progenitors they arose from.

Collectively, the data support a model whereby diverse monocyte responses

are achieved by selective recruitment, expansion, or activation of functionally specialized monocyte subtypes rather than by functional adaptation of more generic cells responding to different stimuli. Deeper evaluation of the responses of individual monocytes to diverse stimuli will be necessary to define the limits of their preprogramming and the extent to which they may have retained a degree of plasticity, but the insights from this study have broad implications for our understanding of monocyte and macrophage responses to pathogens and other threats.

In light of these findings, it will be important to reexamine other monocyte categorizations to determine how the 4 monocyte classes identified in the current study relate to previously defined subsets. In recent years, it has become apparent that monocyte diversity extends beyond their categorization as classical and nonclassical monocytes,^{2,4} and notably, Rhee et al detected both classical and nonclassical subsets among all 4 monocyte classes, albeit with differing relative frequencies.

Other recent studies have also suggested that monocyte heterogeneity is, at least in part, defined during their differentiation.²⁻⁷ For instance, granulocyte-monocyte progenitors (GMPs) and monocyte-dendritic cell (DC) progenitors (MDPs) independently produce classical monocyte subsets with neutrophil-like and DC-like gene expression signatures, respectively, and monocyte-derived DCs arise from MDPs but not GMPs.⁷ It will be interesting to determine whether the 4 monocyte subtypes identified in the current study arise via previously defined progenitor pathways (eg, GMP versus MDP) or via as yet unrecognized routes.

The study also casts myeloid progenitors as key players in shaping innate immune responses, which raises the possibility that the balance of myeloid progenitor subsets may contribute to the diverse responses of individuals or the dynamics of monocyte responses over a lifetime. Similarly, understanding the functional heterogeneity of monocytes and the myeloid progenitors that produce them could reveal new candidates for more precise therapeutic targeting to manipulate immune responses.

Conflict-of-interest disclosure: The author declares no competing financial interests. ■

REFERENCES

1. Rhee C, Scadden EW, Wong LP, et al. Limited plasticity of monocyte fate and function associated with epigenetic scripting at the level of progenitors. *Blood*. 2023;142(7):658-674.
2. Williams M, Mildner A, Yona S. Developmental and functional heterogeneity of monocytes. *Immunity*. 2018;49(4):595-613.
3. Ng LG, Liu Z, Kwok I, Ginhoux F. Origin and heterogeneity of tissue myeloid cells: a focus on GMP-derived monocytes and neutrophils. *Annu Rev Immunol*. 2023;41:375-404.
4. Wolf AA, Yanez A, Barman PK, Goodridge HS. The ontogeny of monocyte subsets. *Front Immunol*. 2019;10:1642.
5. Weinreb C, Rodriguez-Fraticelli A, Camargo FD, Klein AM. Lineage tracing on transcriptional landscapes links state to fate during differentiation. *Science*. 2020; 367(6479):eaaw3381.
6. Ikeda N, Kubota H, Suzuki R, et al. The early neutrophil-committed progenitors aberrantly differentiate into immunoregulatory monocytes during emergency myelopoiesis. *Cell Rep*. 2023;42(3):112165.
7. Yanez A, Coetzee SG, Olsson A, et al. Granulocyte-monocyte progenitors and monocyte-dendritic cell progenitors independently produce functionally distinct monocytes. *Immunity*. 2017;47(5):890-902.e4.

<https://doi.org/10.1182/blood.2023021298>

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