

Considering the rarity of NLPHL, the size of the cohort is respectable, giving enough power for the necessary statistical analyses. Owing to the fact that patients with NLPHL have 100% overall survival and similar PFS as patients with classic Hodgkin lymphoma, except a tendency to poorer PFS for early-stage patients who were not irradiated, the authors draw the conclusion that classic Hodgkin lymphoma therapy is a suitable option for patients with early-stage NLPHL. The necessity of radiotherapy for early stages, irrespective of interim PET result, is noted and mentioned in the conclusion. There also was an encouraging absence of histological transformation.

The findings in the article are novel, describing modern therapy not previously evaluated for NLPHL. However, there are caveats, the most important being that NLPHL is already treated as an indolent B-cell lymphoma in many countries with a rituximab- and radiotherapy-based approach, using primary chemotherapy only for a limited number of patients. This kind of approach is significantly less toxic than treating all with chemotherapy. Further, as the authors point out in the conclusion, the optimal chemotherapy approach still needs to be defined. ABVD (doxorubicin, bleomycin, vinblastine, and dacarbazine) might be the most proven owing to tradition, but biologically there is no rationale for using Hodgkin lymphoma-directed chemotherapy instead of chemotherapy with better results for indolent B-cell lymphomas.

In conclusion, the results of this study give important further evidence for primary therapy of early-stage NLPHL and advance the field. At the same time, the international trend is to treat more like indolent B-cell lymphoma; however, there is a lack of solid evidence of its superiority. There is certainly a need for a randomized trial specifically for NLPHL, something that would take a substantial international effort to achieve. Also, standard and experimental indolent B-cell lymphoma-directed approaches beyond single rituximab and radiotherapy, such as R-bendamustine, R2, BTK- and PI3K-inhibitors, bispecific antibodies, and chimeric antigen receptor (CAR) T cells should ideally be tested on relapsing NLPHL. This could be achieved by including patients with NLPHL

in relapsed/refractory indolent B-cell lymphoma trials.

*You are Mr. Owl. I am Ms. Hummingbird.
We maybe came from different
species but as long as you're a
bird, I'm a bird too.*
—GLAD MUNAISECHE

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REFERENCES

1. Eichenauer DA, Bühnen I, Baues C, et al. Interim PET-guided treatment for early-stage NLPHL: a subgroup analysis of the randomized GHSG HD16 and HD17 studies. *Blood*. 2023;142(6):553-560.
2. Campo E, Jaffe ES, Cook JR, et al. The International Consensus Classification of Mature Lymphoid Neoplasms: a report from the Clinical Advisory Committee. *Blood*. 2022;140(11):1229-1253.
3. Alaggio R, Amador C, Anagnostopoulos I, et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Lymphoid Neoplasms. *Leukemia*. 2022;36(7):1720-1748.
4. Eichenauer DA, Plutschow A, Fuchs M, et al. Long-term follow-up of patients with nodular lymphocyte-predominant Hodgkin lymphoma treated in the HD7 to HD15 trials: a report from the German Hodgkin Study Group. *J Clin Oncol*. 2020;38(7):698-705.

5. Eichenauer DA, Plutschow A, Fuchs M, et al. Rituximab in newly diagnosed stage IA nodular lymphocyte-predominant Hodgkin lymphoma: long-term follow-up of a phase 2 study from the German Hodgkin Study Group. *Leukemia*. 2020;34(3):953-956.
6. Fanale MA, Cheah CY, Rich A, et al. Encouraging activity for R-CHOP in advanced stage nodular lymphocyte-predominant Hodgkin lymphoma. *Blood*. 2017;130(4):472-477.
7. Della Pepa R, Picardi M, Giordano C, et al. Rituximab in a risk-adapted treatment strategy gives excellent therapeutic results in nodular lymphocyte-predominant Hodgkin lymphoma. *Br J Haematol*. 2018;182(5):715-718.
8. Garciaz S, Harel S, Amorim S, Bouabdallah R, Thieblemont C, Brice P. RituximabABV(D) for patients with nodular lymphocyte predominant Hodgkin lymphoma ineligible for radiation therapy. *Br J Haematol*. 2016;175(4):735-737.
9. Molin D, Linderöth J, Wahlin BE. Nodular lymphocyte predominant Hodgkin lymphoma in Sweden between 2000 and 2014: an analysis of the Swedish Lymphoma Registry. *Br J Haematol*. 2017;177(3):449-456.
10. Eichenauer DA, Bühnen I, Plutschow A, et al. Phase II study of fixed-duration single-agent ibrutinib in relapsed nodular lymphocyte predominant Hodgkin lymphoma: a report from the German Hodgkin Study Group. *Hematol Oncol*. 2022;40(4):801-804.

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LYMPHOID NEOPLASIA

Comment on [Dreval et al](#), page 561

An AID to follicular lymphoma transformation

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In this issue of *Blood*, Dreval et al¹ attempted to unravel the basis of follicular lymphoma (FL) transformation using whole genome sequencing, comparing cases with and without histologic transformation (HT and no HT), with de novo diffuse large B-cell lymphoma (DLBCL). FL shares many common genetic events with the germinal center subtype (GCB) of DLBCL; thus, the authors reasoned that DNA from de novo DLBCL would be enriched for mutations associated with HT, and an ideal group to mine for predictive genes. To sort these subtypes, the authors employed a multistep process defining significantly mutated genes and activation-induced cytidine deaminase (AID)-dependent mutational signatures within the cohort and then searched for differences between lymphoma subtypes. In doing so, they discovered 2 genetically distinct subgroups of FL, DLCL-like FL (dFL) and constrained FL (cFL), that showed a remarkable 10-year difference in time to transformation.

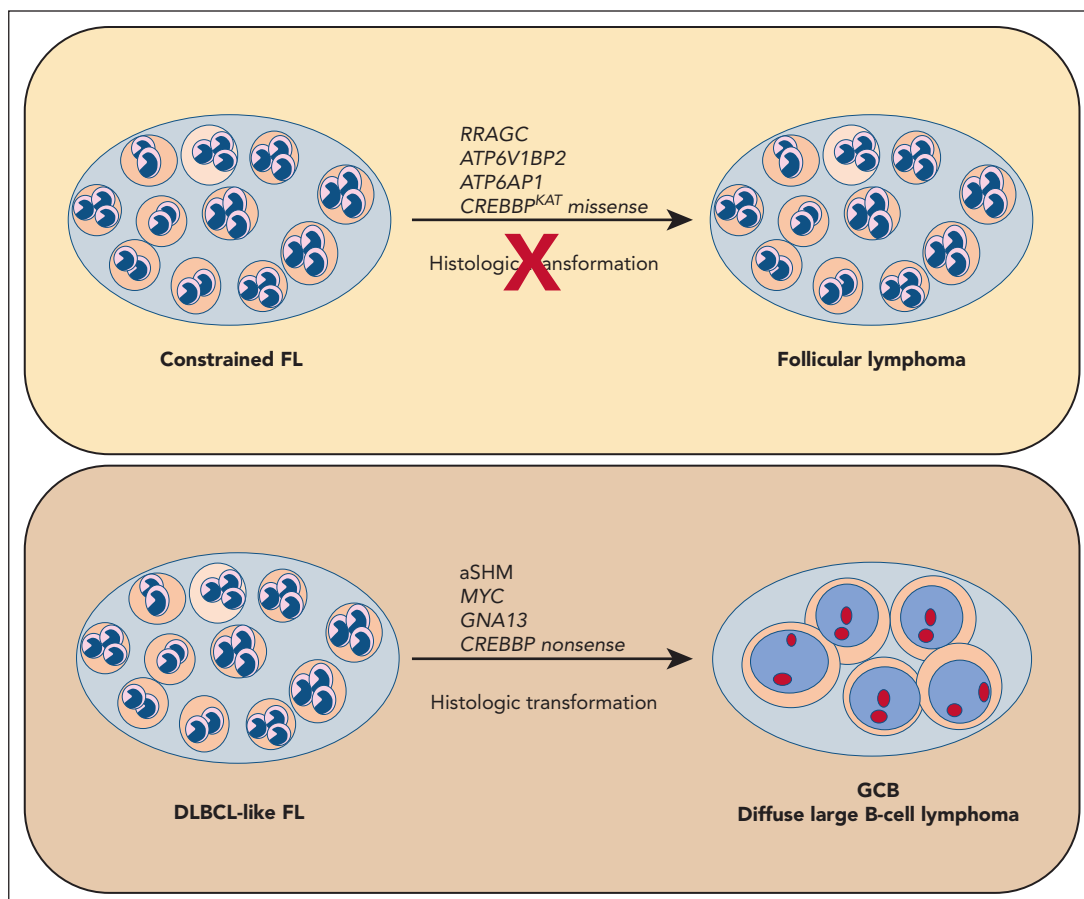
These 2 FL subtypes demonstrated differences in mutation signatures, mutated gene frequencies, gene expression, and translocations. For instance, the cFL samples showed a lower mutational burden for hotspots of aberrant somatic hypermutation (aSHM) induced by AID, greater enrichment of mutations in *RRAGC*, *ATP6V1BP2*, and *ATP6AP1*, and missense mutations in *CREBBP* (see figure). cFL cases had lower gene expression of *MYC*, and lower frequency of *MYC* translocations. Conversely, the dFL cases had a higher incidence of aSHM, as evidenced by mutations in the transcription start sites of *BCL6*, *BCL7A*, *RHOH*, and *ZFPL36L1*. Moreover, dFL shared a similar mutation pattern with DLCL, including a higher incidence of *GNA13* mutations and higher gene expression of *MYC* compared with cFL.

The overall similarity of mutation patterns between cFL, dFL, and GCB DLBCL underscores the genetic relationship

between these diseases and further supports the notion that FL represents the indolent counterpart of the *EZH2* mutant, *BCL2* translocated genetic subtype of DLBCL.² However, a notable finding was the enrichment of missense mutations in the lysine acetyltransferase (KAT) domain of *CREBBP* within the cFL cases vs nonsense and frameshift mutations in the dFL and DLBCL cases. The authors postulate that the missense mutations in *CREBBP* (and higher gene expression) in cFL are incompatible with HT, and possibly limit other mechanisms of transformation, including aSHM and higher *MYC* activity. Previous studies using mouse models of FL/DLCL noted similar findings, suggesting that higher *MYC* expression and more aggressive disease were associated with complete inactivation of *Crebbp*.³ The authors were able to demonstrate that the presence or absence of *CREBBP* KAT domain missense mutations alone was able to maintain the significant association with

time to transformation in 2 separate clinical cohorts.^{4,5} Thus, *CREBBP* missense mutations may reflect the hallmark genetic feature of truly indolent FL and could be studied as a prospective biomarker for clinical decisions for treatment vs watch and wait.

Other tools to predict poor prognosis FL have been developed, including the FL international prognostic index (FL-IPI),⁶ the m7-FLIPI⁷ (a clinical and genetic multivariate risk model), and progression of disease within 2 years (POD24).⁸ How does the current study of genetic subtypes compare? A direct comparison is difficult because the above risk models predict progression-free survival, whereas the new genetic subtypes predict time to transformation. Although no difference in FL-IPI scores or POD24 status was observed between cFL and dFL, Dreval et al demonstrate that cFL/dFL designation is an independent risk model and complements both the FL-IPI and POD24



Whole genome sequencing identified 2 subtypes of FL with differing risks of histologic transformation. Aberrant somatic hypermutation (aSHM) induced by AID was associated with increased risk of transformation and increased number of mutations shared with GCB DLCL. This subset was named DLBCL-like FL (dFL). Conversely, the cohort with lower risk of transformation showed less aSHM and fewer overall mutations, and enrichment for missense mutations of *CREBBP*, *ATP6V1BP2*, *ATP6AP1*, and *RRAGC*. This subset was named constrained FL (cFL).

in identifying high-risk patients. Interestingly, the dFL subtype showed a near 3-fold enrichment of grade 3A FL cases. This finding suggests value in the traditional histologic grading system based on centroblasts per high-power field and supports a potential difference in the mutation patterns among FL grades.

Other large-scale sequencing studies in FL have also recently been published.⁵ Crouch et al used a panel of nearly 300 genes and reported 3 genetic subtypes that differed significantly with regard to prevalence of aSHM, but the subtypes did not correlate to FLIPI or histologic transformation. However, differences in aSHM were observed between subgroups and CREBBP mutations clustered with STAT6 and TNFRSF14 mutations, both of which trended toward significance in the cFL subtype reported by Dreval et al. Crouch et al conclude that the genetic subtypes they found had limited prognostic clinical significance. How then do we reconcile these different conclusions? First, the approaches were different. Crouch et al used panel deep sequencing (~500×) of formalin-fixed, paraffin-embedded FL biopsies, whereas Dreval et al used whole genome sequencing (~50×) with paired normal from newly sequenced and historic cohorts of FL and DLBCL. Thus, the patient populations, controls, and genomic features detected for the 2 models were different. Second, of the overlapping genetic features, CREBBP mutations were all treated similarly by Crouch et al, whereas Dreval et al distinguished missense mutations within the KAT domain from other mutations. This small but important distinction appears to be a critical nuance in predicting transformation of FL. The distinction of domain-specific genetic features is also critical for predicting genetic subtypes of DLBCL.² However, like the FL predictor, the full complement of genetic features is required to achieve maximum sensitivity, specificity, and accuracy. Future studies with large replication cohorts will be needed to establish the minimum set of genetic alterations required to appropriately predict genetic subtypes of lymphoma.

As machine learning approaches continue to gain traction and offer new insights into the heterogeneity of FL, it will be critical to thoroughly annotate the research cohorts being studied. Roughly 20% of transformed FL cases are of the

activated B-cell (ABC) type by gene expression and have a lower incidence of BCL2 rearrangement, compared with GCB cases.⁴ Prior studies also have identified subsets of FL that more closely resemble ABC-DLBCL, with high expression of interferon regulatory factor 4 (IRF4) and low expression of CD10; high expression of IRF4 is a risk for early transformation.^{4,9} Notably, such cases are generally negative for the BCL2 translocation and should be distinguished from conventional FL. Recognizing that FL is >1 disease, we must ensure that future models take this heterogeneity into consideration.¹⁰

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REFERENCES

- Dreval K, Hilton LK, Cruz M, et al. Genetic subdivisions of follicular lymphoma defined by distinct coding and noncoding mutation patterns. *Blood*. 2023;142(6):561-573.
- Wright GW, Huang DW, Phelan JD, et al. A probabilistic classification tool for genetic subtypes of diffuse large B cell lymphoma with therapeutic implications. *Cancer Cell*. 2020;37(4):551-568.e14.
- Garcia-Ramirez I, Tadros S, Gonzalez-Herrero I, et al. Crebbp loss cooperates with Bcl2 overexpression to promote lymphoma in mice. *Blood*. 2017;129(19):2645-2656.
- Kridel R, Mottok A, Farinha P, et al. Cell of origin of transformed follicular lymphoma. *Blood*. 2015;126(18):2118-2127.
- Crouch S, Painter D, Barrans SL, et al. Molecular subclusters of follicular lymphoma: a report from the United Kingdom's Haematological Malignancy Research Network. *Blood Adv*. 2022;6(21):5716-5731.
- Solal-Celigny P, Roy P, Colombat P, et al. Follicular lymphoma international prognostic index. *Blood*. 2004;104(5):1258-1265.
- Pastore A, Jurinovic V, Kridel R, et al. Integration of gene mutations in risk prognostication for patients receiving first-line immunochemotherapy for follicular lymphoma: a retrospective analysis of a prospective clinical trial and validation in a population-based registry. *Lancet Oncol*. 2015;16(9):1111-1122.
- Casulo C, Byrtek M, Dawson KL, et al. Early relapse of follicular lymphoma after rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone defines patients at high risk for death: an analysis from the National LymphoCare Study. *J Clin Oncol*. 2015;33(23):2516-2522.
- Karube K, Guo Y, Suzumiya J, et al. CD10-MUM1+ follicular lymphoma lacks BCL2 gene translocation and shows characteristic biologic and clinical features. *Blood*. 2007;109(7):3076-3079.
- Campo E, Jaffe ES, Cook JR, et al. The international consensus classification of mature lymphoid neoplasms: a report from the Clinical Advisory Committee. *Blood*. 2022;140(11):1229-1253.

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MYELOID NEOPLASIA

Comment on [Qiu et al](#), page 574

Food for thought (and CML survival)

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In this issue of *Blood*, [Qiu et al](#)¹ shed light on the metabolic changes that arise in a population of chronic myelogenous leukemia (CML) stem cells in a mouse model that allow them to better adapt to continued tyrosine kinase inhibitor (TKI) treatment through HIF1A activity.

CML is a myeloproliferative disease, characterized by leukocytosis and uncontrolled proliferation of granulocytes. The hallmark of CML is the Philadelphia chromosome, in which a reciprocal translocation of t(9;22) results in the BCR-ABL fusion gene and constitutively active tyrosine kinase activity of the resulting oncoprotein. The incorporation of small-molecule

TKIs as the first-line treatment for CML has revolutionized prognosis for patients, elevating median survival from a dismal 3 to 5 years from initial diagnosis to that of the general population.² Even so, cases of TKI monotherapies unable to completely eradicate CML persist; imatinib reports molecular relapse in 30% to 60% of patients, and 25% of patients have TKI intolerance.^{3,4}