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current approach in management of SCA, sickle cell disease-associated cardiomyopathy.

 risk factor evaluation in patients.
Gupta A, Fei YD, Kim TY, et al. IL-18 mediates sickle cell cardiomyopathy and ventricular arrhythmias. Blood. 2021;137(9): 1208-1218.

> Bakeer N, James J, Roy S, et al. Sickle cell anemia mice develop a unique cardiomyopathy with restrictive physiology. *Proc Natl Acad Sci U S A*. 2016;113(35): E5182-E5191.

Circ Cardiovasc Imaging. 2014;7(3):430-437.

- Indik JH, Nair V, Rafikov R, et al. Associations of prolonged QTc in sickle cell disease. *PLoS* One. 2016;11(10):e0164526.
- Niss O, Fleck R, Makue F, et al. Association between diffuse myocardial fibrosis and diastolic dysfunction in sickle cell anemia. *Blood*. 2017;130(2):205-213.
- Maron BJ, Rowin EJ, Maron MS. Evolution of risk stratification and sudden death prevention in hypertrophic cardiomyopathy: twenty years with the implantable cardioverter-defibrillator. *Heart Rhythm*. 2021;18(6):1012-1023.

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suggesting the need for careful cardiac risk factor evaluation in patients.

1. d'Humières T, Saba J, Savale L, et al.

Determinants of ventricular arrhythmias in

 Nze C, Fortin B, Freedman R, et al. Sudden death in sickle cell disease: current experience.

Br J Haematol. 2020;188(4):e43-e45.

3. Manci EA, Culberson DE, Yang YM, et al.

4. Gladwin MT, Sachdev V. Cardiovascular

abnormalities in sickle cell disease.

J Am Coll Cardiol. 2012;59(13):

5. Desai AA, Patel AR, Ahmad H, et al. Mechanistic insights and characterization of

Causes of death in sickle cell disease: an

autopsy study. Br J Haematol. 2003;123(2):

sickle cell anemia: toward better prevention

of sudden cardiac death. Blood. 2023;142(5):

REFERENCES

409-420

359-365.

1123-1133.

Evolving beyond morphology in pediatric PTLD

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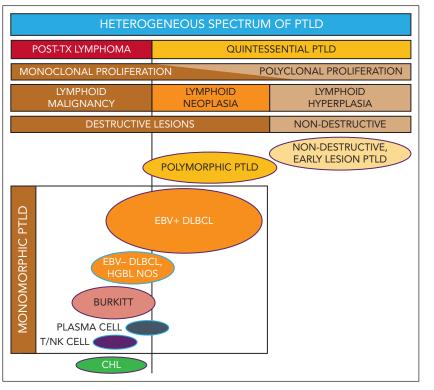
In this issue of *Blood*, Salmerón-Villalobos et al have elucidated the molecular code of monomorphic posttransplant lymphoproliferative disorders (PTLDs) in the pediatric population.¹ Through an integrated molecular approach including fluorescence in situ hybridization, copy number arrays, and targeted gene sequencing, the study establishes the genetic landscape of monomorphic PTLD with diffuse large B-cell lymphoma (DLBCL) and Burkitt lymphoma (BL) histology. By comparing pediatric cases of PTLD-DLBCL and PTLD-BL to their counterparts occurring as de novo lymphomas in immuno-competent children and PTLD-DLBCL in adults, this study seeks to better understand the translational biology of pediatric monomorphic B-cell PTLD. It addresses a critical gap in knowledge that may enhance our understanding of the classification of PTLD and expected patterns of treatment response.

Children with advanced-stage, monomorphic PTLD with DLBCL histology can present with clinically aggressive disease including multifocal lymphoid masses, diffuse extranodal involvement, and an alarming disease burden on imaging studies. Selection of the optimal treatment regimen for these patients, particularly those with stage III disease, remains a formidable challenge. Some patients like this are cured with gentle therapeutic approaches, such as reduction of immune suppression, rituximab monotherapy, or low-dose chemoimmunotherapy (as reaffirmed within this cohort). Others, though, resemble clinical scenarios akin to de novo pediatric mature B-cell lymphomas and require intensive multiagent chemoimmunotherapy. Such polarizing heterogeneity in pediatric monomorphic PTLD-DLBCL renders it a high-stakes clinical dilemma, especially considering the potential life-threatening and transplant-threatening complications of intensive chemotherapy.

PTLD includes a broad spectrum of lymphoproliferation (see figure) ranging from nondestructive, early lesion PTLD (characterized as polyclonal, reactive B-cell hyperplasia) to polymorphic PTLD (polymorphic, often monoclonal, neoplastic, destructive lesions) to monomorphic and classical Hodgkin lymphoma PTLD (demonstrating transformation to or toward malignant lymphoma).² It is challenging to unify such a heterogeneous category of disease processes that are characterized by drastically different clinical patterns, therapeutic approaches, and survival outcomes.

Nondestructive and polymorphic PTLDs are prototypical Epstein-Barr virus (EBV)driven PTLDs. Such quintessential PTLD in children is conceptually built on a framework in which acquired immune suppression in patients that are often EBV immune-naive creates susceptibility to varying degrees of EBV-driven lymphoproliferation.³ For such patients, restoration of the immune response and EBV-directed therapeutic strategies are often curative.

Monomorphic PTLD is a problematic category because it encompasses an expansive array of posttransplant lymphoid neoplasia that does not neatly fit this conceptual framework. Some patients with EBV⁺ PTLD-DLBCL have a favorable response to less intensive therapy such as rituximab and thus belong in the spectrum of quintessential PTLD. In contrast, others have relapsing or refractory disease after standard or even novel EBV-directed or low-dose chemoimmunotherapy strategies. For these latter patients, EBV may represent only one of multiple pathogenic factors driving lymphomagenesis. Such patients, plus many with EBV-negative PTLD-DLBCL, are similar to patients with PTLD-BL-they typically require aggressive multiagent chemotherapy for curative



Reconceptualizing the framework for categorizing the heterogeneous spectrum of lymphoproliferation in pediatric PTLD. Traditional PTLD classification is based on morphological features stratified as nondestructive, polymorphic, monomorphic, and classical Hodgkin lymphoma (CHL) PTLD. This revised stratification highlights the heterogeneity of monomorphic PTLD and demarcates a threshold beyond which quintessential EBV-driven lymphoid hyperplasia or neoplasia transforms into malignant lymphoma. EBV⁺ DLBCL in particular straddles the threshold between quintessential PTLD and posttransplant lymphoma. Ovals with purple borders indicate entities that are typically EBV⁺, and ovals with blue borders indicate entities that are often EBV⁻. HGBL NOS, high-grade B-cell lymphoma, not otherwise specified and other gray-zone, Burkitt-like mature B-cell lymphomas; PLASMA CELL, plasma cell neoplasm; T/NK CELL, T/NK-cell lymphoma; TX, transplant.

outcomes.^{4,5} Additionally, other rare monomorphic entities, such as PTLD with any mature T-cell or natural killer (NK)-cell lymphoma histology or classical Hodgkin lymphoma PTLD, also require treatment with disease-specific chemotherapy regimens typically given to patients who are immunocompetent.

Successive generations of international clinical trials for pediatric PTLD have repeatedly demonstrated an event-free survival of approximately 70% after treatment with low-dose chemotherapy with or without rituximab.⁶⁻⁸ Data from the Berlin-Frankfurt-Münster group demonstrated that 53% of patients could achieve durable complete remission with rituximab alone.⁹ Throughout these studies, polymorphic PTLD and monomorphic DLBCL have shown similar outcomes, and the only prognostic factors identified were the presence of stage IV disease and BL histology.^{7,8} Despite the efficacy of rituximab in a significant subset of patients, and despite novel development of EBV-specific cytotoxic T-lymphocyte immunotherapy, eventfree survival for pediatric PTLD over the past 20 years has plateaued at ~70% after treatment with milder, rituximab or low-dose chemoimmunotherapy-based approaches.¹⁰ Identification of patients that require intensive multiagent chemoimmunotherapy has remained elusive with neither traditional PTLD morphological or histological classification or clinical criteria reliably identifying this high-risk subset.

This study examined monomorphic PTLD to try to identify which patients can be successfully treated with low-intensity immunotherapy-based interventions and which patients require a full-intensity chemoimmunotherapy approach. Specifically, it demonstrated that pediatric PTLD-DLBCL is genetically less complex and biologically distinct—from both disease in adults as well as de novo pediatric DLBCL—and that somatic alterations in genes involved in cell cycle and Notch pathways were associated with worse outcomes.¹ In contrast, they also showed that the mutational profile of PTLD-BL shares biological overlap with de novo EBV-positive BL, thus providing translational evidence supporting the widely accepted practice of treating PTLD-BL with the intensive chemotherapy regimens used for patients that are immunocompetent.^{5,8} BL, after all, is characterized by its principal molecular feature—C-MYC rearrangement. Results from this study offer evidence that a molecular classification system for pediatric PTLD may have potential to prospectively provide more precise risk stratification and, ideally, tailor therapy.

PTLD classification systems built upon morphological features are generally effective but have some important shortcomings, specifically limitations that pertain to monomorphic DLBCL. These data suggest the need to expand the PTLD classification to integrate molecular characteristics with traditional morphology. Such a new classification should consider distinguishing 2 overarching categories-one being the classic spectrum of quintessential PTLD including nondestructive, polymorphic, and monomorphic EBV⁺ PTLD-DLBCL, and the second being a heterogenous group categorized as posttransplant lymphomas considered analogous to those occurring in the setting of other underlying (inherited or acquired) immunodeficiency syndromes (see figure). Within this framework, the biologically and clinically heterogeneous entity of PTLD-DLBCL would straddle both categories. Next-generation targeted sequencing studies identified worse outcomes for PTLD-DLBCL in association with somatic mutations in cell cycle and Notch molecular pathways, which suggests a role for molecular diagnostics to categorize individual cases with greater precision. Future studies are needed to prospectively evaluate molecular biomarkers in larger cohorts for clinical validation. Ideally, a risk-stratification system for pediatric PTLD would reliably identify those patients that may be effectively treated with milder, EBV-directed or rituximabbased therapies, thus sparing them the severe toxicities of intensive multiagent chemotherapy. It should simultaneously be capable of identifying the high-risk patients for whom the risks of intensive therapy are justified by significantly improved chances for cure.

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REFERENCES

- Salmerón-Villalobos J, Castrejón-de-Anta N, Guerra-García P, et al. Decoding the molecular heterogeneity of pediatric monomorphic post-solid organ transplant lymphoproliferative disorders. *Blood*. 2023; 142(5):434-445.
- Natkunam Y, Gratzinger D, Chadburn A, et al. Immunodeficiency-associated lymphoproliferative disorders: time for reappraisal? *Blood*. 2018;132(18):1871-1878.
- Baker A, Frauca Remacha E, Torres Canizales J, et al. Current practices on diagnosis, prevention and treatment of posttransplant lymphoproliferative disorder in pediatric patients after solid organ transplantation: results of ERN TransplantChild Healthcare Working Group Survey. Children. 2021;8(8):661.
- Afify ZAM, Taj MM, Orjuela-Grimm M, et al. Multicenter study of pediatric Epstein-Barr virus-negative monomorphic post solid organ transplant lymphoproliferative disorders. *Cancer.* 2023;129(5):780-789.
- Afify Z, Orjuela-Grimm M, Smith CM, et al. Burkitt lymphoma after solid-organ transplant: treatment and outcomes in the paediatric PTLD collaborative. *Br J Haematol.* 2023;200(3):297-305.
- 6. Gross TG, Bucuvalas JC, Park JR, et al. Low-dose chemotherapy for Epstein-Barr

LYMPHOID NEOPLASIA

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virus-positive posttransplantation lymphoproliferative disease in children after solid organ transplantation. *J Clin Oncol.* 2005;23(27):6481-6488.

- Gross TG, Orjuela MA, Perkins SL, et al. Lowdose chemotherapy and rituximab for posttransplant lymphoproliferative disease (PTLD): a Children's Oncology Group Report. *Am J Transplant*. 2012;12(11):3069-3075.
- Maecker B, Jack T, Zimmermann M, et al. CNS or bone marrow involvement as risk factors for poor survival in posttransplantation lymphoproliferative disorders in children after solid organ transplantation. *J Clin Oncol.* 2007;25(31):4902-4908.
- Maecker-Kolhoff B, Beier R, Zimmermann M, et al. Response-adapted sequential immunochemotherapy of post-transplant lymphoproliferative disorders in pediatric solid organ transplant recipients: results from the prospective Ped-PTLD 2005 trial. Br J Haematol. 2015;171(suppl 1):34.
- Rubinstein J, Toner K, Gross T, Wistinghausen B. Diagnosis and management of post-transplant lymphoproliferative disease following solid organ transplantation in children, adolescents, and young adults. Best Pract Res Clin Haematol. 2023;36(1):101446.

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Karyotype and outcome in CLL

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In this issue of *Blood*, Fürstenau et al¹ describe the spectrum of conventional karyotypes and their prognostic impact in chronic lymphocytic leukemia (CLL) from analysis of 895 patients recruited to the GAIA/CLL13 study.

The importance of karyotype in CLL was first recognized by Catovsky,² Juliusson,³ and others, in the 1980s, when the ability to generate metaphases for chromosome banding analysis was much less efficient. Fluorescence in situ hybridization (FISH) in CLL, published by Döhner et al⁴ in 2000, described the impact of 4 genomic abnormalities, 13q-, +12, 11q-, and 17p-, and had a major influence lasting >20 years. FISH has been the only chromosome evaluation used in most clinical trials, and it is the only technique recommended in many CLL guidelines, including the International Workshop on CLL (iwCLL).⁵ Although these 4 gene targets are both common and important, it became clear with improvements in standard karyotyping (96.7% success in this article¹ using CpG + interleukin-2) as well as the advent of genomic microarrays ("molecular karyotyping"),⁶ that there is a wide range of chromosomal aberrations that occur commonly in CLL; and for many of these, we know little of their biological or prognostic relevance. Recent interest has focused on "complex karyotype" (CKT) with simple addition of the number of aberrations. Initially, \geq 3 aberrations were used as a definition, but more recently, Baliakis et al⁷ showed \geq 5 aberrations have much more significance. There has been little focus on the individual aberrations and to what extent each contributed to the prognosis. Many patients with "CKT" have a 17p deletion as one of these aberrations, and it has been difficult to discriminate a "CKT effect" from that of 17p deletion. This study excludes 17p deletion and TP53 mutation and clearly shows an impact of CKT separated from TP53 dysfunction. As with virtually all clinical trials, only patients with progressive CLL who required treatment were recruited.

The GAIA/CLL13 clinical trial compared chemoimmunotherapy (CIT) (2 regimens: fludarabine, cyclophosphamide, and rituximab [FCR] for patients ≤65 years old and bendamustine, and rituximab [BR] for >65 years old) with 3 venetoclax regimens: 2 "doublets" of venetoclax with antibody, rituximab or obinutuzumab (RV or GV, respectively), and the "triplet" obinutuzumab, ibrutinib, and venetoclax (GIV). Therapy duration differed: CIT for 6 months; RV and GV for 12 months; and for GIV, ibrutinib monotherapy was continued to 36 months in those who did not achieve undetectable minimal residual disease (uMRD). In this study, the median observation time was 38.8 months.

Fürstenau et al initially focus on CKT status using a stratification from "non-complex" (nCKT) with \leq 2 aberrations to "complex" with ≥ 3 aberrations. The latter is subdivided into "intermediate CKT" (iCKT) with 3 to 4 aberrations and "high CKT" (hCKT) with ≥ 5 aberrations (see figure). hCKT proportions were low in all treatment arms, although slightly higher in CIT, with 7.2%, 4.3%, 5.0%, and 2.7% in CIT, RV, GV, and GIV, respectively. CKT did not influence overall response rate, whereas uMRD rates were lower with CKT when using CIT, but not with venetoclax regimens. Progression-free survival (PFS) with CIT was adverse with both iCKT and hCKT, whereas in the pooled venetoclax arms, only hCKT had worse outcome. For both CIT and venetoclax, this only occurred for those with unmutated immunoglobulin heavy chain variable region (IGHV) genes (unmutated CLL [U-CLL]); in mutated IGHV (mutated CLL [M-CLL]), CKT had no statistical effect.

A small group of ~2.3% formally with CKT have an outstandingly good outcome: patients with "triple trisomy" (often +12, +18, and +19, or more precisely +12 with +18 and/or +19; and all M-CLL). This