

Integrating novel agents into the treatment of advanced mycosis fungoides and Sézary syndrome

Michael S. Khodadoust,^{1,2} Eric Mou,³ and Youn H. Kim^{1,2}

¹Division of Oncology, Stanford University, Stanford, CA; ²Department of Dermatology, Stanford University, Stanford, CA; ³Division of Hematology, Oncology, and Blood & Marrow Transplantation, University of Iowa, Iowa City, IA

Agents targeting the unique biology of mycosis fungoides and Sézary syndrome are quickly being incorporated into clinical management. With these new therapies, we are now capable of inducing more durable responses and even complete remissions in advanced disease, outcomes which were exceedingly rare with prior therapies. Yet, even this new generation of therapies typically produce objective responses in only a minority of patients. As our therapeutic options

increase, we are now challenged with selecting treatments from a growing list of options. To gain the full benefit of these novel agents, we must develop strategies to match treatments for the patients most likely to benefit from them. Here, we consider both the current approaches to treatment selection based on clinical features and the future of molecular biomarker-guided therapy for patients with this heterogeneous disease.

Introduction

Mycosis fungoides (MF) and Sézary syndrome (SS) are the most common subtypes of cutaneous T-cell lymphoma (CTCL). Although the skin is the primary site of involvement, malignant T cells may expand in lymph node, visceral, and blood compartments. Most patients with MF or SS have a chronically relapsing disease course requiring multiple lines of therapy because individual treatments seldom result in lasting remission. Therefore, the goals of therapy include not just overall disease control but also symptom palliation and mitigation of treatment-related toxicities.

Traditionally, clinical stage has been a primary consideration in treatment selection. In early-stage MF, improved quality of life can be achieved with skin-directed therapies or milder systemic agents with less therapy-related morbidity. In contrast, patients with advanced-stage MF or SS are faced with a comparatively worse prognosis, necessitating consideration for more intensive systemic therapies that risk increased toxicity. For patients with relapsed or refractory advanced disease, allogeneic stem cell transplantation (allo-SCT) is potentially curative, though the significant risk of transplant-related morbidity and mortality warrants individualized consideration of its timing of and selection for use.

Historically, most systemic therapies utilized in CTCL were originally developed for other malignancies and subsequently adopted based on limited data. More recently, however, a wave of novel therapeutic approaches has emerged, built upon rationally selected targets for MF and SS. Although these novel agents represent a major advancement, they pose new

challenges in matching patients to the therapies from which they are most likely to benefit. Here, we discuss the landscape of both established and novel therapeutics for patients with advanced-stage MF and SS requiring systemic therapy, focusing on the factors that currently guide treatment prioritization, as well as emerging strategies directed at further honing therapeutic precision amidst disease heterogeneity.

Traditional treatment options

Although treatment patterns are highly heterogeneous, in patients with advanced-stage MF and SS, systemic therapies are often considered. Numerous treatments of varying modalities and intensities have been explored, beginning with cytotoxic chemotherapy, extracorporeal photopheresis (ECP), interferon, methotrexate, bexarotene, and vorinostat. More recently, trials establishing the efficacy of romidepsin and pralatrexate heralded a shift toward exploring increasingly targeted, single-agent infusional drugs in CTCL, representing a bridge between historical and novel eras.^{1,2}

As biologic and novel therapies have emerged over the past decade, accumulating evidence suggests that traditional cytotoxic chemotherapy is associated with comparatively greater risk and limited benefit in the routine treatment of MF and SS.^{3,4} A large international study of CTCL treatment patterns found the use of single-agent or combination chemotherapy to be independently associated with increased mortality.⁵ Thus, for patients requiring systemic therapy, we prioritize the use of nonchemotherapeutic drugs, reserving chemotherapy for later lines of treatment or specific situations detailed below. The true efficacies of traditional therapies are often difficult to discern given that data supporting their use were derived primarily from

phase 2 studies. Subsequent phase 3 trials comparing historical systemic therapies to novel agents have helped contextualize their efficacy and safety profiles.

Novel systemic therapies

Brentuximab vedotin (BV), an anti-CD30 monoclonal antibody conjugated to the microtubule toxin monomethyl auristatin E, was approved for the treatment of CTCL on the basis of the randomized phase 3 ALCANZA trial.⁶ The study included 131 patients with CD30-positive (defined as $\geq 10\%$ expression) MF having received at least 1 prior systemic therapy and compared BV with physician's choice of methotrexate or bexarotene. Patients with SS were excluded. In the MF cohort, the primary end point of overall response rate sustained for at least 4 months (ORR4) favored BV as compared with the control arm (50% vs 10%). Updated trial analysis showed that BV also resulted in longer time to next treatment (median, 13.4 vs 5.6 months) and progression-free survival (PFS) (median, 16.1 vs 3.5 months).⁷ Peripheral neuropathy was more common in the BV arm, affecting 66% of patients overall (9% grade 3), and led to permanent discontinuation in 14% of cases.

Mogamulizumab, a defucosylated humanized IgG1 κ monoclonal antibody targeting chemokine receptor 4 (CCR4), was approved for the treatment of CTCL on the basis of the randomized phase 3 MAVORIC trial.⁸ The study included 372 patients with stage IB-IVB MF/SS having received at least 1 prior systemic therapy and compared mogamulizumab with vorinostat. Patients with large cell transformation (LCT) were excluded. The primary end point of PFS favored mogamulizumab (median, 7.7 vs 3.1 months), as did overall response rate (ORR, 28% vs 5%). The most common adverse events were infusion-related reactions, diarrhea, fatigue, and drug-associated rash. The latter occurred in 24% of patients and led to treatment discontinuation in 7% of cases.

The ALCANZA and MAVORIC trials led to the Food and Drug Administration approval of both BV and mogamulizumab in patients with CTCL having received at least 1 prior systemic therapy. The approval of these drugs as second-line or subsequent treatments raises important questions regarding frontline therapy in advanced MF and SS. Should BV and mogamulizumab be uniformly relegated for use behind the same traditional therapies against which they have demonstrated superiority? Are there scenarios in which they might be appropriate for use upfront? Putting the available data into practical context, we believe that the frontline use of either BV or mogamulizumab may be preferred in selected patients. Specifically, these agents should be considered as initial treatment in patients with advanced disease in need of rapid clinical response and whose clinical phenotype would be expected to respond especially favorably, discussed in further detail below.

Clinical factors guiding prioritization of systemic therapy in advanced MF and SS

Although many patients with advanced MF or SS initially respond to a given therapy, durations of response are generally limited. Thus, treatment selection is a repeatedly revisited task

which requires an individualized approach, accounting for disease-related factors, notable drug toxicities, and whether future allo-SCT is planned.

Compartmental burden of disease

Because systemic treatments often exhibit differential efficacy across skin, blood, and lymph nodes, 1 useful framework to guide therapeutic prioritization is through an assessment of compartmental disease burden. More recent trials investigating single-agent infusional drugs have placed increased emphasis on describing their compartment-specific activity, allowing for a more nuanced approach to initial therapy selection (Figure 1).

Patients with significant blood burden (eg, B2) may particularly benefit from romidepsin or mogamulizumab. In this context, romidepsin has shown blood compartment response rates of up to 54%.⁹ In the MAVORIC study, mogamulizumab was associated with a blood ORR of 68%, a median time to response of 1.1 months, and median response duration of 26 months.⁸ Furthermore, a posthoc analysis found that the subset of patients eventually achieving long-term (≥ 12 months) responses to mogamulizumab were more likely to have SS or blood involvement.¹⁰ We find that mogamulizumab is often better tolerated than romidepsin, especially in patients with advanced age or decreased performance status, and thus consider it a frontline option in this setting. Though limited by selective availability and longer time to response, ECP is also effective in patients with erythrodermic MF or SS, especially when used earlier in a patient's treatment course and in those with lower blood burden.^{11,12} In contrast, data regarding the utility of BV in this setting are limited because patients with SS were excluded from the ALCANZA study.^{13,14}

Patients with erythrodermic skin involvement frequently have accompanying blood involvement, and so their treatment follows similar principles as described above. In patients with extensive skin tumors, however, the approach differs. Both romidepsin and pralatrexate have shown significant activity in patients with tumor-stage disease, with ORRs of 45% and 67%, respectively.^{2,15} However, based on the strength of data from the ALCANZA study, in which patients with tumor-stage MF treated with BV achieved striking response rates as compared with the control arm (ORR4, 63% vs 5%), we consider BV a frontline treatment option for these patients.⁶ In contrast, mogamulizumab is less effective in this context because only 16% of patients with tumor-stage MF in the MAVORIC study achieved a response.⁸

In patients with lymph node involvement, both romidepsin and pralatrexate are active, based on analyses of both specific CTCL cohorts and the landmark trials in nodal peripheral T-cell lymphomas (PTCLs).^{9,16} Similarly, BV is effective in treating nodal disease based on data from trials of both CTCL and nodal PTCLs.^{13,17,18} In the ALCANZA study, patients with extracutaneous involvement achieved an ORR4 of 46% vs 9% in the control arm.⁶ Thus, we view BV, romidepsin, and pralatrexate all as useful treatment options in these patients. In contrast, mogamulizumab is less effective, with only 17% of patients enrolled in the MAVORIC trial achieving a nodal response.⁸

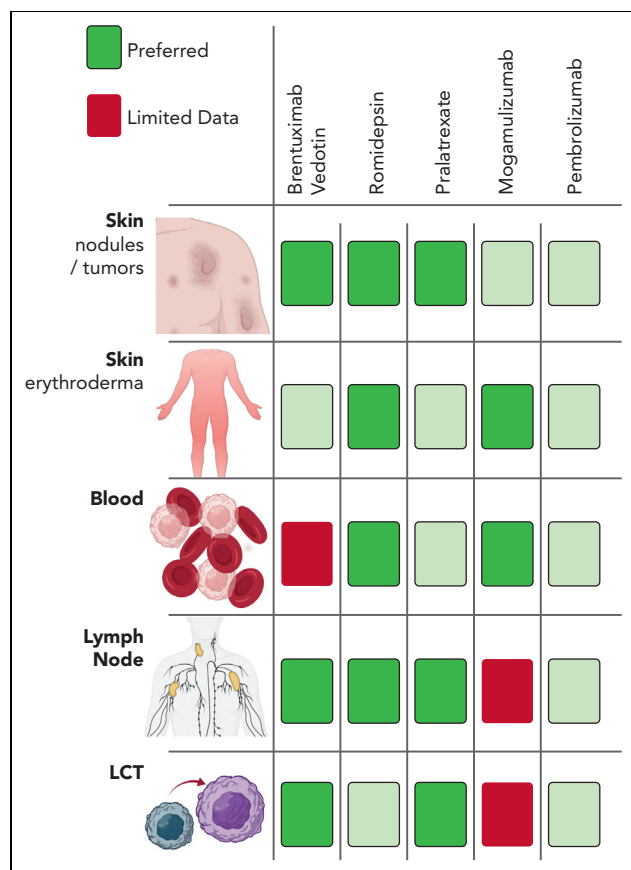


Figure 1. Disease compartment driven drug selection. Relative activity of single-agent infusional therapies in various disease compartments in cutaneous T-cell lymphomas.

In patients with extensive nodal or visceral involvement, the historically high ORRs associated with single-agent cytotoxic chemotherapies such as gemcitabine or liposomal doxorubicin provide an avenue for rapid disease control.¹⁹⁻²¹ Multiagent chemotherapy still holds an important role in managing patients with urgent need of clinical response and in obtaining maximal clinical response of extracutaneous compartments before allo-SCT. Given the limited response duration and risk of cumulative toxicity associated with chemotherapy, polychemotherapy is best used with a plan to transition to an alternative noncytotoxic agent or to proceed to allo-SCT once maximal disease response is achieved.

LCT

LCT is a consistent independent adverse prognostic factor in MF and SS.²² Historically, patients with LCT who required systemic treatment commonly received multiagent cytotoxic chemotherapy.²³ Subsequent prospective studies of single-agent chemotherapies including gemcitabine, pentostatin, and liposomal doxorubicin specifically included patients with LCT, as did the pivotal PROPEL study which investigated pralatrexate in relapsed/refractory T-cell lymphomas.^{16,20,21,24} Taken together, these therapies result in ORRs of 25% to 60% and median response durations of <6 months.

LCT is now emerging as a predictive factor in the selection of novel agents. Although CCR4 is commonly expressed in MF or SS

with LCT,²⁵ mogamulizumab has not shown encouraging activity in this patient population,²⁶ and patients with LCT were excluded from the MAVORIC study.⁸ In contrast, patients with LCT were included in the ALCANZA study, and BV was more effective in patients with LCT than in those without LCT (ORR4, 65% vs 39%), with a median PFS of 15.5 months.²⁷ Together with emerging data confirming its effectiveness in typical clinical practice, BV should be considered a preferred option for patients with LCT.¹⁷

Planned allo-SCT

Despite the introduction of highly effective novel treatments, allo-SCT remains the only curative treatment in advanced MF and SS. However, because of high morbidity and mortality, only a small minority of patients undergo transplantation. Criteria for transplantation remain controversial and vary across experienced CTCL centers, with SCT usually reserved for patients with high-risk or multiple relapsed/refractory advanced-stage disease. Traditional transplantation regimens are associated with a 5-year PFS of 17% to 26% and overall survival of 32% to 38%.^{28,29} The introduction of novel reduced intensity conditioning regimens that include total skin electron beam therapy have improved outcomes, with a 5-year overall survival of >50% because of decreased peritransplant mortality.^{30,31} These treatment protocols may extend the possibility of allo-SCT to older and higher risk patients earlier in their disease course. A large international study of prognostic factors in advanced-stage MF and SS (retro-CLIP) identified 3 risk groups with distinct outcomes based on clinical characteristics (age >60, stage IV, LCT, and elevated lactate dehydrogenase). Patients in the high-risk group, with 5-year overall survival of 28%, may generally be appropriate for allo-SCT, if eligible.²² Improved prognostic models based on molecular features are needed to further stratify decision-making in intermediate- and low-risk patients. Ultimately, the decision to undergo transplantation is individualized and requires weighing multiple factors including the prospect of durable remission, the risk of graft-versus-host disease, the possibility of post-transplant disease relapse, and remaining treatment options if transplantation is not pursued.

Although BV can effectively serve as bridging therapy without enhancing pretransplant toxicity,³² mogamulizumab should be used cautiously. In patients with adult T-cell leukemia/lymphoma, the use of mogamulizumab before transplant was associated with increased risk of severe acute graft-versus-host disease and nonrelapse mortality because of depletion of CCR4-expressing regulatory T cells.³³ This association has not been established in patients with CTCL, with 1 study showing no clear evidence of increased risk of acute graft-versus-host disease in patients who had received mogamulizumab ≥6 months before transplantation.³⁴

Maximizing therapeutic benefit of novel systemic agents

Decisions regarding treatment duration must balance conflicting goals of maximizing efficacy and minimizing toxicity. Because responses to traditional treatments are usually partial and short-lived, modern therapies are typically continued until progression. Although only occurring in a minority of patients, treatment with the latter may result in deeper, even complete responses.^{6,10,35,36} Thus, some patients may continue infusional treatments for years, accumulating toxicity and cost, and

interfering with quality of life. Although prospective trials are ongoing, limited evidence supports strategies including dose-reduction or extension of dosing interval between infusions, in patients receiving continuous therapies.³⁷⁻³⁹

BV is associated with cumulative neurotoxicity which can be mitigated by limiting treatment to 16 cycles. Even so, most patients may still experience some degree of neuropathy, which can be permanent.⁴⁰ Thus, for patients with a complete or good partial response, the emergence of neuropathy should prompt consideration for reduction in dose or a treatment holiday to ensure resolution. Upon disease relapse, such patients typically can be safely and effectively treated again with BV, thereby extending its utility while simultaneously reducing the risk of disabling neuropathy.⁴¹ Prospective evaluation of reduced-dose BV is ongoing, including 1 study showing a 42% ORR in patients with MF treated with BV at 0.9 mg/kg (50% of the dose used in ALCANZA), which may allow for longer treatment courses, maintained clinical benefit, and reduced neurotoxicity.^{39,42}

Mogamulizumab can be continued long-term, but a symptomatic, drug-associated rash may be observed in a significant fraction of patients.^{8,43-45} In addition, ongoing treatment is costly and inconvenient, requiring infusions every 2 weeks. Like BV, we have found that patients discontinuing mogamulizumab in the setting of a good response can be successfully retreated after relapse.⁴⁴ For patients with a complete response to mogamulizumab, we believe that a fixed duration of treatment may be considered to reduce the risk of mogamulizumab-associated rash and costs associated with indefinite maintenance.

Combination therapies

Additional efforts have been sought to combine treatments with nonoverlapping toxicity profiles to increase efficacy. ECP-based combinations with bexarotene or interferon have been extensively investigated, which augments response rates compared with ECP alone.⁴⁶ Romidepsin-based combinations with either pralatrexate or azacitidine have shown encouraging activity in patients with PTCL,^{47,48} and in those with CTCL. Emerging trials include combinations with pembrolizumab (NCT03278782) or lenalidomide/carfilzomib.⁴⁹ Combinations based on novel therapies are also being explored, including mogamulizumab or BV combined with total skin electron beam therapy (NCT04256018, NCT05357794), ECP (NCT04930653, NCT04676087), romidepsin (NCT02616965), lenalidomide (NCT03302728, NCT03409432), nivolumab (NCT02581631, NCT01703949), or with each other (NCT05414500). Combination strategies may yield more reliable responses across multiple disease compartments but require further study to show an advantage over using these therapies sequentially.⁵⁰

Improving biomarker-guided treatment approaches with emerging therapies

Decoding the molecular drivers of CTCL has catalyzed research into personalized treatment for this markedly heterogeneous disease, fueling both the discovery of new targeted treatments and the need for novel biomarkers predictive of therapeutic

benefit. Emerging therapies can be broadly categorized into 3 groups, each associated with its own class of biomarker: (1) Targeting surface molecules, (2) Targeting disrupted cellular pathways, and (3) Targeting the host immune system (Figure 2).

Surface targets in CTCL

In contrast to B-cell malignancies, in which lineage molecules can be relatively safely targeted, targeting entire T-cell lineage markers is often hindered by unacceptable toxicity.⁵¹⁻⁵³ Efforts in identifying more specific T-cell lymphoma surface markers including CCR4, CD30, CD25,⁵⁴ CD70,⁵⁵ CD37,⁵⁶ TRBC1/2,⁵⁷ and KIR3DL2 have produced a stream of antibody and cellular-based therapies with lesser toxicity. For example, in a phase 1 or 2 trial of patients with SS, the anti-KIR3DL2 antibody, lacutamab, was active (ORR, 43%) with few adverse events.⁵⁸

BV represents a cautionary example in the application of predictive biomarkers for a biomarker-specified treatment in CTCL. In the ALCANZA study, positive CD30 expression was defined as having at least 1 biopsy with $\geq 10\%$ CD30-positive malignant cells, but this cutoff was not based on data for superior outcomes in patients with higher CD30 expression. It is becoming clear that in CTCL, as with other lymphoid malignancies, CD30 expression is not a reliable predictive biomarker of response to BV.^{14,35,59,60} Remarkably, responses can be observed in CTCL even in the absence of immunohistochemical detection of CD30.^{14,61,62} Several hypotheses have been posited for this discordance, with one of the best validated explanations identifying a high inpatient variability of CD30 expression.²⁷ A biopsy of a single site may poorly represent CD30 expression in other areas, inappropriately selecting out patients who may nevertheless significantly benefit from BV.

Although the high interlesional variability of CTCL has been best demonstrated in the setting of CD30 expression based on the high molecular intratumoral heterogeneity of CTCL,⁶³⁻⁶⁶ it is likely that other putative biomarkers for novel therapies will similarly suffer sampling biases, limiting their utility. Ideally, future targeted therapies will require sampling of multiple lesions. Acknowledging the limitations of target detection, we believe that target expression should not necessarily be required as part of eligibility criteria for future trials but rather explored via planned analysis to determine whether expression correlates with clinical responses.

The revolution of cellular-based therapies such as chimeric antigen receptor T-cell therapy is now entering clinical exploration in CTCL. The adoption of cellular therapies into the treatment of T-cell lymphoma has been hampered by the challenges imposed by T-cell biology. T-cell based therapies, such as chimeric antigen receptor T-cell therapy, must circumvent fratricide whereby the therapeutic T cells are impeded by self-targeting. Additionally, it is unclear if any CTCL surface target exists that can yield durable responses comparable to those seen by CD19-directed T cells in B-cell malignancies, without additionally causing lasting T-cell aplasia. Although several cellular therapies are being studied in CTCL, efficacy data are limited. Notably, CD70-directed allogeneic chimeric antigen receptor T-cell therapy represents one example showing early promise in patients with CTCL.⁶⁷

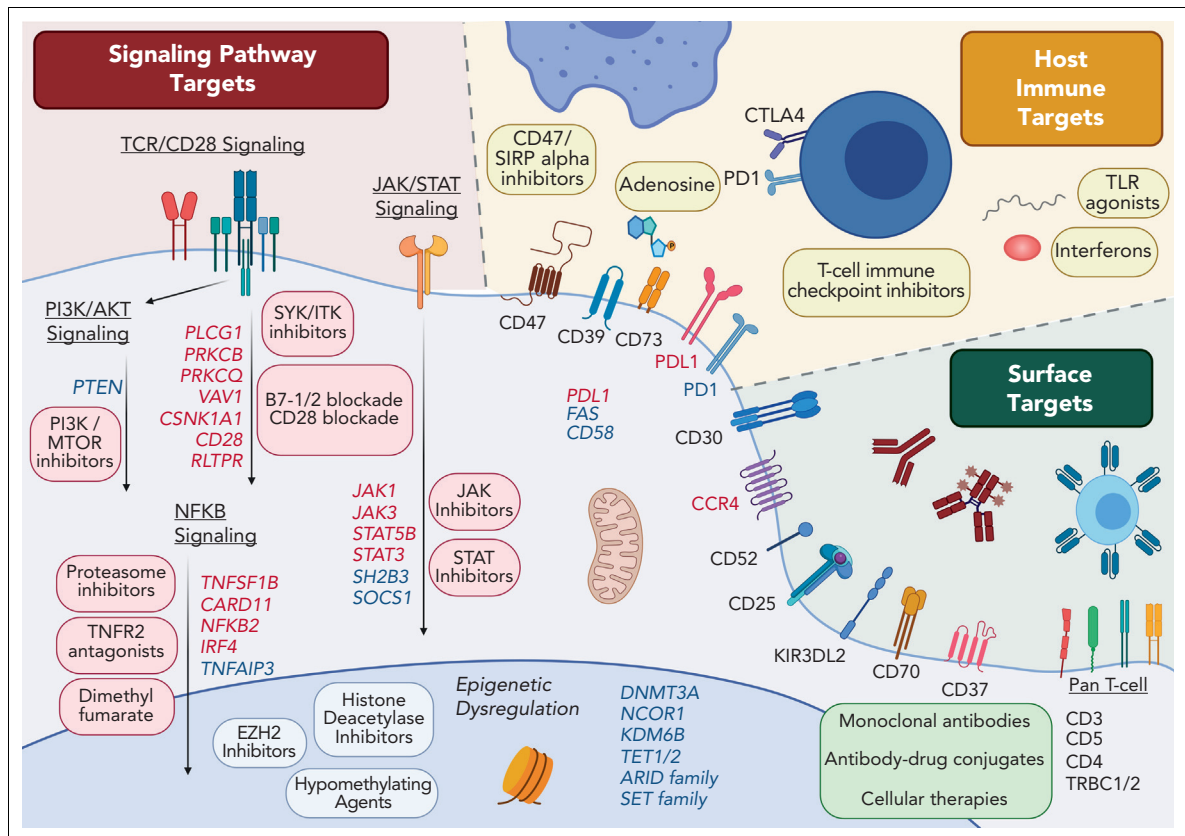


Figure 2. Therapeutic targets in cutaneous T-cell lymphoma. Genes recurrently affected by gain-of-function genomic variants (red) or loss-of-function variants (blue) are shown with each altered pathway.

Targeting disrupted pathways in CTCL

It was hoped that increased understanding of the molecular pathogenesis of CTCL would lead to a new era of precision medicine. However, the marked genomic heterogeneity of CTCL has complicated these efforts.^{63,68-77} Even the most frequently mutated gene in CTCL, *TP53*, is mutated in only 15% of patients. Although recurrent sequence variants are rare, structural variants and copy number alterations are more stereotyped in CTCL.^{70,77,78} In addition, structural variants appear to vary by disease stage, with more frequent aberrations (including deletions of 17p and 10q and amplification of 17q) in patients with SS than in those with MF.⁷⁹ Thus, efforts to personalize treatment in CTCL may require improved availability of assays to detect key copy number variants, rather than the current generation of targeted cancer sequencing panels focusing on sequence variants. Despite the genomic heterogeneity of CTCL, a clear pattern has emerged with recurrent clustering of genomic alterations within key pathways. These include activation of T-cell receptor signaling, T-cell receptor costimulation, JAK-STAT signaling, and the NF-κB pathway. As in other lymphomas, there is also frequent disruption of cell cycle and epigenetic regulators. Although the detection of these pathway alterations may be used to select targeted therapies, they are so frequently affected by various mechanisms that their predictive utility may be limited. For example, 1 case series found deletions or sequence variants in the *ARID* or *SMARCB1* gene families in 92% of patients with SS,⁷² whereas response rates in patients

with SS to the histone deacetylase inhibitors vorinostat or romidepsin range from 2% to 34%.^{8,9}

JAK inhibition represents the most practical initial candidate for a personalized treatment because of the wide availability of Food and Drug Administration–approved inhibitors. Single nucleotide variants in *JAK1*, *JAK3*, *STAT3*, or *STAT5B* cumulatively occur in 10% to 15% of patients with CTCL.^{72,80} A trial of ruxolitinib (a dual JAK1/JAK2 inhibitor) in T-cell lymphoma provides support for the concept that lymphomas with mutations promoting JAK/STAT signaling may respond better to JAK inhibitors.⁸¹ Likely because of the rarity of JAK and STAT mutations, no patients with CTCL were enrolled in cohort consisting the JAK/STAT mutation, highlighting the challenges of studying precision targeted strategies in prospective trials of rare diseases.

Targeting the host immune system in CTCL

Immune therapies play an increasingly important role in CTCL. In particular, programmed cell death protein 1 (PD1) targeting therapies are among the few treatments shown to induce lasting responses in patients.⁸² Pembrolizumab, a monoclonal antibody targeting PD1, was explored in a phase 2 study of 24 patients with relapsed or refractory stage IB-IV MF/SS,⁸³ showing an ORR of 38%. Responses were observed across all disease stages; at a median follow-up of 58 weeks, median response duration was not reached, and 1-year PFS was 65%.

The future of PD1/ programmed cell death ligand 1 (PDL1) inhibition in CTCL depends on improved selection of those patients who are most likely to respond. Thus far, biomarkers for immune checkpoint response established in other malignancies do not appear to validate in CTCL. PDL1 expression has not correlated with response in the small studies to-date.^{82,83} However, structural variants of PDL1 occurring in a subset of patients with LCT may predict clinical benefit to immune checkpoint inhibitors.⁸⁴ It is possible that some patients with LCT may show a more immunogenic phenotype, perhaps because of the higher tumor mutational burden seen in this setting.^{75,85} Other genomic events that promote immune evasion, such as *CD58* and *FAS* mutations, are rarely found in CTCL but warrant study as potential immunotherapy biomarkers.

Immune profiling may reveal predictive biomarkers for other classes of immune mediated therapies being explored in CTCL including Toll-like receptor agonists, interferons, interleukins, and other immune checkpoint inhibitors. CD47 inhibitors have shown promise in CTCL with a response rate of 26%.⁸⁶ Phenotyping of the macrophage component of the skin microenvironment is being explored as a biomarker for CD47 therapy. Despite single-agent activity, the future of CD47 blockade in CTCL is likely to be a part of combination therapies. The combination of CD47 inhibition with tumor-targeting monoclonal antibodies appears particularly promising.^{87,88}

The most accurate predictive biomarkers for immunotherapies in CTCL would likely include not only tumor-intrinsic features, but also features of the tumor microenvironment and the host (eg, the microbiome).^{89,90} Components of the tumor microenvironment, such as tumor-associated macrophages, myeloid-derived suppressor cells, regulatory T cells, and cancer-associated fibroblasts, have been shown to contribute to disease pathogenesis by maintaining a strongly immune suppressive environment.⁹¹ Tumor microenvironment-based biomarkers of immunotherapy in CTCL may require not only quantification of these components but also more complex analysis of their spatial organization.⁹² Dysregulation of immune suppressive metabolic pathways should also be explored. In particular, overexpression of CD39 and CD73 in SS points to dysregulation of the adenosine pathway as a target for future immune based therapies.^{93,94}

An additional challenge in adopting immunotherapy in CTCL treatment, and particularly in the targeting of T-cell immune checkpoints such as PD1, is the threat of "hyperprogression" or an acceleration of disease growth. Blockade of PD1 signaling may release malignant T cells from negative regulators of T-cell activation resulting in rapid proliferation. Although PD1 is commonly expressed in advanced-stage CTCL,^{95,96} the *PDCD1* gene is paradoxically often deleted in advanced disease.⁷⁹ Multiple studies have shown the potential for deletion of PD1

to promote lymphoma growth,^{79,97} and in vitro blockade of PD1 signaling to stimulate CTCL proliferation.⁹⁸ However, as opposed to studies of other T-cell lymphomas,^{99,100} there have not yet been well-documented cases of hyperprogression in CTCL after PD1 blockade, even in lymphomas with PD1 deletion.⁸³ Disease flaring was observed in patients treated with pembrolizumab with high PD1 expression but typically remitted without intervention and could evolve into clinical responses.⁸³ Nevertheless, until there is more experience with immune checkpoint inhibitors, we cannot exclude their potential for hyperprogression in a subset of patients.

Conclusion

The escalating pace of drug discovery in CTCL continues to expand our therapeutic options. As we study these new treatments, we may need to adjust our measurement of clinical success. The combination of highly targeted therapies and a highly heterogeneous disease means that efficacy may not always be reflected in improved response rates in unselected patients. Rather, successful treatments may be manifested by extraordinary, durable responses in a subset of patients with susceptible disease. Though currently predominantly being guided by clinical factors, molecular biomarkers must become a part of our treatment paradigms if we are to take the leap from incremental gains to long-term improvements in outcomes for patients with advanced MF and SS.

Acknowledgments

Figures were created with [Biorender.com](https://biorender.com).

Authorship

Contribution: M.S.K., E.M., and Y.H.K. designed and wrote the paper.

Conflict-of-interest disclosure: M.S.K. has received research funding from CRISPR Therapeutics and Nutcracker Therapeutics, consulting fees from Myeloid Therapeutics, and advisory board participation from Daiichi Sankyo. Y.H.K. has received research funding from Kyowa Kirin, Innate Pharma, Corvus Pharmaceuticals, Galderma Pharmaceuticals, CRISPR Therapeutics, Trillium Therapeutics, and advisory board participation from Kyowa Kirin, Galderma Pharmaceuticals, and Secura Bio. E.M. declares no competing financial interests.

ORCID profiles: M.S.K., 0000-0001-9061-7351; E.M., 0000-0002-9865-4517; Y.H.K., 0000-0003-2686-5873.

Correspondence: Michael S. Khodadoust, 1701 Page Mill Rd, Palo Alto, CA 94304; email: mkhodado@stanford.edu; and Youn H. Kim, 780 Welch Rd, CJ220D, Stanford, CA 94305; email: younkim@stanford.edu.

Footnote

Submitted 27 September 2021; accepted 31 October 2022; prepublished online on *Blood* First Edition 15 November 2022. <https://doi.org/10.1182/blood.2020008241>.

REFERENCES

- Whittaker SJ, Demierre MF, Kim EJ, et al. Final results from a multicenter, international, pivotal study of romidepsin in refractory cutaneous T-cell lymphoma. *J Clin Oncol*. 2010;28(29):4485-4491.
- Horwitz SM, Kim YH, Foss F, et al. Identification of an active, well-tolerated dose of pralatrexate in patients with relapsed or refractory cutaneous T-cell lymphoma. *Blood*. 2012;119(18):4115-4122.
- Hughes CF, Khot A, McCormack C, et al. Lack of durable disease control with chemotherapy for mycosis fungoides and Sézary syndrome: a comparative study of systemic therapy. *Blood*. 2015;125(1):71-81.

4. Hanel W, Briski R, Ross CW, et al. A retrospective comparative outcome analysis following systemic therapy in Mycosis fungoides and Sézary syndrome. *Am J Hematol*. 2016;91(12):E491-E495.
5. Quaglino P, Maule M, Prince HM, et al. Global patterns of care in advanced stage mycosis fungoides/Sézary syndrome: a multicenter retrospective follow-up study from the Cutaneous Lymphoma International Consortium. *Ann Oncol*. 2017;28(10):2517-2525.
6. Prince HM, Kim YH, Horwitz SM, et al. Brentuximab vedotin or physician's choice in CD30-positive cutaneous T-cell lymphoma (ALCANZA): an international, open-label, randomised, phase 3, multicentre trial. *Lancet*. 2017;390(10094):555-566.
7. Horwitz SM, Scarisbrick JJ, Dummer R, et al. Randomized phase 3 ALCANZA study of brentuximab vedotin vs physician's choice in cutaneous T-cell lymphoma: final data. *Blood Adv*. 2021;5(23):5098-5106.
8. Kim YH, Bagot M, Pinter-Brown L, et al. Mogamulizumab versus vorinostat in previously treated cutaneous T-cell lymphoma (MAVORIC): an international, open-label, randomised, controlled phase 3 trial. *Lancet Oncol*. 2018;19(9):1192-1204.
9. Kim EJ, Kim YH, Rook AH, et al. Clinically significant responses achieved with romidepsin across disease compartments in patients with cutaneous T-cell lymphoma. *Leuk Lymphoma*. 2015;56(10):2847-2854.
10. Kim YH, Khodadoust MS, de Masson A, et al. Patient characteristics of long-term responders to mogamulizumab: results from the MAVORIC study. *Blood*. 2020;136(Supplement 1):35.
11. Gao C, McCormack C, van der Weyden C, et al. Prolonged survival with the early use of a novel extracorporeal photopheresis regimen in patients with Sézary syndrome. *Blood*. 2019;134(16):1346-1350.
12. Zic JA. Extracorporeal photopheresis in the treatment of mycosis fungoides and Sézary Syndrome. *Dermatol Clin*. 2015;33(4):765-776.
13. Lewis DJ, Haun PL, Samimi SS, et al. Brentuximab vedotin for relapsed or refractory Sézary syndrome. *JAMA Dermatol*. 2021;157(3):317-321.
14. Kim YH, Tavallaee M, Sundram U, et al. Phase II investigator-initiated study of brentuximab vedotin in mycosis fungoides and Sézary syndrome with variable CD30 expression level: a multi-institution collaborative project. *J Clin Oncol*. 2015;33(32):3750-3758.
15. Foss F, Duvic M, Lerner A, Waksman J, Whittaker S. Clinical efficacy of romidepsin in tumor stage and folliculotropic mycosis fungoides. *Clin Lymphoma Myeloma Leuk*. 2016;16(11):637-643.
16. Foss F, Horwitz SM, Coiffier B, et al. Pralatrexate is an effective treatment for relapsed or refractory transformed mycosis fungoides: a subgroup efficacy analysis from the PROPEL study. *Clin Lymphoma Myeloma Leuk*. 2012;12(4):238-243.
17. O'Donnell M, Zaya R, Correia E, et al. Disease characteristics, prognosis, and response to therapy in patients with large-cell transformed mycosis fungoides: A single-center retrospective study. *J Am Acad Dermatol*. 2022;86(6):1285-1292.
18. Horwitz SM, Advani RH, Bartlett NL, et al. Objective responses in relapsed T-cell lymphomas with single-agent brentuximab vedotin. *Blood*. 2014;123(20):3095-3100.
19. Zinzani PL, Baliva G, Magagnoli M, et al. Gemcitabine treatment in pretreated cutaneous T-cell lymphoma: experience in 44 patients. *J Clin Oncol*. 2000;18(13):2603-2606.
20. Duvic M, Talpur R, Wen S, Kurzrock R, David CL, Apisarnthanarax N. Phase II evaluation of gemcitabine monotherapy for cutaneous T-cell lymphoma. *Clin Lymphoma Myeloma*. 2006;7(1):51-58.
21. Dummer R, Quaglino P, Becker JC, et al. Prospective international multicenter phase II trial of intravenous pegylated liposomal doxorubicin monotherapy in patients with stage IIB, IVA, or IVB advanced mycosis fungoides: final results from EORTC 21012. *J Clin Oncol*. 2012;30(33):4091-4097.
22. Scarisbrick JJ, Prince HM, Vermeer MH, et al. Cutaneous lymphoma international consortium study of outcome in advanced stages of mycosis fungoides and Sézary syndrome: effect of specific prognostic markers on survival and development of a prognostic model. *J Clin Oncol*. 2015;33(32):3766-3773.
23. Vergier B, de Muret A, Beylot-Barry M, et al. Transformation of mycosis fungoides: clinicopathological and prognostic features of 45 cases. French Study Group of Cutaneous Lymphomas. *Blood*. 2000;95(7):2212-2218.
24. Tsimberidou AM, Giles F, Duvic M, Fayad L, Kurzrock R. Phase II study of pentostatin in advanced T-cell lymphoid malignancies: update of an M.D. Anderson Cancer Center series. *Cancer*. 2004;100(2):342-349.
25. Jones D, O'Hara C, Kraus MD, et al. Expression pattern of T-cell-associated chemokine receptors and their chemokines correlates with specific subtypes of T-cell non-Hodgkin lymphoma. *Blood*. 2000;96(2):685-690.
26. Zinzani PL, Karlin L, Radford J, et al. European phase II study of mogamulizumab, an anti-CCR4 monoclonal antibody, in relapsed/refractory peripheral T-cell lymphoma. *Haematologica*. 2016;101(10):e407-e410.
27. Kim YH, Prince HM, Whittaker S, et al. Response to brentuximab vedotin versus physician's choice by CD30 expression and large cell transformation status in patients with mycosis fungoides: An ALCANZA sub-analysis. *Eur J Cancer*. 2021;148:411-421.
28. Domingo-Domenech E, Duarte RF, Boumedil A, et al. Allogeneic hematopoietic stem cell transplantation for advanced mycosis fungoides and Sézary syndrome. An updated experience of the Lymphoma Working Party of the European Society for Blood and Marrow Transplantation. *Bone Marrow Transplant*. 2021;56(6):1391-1401.
29. Lechowicz MJ, Lazarus HM, Carreras J, et al. Allogeneic hematopoietic cell transplantation for mycosis fungoides and Sézary syndrome. *Bone Marrow Transplant*. 2014;49(11):1360-1365.
30. Weng WK, Arai S, Rezvani A, et al. Nonmyeloablative allogeneic transplantation achieves clinical and molecular remission in cutaneous T-cell lymphoma. *Blood Adv*. 2020;4(18):4474-4482.
31. Morris S, Palanicanwader R, Grandi V, et al. Non-Myeloablative Allogeneic Stem Cell Transplantation using TSEB TLI and ATG for Mycosis Fungoides and Sézary Syndrome. Medium term results from a large single centre cohort. World Congress of Cutaneous Lymphomas; 2020.
32. Garciaz S, Loschi M, De Masson A, et al. Brentuximab vedotin as a bridge to allogeneic stem-cell transplantation for refractory or relapsing patients with CD30 positive anaplastic or T-cell non-Hodgkin lymphomas: a study on behalf of the SFGM-TC. *Leuk Lymphoma*. 2019;60(11):2802-2805.
33. Fuji S, Inoue Y, Utsunomiya A, et al. Pretransplantation Anti-CCR4 antibody mogamulizumab against adult T-Cell leukemia/lymphoma is associated with significantly increased risks of severe and corticosteroid-refractory graft-versus-host disease, nonrelapse mortality, and overall mortality. *J Clin Oncol*. 2016;34(28):3426-3433.
34. Dai J, Almazan TH, Hong EK, et al. Potential association of anti-CCR4 antibody mogamulizumab and graft-vs-host disease in patients with mycosis fungoides and Sézary syndrome. *JAMA Dermatol*. 2018;154(6):728-730.
35. Papadavid E, Kapniari E, Pappa V, et al. Multicentric EORTC retrospective study shows efficacy of brentuximab vedotin in mycosis fungoides and Sézary syndrome patients with variable CD30 positivity. *Br J Dermatol*. 2021;185(5):1035-1044.
36. Duvic M, Bates SE, Piekarz R, et al. Responses to romidepsin in patients with cutaneous T-cell lymphoma and prior treatment with systemic chemotherapy. *Leuk Lymphoma*. 2018;59(4):880-887.
37. Martinez-Escalera ME, Kuzel TM, Kaplan JB, et al. Durable responses with maintenance dose-sparing regimens of romidepsin in cutaneous T-cell lymphoma. *JAMA Oncol*. 2016;2(6):790-793.
38. Foss FM, Parker TL, Girardi M, Li A. Clinical activity of pralatrexate in patients with cutaneous T-cell lymphoma treated with varying doses of pralatrexate. *Clin*

Lymphoma Myeloma Leuk. 2018;18(11):e445-e447.

39. Stranzenbach R, Dippel E, Schlaak M, Stadler R. Brentuximab vedotin in CD30. *Br J Dermatol.* 2017;177(6):1503-1509.
40. Corbin ZA, Nguyen-Lin A, Li S, et al. Characterization of the peripheral neuropathy associated with brentuximab vedotin treatment of Mycosis Fungoides and Sézary Syndrome. *J Neuro Oncol.* 2017;132(3):439-446.
41. Bartlett NL, Chen R, Fanale MA, et al. Retreatment with brentuximab vedotin in patients with CD30-positive hematologic malignancies. *J Hematol Oncol.* 2014;7:24.
42. Khan N, Noor S, Geller S, et al. A phase II trial of reduced dose brentuximab vedotin for cutaneous T-cell lymphomas. *Hematol Oncol.* 2021;39(S2).
43. Musiek ACM, Whittaker S, Horwitz SM, et al. Characterization and outcomes in patients with mogamulizumab-associated skin reactions in the MAVORIC trial. *Blood.* 2020;136(Supplement 1):23-24.
44. Hirotsu KE, Neal TM, Khodadoust MS, et al. Clinical characterization of mogamulizumab-associated rash during treatment of mycosis fungoides or Sézary syndrome. *JAMA Dermatol.* 2021;157(6):700-707.
45. Wang JY, Hirotsu KE, Neal TM, et al. Histopathologic characterization of mogamulizumab-associated rash. *Am J Surg Pathol.* 2020;44(12):1666-1676.
46. Knobler R, Arenberger P, Arun A, et al. European dermatology forum - updated guidelines on the use of extracorporeal photopheresis 2020 - part 1. *J Eur Acad Dermatol Venereol.* 2020;34(12):2693-2716.
47. Amengual JE, Lichtenstein R, Lue J, et al. A phase 1 study of romidepsin and pralatrexate reveals marked activity in relapsed and refractory T-cell lymphoma. *Blood.* 2018;131(4):397-407.
48. Falchi L, Ma H, Klein S, et al. Combined oral 5-azacytidine and romidepsin are highly effective in patients with PTCL: a multicenter phase 2 study. *Blood.* 2021;137(16):2161-2170.
49. Mehta-Shah N, Lunning MA, Moskowitz AJ, et al. Romidepsin and lenalidomide-based regimens have efficacy in relapsed/refractory lymphoma: combined analysis of two phase I studies with expansion cohorts. *Am J Hematol.* 2021;96(10):1211-1222.
50. Fong S, Hong EK, Khodadoust MS, et al. Low-dose total skin electron beam therapy combined with mogamulizumab for refractory mycosis fungoides and Sézary syndrome. *Adv Radiat Oncol.* 2021;6(3):100629.
51. Frankel AE, Woo JH, Ahn C, et al. Resimmune, an anti-CD3ε recombinant immunotoxin, induces durable remissions in patients with cutaneous T-cell lymphoma. *Haematologica.* 2015;100(6):794-800.
52. Kim YH, Duvic M, Obtiz E, et al. Clinical efficacy of zanolimumab (HuMax-CD4): two phase 2 studies in refractory cutaneous T-cell lymphoma. *Blood.* 2007;109(11):4655-4662.
53. Thursky KA, Worth LJ, Seymour JF, Miles Prince H, Slavin MA. Spectrum of infection, risk and recommendations for prophylaxis and screening among patients with lymphoproliferative disorders treated with alemtuzumab*. *Br J Haematol.* 2006;132(1):3-12.
54. Prince HM, Duvic M, Martin A, et al. Phase III placebo-controlled trial of denileukin diftitox for patients with cutaneous T-cell lymphoma. *J Clin Oncol.* 2010;28(11):1870-1877.
55. Bagot M, Maerevoet M, Zinzani PL, et al. Argx-110 for treatment of CD70-positive advanced cutaneous T-cell lymphoma in a phase 1/2 clinical trial. *Blood.* 2018;132(Supplement 1):1627.
56. Sawas A, Savage KJ, Perez RP, et al. A phase 1 study of the anti-CD37 antibody-drug conjugate AGS67E in advanced lymphoid malignancies. Interim results. *Hematol Oncol.* 2017;35(S2):49.
57. Maciocia PM, Wawrzyniecka PA, Philip B, et al. Targeting the T cell receptor β-chain constant region for immunotherapy of T cell malignancies. *Nat Med.* 2017;23(12):1416-1423.
58. Bagot M, Porcu P, Marie-Cardine A, et al. IPH4102, a first-in-class anti-KIR3DL2 monoclonal antibody, in patients with relapsed or refractory cutaneous T-cell lymphoma: an international, first-in-human, open-label, phase 1 trial. *Lancet Oncol.* 2019;20(8):1160-1170.
59. Jagadeesh D, Horwitz SM, Bartlett NL, et al. Response to brentuximab vedotin by CD30 expression: Results from five trials in PTCL, CTCL, and B-cell lymphomas. *J Clin Oncol.* 2019;37(15_suppl):7543.
60. Advani RH, Horwitz SM, Iyer SP, et al. Response to A+CHP by CD30 expression in the ECHELON-2 trial. *J Clin Oncol.* 2019;37(15_suppl):7538.
61. Goyal A, Hordinsky M, Lazaryan A. Impressive response of CD30-negative, treatment-refractory mycosis fungoides to brentuximab vedotin. *Dermatol Ther.* 2019;32(2):e12835.
62. Zhang C, Chairatchaneeboon M, Haun P, Landsburg D, Kim EJ. Treatment of CD30-negative refractory mycosis fungoides with brentuximab vedotin. *JAMA Dermatol.* 2018;154(1):109-110.
63. Iyer A, Hennessey D, O'Keefe S, et al. Clonotypic heterogeneity in cutaneous T-cell lymphoma (mycosis fungoides) revealed by comprehensive whole-exome sequencing. *Blood Adv.* 2019;3(7):1175-1184.
64. Gaydosik AM, Tabib T, Geskin LJ, et al. Single-cell lymphocyte heterogeneity in advanced cutaneous T-cell lymphoma skin tumors. *Clin Cancer Res.* 2019;25(14):4443-4454.
65. Iyer A, Hennessey D, O'Keefe S, et al. Branched evolution and genomic intratumor heterogeneity in the pathogenesis of cutaneous T-cell lymphoma. *Blood Adv.* 2020;4(11):2489-2500.
66. Herrera A, Cheng A, Mimitou EP, et al. Multimodal single-cell analysis of cutaneous T cell lymphoma reveals distinct sub-clonal tissue-dependent signatures. *Blood.* 2021;138(16):1456-1464.
67. Iyer SP, Alejandro Sica R, Joy Ho P, et al. The COBALT-LYM study of CTX130: A Phase 1 dose escalation study of CD70-targeted allogeneic CRISPR-CAS9-engineered CAR T cells in patients with relapsed/refractory (R/R) T-cell malignancies. *European Hematology Association;* 2022.
68. Bastidas Torres AN, Cats D, Mei H, et al. Genomic analysis reveals recurrent deletion of JAK-STAT signaling inhibitors HNRNP and SOCS1 in mycosis fungoides. *Genes Chromosomes Cancer.* 2018;57(12):653-664.
69. Chang LW, Patrone CC, Yang W, et al. An integrated data resource for genomic analysis of cutaneous T-cell lymphoma. *J Invest Dermatol.* 2018;138(12):2681-2683.
70. Choi J, Goh G, Walradt T, et al. Genomic landscape of cutaneous T cell lymphoma. *Nat Genet.* 2015;47(9):1011-1019.
71. da Silva Almeida AC, Abate F, Khiabani H, et al. The mutational landscape of cutaneous T cell lymphoma and Sézary syndrome. *Nat Genet.* 2015;47(12):1465-1470.
72. Kiel MJ, Sahasrabudhe AA, Rolland DCM, et al. Genomic analyses reveal recurrent mutations in epigenetic modifiers and the JAK-STAT pathway in Sézary syndrome. *Nat Commun.* 2015;6:8470.
73. McGirt LY, Jia P, Baerenwald DA, et al. Whole-genome sequencing reveals oncogenic mutations in mycosis fungoides. *Blood.* 2015;126(4):508-519.
74. Park J, Yang J, Wenzel AT, et al. Genomic analysis of 220 CTCLs identifies a novel recurrent gain-of-function alteration in RLTPR (p.Q575E). *Blood.* 2017;130(12):1430-1440.
75. Ungewickell A, Bhaduri A, Rios E, et al. Genomic analysis of mycosis fungoides and Sézary syndrome identifies recurrent alterations in TNFR2. *Nat Genet.* 2015;47(9):1056-1060.
76. Woollard WJ, Pullabhatla V, Lorenc A, et al. Candidate driver genes involved in genome maintenance and DNA repair in Sézary syndrome. *Blood.* 2016;127(26):3387-3397.
77. Wang L, Ni X, Covington KR, et al. Genomic profiling of Sézary syndrome identifies alterations of key T cell signaling and differentiation genes. *Nat Genet.* 2015;47(12):1426-1434.
78. Gug G, Huang Q, Chiticariu E, Solovan C, Baudis M. DNA copy number imbalances in primary cutaneous lymphomas. *J Eur Acad Dermatol Venereol.* 2019;33(6):1062-1075.

79. Park J, Daniels J, Wartewig T, et al. Integrated genomic analyses of cutaneous T-cell lymphomas reveal the molecular bases for disease heterogeneity. *Blood*. 2021;138(14):1225-1236.
80. Pérez C, González-Rincón J, Onaindia A, et al. Mutated JAK kinases and deregulated STAT activity are potential therapeutic targets in cutaneous T-cell lymphoma. *Haematologica*. 2015;100(11):e450-e453.
81. Moskowitz AJ, Ghione P, Jacobsen E, et al. A phase 2 biomarker-driven study of ruxolitinib demonstrates effectiveness of JAK/STAT targeting in T-cell lymphomas. *Blood*. 2021;138(26):2828-2837.
82. Lesokhin AM, Ansell SM, Armand P, et al. Nivolumab in patients with relapsed or refractory hematologic malignancy: preliminary results of a phase Ib study. *J Clin Oncol*. 2016;34(23):2698-2704.
83. Khodadoust MS, Rook AH, Porcu P, et al. Pembrolizumab in relapsed and refractory mycosis fungoides and Sézary syndrome: a multicenter phase II study. *J Clin Oncol*. 2020;38(1):20-28.
84. Beygi S, Fernandez-Pol S, Duran G, et al. Pembrolizumab in mycosis fungoides with PD-L1 structural variants. *Blood Adv*. 2021; 5(3):771-774.
85. Argyropoulos KV, Pulitzer M, Maura F, et al. Targeted genomic analysis of cutaneous T cell lymphomas identifies a subset with aggressive clinicopathological features. *Blood Cancer J*. 2020;10:116.
86. Ansell SM, Maris MB, Lesokhin AM, et al. Phase I study of the CD47 blocker TTI-621 in patients with relapsed or refractory hematologic malignancies. *Clin Cancer Res*. 2021;27(8):2190-2199.
87. Jain S, Van Scoyk A, Morgan EA, et al. Targeted inhibition of CD47-SIRPα requires Fc-FcγR interactions to maximize activity in T-cell lymphomas. *Blood*. 2019;134(17): 1430-1440.
88. Advani R, Volkmer JP, Chao MP. CD47 blockade and rituximab in non-Hodgkin's lymphoma. *N Engl J Med*. 2018;379(18): 1711-1721.
89. Baruch EN, Youngster I, Ben-Betzalel G, et al. Fecal microbiota transplant promotes response in immunotherapy-refractory melanoma patients. *Science*. 2021; 371(6529):602-609.
90. Davar D, Dzutsev AK, McCulloch JA, et al. Fecal microbiota transplant overcomes resistance to anti-PD-1 therapy in melanoma patients. *Science*. 2021;371(6529):595-602.
91. Liu Z, Wu X, Hwang ST, Liu J. The role of tumor microenvironment in mycosis fungoides and Sézary syndrome. *Ann Dermatol*. 2021;33(6):487-496.
92. Phillips D, Matusiak M, Gutierrez BR, et al. Immune cell topography predicts response to PD-1 blockade in cutaneous T cell lymphoma. *Nat Commun*. 2021;12:6726.
93. Sonigo G, Bozonnet A, Dumont M, et al. Involvement of the CD39/CD73/adenosine pathway in T-cell proliferation and NK cell-mediated antibody-dependent cell cytotoxicity in Sézary syndrome. *Blood*. 2022;139(17):2712-2716.
94. Yakymiv Y, Marchisio S, Ortolan E, et al. CD39/CD73 dysregulation and adenosine metabolism contribute to T cell immunosuppression in patients with Sézary syndrome. *Blood*. 2022;141(1):111-116.
95. Samimi S, Benoit B, Evans K, et al. Increased programmed death-1 expression on CD4+ T cells in cutaneous T-cell lymphoma: implications for immune suppression. *Arch Dermatol*. 2010;146(12):1382-1388.
96. Cetinözman F, Jansen PM, Vermeer MH, Willemze R. Differential expression of programmed death-1 (PD-1) in Sézary syndrome and mycosis fungoides. *Arch Dermatol*. 2012;148(12): 1379-1385.
97. Wartewig T, Kurgis Z, Keppler S, et al. PD-1 is a haploinsufficient suppressor of T cell lymphomagenesis. *Nature*. 2017;552(7683): 121-125.
98. Saulite I, Ignatova D, Chang YT, et al. Blockade of programmed cell death protein 1 (PD-1) in Sézary syndrome reduces Th2 phenotype of non-tumoral T lymphocytes but may enhance tumor proliferation. *Oncol Immunology*. 2020;9(1):1738797.
99. Rauch DA, Conlon KC, Janakiram M, et al. Rapid progression of adult T-cell leukemia/lymphoma as tumor-infiltrating Tregs after PD-1 blockade. *Blood*. 2019;134(17): 1406-1414.
100. Bennani NN, Pederson LD, Atherton P, et al. A phase II study of nivolumab in patients with relapsed or refractory peripheral T-cell lymphoma [abstract]. *Blood*. 2019; 134(suppl 1). Abstract 467.

© 2023 by The American Society of Hematology