

The hypothesis that the propensity of subset 8 CLL to undergo Richter transformation is due to the epigenetic derepression of a particular set of genes caused by overactivation of a limited set of transcription factors is intriguing; however, important questions remain to be answered. Further research is necessary to elucidate the mechanistic connection between the BCR of subset 8 and the implicated transcription factors. This might help to identify druggable targets that could be exploited to improve the dismal clinical prognosis of patients with subset 8 and to potentially prevent Richter transformation. The current study did not find examples of distinct chromatin regulation in other CLL subsets, but this may be due to the relatively small sample sizes and the resulting low statistical power. Future studies with more patients and broadened scope by including cases from rarer CLL subsets might yield additional insights. In any case, Tsagiopoulou et al have to be applauded for their remarkable achievement outlining how a comprehensive “omics” approach linking immunogenetic features with (epi)genomic and transcriptional characterization can be instrumental in illuminating complex diseases mechanisms.

**Conflict-of-interest disclosure:** The authors declare no competing financial interests. ■

## REFERENCES

1. Tsagiopoulou M, Chapaprieta V, Russiñol N, et al. Chromatin activation profiling of stereotyped chronic lymphocytic leukemias reveals a subset 8-specific signature. *Blood*. 2023;141(24):2955-2960.
2. Rossi D, Spina V, Cerri M, et al. Stereotyped B-cell receptor is an independent risk factor of chronic lymphocytic leukemia transformation to Richter syndrome. *Clin Cancer Res*. 2009; 15(13):4415-4422.
3. Jaramillo S, Agathangelidis A, Schneider C, et al. Prognostic impact of prevalent chronic lymphocytic leukemia stereotyped subsets: analysis within prospective clinical trials of the Germann CLL Study Group (GCLLSG). *Haematologica*. 2019;105(11): 2598-2607.
4. Agathangelidis A, Chatzidimitriou A, Chatzikonstantinou T, et al. Immunoglobulin gene sequence analysis in chronic lymphocytic leukemia: the 2022 update of the recommendations by ERIC, the European Research Initiative on CLL. *Leukemia*. 2022; 36(8):1961-1968.
5. Maity PC, Bilal M, Koning MT, et al. IGLV3-21\*01 is an inherited risk factor for CLL through the acquisition of a single-point

mutation enabling autonomous BCR signaling. *Proc Natl Acad Sci U S A*. 2020; 117(8):4320-4327.

6. Nadeu F, Royo R, Clot G, et al. IGLV3-21<sup>R110</sup> identifies an aggressive biological subtype of chronic lymphocytic leukemia with intermediate epigenetics. *Blood*. 2021; 137(21):2935-2946.
7. Parry EM, Ten Hacken E, Wu CJ. Richter syndrome: novel insights into the biology of transformation. *Blood*. Published online 9 February 2023. <https://doi.org/10.1182/blood.2022016502>
8. Gounari M, Ntoufa S, Apollonio B, et al. Excessive antigen reactivity may underlie

the clinical aggressiveness of chronic lymphocytic leukemia stereotyped subset #8. *Blood*. 2015;125(23):3580-3587.

9. Broséus J, Hergalant S, Vogt J, et al. Molecular characterization of Richter syndrome identifies de novo diffuse large B-cell lymphomas with poor prognosis. *Nat Commun*. 2023;14(1):309.
10. Beekman R, Chapaprieta V, Russiñol N, et al. The reference epigenome and regulatory chromatin landscape of chronic lymphocytic leukemia. *Nat Med*. 2018;24(6):868-880.

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## PLATELETS AND THROMBOPOIESIS

Comment on *Kaiser et al*, page 2973

# The hemorrhage risk of dasatinib therapy

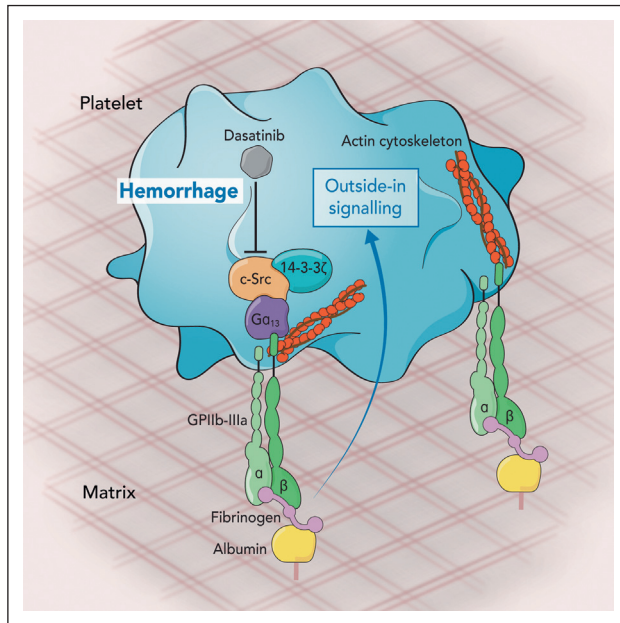
**Giulia Pontarollo and Christoph Reinhardt** | Johannes Gutenberg University Medical Center

**In this issue of *Blood*, Kaiser et al<sup>1</sup> pinpoint the glycoprotein IIb (GPIIb)/Gα<sub>13</sub>/Src tyrosine kinase (c-Src)/14-3-3ζ signaling axis as a requirement for platelet migration and shed light on how the antineoplastic oral tyrosine kinase inhibitor dasatinib increases the risk of inflammation-associated mucosal hemorrhage.**

Platelets are highly dynamic, migratory anucleate blood cells that not only are involved in blood clot formation but also play a key role in first-line immune responses. To accomplish both functions, platelets restructure their cytoskeleton when they adhere to matrix molecules following vascular injury.<sup>2</sup> This phenomenon mainly involves myosin heavy chain 9 (MYH9) and actin-related protein complex 2/3 (Arp2/3).<sup>3,4</sup> So far, the signaling pathways that regulate platelet migration up- and downstream of these mechanisms have remained elusive. In this issue of *Blood*, Kaiser and colleagues reveal how the sarcoma family kinase c-Src and the Src family kinase-binding protein 14-3-3ζ, situated downstream of the fibrinogen receptor GPIIb/IIIa, coupled to Gα<sub>13</sub>, is required for platelet polarization and migration.<sup>1</sup> In addition to further defining this downstream signaling pathway, Kaiser et al elucidate an important side effect of the antineoplastic oral tyrosine kinase inhibitor dasatinib. Dasatinib is an inhibitor of B-cell receptor-ABL-tyrosine kinase and of the sarcoma family kinase c-Src, used clinically to treat

chronic myeloid leukemia (CML) and Philadelphia chromosome-positive acute lymphoblastic leukemia (ALL). They found that dasatinib-treatment causes mucosal bleeding through the blockade of platelet migration.

The fibrin(ogen) receptor GPIIb/IIIa regulates platelet adherence to extracellular matrices. Kaiser and colleagues found that both genetic and pharmacologic targeting of the Arp2/3 complex abrogates the polarized platelet phenotype and the ability of platelets to form lamellipodia and migrate on albumin/fibrin(ogen) hybrid matrices, highlighting the importance of Arp2/3 for platelet polarization and migration.<sup>5</sup> Intriguingly, although platelet size and circularity was reduced, inhibition of Arp2/3 did not affect clot retraction. Previous work by Gaertner and coworkers has demonstrated the critical involvement of MYH9 in platelet migration.<sup>6</sup> In this issue of *Blood*, Kaiser et al show that coordinated myosin II function is a requirement for motility (retraction of lamellipodia) but is not necessary for platelet polarization, which



Platelet migration on fibrinogen surfaces measured *ex vivo*, after derivatization of albumin-coated matrices. The formation of lamellipodia is accomplished through the integrin receptor GPIIb/IIIa. Fibrinogen binding to GPIIb/IIIa triggers an outside-in signaling mediated by the kinases c-Src and 14-3-3ζ through the Gα<sub>13</sub> that results in the reorganization of actin cytoskeleton and subsequent platelet migration. Dasatinib, a kinase inhibitor therapeutically used to treat chronic myeloid leukemia, inhibits platelet migration and is therefore associated with hemorrhage events observed during the treatment of leukemia patients. Professional illustration by Somersault18:24.

is mainly mediated by dynamic actin waves along the leading edge of lamellipodia. Moreover, the authors identified phospholipase C as the upstream regulator of Arp2/3-dependent platelet migration. Inhibitor experiments with pyrazolopyrimidine, which blocks the action of potent platelet agonists, suggested the involvement of Src family kinases in platelet polarization and migration. A subsequent screening of different kinase inhibitors identified c-Src as the most sensitive Src family kinase promoting platelet migration on fibrinogen matrices. In addition to c-Src, the sarcoma family kinase-binding protein 14-3-3ζ is involved in GPIIb/IIIa-mediated outside-in signaling. Indeed, colocalization of GPIIb to 14-3-3ζ, and inhibitor screening with the 14-3-3ζ-inhibitor 3',4',7' trihydroxyisoflavone, univocally demonstrated the involvement of the Src family kinase binding protein 14-3-3ζ in platelet migration. Stimulation with soluble agonists such as ADP and thromboxane play a role in platelet recruitment prior to adhesion but have no effect once migration is initiated.

Most importantly, the authors found that platelet migration, as well as polarization and lamellipodium formation,<sup>7</sup> was impaired at low doses of the c-Src inhibitor dasatinib, whereas retraction of cross-linked fibrin, platelet degranulation, and

thrombus formation were only impaired at higher doses (see figure). Indeed, treatment with low doses of dasatinib reduced platelet area, circularity, and aspect ratio, with an abnormal presence of filopodia, through inhibition of Src tyrosine 418 phosphorylation. Since the occurrence of hemorrhage due to dasatinib treatment is associated with mucosal inflammation<sup>8</sup> and platelet GPIIb/IIIa function is known to be crucial to prevent bleeding when vascular integrity is jeopardized in the inflamed vasculature,<sup>9</sup> a mouse model of acute lung injury was used to better explore the underlying molecular mechanisms. This *in vivo* model of lipopolysaccharide (LPS)-induced mucosal inflammation confirmed the clinical observation of enhanced inflammatory hemorrhage in patients treated with dasatinib.<sup>10</sup> Importantly, neither systemic platelet counts nor the number of recruited platelets or neutrophils were changed. Furthermore, in an LPS-induced sepsis mouse model, 4-dimensional confocal intravital microscopy of mesenteric vessels revealed fewer migrating platelets in the dasatinib-treated group. Importantly, Kaiser et al confirmed the translational relevance of their findings by a pilot study, analyzing the platelets of patients with CML treated with dasatinib or bosutinib compared with those receiving imatinib, which does

not inhibit c-Src. Although platelet spreading on fibrinogen matrices was unaffected, the migratory capacity of platelets from patients with CML treated with c-Src inhibitor was vastly reduced.

Altogether, this translational study by Kaiser et al demonstrate that the GPIIb/Gα<sub>13</sub>/c-Src/14-3-3ζ signaling axis is essential for platelet migration, but in CML and ALL therapies the blockade of this pathway comes with the risk of inflammatory hemorrhage due to impaired platelet migration.

**Conflict-of-interest disclosure:** The authors declare no competing financial interests. ■

## REFERENCES

- Kaiser R, Anjum A, Kammerer L, et al. Mechanosensing via a GPIIb/Src/14-3-3ζ axis critically regulates platelet migration in vascular inflammation. *Blood*. 2023;141(24):2973-2992.
- Calaminus SDJ, Thomas S, McCarty OJT, Machesky LM, Watson SP. Identification of a novel, actin-rich structure, the actin nodule, in the early stages of platelet spreading. *J Thromb Haemost*. 2008;6(11):1944-1952.
- Baumann J, Sachs L, Otto O, et al. Reduced platelet forces underlie impaired hemostasis in mouse models of MYH9-related disease. *Sci Adv*. 2022;8(20):eabn2627.
- Falet H, Hoffmeister KM, Neujahr R, et al. Importance of free actin filament barbed ends for Arp2/3 complex function in platelets and fibroblasts. *Proc Natl Acad Sci U S A*. 2002;99(26):16782-16787.
- Nicolai L, Schiefelbein K, Lipsky S, et al. Vascular surveillance by haptotactic blood platelets in inflammation and infection. *Nat Commun*. 2020;11(1):5778.
- Gaertner F, Ahmad Z, Rosenberger G, et al. Migrating platelets are mechanoscavengers that collect and bundle bacteria. *Cell*. 2017;171(6):1368-1382.e23.
- Vielreicher M, Harms G, Butt E, Walter U, Obergfell A. Dynamic interaction between Src and C-terminal Src kinase in integrinIIbβ3-mediated signaling to the cytoskeleton. *J Biol Chem*. 2007;282(46):33623-33631.
- Apperley JF, Cortes JE, Kim D-W, et al. Dasatinib in the treatment of chronic myeloid leukemia in accelerated phase after imatinib failure: the START a trial. *J Clin Oncol*. 2009;27(21):3472-3479.
- Deppermann C. Platelets and vascular integrity. *Platelets*. 2018;29(6):549-555.
- Kaiser R, Escaig R, Kranich J, et al. Procoagulant platelet sentinels prevent inflammatory bleeding through GPIIb/IIIa and GPVI. *Blood*. 2022;140(2):121-139.

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