

TO THE EDITOR:

CREBBP alterations are associated with a poor prognosis in de novo AML

Adam J. Lamble,¹ Kohei Hagiwara,² Robert B. Gerbing,³ Jenny L. Smith,⁴ Pandurang Kolekar,² Rhonda E. Ries,⁴ Edward A. Kolb,⁵ Todd A. Alonzo,^{3,6} Xiaotu Ma,² and Soheil Meshinchi⁴

¹Division of Hematology and Oncology, Seattle Children's Hospital, Seattle, WA; ²Department of Computational Biology, St. Jude Children's Research Hospital, Memphis, TN; ³Children's Oncology Group, Monrovia, CA; ⁴Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, WA; ⁵Division of Oncology, Nemours Alfred I. duPont Hospital for Children, Wilmington, DE; and ⁶Keck School of Medicine, University of Southern California, Los Angeles, CA

The *cyclic adenosine monophosphate response element-binding protein (CREBBP)* gene is located on chromosome 16p13 and encodes a histone acetyltransferase having the same name that is involved in transcriptional regulation and cell cycle control.^{1,2} The translocation t(8;16)(p11;p13)[KAT6A::CREBBP] results in the disruption of *CREBBP* as well as its fusion to *KAT6A*, another gene important in transcription control. This fusion is sufficient for leukemogenesis and leads to a rare but well described type of acute myeloid leukemia (AML) with consistent biologic characteristics and a distinct gene expression profile.³⁻⁸ Although generally associated with inferior outcomes among adults, including a recent adjustment made by the European LeukemiaNet toward the adverse-risk group, there are variable reports regarding the prognostic significance of this fusion among pediatric patients.^{4,7,9} This prognostic variability is partially related to the high association of *KAT6A::CREBBP* with congenital leukemia as well as the phenomenon of spontaneous regression in a majority of these patients.^{4,10-17} Because of the poor outcomes associated with this alteration on North American protocols, patients aged >90 days with *KAT6A::CREBBP* have been reclassified as high risk in the active Phase 3 Children's Oncology Group (COG) trial for de novo AML, AAML1831 (NCT04293562). Little is known about the prognostic significance of *CREBBP* sequence variants, including pathogenic single nucleotide variants (SNVs) and insertion/deletions (indels). In this letter, we report the prevalence and prognostic significance of all alterations involving *CREBBP* in pediatric patients with AML and their impact on co-occurring lesions, specifically t(8;21)[*RUNX1::RUNX1T1*].

CREBBP variant status was determined in a total of 2216 patients enrolled in 4 successive COG trials for de novo pediatric AML (NCT00003790, NCT00070174, NCT01407757, and NCT01371981) and associated with comprehensive clinical and cytogenetic information. Fusions involving *CREBBP* were prospectively obtained via conventional cytogenetics and retrospectively confirmed via RNA sequencing. Indels and SNVs were retrospectively interrogated via next generation sequencing¹⁸ and their pathogenicities were established in accordance with American College of Medical Genetics and Genomics and the Association for Molecular Pathology guidelines. For patients with transcriptome data available, unsupervised clustering was done and their results were compared with those of patients without *CREBBP* alterations. The research was approved by the appropriate review boards and conducted in accordance with the Declaration of Helsinki. The Kaplan-Meier method was used to determine overall survival (OS) and event-free survival (EFS). The significance of predictor variables was tested using the log-rank

statistic. The significance of observed difference in proportions was analyzed using Fisher exact test, and observed differences in medians were analyzed using the Kruskal-Wallis test. The Cox proportional hazards model was used to estimate a hazard ratio for EFS in a multivariable analysis.

We identified 52 (2.3%) patients with a pathogenic alteration involving *CREBBP*, which is slightly higher than the incidence in publicly available adult AML databases (1.1%).^{19,20} This higher incidence is largely driven by fusions involving *CREBBP (CREBBP/fus)*, which made up 31% (n = 16) of our cohort (compared with 12.5% in the adult database). The remaining 36 patients (69%) in our cohort had a *CREBBP* mutation (*CREBBP/mut*; supplemental Table 1, available on the *Blood* website). Eighteen of these were due to an indel (*CREBBP/indel*), including 15 leading to frameshift mutations and 3 leading to deleterious inframe insertions or deletions (Figure 1A). The other 18 were due to a SNV (*CREBBP/SNV*), including 17 deleterious missense or nonsense mutations and 1 splice-site mutation (Figure 1A). A single patient had 2 pathogenic missense mutations. Patients with *CREBBP/fus* were younger than patients with *CREBBP/indel* or *CREBBP/SNV* (median ages of 2.6 vs 8.7 vs 7.4 years, respectively; *P* = .056), including 4 patients with a *CREBBP/fus* diagnosed in the first 90 days of life. There was a higher prevalence of *RUNX1::RUNX1T1* in patients with *CREBBP/indel* (n = 8) than in patients with *CREBBP/SNV* (n = 1) (44.4% vs 5.6%, respectively; *P* = .018; Figure 1A). In contrast, there was no association between *CREBBP/SNV* and a specific cytogenetic abnormality. There was a similar prevalence of cooccurring contemporarily defined high-risk lesions among patients with *CREBBP/indel* (n = 4; *CBFA2T3::GLIS2*, *KMT2A::AFF1*, *MLL10::PICALM*, and *NUP98::HOXA9*) and those with *CREBBP/SNV* (n = 5; *CBFA2T3::GLIS2*, *FUS::ERG*, *NUP98::KDM5A*, *ETV6::MNX1*, and high allelic ratio *FLT3-ITD*). There was a paucity of cooccurring genomic mutations in patients with *CREBBP/fus* (Figure 1A). In contrast, genomic mutations most frequently cooccurring in patients with *CREBBP/mut* included mutations in *RAS* (25%), *KIT* (13.9%), and *WT1* (11.1%). The frequency of these mutations were the same between patients with *CREBBP/SNV* and patients with *CREBBP/indel* and had a similar distribution to that of patients without *CREBBP* mutations. Transcriptome data were analyzed, and, consistent with previous reports, unsupervised clustering showed tight clustering of *CREBBP/fus* as well as a similar but distinct gene expression profile to that of *KMT2A* rearrangements.^{3,4} *CREBBP/indel* clustered together, which was likely driven by the cooccurrence of *RUNX1::RUNX1T1*, given its strong gene expression profile.²¹ In contrast, *CREBBP/SNV* did not show a specific clustering pattern (Figure 1B).

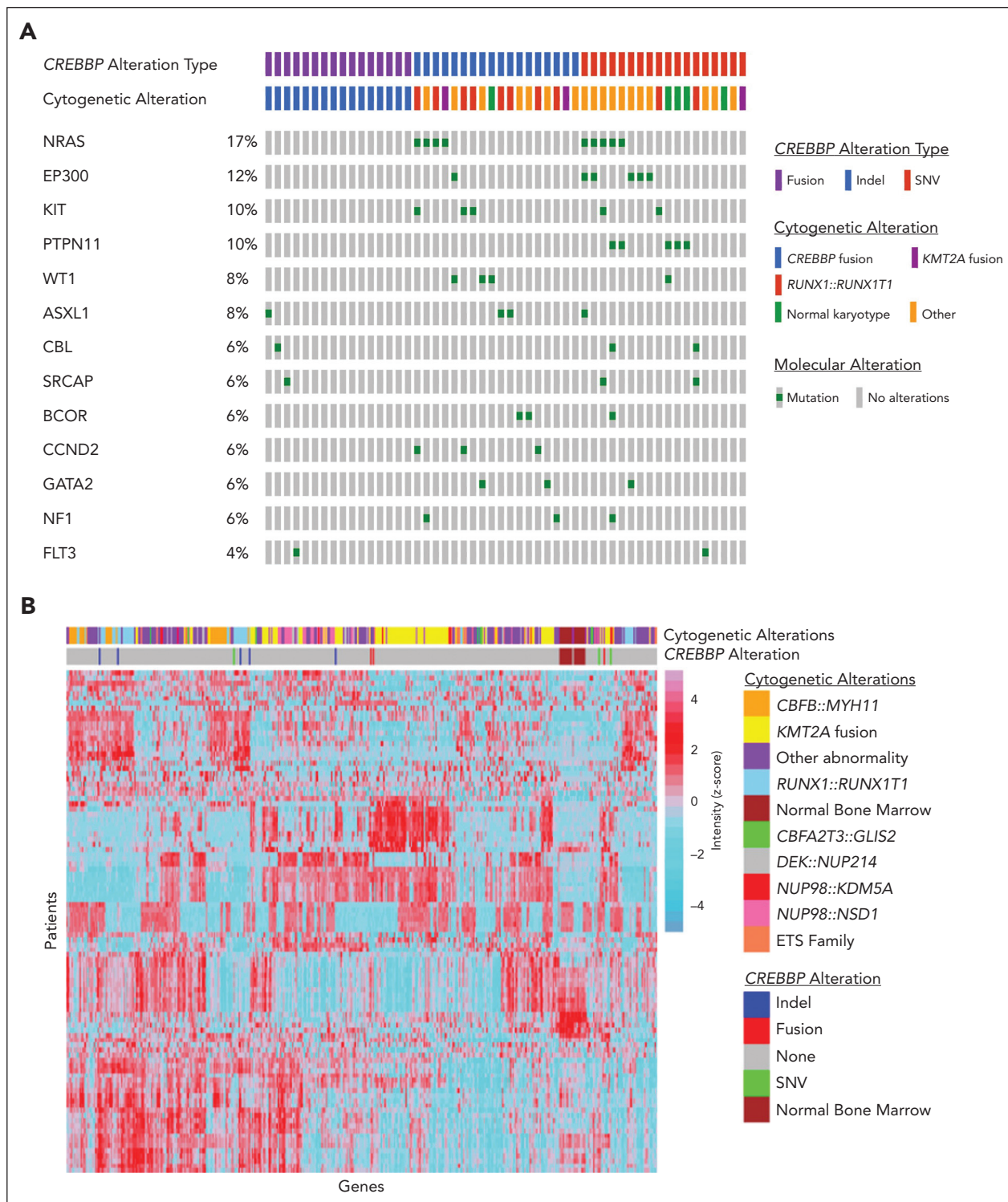


Figure 1. Molecular and transcriptome profiles based on CREBBP status. (A) Cytomolecular status and cooccurring mutational profile of patients based on CREBBP alterations, (B) Unsupervised clustering was performed across the entire transcriptome patient data set ($n = 1079$). Variance stabilizing transformation was performed (R function, vst), and scaled values were used to calculate the variance for each gene across all samples (R function, var). The genes were sorted based on variance, and the top 100 genes with highest variance were clustered (R function, ward.D2) and presented visually in a heatmap generated using pheatmap function in R.

Evaluation of outcomes based on the type of alteration demonstrated a similar EFS between patients with CREBBP/fus and patients with CREBBP/mut (5-year EFS was 33.3% vs 22.2%, respectively; $P = .801$; Figure 2A). Patients with CREBBP/mut

had a significantly worse EFS compared with patients without (5-year EFS was 22.2% vs 45.2%, respectively; $P = .005$; Figure 2A), and this inferior EFS is similar to that of patients with contemporarily defined high-risk but without CREBBP/mut

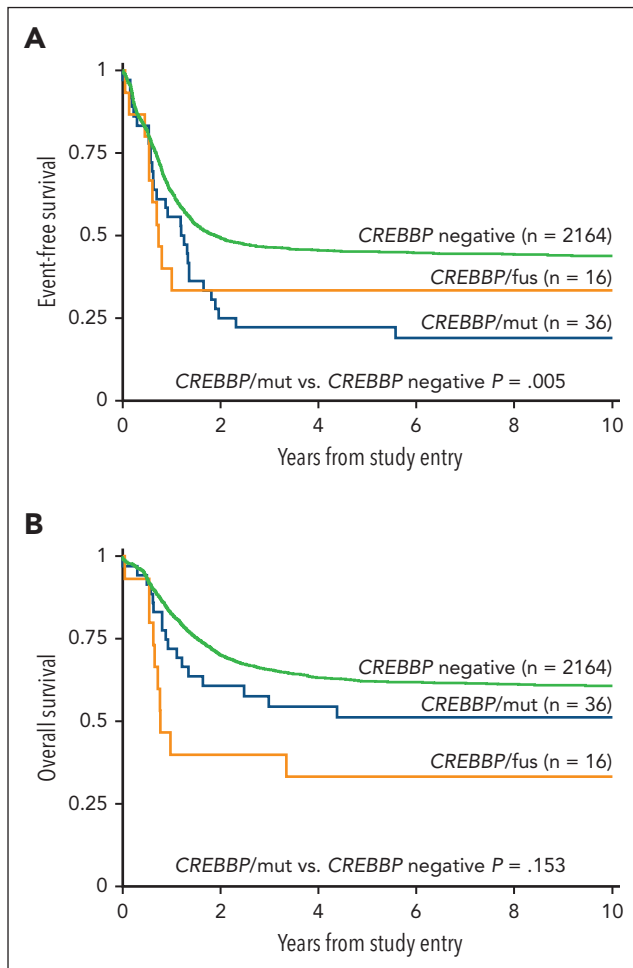


Figure 2. Correlation of clinical outcomes with CREBBP status. (A) EFS based on CREBBP status, (B) OS based on CREBBP status.

(5-year EFS is 25.2%; $P = .929$). Furthermore, this poor EFS was maintained in patients with *CREBBP*/indel with cooccurring *RUNX1::RUNX1T1*, which was significantly worse than that of patients with *RUNX1::RUNX1T1* but without *CREBBP*/indel (5-year EFS was 12.5% vs 67.7%, respectively; $P = .001$). When patients with cooccurring high-risk lesions were excluded from the analysis, the remaining patients with *CREBBP*/mut ($n = 27$) who would otherwise be considered low-risk maintained their poor EFS (5-year EFS was 29.6%). We performed a Cox regression analysis and found *CREBBP*/mut was an independent predictor of an inferior EFS in the presence of contemporarily defined cooccurring low- or high-risk cytogenetic alterations (hazard ratio, 1.71; 95% confidence interval, 1.18-2.47; $P = .005$; supplemental Table 2). Despite their poor EFS, patients with *CREBBP*/mut had a comparable OS with those without *CREBBP* disruptions (5-year OS was 51.4% vs 62.4%, respectively; $P = .153$, Figure 2B), demonstrating that these patients could successfully be salvaged following relapse. In contrast, patients with *CREBBP*/mut had a comparable OS with those without *CREBBP* disruptions (5-year OS was 51.4% vs 62.4%, respectively; $P = .153$, Figure 2B), demonstrating that these patients could successfully be salvaged following relapse. In contrast, patients with *CREBBP*/fus had an inferior OS (5-year OS was 33.3%; $P = .002$). Strikingly, all patients with *CREBBP*/fus that experienced a relapse ultimately died from their disease. Of the 5 long-term survivors with *CREBBP*/fus, 2 were diagnosed within the first 90 days of life and might have ultimately benefited from spontaneous regression.

We report the prevalence of *CREBBP*/fus in a large cohort of patients with pediatric AML with an incidence similar to that reported in a prior analysis.⁴ This is also the first report, to our knowledge, to comprehensively report the prevalence of *CREBBP*/mut in pediatric AML. Furthermore, we show that these patients have a poor EFS, regardless of the alteration type. This was especially notable in those patients with *CREBBP*/indel with a cooccurring *RUNX1::RUNX1T1*. The favorable EFS typically conferred by *RUNX1::RUNX1T1* was abrogated by the cooccurrence of *CREBBP*/indel. Finally, the presence of a *CREBBP*/mut maintained independent prognostic significance for an inferior EFS, within the context of a Cox regression analysis.

Translocations between *CREBBP* and *KAT6A* in patients aged >90 days are considered as high risk in the active COG phase 3 trial. Survival of patients with relapsed *CREBBP*/fus AML is dismal and warrants novel interventions. Histone deacetylase inhibitors have shown preclinical promise in *CREBBP* mutated tumors, and further studies in AML are warranted.^{22,23} Given the comparably poor EFS and high salvage rates associated with *CREBBP*/mut AML, intensification of upfront treatment, including hematopoietic stem cell transplant, should be considered in this population.

Acknowledgments

This work was supported by grants from the St Baldrick's Foundation; the National Cancer Institute's National Clinical Trials Network Operations Center (U10CA180886) and National Clinical Trials Network Statistics & Data Center (U10CA180899). The authors also thank Bayer, Jazz Pharmaceuticals, and Astellas Pharma Global Development for their support of CCG2961, AAML03P1, AAML0531, and AAML1031.

The content of this article is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Authorship

Contribution: A.J.L., R.E.R., J.L.S., and S.M. designed the research; A.J.L., K.H., R.E.R., J.L.S., S.M., T.A.A., R.B.G., P.K., E.A.K., and X.M. analyzed the data and wrote the paper.

Conflict-of-interest disclosure: E.A.K. reports funding from Roche/Genentech for travel and accommodation expenses. The remaining authors declare no competing financial interests.

ORCID profile: P.K., 0000-0003-0044-0076.

Correspondence: Adam J. Lamble, Division of Hematology/Oncology, University of Washington, Seattle Children's Hospital, Seattle, WA; email: adam.lamble@seattlechildrens.org.

Footnotes

Submitted 24 June 2022; accepted 4 January 2023; prepublished online on *Blood* First Edition 12 January 2023.

The online version of this article contains a data supplement.

REFERENCES

- Chan HM, La Thangue NB. p300/CBP proteins: HATs for transcriptional bridges and scaffolds. *J Cell Sci*. 2001;114(Pt 13):2363-2373.
- Shiama N. The p300/CBP family: integrating signals with transcription factors and chromatin. *Trends Cell Biol*. 1997;7(6):230-236.

3. Camos M, Esteve J, Jares P, et al. Gene expression profiling of acute myeloid leukemia with translocation t(8;16)(p11;p13) and MYST3-CREBBP rearrangement reveals a distinctive signature with a specific pattern of HOX gene expression. *Cancer Res.* 2006;66(14):6947-6954.
4. Coenen EA, Zwaan CM, Reinhardt D, et al. Pediatric acute myeloid leukemia with t(8;16)(p11;p13), a distinct clinical and biological entity: a collaborative study by the International-Berlin-Frankfurt-Munster AML-study group. *Blood.* 2013;122(15):2704-2713.
5. Diaz-Beya M, Navarro A, Ferrer G, et al. Acute myeloid leukemia with translocation (8;16)(p11;p13) and MYST3-CREBBP rearrangement harbors a distinctive microRNA signature targeting RET proto-oncogene. *Leukemia.* 2013;27(3):595-603.
6. Gervais C, Murati A, Helias C, et al. Acute myeloid leukaemia with 8p11 (MYST3) rearrangement: an integrated cytologic, cytogenetic and molecular study by the groupe francophone de cytogenetique hematologique. *Leukemia.* 2008;22(8):1567-1575.
7. Haferlach T, Kohlmann A, Klein HU, et al. AML with translocation t(8;16)(p11;p13) demonstrates unique cytomorphological, cytogenetic, molecular and prognostic features. *Leukemia.* 2009;23(5):934-943.
8. Xie W, Hu S, Xu J, Chen Z, Medeiros LJ, Tang G. Acute myeloid leukemia with t(8;16)(p11.2;p13.3)/KAT6A-CREBBP in adults. *Ann Hematol.* 2019; 98(5):1149-1157.
9. Dohner H, Wei AH, Appelbaum FR, et al. Diagnosis and management of AML in adults: 2022 recommendations from an international expert panel on behalf of the ELN. *Blood.* 2022;140(12):1345-1377.
10. Barrett R, Morash B, Roback D, et al. FISH identifies a KAT6A/CREBBP fusion caused by a cryptic insertional t(8;16) in a case of spontaneously remitting congenital acute myeloid leukemia with a normal karyotype. *Pediatr Blood Cancer.* 2017;64(8):e26450.
11. Classen CF, Behnisch W, Reinhardt D, Koenig M, Moller P, Debatin KM. Spontaneous complete and sustained remission of a rearrangement CBP (16p13)-positive disseminated congenital myelosarcoma. *Ann Hematol.* 2005;84(4):274-275.
12. Dinulos JG, Hawkins DS, Clark BS, Francis JS. Spontaneous remission of congenital leukemia. *J Pediatr.* 1997;131(2):300-303.
13. Hanada T, Ono I, Minosaki Y, Moriyama N, Nakahara S, Ohtsu A. Translocation t(8;16)(p11;p13) in neonatal acute monocytic leukaemia. *Eur J Pediatr.* 1991;150(5):323-324.
14. Liu M, Ren Y, Wang X, et al. Two rare cases of acute myeloid leukemia with t(8;16)(p11.2;p13.3) and 1q duplication: case presentation and literature review. *Mol Cytogenet.* 2020;13(1):1-9.
15. Wong KF, Yuen HL, Siu LL, Pang A, Kwong YL. t(8;16)(p11;p13) predisposes to a transient but potentially recurring neonatal leukemia. *Hum Pathol.* 2008;39(11):1702-1707.
16. Wu X, Sulavik D, Roulston D, Lim MS. Spontaneous remission of congenital acute myeloid leukemia with t(8;16)(p11;13). *Pediatr Blood Cancer.* 2011;56(2):331-332.
17. Andrade FG, Noronha EP, Baseggio RM, et al. Identification of the MYST3-CREBBP fusion gene in infants with acute myeloid leukemia and hemophagocytosis. *Rev Bras Hematol Hemoter.* 2016;38(4):291-297.
18. Hagiwara K, Ding L, Edmonson MN, et al. RNAIndel: discovering somatic coding indels from tumor RNA-Seq data. *Bioinformatics.* 2020;36(5): 1382-1390.
19. Cancer Genome Atlas Research N, Ley TJ, Miller C, et al. Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. *N Engl J Med.* 2013;368(22):2059-2074.
20. Tyner JW, Tognon CE, Bottomly D, et al. Functional genomic landscape of acute myeloid leukaemia. *Nature.* 2018;562(7728):526-531.
21. Ross ME, Mahfouz R, Onciu M, et al. Gene expression profiling of pediatric acute myelogenous leukemia. *Blood.* 2004;104(12):3679-3687.
22. Hellwig M, Merk DJ, Lutz B, Schuller U. Preferential sensitivity to HDAC inhibitors in tumors with CREBBP mutation. *Cancer Gene Ther.* 2020; 27(5):294-300.
23. Mondello P, Tadros S, Teater M, et al. Selective inhibition of HDAC3 targets synthetic vulnerabilities and activates immune surveillance in lymphoma. *Cancer Discov.* 2020;10(3):440-459.

<https://doi.org/10.1182/blood.2022017545>

© 2023 by The American Society of Hematology