Lulla PD. CAR T cells and autologous transplantation can coexist for DLBCL. *Blood*. 2021;139(9):1266-1267.

Page 1266: In the paragraph that begins with "Results from," the passage "In practice, however, we will still see patients who have received 1 to 2 cycles of salvage chemotherapy (not allowed in ZUMA7 or TRANSFORM) before consideration of a cell therapy, who do not fit the early chemotherapy failure criteria in those trials (ie, refractory to or relapsed within 12 months of first-line chemotherapy), or who cannot readily access a CAR T-cell center. Indeed, the EFS was not different between the standard of care and CAR T cells in the BELINDA trial, which had a design similar to those of ZUMA-7 and TRANSFORM except that it permitted salvage chemotherapy prior to administration of another potent CD19 CAR T-cell product, tisagenlecleucel.⁸" should read, "In practice, however, we will still see patients who have received salvage chemotherapy (not allowed in ZUMA7 and only 1 cycle in TRANSFORM, which is not typical salvage) before consideration of a cell therapy, who do not fit the early chemotherapy failure criteria in those trials (ie, refractory to or relapsed within 12 months of first-line chemotherapy), or who cannot readily access a CAR T-cell center. Indeed, the EFS was not different between the standard of care and CAR T cells in the BELINDA trial, which had a design similar to those of ZUMA-7 and only 1 cycle in TRANSFORM, which is not typical salvage) before consideration of a cell therapy, who do not fit the early chemotherapy failure criteria in those trials (ie, refractory to or relapsed within 12 months of first-line chemotherapy), or who cannot readily access a CAR T-cell center. Indeed, the EFS was not different between the standard of care and CAR T cells in the BELINDA trial, which had a design similar to those of ZUMA-7 and TRANSFORM except that it permitted standard salvage chemotherapy (~48% of patients received 2 or more salvage cycles) prior to administration of another potent CD19 CAR T-cell product, tisagenlecleucel.⁸" The errors have been corrected in the online version of the article.

Errata

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Sangaletti S, Tripodo C, Chiodoni C, et al. Neutrophil extracellular traps mediate transfer of cytoplasmic neutrophil antigens to myeloid dendritic cells toward ANCA induction and associated autoimmunity. *Blood.* 2012;120(15):3007-3018.

Page 3011: In the left column of Figure 2C, the image for necrotic polymorphonuclear leukocytes (PMNs) stained with PKH-26 was inadvertently duplicated for apoptotic PMNs. In the legend to Figure 2, the sentence "Scale bars represent 5 μ m" should not appear in the description of panel A; the sentence "Scale bars represent 30 μ m" should not appear in the description of D and E, and the sentence "The scale bar represents 30 μ m" should be added to the description of panel F. The corrected Figure 2 and legend are shown below.

Check for updates



Figure 2. (A) NETs persistently interact with mDCs. In this live imaging experiment, agar-PMNs were seeded onto culture dishes and allowed to adhere 30 minutes before adding mDCs previously labeled with the membrane vital dye PKH-26. The cocultures were also performed in the presence of the DNA dye SYTOX green, which stains only PMNs undergoing NETosis. The coculture was maintained o/n in a humidified chamber under confocal microscope, acquiring a picture every 10 minutes to highlight any interaction between PMNs and mDCs. PMNs in the cellular mixture form NETs (turquoise arrows) and NETs interact with mDCs (red arrows). Black arrows indicate naive neutrophils. (B) Uploading of mDCs with NET components. After NET interaction, mDCs are uploaded with the neutrophil protein PR3 and MPO. In this live imaging experiment, labeled PKH-26 mDCs were added to agar-PMN in the presence of a mAb to PR3 or MPO directly conjugated with Alexa-488 dye. After 18 hours, cells were fixed with PFA 4% and observed under a confocal microscope. Scale bars represent 10 μm. One representative experiment of 5 performed. (C) Uploading of PR3 and MPO by mDCs tested in the presence of different type of PMN death. Apoptotic PMNs, generated by Fas triggering Ab and necrotic PMNs obtained by freeze and thaw, were compared with NETotic PMNs for their capability to transfer PR3 and MPO to mDCs in coculture experiments. Representative confocal IF analysis showing that necrotic PMNs almost failed to transfer neutrophil PR3 or MPO to mDCs contrary, PR3 and MPO were detectable as dotted green fluorescence in mDCs that interacted with NETotic PMNs and, to a lesser extent, in mDCs cultured with apoptotic PMNs. (D-E) Software-assisted micrograph quantification of mDCs uploading of PR3 (D) and MPO (E) performed on a total of 200 cells that were imaged by confocal microscopy. DCs cultured with naive PMNs, maintained alive by incubating them in the presence of 30% of FCS and DCs alone, were the negative controls for PR3 transfer and MPO. (F) Appearance

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