Inflammation and myeloid malignancy: quenching the flame

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Chronic inflammation with aging ("inflammaging") plays a prominent role in the pathogenesis of myeloid malignancies. Aberrant inflammatory activity affects many different cells in the marrow, including normal blood and stromal marrow elements and leukemic cells, in unique and distinct ways. Inflammation can promote selective clonal expansion through differential immune-mediated suppression of normal hematopoietic cells and malignant clones. We review these complex roles, how they can be understood by separating cell-intrinsic from extrinsic effects, and how this informs future clinical trials.

Introduction

Aging and chronic inflammation are tightly linked, and the term "inflammaging" has been coined to describe these intertwined processes.¹ Chronic exposure to exogenous pathogen associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs), endogenous proteins released from damaged cells, produces an "immune biography" that installs an environment of sterile inflammation.² The result is reduced adaptive immunity,³ enhanced proinflammatory reactions,⁴ and predisposition to clonal disorders and malignancies.^{5,6} Inflammaging also drives hematopoietic stem cell (HSC) dysfunction, causing a loss of HSC quiescence,⁷ reduced selfrenewal capacity,^{8,9} and myeloid differentiation bias.^{10,11} Inflammation drives pathogenesis in myeloid malignancies across the disease spectrum, including myeloproliferative neoplasms (MPNs),¹² myelodysplastic syndromes (MDS),^{13,14} and acute myeloid leukemia (AML).¹⁵

There is also evidence that clonal hematopoiesis (CH) causes inflammation, and inflammaging also likely provides a permissive environment for clonal expansion.¹⁶ In addition, chronic inflammatory conditions (eg, cardiovascular¹⁷ and rheumatologic¹⁸ diseases) are strongly associated with CH, though whether inflammation can initiate CH is less clear. Although these are distinct disease states (eg, MDS causes ineffective hematopoiesis, whereas AML is proliferative), there are commonalities in the pathways involved, and inflammation can drive disease progression.¹⁹ Beyond the marrow, systemic inflammatory disorders are also increasingly associated with myeloid malignancy (eg, vacuoles, E1-enzyme, X-linked, autoinflammatory, somatic [VEXAS] syndrome and MDS).²⁰ Approaches targeting inflammation are being rapidly translated into clinical trials.^{9,21,22}

In this article, we outline the complex role of inflammation in myeloid malignancies and, using examples from preclinical

models and the framework of cell-intrinsic vs extrinsic effects, emphasize that inflammation is not a monolithic entity in myeloid malignancies, but rather has a cytokine, signaling pathway, and cell context-dependent nature that is dynamic throughout the disease course. Understanding these dependencies will be crucial to successfully modulating inflammation for clinical benefit in myeloid malignancies.

Inflammation in myeloid malignancies: context matters

Here we provide an overview of the specific inflammatory pathways involved in myeloid malignancies, demonstrating how the malignant cells, the marrow stroma, and immune cells interact to establish the inflammatory environment (Figure 1A). This is nuanced by the fact that malignant hematopoietic stem and progenitor cells (HSPCs) can differentiate into mature immune cells (eg, natural killer [NK], myeloid derived suppressor cells [MDSCs] and others).²³⁻²⁵ Further, preleukemic mutations (eg, *DNMT3A, TET2*) that alter inflammatory signaling are transmitted to the entirety of myeloid and lymphoid cells in patients with MDS.^{23,26,27} Differentiated malignant cells can then elaborate inflammatory cytokines such as tumor necrosis factor (TNF), interleukin-6 (IL-6), and interferon-γ (IFN-γ).^{9,28}

Innate immune signaling is frequently aberrantly activated through Toll-like receptor (TLR) signaling in malignant myeloid cells.²⁹ TLRs, which recognize PAMPs and DAMPs, produce an inflammatory response upon activation in hematopoietic cells.^{29,30} In malignant myeloid cells, TLRs³¹ and downstream effectors (eg, *MYD88*,³² *IRAK1*,³³ *TRAF6*³⁴) are often expressed at higher levels or in alternate isoforms (eg, *IRAK4L*³⁵), and their intrinsic regulators (eg, *miR-145*, *miR-146a*³⁶) are downregulated. This activated TLR axis results in the secretion of several cytokines (eg, IL-1,³⁷ IL-6,³⁶ IL-8,³² TNF,³⁸ IFN-γ, granulocyte



Figure 1. The role of inflammation in myeloid malignancies. (A) Intrinsic leukemic stem cell (LSC) signaling as well as interactions with the microenvironment result in inflammation and a permissive environment for further clonal expansion. Cytokines secreted from the LSC or their progeny may promote clonal expansion and/or suppress normal HSCs. Immune and stromal cells can also respond to the LSC by secreting cytokines, which can have differential effect on the HSC and LSC. (B) Activation of inflammatory pathways in LSCs can be divided into cell-intrinsic and cell-extrinsic. Cell-intrinsic alterations can drive increased intracellular signalling through activated intracellular pathways, causing proliferative signalling and cytokine secretion. Cell-extrinsic factors refer to cytokines or other factors secreted by LSCs, immune cells, or the stroma which can (i) suppress normal HSCs. (ii) suppress the LSCs, or (iii) activate the LSCs to different degrees after interacting with their cellular receptors. MDSC, myeloid-derived stromal cell; MSC, mesenchymal stromal cell. Illustration by Vicky Earle.

colony stimulating factor³⁹) from malignant myeloid cells. IL-1 receptor signalling is linked with TLRs through the shared cytoplasmic Toll-IL-1-receptor domain and is also upregulated by TLR activation.⁴⁰ Overexpression of the IL-1 receptor accessory protein on HSCs has also been shown to increase AML cell viability and clonal output in patients.⁴¹

Another key innate immune component in malignant myeloid cells are nucleotide-binding domain and leucine-rich repeat (NLR) receptors. NLRs associate with nucleotide-binding domains to form the inflammasome complex, producing proinflammatory, lytic cell death (pyroptosis) on activation by PAMPs or DAMPs.⁴² The alarmin \$100A9, a DAMP secreted by stromal cells and found at high levels in MDS marrows,⁴³ can bind NLRs and activate the NLRP3 inflammasome, which drives clonal expansion and pyroptosis by activating caspase-1, thereby generating mature IL-1β and IL-18 in patients with MDS.⁴² The CD33 receptor, expressed on malignant cells and MDSCs, senses \$100A8/9 and can activate the inflammasome.⁴⁴ Overactivity of TLRs and the inflammasome, with other factors, results in a heightened inflammatory milieu in the MDS/AML marrow microenvironment.^{24,45-47} This inflammatory signaling generally promotes differential fitness of the malignant MDS/AML LSC over normal HSPCs, although this effect varies depending on the disease context.47

Our understanding of the role for cellular immunity in myeloid malignancies continues to evolve. MDS/AML demonstrate relatively low rates of somatic mutations,⁴⁸ and generating robust

T-cell responses to malignant myeloid cells with low neoantigen burdens can be challenging.49,50 However, the curative graftversus-leukemia effect from allogeneic hematopoietic stem cell transplant is mediated through T and NK cells,^{51,52} and inflammation may promote the graft-versus-leukemia effect.⁵³ Other new dimensions in this area are the advent of cellular thera- $\mathsf{pies}^{\mathsf{54-56}}$ and the recognition of the role for regulatory T cells (Tregs) in the MDS/AML marrow microenvironment.^{57,58} Tregs can facilitate or suppress CD8⁺ T-cell-mediated control of malignant clones.^{57,58} Tregs in lower risk MDS have more proinflammatory immune responses and effector-type cells, 57,58 whereas higher risk MDS has expanded Treg and MDSC compartments.^{57,59} Higher Treg infiltrates are correlated with worse outcomes in MDS/AML, possibly because of suppression of T cell-mediated immune surveillance.^{57,59-61} It is also becoming evident that macrophage and NK cells are crucial to immune surveillance of malignant myeloid cells. Overexpression of the macrophage inhibitory molecule CD47 on LSCs has been shown to drive disease progression,⁶² whereas MDS clone-derived mesenchymal stromal cells can inhibit NK-cell function and promote malignant clonal expansion.^{17,22,26,63}

Inflammation in myeloid malignancies can also occur outside the marrow space, with up to 7.4% of patients with MDS meeting criteria for a systemic autoinflammatory disease.⁶⁴ Autoinflammatory features are associated with acquired transcription factor mutations and abnormal karyotype.^{64,65} Recently, a new systemic autoinflammatory disease associated with low-grade MDS was described and named vacuoles, E1-enzyme, X-linked, autoinflammatory, somatic (VEXAS) syndrome, which arises from somatic mutations in the ubiquitin-activating enzyme UBA1.²⁰

Cell-intrinsic vs cell-extrinsic effects

One way to conceptualize inflammation is by separating cellintrinsic from cell-extrinsic effects, each of which may differentially influence the fate of individual malignant cells (Figure 1B). In the context of therapeutic targeting of inflammation, we refer to cellintrinsic effects as those that originate and act principally within the malignant cell (eg, somatic mutations, cytogenetic, epigenetic changes).⁶⁶ In contrast, we refer to cell-extrinsic effects as those that impinge on the malignant cell through an external signal such as cytokines. Extrinsic factors may arise from activated signals in the malignant cell, from normal hematopoietic cells, or cells in the marrow microenvironment. These signals may impinge on all these various cell types, thereby having a major impact on the cellular environment as well as the malignant cell. Thus, although therapeutic targeting of cell extrinsic factors, such as cytokines, may relieve cytopenias or dampen proliferative signals in malignant cells, the intrinsic drivers of myeloid malignancy would likely not be affected, and such an approach would not likely eliminate the malignant cell as a result. The corollary is that targeting tumor cell intrinsic drivers, although more likely to suppress the malignant clone, may also have wider toxicity issues unless the target is tumor specific, as these pathways have significant autonomous effects on the cell independent of activating cytokines. Though targeting cytokines may also have systemic effects, these are less likely to cause cellular toxicity.

Although from the standpoint of therapy separating intrinsic from extrinsic factor targeting provides a convenient conceptual framework, these effects are inextricably linked and interwoven. For example, spliceosome mutations in MDS induce innate immune signalling by altering splice isoform expression of intermediaries such as IRAK4,³⁵ MAP3K7, and CASP8.⁶⁷ Somatic TET2 mutations, frequently seen in CH,⁶⁸ also activate the NLRP3 inflammasome.⁶⁹ Loss of the intrinsic TRAF6 regulator miR-146a alters stem cell activity and promotes myeloid malignancy in mice,⁵ though this has recently been shown to also act as a tumor suppressor through MYC in myeloid malignancy.70 However, all the intrinsic perturbations described previously result in the release of one or more cytokines such as TNF, IL-6, IL-1, or others. Articulating these mechanistic distinctions are important to establish specific goals for therapeutic approaches (eg, reducing clonal expansion vs relieving cytopenias) and avoiding unexpected adverse events (eg, by blocking cytokines that suppress some malignant clones). Thus, inflammation can be targeted at multiple levels including the cytokine, receptor, and downstream pathways, but depending on the target or cell type affected, the overall global effect may be different.

Differential effects: the pathways and cytokines matter

In myeloid malignancies, multiple inflammatory pathways can be simultaneously activated, producing a multitude of signals that affect marrow cells in a variety of context-dependent and cellspecific ways. For example, activation of one inflammatory pathway might drive malignant clonal expansion, whereas a different cytokine induces global marrow dysfunction by suppressing HSPCs and LSCs, and another is simply a bystander. The role these pathways play can also shift with disease progression, for example, by exacerbating cytopenias in MDS²⁴ but regulating proliferation in AML.⁷¹ It is important to note that, although inflammation can drive clonal expansion, it is also part of the immune response that can suppress clonal expansion, and the multiple types of secreted cytokines can have differential effects on malignant and normal cells.

A diseased stroma and microenvironment, resulting from the influence of malignant cells or inflammaging, alters the cytokine milieu and promotes the relative fitness of malignant over normal cells.^{7,43} These cytokines can then differentially suppress normal hematopoiesis, facilitating the emergence of the malignant clone, and affect the marrow niche to different degrees.^{72,73} For normal HSPCs, chronic inflammation can permanently imprint HSPC differentiation programs, even after removal of the stimulus, suggesting that cell-intrinsic factors can perpetuate inflammatory phenotypes in the context of normal hematopoiesis.^{10,11} Cytokines can also simultaneously stimulate multiple pathways. In a mouse bone marrow failure model initiated by constitutive TIRAP expression, IFN-γ suppresses erythropoiesis and megakaryopoiesis possibly through direct interaction with the thrombopoietin receptor,^{28,74} but inhibits myelopoiesis by releasing high mobility group box 1 (HMGB1), which suppresses the endothelial niche.²⁸ Extrinsic stimuli such as infection also can drive IFN-y-induced CH proliferation in DNMT3A-deficient HSPCs.¹⁶

The discordant effects of inflammatory signaling is exemplified by IL-1, which expands myeloid progenitors in an AML xenograft, but suppresses normal progenitors.⁷⁵ Heterogeneity can also emerge during malignant clonal evolution. Recent data show that expression of inflammatory signalling genes (BST2, IFITM1, IFITM3) can promote clonal expansion of TP53-mutant LSCs in patients with MPNs, and appears necessary for AML transformation while simultaneously suppressing antecedent TP53-wild type clones present before AML transformation.¹² In contrast, secreted IL-10 and TGF- β from MDSCs suppress both malignant and normal hematopoiesis in the MDS context, producing global marrow dysfunction and cytopenias in mice.²⁴ However, reducing TGF-B activity by blocking the central adaptor protein Disabled-2 accelerates leukemic progression by increasing stem cell activity at the expense of mature progenitors in a mouse AML xenograft model.⁷¹ A TGF-β superfamily ligand trap that reduces downstream SMAD signalling (luspatercept) improves anemia in patients with low-risk MDS,⁷⁶ and no similar risk has been seen with this drug, though follow-up is short.⁷¹ The differences seen may also be due to the fact that Disabled-2 blockade also affects other intrinsic signaling pathways independent of TGF- β , with blockade at the receptor level being more specific.

In another MDS mouse model, IFN- γ depletion caused progression from marrow failure to a myeloproliferation.²⁸ Interestingly IFN- γ is also expressed at higher levels in MDS compared with AML patient samples.²⁸ In a mouse model of TRAF6-driven MDS, which results in marrow failure or acute leukemia, IL-6 deletion blocked marrow failure but not leukemic progression.³⁶ In contrast, inhibiting IL-6 prolonged survival in an AML xenograft model,⁷⁷ but had no effect in a mouse MPN model.⁷⁸ It may be that IL-6 blockade can prevent AML-induced anemia, prolonging survival, but in the MDS or MPN context, suppression of IL-6 may

Target	Molecule	Mechanism	Combination	Disease	Phase	Registration or PMID	Status
Cell-intrinsic targets							
TLR2	OPN-305	Anti-TLR2 MAB	HMA	MDS	1/2	NCT02363491	Completed
TLR4	Bortezomib	Proteasome inhibitor	NA	MDS	2	NCT01891968	Completed
TLR9	GNKG168	Small molecule agonist	None	AML/ALL	1	NCT01743807	Terminated
IRAK1	IRAK-Inh	Small molecule inhibitor	NA	MDS	Preclinical	23845443	NA
IRAK4	CA-4948	Inhibits long isoform	HMA or Venetoclax	MDS/AML	1/2a	NCT04278768	Recruiting
TRAF6	NSC697923	UBE2N cofactor inhibitor	NA	MDS/AML	Preclinical	Blood. 2016;128(22):579	NA
NLRP3	MCC-950	NACTH ATPase domain inhibitor	NA	MDS	Preclinical	31086327	NA
	MNS	Blocks cysteine modification	NA	MDS	Preclinical	24265316	NA
	CY-09	Blocks NLRP3 activation	NA	MDS	Preclinical	33765556	NA
	Ibrutinib	BTK inhibitor	HMA	MDS	1b	NCT02553941	Active
Caspase-1	VX-765	Peptidomimetic drug	NA	Epilepsy	2	NCT01048255	Completed
				Psoriasis		NCT00205465	Completed
cGAS- STING	Pending	NA	NA	MDS/AML	Preclinical	33329537	NA
Cell-extrinsic targets							
TGF-β	Luspatercept	Ligand trap	None	MDS	3	31914241	Published
IL-1β	Canakinumab	MAB	HMA	MDS	2	NCT04239157	Recruiting
IL-6	Siltuximab	MAB	NA	MDS/AML	Preclinical	32269167	NA
	Tocilizumab						
HMGB1	CX-01	Small molecule inhibitor	HMA	MDS/AML	1	NCT02995655	Completed
S100A8/9	Pending	NA	NA	MDS/AML	Preclinical	27666011	NA

BTK, Bruton's tyrosine kinase; cGAS-STING, cyclic GMP-AMP synthase stimulator of interferon genes; HMA, hypomethylating agent; MAB, monoclonal antibody; NA, not available; TLR, toll-like receptor.

Current from 30 November 2021.

repress an inhibitory signal resulting in myeloproliferation and disease progression. These findings emphasize that inflammation likely suppresses some malignant clones in addition to normal hematopoiesis,⁹ and relieving cytokine-induced suppression may permit expansion and evolution of independent preleukemic or leukemic clones.⁷⁹

How can we target inflammation in myeloid malignancies?

Several agents are under investigation that target cell-intrinsic innate immune pathways, such as the TLR axis (Table 1). One example is the *IRAK4*-long isoform (*IRAK4L*). *U2AF1* mutations in AML generate *IRAK4L*, which hyperactivates NF- κ B, driving proliferation in a cell-intrinsic manner.³⁵ An IRAK4L inhibitor (CA-4948) blocked leukemic proliferation in an AML model, and

is in early-phase trials (NCT04278768), with early results showing tolerability.^{35,80,81} Other IRAK inhibitors (eg, IRAK1) are being developed.^{33,82} Inhibitors of UBE2N/Ubc13, an essential E2 ubiquitin-conjugating enzyme required for TRAF6 signalling, are under investigation.⁸³ Blocking NLRP3 inflammasome signaling is another appealing cell-intrinsic approach, given its importance in MDS/AML pathogenesis. Many early-stage NLRP3 inhibitors have been developed targeting the NACHT ATPase domain (MCC-950),⁸⁴ ATPase activity by cysteine modification (MSN),⁸⁵ and ATPase activity by blocking NLRP3 activation (CY-09) and pyroptosis, though few clinical data are available about these drugs.⁸⁶ Whether inhibition of pyroptosis is a targetable strategy in MDS is open to debate, as in another mouse model, genetic targeting of caspase-1 had no impact on marrow failure.²⁸ The cyclic GMP-AMP synthase stimulator of interferon genes pathway is another overactive intrinsic pathway with inhibitors in early development.⁸⁷ Targeting receptors may block cell-extrinsic or cell-intrinsic effects by blocking autocrine activation. Direct TLR2 inhibition (OPN-305, NCT03337451) has undergone a phase 1/2 trial, for which results are awaited.⁸⁸ It has also been suggested that targeting CD33 as a ligand for S100A8/9 may be an approach to block the inflammasome. Although CD33 has been targeted previously in AML, the rationale of this approach was not to explicitly block the CD33 and S100A8/9 interaction, but rather to eradicate blast cells.⁸⁹

Cell-extrinsic approaches are more mature in their development. Luspatercept, a ligand trap for TGF- β superfamily members, was approved after a phase 3 trial demonstrated reduced transfusion burden and good tolerance for lower risk MDS with ring sideroblasts or SF3B1 mutations.⁹⁰ Although luspatercept attenuates anemia, it does not alter disease course or prevent progression, suggesting it primarily alleviates suppression of normal erythropoiesis or promotes maturation in the context of ineffective erythropoiesis. Recent evidence suggests other lineages are also improved, pointing toward a global cell-extrinsic and stroma-modulating activity of luspatercept.^{91,92} One earlyphase trial attempted TNF- α blockade, though this has not entered later phase trials.⁹³ Surprisingly, given its importance in MDS/AML pathogenesis, few approaches targeting IL-1 receptor accessory protein have been tried.^{41,91,92,94} A phase 2 trial of canakinumab, an IL-1β-blocking monoclonal antibody that is well tolerated in other inflammatory conditions, has recently opened (NCT04239157), and the results are highly anticipated. Interestingly, an exploratory analysis suggested that the presence of CH predicts for cardiovascular benefit with canakinumab.⁹⁵ Another promising cytokine target is IL-6. Several studies have demonstrated IL-6 blockade might improve AML-induced anemia and possibly survival in mice, 9,77 though accelerated AML progression has also been observed.³⁶ Other cell-extrinsic approaches in early-stage investigation include targeting DAMPs such as S100A8/9^{43,96} and HMGB1.97

Immune-mediated therapies in MDS/AML are also currently used. Immunosuppressive therapy with horse antithymocyte globulin and cyclosporine produced an overall response rate of 48.8% in patients with hypoplastic MDS,^{98,99} and alemtuzumab up to 60% in younger, less transfused HLA-DR-15⁺ patients.¹⁰⁰ Although these approaches improve cytopenias and modulate the immunome, only hematopoietic stem cell transplant is disease-modifying.¹⁰¹⁻¹⁰³ Immunotherapeutic approaches have been tried in MDS/AML (eg, checkpoint inhibitors), with disappointing overall response rates.¹⁰⁴ However, targeting macrophage activity with an anti-CD47 antibody (magrolimab) and 5-azacitidine is promising, particularly in *TP53*-mutated MDS, and results are eagerly awaited.¹⁰⁵

Future directions

Inflammation plays a dynamic and heterogeneous role in the pathogenesis of myeloid malignancies that is both context- and patient-dependent. It can simultaneously drive or suppress clonal expansion and alter global marrow function through cell-extrinsic

and cell-intrinsic mechanisms. There remain many burning questions around which patients might benefit from these approaches and to what degree. For example, should we be targeting different inflammatory pathways in different disease states, or should these be patient-tailored? Eventually, a "precision medicine" approach might define patients who will benefit from specific immune-targeting therapies, based on disease features or overactive pathways. One other major conceptual question that remains unanswered is whether modulating inflammation can ever modify the disease course in myeloid malignancies. We suggest that blocking cell-extrinsic pathways primarily alleviates global marrow suppression, whereas targeting cell-intrinsic pathways is more likely to be disease-modifying, though possibly with more offtarget effects. Further, is targeting one pathway enough, or do we need multitargeting approaches? To move the field forward, we must think beyond inflammation as a general concept and focus on both the inflammatory context and actions of specific pathways and cytokines.

Acknowledgments

R.J.S. received funding from the Leukemia Lymphoma Society of Canada, Canadian Institutes of Health Research (202002LFC-439884), and the Clinician Investigator Program of the University of British Columbia. Work in the laboratory of A.K. is funded by grants from the Canadian Institutes of Health Research, the Terry Fox Research Institute, the Canadian Cancer Society Research Institute, Genome BC, Genome Canada, the Leukemia and Lymphoma Society of Canada, and the BC Cancer Foundation through the Leukemia and Myeloma Program. A.K. is the recipient of the BC Cancer Foundation John Auston Clinical Scientist Award. U.P. received funding from the Jose Carreras Leukemia Fund, the German Ministry of Education and Health, and the Jackstädt Foundation.

Authorship

Contribution: R.J.S., U.P., and A.K. contributed equally to conceiving, constructing, reviewing, and approving the final manuscript.

Conflict-of-interest disclosure: U.P. has received honoraria from Novartis and Curis. A.K. has received a grant-in-aid from AstraZeneca and an honorarium from Jazz Pharmaceuticals. R.J.S. declares no competing financial interests.

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Footnotes

Submitted 14 December 2021; accepted 22 March 2022; prepublished online on *Blood* First Edition 25 April 2022. DOI 10.1182/blood. 2021015162.

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REFERENCES

- Franceschi C, Bonafè M, Valensin S, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. Ann N Y Acad Sci. 2000;908:244-254.
- Crişan TO, Netea MG, Joosten LA. Innate immune memory: implications for host responses to damage-associated molecular patterns. Eur J Immunol. 2016;46(4): 817-828.
- de Mol J, Kuiper J, Tsiantoulas D, Foks AC. The dynamics of B cell aging in health and disease. Front Immunol. 2021;12:733566.
- Boren E, Gershwin ME. Inflamm-aging: autoimmunity, and the immune-risk phenotype. Autoimmun Rev. 2004;3(5): 401-406.
- Crusz SM, Balkwill FR. Inflammation and cancer: advances and new agents. Nat Rev Clin Oncol. 2015;12(10):584-596.
- Avagyan S, Henninger JE, Mannherz WP, et al. Resistance to inflammation underlies enhanced fitness in clonal hematopoiesis. *Science*. 2021;374(6568):768-772.
- Agarwal P, Isringhausen S, Li H, et al. Mesenchymal niche-specific expression of Cxcl12 controls quiescence of treatmentresistant leukemia stem cells. *Cell Stem Cell*. 2019;24(5):769-784.e6.
- de Haan G, Lazare SS. Aging of hematopoietic stem cells. *Blood.* 2018; 131(5):479-487.
- Grants JM, Wegrzyn J, Hui T, et al. Altered microRNA expression links IL6 and TNFinduced inflammaging with myeloid malignancy in humans and mice. *Blood*. 2020; 135(25):2235-2251.
- Mitroulis I, Ruppova K, Wang B, et al. Modulation of myelopoiesis progenitors is an integral component of trained immunity. *Cell.* 2018;172(1-2):147-161.e12.
- 11. Oduro KA Jr, Liu F, Tan Q, et al. Myeloid skewing in murine autoimmune arthritis occurs in hematopoietic stem and primitive progenitor cells. *Blood.* 2012;120(11): 2203-2213.
- Rodriguez-Meira A, Rahman H, Norfo R, et al. Single-cell multi-omics reveals the genetic, cellular, and molecular landscape of *TP53* mutated leukemic transformation in MPN. *Blood.* 2021;128(suppl 1):3.
- Trowbridge JJ, Starczynowski DT. Innate immune pathways and inflammation in hematopoietic aging, clonal hematopoiesis, and MDS. J Exp Med. 2021;218(7): e20201544.
- Sallman DA, List A. The central role of inflammatory signaling in the pathogenesis of myelodysplastic syndromes. *Blood*. 2019;133(10):1039-1048.
- Récher C. Clinical implications of inflammation in acute myeloid leukemia. Front Oncol. 2021;11:623952.
- Hormaechea-Agulla D, Matatall KA, Le DT, et al. Chronic infection drives Dnmt3a-lossof-function clonal hematopoiesis via IFNγ

signaling. Cell Stem Cell. 2021;28(8): 1428-1442.e6.

- Jaiswal S, Natarajan P, Silver AJ, et al. Clonal hematopoiesis and risk of atherosclerotic cardiovascular disease. N Engl J Med. 2017;377(2):111-121.
- Savola P, Lundgren S, Keränen MAI, et al. Clonal hematopoiesis in patients with rheumatoid arthritis. *Blood Cancer J.* 2018; 8(8):69.
- Stratmann S, Yones SA, Garbulowski M, et al. Transcriptomic analysis reveals proinflammatory signatures associated with acute myeloid leukemia progression. *Blood Adv.* 2022;6(1):152-164.
- Beck DB, Ferrada MA, Sikora KA, et al. Somatic mutations in UBA1 and severe adult-onset autoinflammatory disease. N Engl J Med. 2020;383(27):2628-2638.
- Winter S, Shoaie S, Kordasti S, Platzbecker U. Integrating the "immunome" in the stratification of myelodysplastic syndromes and future clinical trial design. J Clin Oncol. 2020;38(15):1723-1735.
- 22. Cook EK, Luo M, Rauh MJ. Clonal hematopoiesis and inflammation: partners in leukemogenesis and comorbidity. *Exp Hematol.* 2020;83:85-94.
- Vercauteren SM, Starczynowski DT, Sung S, et al. T cells of patients with myelodysplastic syndrome are frequently derived from the malignant clone. Br J Haematol. 2012;156(3):409-412.
- Chen X, Eksioglu EA, Zhou J, et al. Induction of myelodysplasia by myeloidderived suppressor cells. J Clin Invest. 2013;123(11):4595-4611.
- Kiladjian JJ, Bourgeois E, Lobe I, et al. Cytolytic function and survival of natural killer cells are severely altered in myelodysplastic syndromes. *Leukemia*. 2006;20(3):463-470.
- Busque L, Sun M, Buscarlet M, et al. Highsensitivity C-reactive protein is associated with clonal hematopoiesis of indeterminate potential. *Blood Adv.* 2020;4(11): 2430-2438.
- Janssen JW, Buschle M, Layton M, et al. Clonal analysis of myelodysplastic syndromes: evidence of multipotent stem cell origin. *Blood*. 1989;73(1):248-254.
- Gopal A, Ibrahim R, Fuller M, et al. TIRAP drives myelosuppression through an Ifnγ-Hmgb1 axis that disrupts the endothelial niche in mice. J Exp Med. 2022;219(3): e20200731.
- Paracatu LC, Schuettpelz LG. Contribution of aberrant Toll like receptor signaling to the pathogenesis of myelodysplastic syndromes. Front Immunol. 2020;11:1236.
- Cluzeau T, McGraw KL, Irvine B, et al. Proinflammatory proteins \$100A9 and tumor necrosis factor-α suppress erythropoietin elaboration in myelodysplastic syndromes. *Haematologica*. 2017;102(12):2015-2020.
- 31. Maratheftis CI, Andreakos E, Moutsopoulos HM, Voulgarelis M. Toll-like receptor-4 is

up-regulated in hematopoietic progenitor cells and contributes to increased apoptosis in myelodysplastic syndromes. *Clin Cancer Res.* 2007;13(4):1154-1160.

- Dimicoli S, Wei Y, Bueso-Ramos C, et al. Overexpression of the toll-like receptor (TLR) signaling adaptor MYD88, but lack of genetic mutation, in myelodysplastic syndromes. *PLoS One.* 2013;8(8):e71120.
- Rhyasen GW, Bolanos L, Fang J, et al. Targeting IRAK1 as a therapeutic approach for myelodysplastic syndrome. *Cancer Cell.* 2013;24(1):90-104.
- Hofmann WK, de Vos S, Komor M, Hoelzer D, Wachsman W, Koeffler HP. Characterization of gene expression of CD34⁺ cells from normal and myelodysplastic bone marrow. *Blood.* 2002;100(10):3553-3560.
- Smith MA, Choudhary GS, Pellagatti A, et al. U2AF1 mutations induce oncogenic IRAK4 isoforms and activate innate immune pathways in myeloid malignancies. *Nat Cell Biol.* 2019;21(5):640-650.
- Starczynowski DT, Kuchenbauer F, Argiropoulos B, et al. Identification of miR-145 and miR-146a as mediators of the 5q- syndrome phenotype. Nat Med. 2010; 16(1):49-58.
- Shi L, Zhao Y, Fei C, et al. Cellular senescence induced by S100A9 in mesenchymal stromal cells through NLRP3 inflammasome activation. Aging (Albany NY). 2019;11(21):9626-9642.
- Pronk CJ, Veiby OP, Bryder D, Jacobsen SE. Tumor necrosis factor restricts hematopoietic stem cell activity in mice: involvement of two distinct receptors. *J Exp Med.* 2011;208(8):1563-1570.
- Boettcher S, Gerosa RC, Radpour R, et al. Endothelial cells translate pathogen signals into G-CSF-driven emergency granulopoiesis. *Blood.* 2014;124(9): 1393-1403.
- Ho TC, Kawano H, LaMere M, et al. IL-1 via IRAK1/4 sustains acute myeloid leukemia stem cells following treatment and relapse. *Blood.* 2021;138(suppl 1):1175.
- Barreyro L, Will B, Bartholdy B, et al. Overexpression of IL-1 receptor accessory protein in stem and progenitor cells and outcome correlation in AML and MDS. *Blood.* 2012;120(6):1290-1298.
- Basiorka AA, McGraw KL, Eksioglu EA, et al. The NLRP3 inflammasome functions as a driver of the myelodysplastic syndrome phenotype. *Blood.* 2016;128(25): 2960-2975.
- Zambetti NA, Ping Z, Chen S, et al. Mesenchymal inflammation drives genotoxic stress in hematopoietic stem cells and predicts disease evolution in human pre-leukemia. *Cell Stem Cell*. 2016; 19(5):613-627.
- Eksioglu EA, Chen X, Heider KH, et al. Novel therapeutic approach to improve hematopoiesis in low risk MDS by targeting MDSCs with the Fc-engineered CD33

antibody BI 836858. *Leukemia.* 2017; 31(10):2172-2180.

- Vogl T, Tenbrock K, Ludwig S, et al. Mrp8 and Mrp14 are endogenous activators of Toll-like receptor 4, promoting lethal, endotoxin-induced shock. *Nat Med.* 2007; 13(9):1042-1049.
- Mundle SD, Venugopal P, Cartlidge JD, et al. Indication of an involvement of interleukin-1 beta converting enzyme-like protease in intramedullary apoptotic cell death in the bone marrow of patients with myelodysplastic syndromes. *Blood*. 1996; 88(7):2640-2647.
- Muto T, Walker CS, Choi K, et al. Adaptive response to inflammation contributes to sustained myelopoiesis and confers a competitive advantage in myelodysplastic syndrome HSCs. *Nat Immunol.* 2020;21(5): 535-545.
- Martincorena I, Campbell PJ. Somatic mutation in cancer and normal cells. *Science*. 2015;349(6255):1483-1489.
- Samstein RM, Lee CH, Shoushtari AN, et al. Tumor mutational load predicts survival after immunotherapy across multiple cancer types. Nat Genet. 2019;51(2):202-206.
- Roerden M, Nelde A, Walz JS. Neoantigens in hematological malignancies–ultimate targets for immunotherapy? *Front Immunol.* 2019;10:3004.
- Biernacki MA, Sheth VS, Bleakley M. T cell optimization for graft-versus-leukemia responses. JCI Insight. 2020;5(9):e134939.
- Van Elssen CHMJ, Ciurea SO. NK cell alloreactivity in acute myeloid leukemia in the post-transplant cyclophosphamide era. *Am J Hematol.* 2020;95(12):1590-1598.
- Ciavattone NG, Wu L, O'Neill R, Qiu J, Davila E, Cao X. MyD88 costimulation in donor CD8⁺ T cells enhances the graftversus-tumor effect in murine hematopoietic cell transplantation. *J Immunol.* 2021;206(4):892-903.
- Ferrari V, Tarke A, Fields H, et al. In vitro induction of neoantigen-specific T cells in myelodysplastic syndrome, a disease with low mutational burden. *Cytotherapy*. 2021; 23(4):320-328.
- Biernacki MA, Foster KA, Woodward KB, et al. CBFB-MYH11 fusion neoantigen enables T cell recognition and killing of acute myeloid leukemia. J Clin Invest. 2020;130(10):5127-5141.
- van der Lee DI, Reijmers RM, Honders MW, et al. Mutated nucleophosmin 1 as immunotherapy target in acute myeloid leukemia. J Clin Invest. 2019;129(2): 774-785.
- Kittang AO, Kordasti S, Sand KE, et al. Expansion of myeloid derived suppressor cells correlates with number of T regulatory cells and disease progression in myelodysplastic syndrome. Oncolmmunology. 2015;5(2):e1062208.
- Kordasti SY, Afzali B, Lim Z, et al. IL-17producing CD4(+) T cells, proinflammatory cytokines and apoptosis are

increased in low risk myelodysplastic syndrome. *Br J Haematol.* 2009;145(1):64-72.

- Sallman DA, McLemore AF, Aldrich AL, et al. TP53 mutations in myelodysplastic syndromes and secondary AML confer an immunosuppressive phenotype. *Blood.* 2020;136(24):2812-2823.
- Lamble AJ, Kosaka Y, Laderas T, et al. Reversible suppression of T cell function in the bone marrow microenvironment of acute myeloid leukemia. *Proc Natl Acad Sci* USA. 2020;117(25):14331-14341.
- Kahn JD, Chamuleau ME, Westers TM, et al. Regulatory T cells and progenitor B cells are independent prognostic predictors in lower risk myelodysplastic syndromes. *Haematologica*. 2015;100(6):e220-e222.
- Pang WW, Pluvinage JV, Price EA, et al. Hematopoietic stem cell and progenitor cell mechanisms in myelodysplastic syndromes. Proc Natl Acad Sci USA. 2013; 110(8):3011-3016.
- Sarhan D, Wang J, Sunil Arvindam U, et al. Mesenchymal stromal cells shape the MDS microenvironment by inducing suppressive monocytes that dampen NK cell function. *JCI Insight*. 2020;5(5):e130155.
- 64. Mekinian A, Grignano E, Braun T, et al. Systemic inflammatory and autoimmune manifestations associated with myelodysplastic syndromes and chronic myelomonocytic leukaemia: a French multicentre retrospective study. *Rheumatology (Oxford).* 2016;55(2): 291-300.
- 65. Watad A, Kacar M, Bragazzi NL, et al. Somatic mutations and the risk of undifferentiated autoinflammatory disease in MDS: an under-recognized but prognostically important complication. *Front Immunol.* 2021;12:610019.
- Sun D, Luo M, Jeong M, et al. Epigenomic profiling of young and aged HSCs reveals concerted changes during aging that reinforce self-renewal. *Cell Stem Cell*. 2014; 14(5):673-688.
- Pollyea DA, Harris C, Rabe JL, et al. Myelodysplastic syndrome-associated spliceosome gene mutations enhance innate immune signaling. *Haematologica*. 2019; 104(9):e388-e392.
- Jaiswal S, Fontanillas P, Flannick J, et al. Age-related clonal hematopoiesis associated with adverse outcomes. N Engl J Med. 2014;371(26):2488-2498.
- Fuster JJ, Zuriaga MA, Zorita V, et al. TET2-loss-of-function-driven clonal hematopoiesis exacerbates experimental insulin resistance in aging and obesity. *Cell Rep.* 2020;33(4):108326.
- Muto T, Guillamot M, Yeung J, et al. TRAF6 functions as a tumor suppressor in myeloid malignancies by directly targeting MYC oncogenic activity. *Cell Stem Cell.* 2022;29(2):298-314.e9.
- Lam J, van den Bosch M, Wegrzyn J, et al. miR-143/145 differentially regulate hematopoietic stem and progenitor activity

through suppression of canonical TGFβ signaling. *Nat Commun.* 2018;9(1):2418.

- Agarwal P, Li H, Choi K, et al. TNF-α-induced alterations in stromal progenitors enhance leukemic stem cell growth via CXCR2 signaling. *Cell Rep.* 2021;36(2):109386.
- Krause DS, Fulzele K, Catic A, et al. Differential regulation of myeloid leukemias by the bone marrow microenvironment. Nat Med. 2013;19(11):1513-1517.
- Alvarado LJ, Huntsman HD, Cheng H, et al. Eltrombopag maintains human hematopoietic stem and progenitor cells under inflammatory conditions mediated by IFN-γ. *Blood.* 2019;133(19):2043-2055.
- Carey A, Edwards DK V, Eide CA, et al. Identification of Interleukin-1 by functional screening as a key mediator of cellular expansion and disease progression in acute myeloid leukemia. *Cell Rep.* 2017;18(13): 3204-3218.
- Cappellini MD, Viprakasit V, Taher AT, et al; BELIEVE Investigators. A phase 3 trial of luspatercept in patients with transfusiondependent β-thalassemia. N Engl J Med. 2020;382(13):1219-1231.
- Zhang TY, Dutta R, Benard B, Zhao F, Yin R, Majeti R. IL-6 blockade reverses bone marrow failure induced by human acute myeloid leukemia. *Sci Transl Med.* 2020; 12(538):eaax5104.
- Baldauf CK, Müller P, Haage TR, et al. Anti-IL-6 cytokine treatment has no impact on elevated hematocrit or splenomegaly in a polycythemia vera mouse model. *Blood* Adv. 2022;6(2):399-404.
- Chen J, Kao YR, Sun D, et al. Myelodysplastic syndrome progression to acute myeloid leukemia at the stem cell level. Nat Med. 2019;25(1):103-110.
- Gummadi VR, Boruah A, Ainan BR, et al. Discovery of CA-4948, an orally bioavailable IRAK4 inhibitor for treatment of hematologic malignancies. ACS Med Chem Lett. 2020;11(12):2374-2381.
- Nowakowski GS, Leslie LA, Younes A, et al. Safety, pharmacokinetics and activity of CA-4948, an IRAK4 inhibitor, for treatment of patients with relapsed or refractory hematologic malignancies: results from the phase 1 study. *Blood.* 2020;136(suppl 1): 44-45.
- Kawano H, Kawano Y, LaMere M, et al. Interleukin-1/Toll-like receptor inhibition can restore the disrupted bone marrow microenvironment in mouse model of myelodysplastic syndromes. *Blood.* 2021; 138(suppl 1):1510.
- Barreyro L, Sampson AM, Bolanos K, et al. Inhibition of UBE2N as a therapeutic approach in myelodysplastic syndromes (MDS) and acute myeloid leukemia (AML). *Blood.* 2016;128(22):579.
- Coll RC, Hill JR, Day CJ, et al. MCC950 directly targets the NLRP3 ATP-hydrolysis motif for inflammasome inhibition. Nat Chem Biol. 2019;15(6):556-559.

- He Y, Varadarajan S, Muñoz-Planillo R, Burberry A, Nakamura Y, Núñez G. 3,4methylenedioxy-β-nitrostyrene inhibits NLRP3 inflammasome activation by blocking assembly of the inflammasome. *J Biol Chem*. 2014;289(2):1142-1150.
- Zhang Y, Lin Z, Chen D, He Y. CY-09 attenuates the progression of osteoarthritis via inhibiting NLRP3 inflammasomemediated pyroptosis. *Biochem Biophys Res Commun.* 2021;553:119-125.
- Liao W, Du C, Wang J. The cGAS-STING pathway in hematopoiesis and its physiopathological significance. *Front Immunol.* 2020;11:573915.
- Reilly M, Miller RM, Thomson MH, et al. Randomized, double-blind, placebo-controlled, dose-escalating phase I, healthy subjects study of intravenous OPN-305, a humanized anti-TLR2 antibody. *Clin Pharmacol Ther.* 2013;94(5):593-600.
- Maakaron JE, Rogosheske J, Long M, Bachanova V, Mims AS. CD33-targeted therapies: beating the disease or beaten to death? J Clin Pharmacol. 2021;61(1):7-17.
- Fenaux P, Platzbecker U, Mufti GJ, et al. Luspatercept in patients with lower-risk myelodysplastic syndromes. N Engl J Med. 2020;382(2):140-151.
- Wobus M, Mies A, Asokan N, et al. Luspatercept restores SDF-1-mediated hematopoietic support by MDS-derived mesenchymal stromal cells. *Leukemia*. 2021;35(10):2936-2947.
- Garcia-Manero G, Mufti GJ, Fenaux P, et al. Neutrophil and platelet increases with luspatercept in lower-risk MDS: secondary endpoints from the MEDALIST trial. *Blood*. 2022;139(4):624-629.

- Boula A, Voulgarelis M, Giannouli S, et al. Effect of cA2 anti-tumor necrosis factoralpha antibody therapy on hematopoiesis of patients with myelodysplastic syndromes. *Clin Cancer Res.* 2006;12(10): 3099-3108.
- Mitchell K, Barreyro L, Todorova TI, et al. IL1RAP potentiates multiple oncogenic signaling pathways in AML. J Exp Med. 2018;215(6):1709-1727.
- 95. Svensson E, Madar A, Campbell CD, et al. TET2-driven clonal hematopoiesis predicts enhanced response to canakinumab in the CANTOS trial: an exploratory analysis [published online ahead of print 6 April 2022]. JAMA Cardiol. doi:10.1161/circ.138. suppl_1.15111
- Chen BY, Song J, Hu CL, et al. SETD2 deficiency accelerates MDS-associated leukemogenesis via S100a9 in NHD13 mice and predicts poor prognosis in MDS. *Blood.* 2020;135(25):2271-2285.
- Kam AYF, Piryani SO, McCall CM, Park HS, Rizzieri DA, Doan PL. Targeting high mobility group box-1 (HMGB1) promotes cell death in myelodysplastic syndrome. *Clin Cancer Res.* 2019;25(13):4155-4167.
- Passweg JR, Giagounidis AA, Simcock M, et al. Immunosuppressive therapy for patients with myelodysplastic syndrome: a prospective randomized multicenter phase III trial comparing antithymocyte globulin plus cyclosporine with best supportive care – SAKK 33/99. J Clin Oncol. 2011; 29(3):303-309.
- Stahl M, DeVeaux M, de Witte T, et al. The use of immunosuppressive therapy in MDS: clinical outcomes and their predictors in a

large international patient cohort. Blood Adv. 2018;2(14):1765-1772.

- 100. Sloand EM, Olnes MJ, Shenoy A, et al. Alemtuzumab treatment of intermediate-1 myelodysplasia patients is associated with sustained improvement in blood counts and cytogenetic remissions. J Clin Oncol. 2010;28(35):5166-5173.
- Garcia-Manero G, Chien KS, Montalban-Bravo G. Myelodysplastic syndromes: 2021 update on diagnosis, risk stratification and management. Am J Hematol. 2020;95(11): 1399-1420.
- 102. Wermke M, Schuster C, Nolte F, et al. Mammalian-target of rapamycin inhibition with temsirolimus in myelodysplastic syndromes (MDS) patients is associated with considerable toxicity: results of the temsirolimus pilot trial by the German MDS Study Group (D-MDS). Br J Haematol. 2016;175(5):917-924.
- 103. Saber W, Horowitz MM. Transplantation for myelodysplastic syndromes: who, when, and which conditioning regimens. *Hematology Am Soc Hematol Educ Program.* 2016;2016:478-484.
- 104. Kapoor S, Champion G, Basu A, Mariampillai A, Olnes MJ. Immune therapies for myelodysplastic syndromes and acute myeloid leukemia. *Cancers* (*Basel*). 2021;13(19):5026.
- 105. Chao MP, Takimoto CH, Feng DD, et al. Therapeutic targeting of the macrophage immune checkpoint CD47 in myeloid malignancies. Front Oncol. 2020;9:1380.

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