#### TO THE EDITOR:

# COVID-19 in patients with CLL: improved survival outcomes and update on management strategies

Lindsey E. Roeker, <sup>1,\*</sup> Toby A. Eyre, <sup>2,\*</sup> Meghan C. Thompson, <sup>1</sup> Nicole Lamanna, <sup>3</sup> Alexander R. Coltoff, <sup>3</sup> Matthew S. Davids, <sup>4</sup> Peter O. Baker, <sup>4</sup> Lori Leslie, <sup>5</sup> Kerry A. Rogers, <sup>6</sup> John N. Allan, <sup>7</sup> Raul Cordoba, <sup>8</sup> Alberto Lopez-Garcia, <sup>8</sup> Darko Antic, <sup>9</sup> John M. Pagel, <sup>10</sup> Nicolas Martinez-Calle, <sup>11</sup> José Antonio García-Marco, <sup>12</sup> Jose-Ángel Hernández-Rivas, <sup>13</sup> Fatima Miras, <sup>14</sup> Catherine C. Coombs, <sup>15</sup> Anders Österborg, <sup>16</sup> Lotta Hansson, <sup>16</sup> Amanda N. Seddon, <sup>17</sup> Javier López Jiménez, <sup>18</sup> Matthew R. Wilson, <sup>19</sup> Dima El-Sharkawi, <sup>20</sup> Daniel Wojenski, <sup>21</sup> Shuo Ma, <sup>21</sup> Talha Munir, <sup>22</sup> Susana Valenciano, <sup>23</sup> Erlene Seymour, <sup>24</sup> Paul M. Barr, <sup>25</sup> Jeffrey Pu, <sup>26</sup> Piers E. M. Patten, <sup>27</sup> Guilherme F. Perini, <sup>28</sup> Scott F. Huntington, <sup>29</sup> Helen Parry, <sup>30</sup> Suchitra Sundaram, <sup>31</sup> Alan Skarbnik, <sup>32</sup> Manali Kamdar, <sup>33</sup> Ryan Jacobs, <sup>34</sup> Harriet Walter, <sup>35</sup> Renata Walewska, <sup>36</sup> Angus Broom, <sup>37</sup> Sonia Lebowitz, <sup>1</sup> Krista M. Isaac, <sup>38</sup> Craig A. Portell, <sup>38</sup> Inhye E. Ahn, <sup>39</sup> Chaitra S. Ujjani, <sup>40</sup> Mazyar Shadman, <sup>40</sup> Sigrid S. Skånland, <sup>41</sup> Elise A. Chong, <sup>42</sup> and Anthony R. Mato<sup>1</sup>

<sup>1</sup>Memorial Sloan Kettering Cancer Center, New York, NY; <sup>2</sup>Oxford University Hospitals, National Health Service (NHS) Foundation Trust, Oxford, United Kingdom; <sup>3</sup>Columbia University Medical Center, New York, NY; <sup>4</sup>Dana-Farber Cancer Institute, Boston, MA; <sup>5</sup>John Theurer Cancer Center, Hackensack University Medical Center, Hackensack, NJ; <sup>6</sup>The Ohio State University, Columbus, OH; <sup>7</sup>New York-Presbyterian Hospital, Weill Cornell Medicine, New York, NY; <sup>8</sup>Fundacion Jimenez Diaz University Hospital, Madrid, Spain; <sup>9</sup>Clinical Center Serbia, University of Belgrade, Belgrade, Serbia; <sup>10</sup>Swedish Cancer Institute, Seattle, WA; <sup>11</sup>Nottingham University Hospitals, NHS Trust, Nottingham, United Kingdom; <sup>12</sup>Hospital Puerta de Hierro Majadahonda, Madrid, Spain; <sup>13</sup>Hospital Infanta Leonor, Madrid, Spain; <sup>14</sup>Hospital Doce de Octubre, Madrid, Spain; <sup>15</sup>The University of North Carolina Cancer Center, Chapel Hill, NC; <sup>16</sup>Karolinska Institute, Solna, Sweden; <sup>17</sup>Rush University Medical Center, Chicago, IL; <sup>18</sup>Hospital Ramon y Cajal, Madrid, Spain; <sup>19</sup>Beatson West of Scotland Cancer Center, Glasgow, United Kingdom; <sup>20</sup>The Royal Marsden Hospital, London, United Kingdom; <sup>21</sup>Robert H. Lurie Comprehensive Cancer Center, Northwestern University, Chicago, IL; <sup>22</sup>St James's University, Detroit, MI; <sup>25</sup>University of Rochester Wilmot Cancer Institute, Rochester, NY; <sup>26</sup>Upstate Cancer Center, Syracuse, NY; <sup>27</sup>Comprehensive Cancer Center, School of Cancer and Pharmaceutical Science, King's College London, United Kingdom; <sup>28</sup>Hospital Israelita Albert Einstein, São Paulo, Brazil; <sup>29</sup>Yale University, New Haven, CT; <sup>30</sup>Medical and Dental Sciences, Institute of Immunology and Immunotherapy, University of Birmingham, Birmingham, United Kingdom; <sup>31</sup>Roswell Park Cancer Institute, <sup>85</sup>Leicester Royal Infirmary, Leicester, United Kingdom; <sup>36</sup>Royal Bournemouth Hospital, Bournemouth, United Kingdom; <sup>37</sup>Western General Hospital, Edinburgh, <sup>95</sup>Leicester Royal Infirmary, Leicester, United Kingdom; <sup>36</sup>Royal Bournemouth Hosp

With enhanced testing availability and evolution of therapeutic strategies, survival of COVID-19–infected patients has improved over time.<sup>1-3</sup> Two large series have reported outcomes for patients with chronic lymphocytic leukemia (CLL) infected with COVID-19 from February through May 2020, reporting case fatality rates (CFRs) of 31% to 33%.<sup>4,5</sup> Whether patients with CLL have experienced improvement in outcomes over time, as observed in the general population, has remained unknown. To understand change in outcomes over time, we present this follow-up study, which builds upon a previously reported cohort with extended follow up and addition of more recently diagnosed cases.

Emergency use authorization has been granted by the US Food and Drug Administration for several agents for treatment of COVID-19,<sup>6-9</sup> and dexamethasone has demonstrated an overall survival (OS) benefit for COVID-19–infected patients requiring oxygen.<sup>10,11</sup> These therapeutic studies have included few patients with hematological malignancies, and disease-specific outcomes have not been presented. Given the possible differences in immune response and risk of infection, understanding the benefit of these therapies in a CLL-specific population is crucial.

Early data from a small series suggest that patients with CLL may not consistently generate anti-SARS-CoV-2 antibodies after infection.<sup>12</sup> This finding, along with previous reports of inadequate

response to vaccines in patients with CLL,<sup>13-19</sup> highlight significant questions regarding COVID-19 vaccine efficacy in this population.

In this retrospective study, investigators from 45 centers identified patients with CLL diagnosed with COVID-19 based on polymerase chain reaction detection of SARS-CoV-2 from 17 February 2020 through 1 February 2021. Institutional review board approvals were granted, and the study was conducted in accordance with the Declaration of Helsinki.

A uniform case report form was used to collect baseline demographics, comorbidities, CLL-directed treatment history, date of COVID-19 diagnosis, and the COVID-19 clinical course and management strategy. Information regarding anti-SARS-CoV-2 serology testing during routine care was collected if performed; the specific antibody tested was not mandated or recorded.

Our primary purpose was to report the CFR for a larger group of patients with CLL who were diagnosed with COVID-19 and had a longer follow-up. We further sought to report the CFR stratified by date (the "early cohort," diagnosed from 17 February through 30 April 2020 and the "later cohort," diagnosed from 1 May 2020 through 1 February 2021; dates selected to mirror population-based studies<sup>1,2</sup>), examine outcomes for patients who received specific COVID-19–directed therapies, and describe serology testing results for those tested in routine clinical care.

OS was estimated using the Kaplan-Meier method.<sup>20</sup> Univariable Cox regression analyses adjusted for potential confounders were performed (Stata, v  $6^{21}$ ) to evaluate the relationship between baseline characteristics and COVID-19–directed therapies and OS.

This analysis included 374 patients with CLL who were diagnosed with COVID-19. With median follow-up of 38 days (range, 1-364 days) for the entire group and 63.5 days for survivors (range, 1-364), the CFR was 28%. Hospital admission was required for 75%, and intensive care unit (ICU) admission was required for 27%. Supplemental oxygen was used for 68% of the patients and mechanical ventilation was necessary for 20%. For patients who were admitted to the hospital, the CFR was 36% (99 of 278), whereas it was 4.3% (4 of 92) for those who were not admitted. Age >75 years and cumulative illness rating scale-geriatric  $(CIRS)^{22} > 6$  were independent predictors of poor survival. Sex, hypogammaglobulinemia, and CLL-directed treatment (including a history of any treatment, current treatment, current Bruton tyrosine kinase inhibitor (BTKi) therapy, and prior lines of therapy) were not associated with survival (supplemental Table 1, available on the Blood Web site).

To examine trends over time, we compared updated information for 254 patients diagnosed from 17 February through 30 April 2020 (early cohort) to data for 120 patients more recently diagnosed from 1 May 2020 through 1 February 2021 (later cohort). Comparison of baseline characteristics and markers of COVID-19 severity in these 2 cohorts are presented in Table 1. A larger proportion of patients in the early cohort were admitted (85% vs 55%) and required ICU admission (32% vs 15%). The CFR in the early cohort was 35% vs 11% in the later cohort (P < .001). For patients requiring hospitalization, the CFR was 40% (86 of 213) in the early cohort and 20% (13 of 65) in the later cohort (P =.003). For those who required oxygen, the CFR was 44% vs 25% (P = .015). The proportion of hospitalized patients requiring ICU-level care was lower in the later cohort (37% in early cohort vs 29% in the later cohort), whereas the CFR remained high for the subset of patients who required ICU-level care (52% vs 50%; P = .89). A difference in management of BTKi-treated patients was observed in the early vs the later cohort. In the early cohort, 76% of patients receiving BTKi had their drug therapy suspended or discontinued. In the later cohort, only 20% of BTKi-treated patients had their therapy suspended or discontinued.

Univariable analyses examined associations between administration of specific COVID-19 therapies and OS in all admitted patients and also the subset of admitted patients who required supplemental oxygen (supplemental Tables 2 and 3). Remdesivir (hazards ratio [HR], 0.48; P = .03) and convalescent plasma (HR, 0.50; P = .04) administration were associated with improved OS, whereas admitted patients who received corticosteroids (HR, 1.73, P = .01) and hydroxychloroquine (HR, 1.53; P = .04) had an increased risk of death.

Supplemental Table 4 describes baseline characteristics and the COVID-19 course for those who did vs did not receive corticosteroids. Corticosteroids were associated with increased risk of death when the data were adjusted for admission status (HR, 1.8; 95%)

confidence interval [CI], 1.2-2.7; P = .007) and the need for mechanical ventilation (HR, 2.0; 95% CI, 1.3-3.1; P = .002), although they were not significantly associated with survival when the data were adjusted for use of supplemental oxygen (HR, 1.4; 95% CI, 0.93-2.2; P = .11). Furthermore, admitted patients treated with corticosteroids in the later cohort did not experience an OS benefit (HR, 2.6; 95% CI, 0.6-11.9; P = .22). Secondary infections were observed in 26% (18 of 69) vs 8% (12 of 154) of those who did vs did not receive corticosteroids and in 26% (26 of 99) vs 8% (6 of 75) of admitted patients who required oxygen and did vs did not receive corticosteroids.

After acute infection, COVID-19 serology was checked in 25% of patients (93 of 374). Of the patients tested, the serology result was positive in 60%, negative in 39%, and equivocal in 1%. The proportions of untreated patients and those treated with BTKi, venetoclax, or hypogammaglobulinemia who developed anti-SARS-CoV-2 antibodies were 74% (29 of 39), 48% (12 of 25), 30% (3 of 10), and 60% (14 of 28), respectively.

Because the CFR of patients with CLL who were diagnosed with COVID-19 in the spring of 2020 was high (31% to 33%),<sup>4,5</sup> we wanted to examine the CFR in a larger cohort with additional follow-up and in a subset of patients diagnosed later in the course of the pandemic. Our findings mirrored population-based studies<sup>1-3</sup> with decreasing CFR (35% in those diagnosed before 1 May 2020 vs 11% in those diagnosed after that date). Improvement in OS was also observed in hospitalized patients and in those who required supplemental oxygen, and the proportion of hospitalized patients who needed ICU-level care declined. These trends suggest that patients in the later cohort experienced a less severe clinical course and that the observed difference in CFR over time may not just be due to more frequent testing and identification of less symptomatic patients. (Figure 1).

Although our data corroborate prior studies that demonstrated the benefit of remdesivir<sup>6</sup> and the lack of benefit of hydroxychloroquine,<sup>23</sup> we interestingly found an OS benefit associated with convalescent plasma<sup>24</sup> and a lack of benefit (significantly inferior OS in admitted patients) with corticosteroids.<sup>10</sup> Regarding convalescent plasma, patients with CLL have known humoral immunodeficiency, and antibody-based therapies may uniquely benefit this population. The RECOVERY trial demonstrated an OS benefit of dexamethasone in COVID-19 patients requiring oxygen (HR, 0.82; 95% CI, 0.72-0.94) and mechanical ventilation (HR, 0.64; 95% CI, 0.51-0.81).<sup>10</sup> In contrast, corticosteroid use was associated with a trend toward inferior OS in patients requiring oxygen and a significant risk of death for intubated patients with CLL. Use of corticosteroids in the earlier cohort may have been reserved for patients with more severe disease, as data regarding use of corticosteroids in COVID-19 were not yet available. Thus, inferior outcomes for steroid-treated patients in this cohort may be an artifact of their use in patients with more severe disease. As RECOVERY trial data were published in July 2020, we hypothesized that patients in the later cohort were more likely to receive corticosteroids in a data-driven, optimal clinical setting. However, corticosteroid use was not associated with improved OS in the later cohort. Although the use of corticosteroids was nonrandomized and is potentially biased by clinical context, the data are hypothesis generating and suggest that COVID-19 directed interventions, particularly immunomodulatory agents, require prospective study, specifically in immunocompromised populations.

### Table 1. Baseline characteristics and COVID-19 management

	Early cohort (n = 254)		Later cohort (n = 120)		Entire cohort (N = 374)	
	Proportion, unless otherwise specified, %	Patients with available data, n	Proportion, unless otherwise specified, %	Patients with available data, n	Proportion, unless otherwise specified, %	Patients with available data, n
Baseline						
Age at CLL diagnosis, median in years (range)	62.5 (31 - 92)	248	59 (29-86)	119	61 (29-92)	367
Age at COVID- 19 diagnosis, median in years (range)	70 (36 - 98)	254	65.5 (29-93)	120	68 (29-98)	374
Male	64	254	65	120	64	374
White	86	249	85	116	85	365
CIRS, <sup>22</sup> median (range)	8 (4-32)	229	8 (4-21)	117	8 (4-32)	346
CLL treatment history		253		119		372
Never treated	44	_	47	_	45	_
Prior therapy	56	—	53	—	55	—
Lines of therapy for previously treated patients, median (range)	1.5 (1-8)	136	1 (1-7)	61	1 (1-8)	197
Receiving therapy at time of COVID-19 diagnosis	39	254	34	119	38	373
Receiving BTK inhibitor at time of COVID-19 diagnosis	29	253	21	119	26	372
Receiving venetoclax at time of COVID-19 diagnosis	8	253	9	119	8	372
COVID-19 management						
Admitted	85	252	55	119	75	371
ICU admission	32	250	15	107	27	357
Imaging performed	89	245	60	108	80	353
Pneumonia on imaging	88	224	62	82	81	306
Supplemental oxygen	78	250	45	112	68	362
Mechanical ventilation	25	246	9	111	20	357
Steroids administered	42	244	39	112	41	356

CIRS, cumulative illness rating scale.



Figure 1. Overall survival from the time of COVID-19 diagnosis. The entire cohort (A) and stratified by timing of diagnosis (B) for patients who required oxygen.

Although these data are not sufficient to change recommendations for the use of corticosteroids given the demonstrated benefit in a prospective clinical trial, they raise a question about the benefit of immunomodulatory or immunosuppressive therapy in a population at increased risk of infection, as demonstrated in CLL-directed therapeutic trials.

Finally, our multicenter series was consistent with a prior singlecenter study,<sup>12</sup> and 60% of patients with CLL developed positive anti-SARS-CoV-2 serology results after polymerase chain reaction diagnosis of COVID-19. That study is the largest reported series of serologic testing for patients with CLL and adds further evidence that antibody production after COVID-19 is not uniform in patients with CLL. Coupled with prior reports of decreased responses to other vaccines,<sup>13-19</sup> further study is ongoing to gain understanding of the immune response to SARS-CoV-2 vaccination in patients with CLL.

Reassuringly, the overall trend in the CFR for patients with CLL mirrors improved OS for patients with COVID-19 in the general population, but the data highlight opportunities for further investigation into optimal management of COVID-19, immune response after infection, and effective vaccination strategy for patients with CLL.

### Acknowledgments

This research was supported, in part, by National Institutes of Health, National Cancer Institute Cancer Center support grant P30 CA008748. L.E.R. was supported by an American Society of Hematology Research Training Award for Fellows outside of the submitted work. I.E.A. receives research support from the Intramural Program of the National Heart, Lung, and Blood, Institute and an American Society of Hematology Scholar Award. T.A.E. was supported by the Oxford National Institute for Health Research (NIHR) Biomedical Research Centre.

## Authorship

Contribution: L.E.R. was a joint senior principal investigator, responsible for the study design, and data collection, coordination, analysis, and interpretation and writing the manuscript; T.A.E. assisted with the study design, site coordination, data

collection and interpretation, and writing and editing the manuscript; M.C.T., N.L., A.R.C., M.S.D., P.O.B., L.L., K.A.R., J.N.A., R.C., A.L.-G., D.A., J.M.P., N.M.-C., J.A.G.-M., J.-A.H.-R., F.M., C.C.C., A.Ö., L.H., A.N.S., J.L.J., M.R.W., D.E.-S., D.W., S.M., T.M., S.V., E.S., P.M.B., J.P., P.E.M.P., G.F.P., S.F.H., H.P., S. Sundaram, A.S., M.K., R.J., H.W., R.W., A.B., S.L., K.M.I., C.A.P., I.E.A., C.S.U., M.S., S. Skånland, and E.A.C. were involved in data collection and interpretation and edited the manuscript; A.R.M. was a joint senior principal investigator and was responsible for study design and data collection, coordination, analysis, and interpretation and writing the manuscript.

Conflict-of-interest disclosure: L.E.R. has served as a consultant for AbbVie, AstraZeneca, Janssen, LOXO Oncology, Pharmacyclics, Pfizer, TG Therapeutics, Vaniam group, and Verastem; holds a minority ownership in Abbott Laboratories; and has received research funding from Aptose Biosciences and Pfizer outside of the submitted work. T.A.E. has received honoraria and advisory board honoraria from Roche, Gilead, Kite, Janssen, AbbVie, Astra-Zeneca, LOXO Oncology, BeiGene, and Incyte; receives research support from Gilead and AstraZeneca; has received travel support from Gilead, Takeda, and AbbVie; and has served on a trial steering committee for LOXO Oncology. M.C.T. has received honoraria from MJH Life Sciences, VJ Heme Onc, and Curio Science. N.L. reports grants from Loxo Oncology; grants and personal fees from and consultancy and board of directors or advisory board membership for AbbVie, AstraZeneca, BeiGene, and Genentech; personal fees from and consultancy and board of directors or advisory committee membership for Celgene, Gilead, Janssen, and Pharmacyclics; and grants from Juno, Octernal, Verastem, TG Therapeutics, MingSight, and Octapharma, outside the submitted work. M.S.D. reports grants and personal fees from AbbVie, Ascentage Pharma, BMS, Genentech, MEI Pharma, Novartis, Pharmacyclics, Takeda, TG Therapeutics, Verastem, and AstraZeneca; personal fees from Adaptive Biotechnologies, BeiGene, Celgene, Eli Lilly, Gilead Sciences, Janssen, Merck, Research to Practice, and Syros Pharmaceuticals; personal fees from Zentalis; and grants from Surface Oncology, outside the submitted work. L.L. has served on speakers' bureaus for Seattle Genetics, Celgene/BMS, KitePharma, BeiGene, Pharmacyclics/Janssen, and AstraZeneca and has participated on advisory boards for Bayer, Seattle Genetics, ADC Therapeutics, AbbVie, Janssen, Pharmacyclics, Kite, and AstraZeneca. K.A.R. receives research funding from Genentech, AbbVie, Janssen, and Novartis; has consulted for Acerta Pharma, Genentech, AbbVie, Pharmacyclics, Innate Pharma, and AstraZeneca; and has received travel funding from AstraZeneca. J.N.A. has received research funding from Genentech, Janssen, and Celgene and has served as a consultant to Pharmacyclics, AbbVie, Genentech, AstraZeneca, Sunesis, and Janssen. R.C. serves as a speaker for Roche, Janssen, BMS, Abb-Vie, and Takeda; serves on advisory boards for Janssen, Celgene, AbbVie, Servier, Kyowa Kirin, and Takeda; and has received travel funding from Roche, Pfizer, Janssen, Celgene, AbbVie, Servier, and Takeda. J.M.P. reports consultancy with LOXO Oncology, AstraZeneca, Gilead, and BeiGene. J.-A.G.-M. has held a consulting or advisory role for AbbVie, AstraZeneca, Janssen, and Roche; has received research funding from AbbVie and Janssen; and has received speakers' fees from AbbVie, Astra-Zeneca, and Janssen. J.-A.H.-R. serves as a consultant for Janssen, AbbVie, Roche, Gilead, Celgene, Amgen, and Takeda; is on the speakers' bureau for Janssen, AbbVie, Roche, Gilead, Celgene, AstraZeneca, Amgen, Takeda, and BeiGene; and receives grants and research support from Celgene. C.C.C. reports personal fees from Loxo Oncology, Novartis, AbbVie, Genentech, MEI Pharma, and Octapharma; and payment to her institution for clinical trials from Loxo Oncology, Gilead, H3 Biomedicine, and Incyte. A.Ö. receives grants for academic research from BeiGene, Kancera; holds stock ownership in Kancera; and has served as a consultant for Sanofi. D.E.-S. has received honoraria from AbbVie, AstraZeneca, Janssen, Roche, and Takeda; has received conference/travel support from AbbVie and Novartis; and has served on advisory boards for AbbVie, ASTEX, AstraZeneca, BeiGene, Janssen, and Kyowa Kirin. L.H. has received research funding from Janssen-Cilag and Gilead Sciences. S.M. served as a consultant or advisor for AstraZeneca, Bei-Gene, Genentech, Pharmacyclics, and Verastem; has served on the speakers' bureau for AstraZeneca, BeiGene, Janssen, and Pharmacyclics; and has received research funding from AbbVie, AstraZeneca, BeiGene, Juno, LOXO Oncology, Novartis, Pharmacyclics, and TG Therapeutics. P.M.B. has consulted for PCYC/Abb-Vie, Genentech, Gilead, Merck, Seattle Genetics, Verastem, AstraZeneca, Celgene, and Morphosys. P.E.M.P. serves as a consultant for AbbVie, AstraZeneca, Atura, Gilead, Janssen, Novartis, Roche, and Tolero Pharmaceuticals; has been a remunerated speaker for AbbVie, AstraZeneca, Gilead, Janssen, Novartis, and Roche; has received travel support from AbbVie, Gilead, Janssen, Novartis, and Roche; and has received research funding from Gilead and Roche. S.F.H. has served as a consultant to Celgene, Bayer, Genentech, Pharmacyclics, Novartis, and AbbVie and has received research funding from DTRM Biopharm, Celgene, and TG Therapeutics. S. Sundaram has served as a consultant for Janssen and received research support from Lymphoma Research Foundation. A.S. has served as a consultant for TG Therapeutics, Alexion, Jazz Pharmaceuticals, Pharmacyclics, AstraZeneca, Janssen, Celgene, AbbVie, Genentech, Kite Pharma, and Novartis and is on the speakers' bureau for TG Therapeutics, Alexion, Jazz Pharmaceuticals, Pharmacyclics, AstraZeneca, Janssen, Celgene, AbbVie, Genentech, Kite Pharma, BeiGene, Verastem, and Seattle Genetics. M.K. has served as a consultant for AZD, Celgene and Pharmacyclics and is on the speaker's bureau for Seattle Genetics. R.W. reports receiving travel support and serving on speakers' bureaus for AbbVie, Janssen, and Gilead. A.B. reports receiving travel support and serving on a speaker's bureau for Gilead. C.S.U. has consulted for Pharmacyclics, AbbVie, and AstraZeneca. M.S. has received research funding from Mustang Bio, Celgene, Pharmacyclics, Gilead, Genentech, AbbVie, TG Therapeutics, BeiGene, Acerta Pharma, and Merck and has served on advisory boards or as a consultant for AbbVie, Genentech, AstraZeneca, Sound Biologics, Verastem, ADC Therapeutics, BMS, and Atara Biotherapeutics. E.A.C. serves on advisory boards for Novartis, Tessa, BMS, and Kite Pharma, and receives research funding from the Lymphoma Research Foundation. A.R.M. has served as a consultant for Celgene, Acerta, and Janssen; has served as a consultant for and received research funding from AbbVie, Loxo, Genentech, Pharmacyclics, AstraZeneca, Sunesis, and Johnson & Johnson; has received research funding from DTRM Biopharma and Gilead; and has served as a consultant for, received research funding from, and is a Data Safety and Monitoring Board member for TG Therapeutics. The remaining authors report no competing financial interests.

ORCID profiles: L.E.R., 0000-0002-3806-059X; T.A.E., 0000-0002-6631-9749; K.A.R., 0000-0001-5748-7874; J.N.A., 0000-0002-2088-0899; R.C., 0000-0002-7654-8836; A.L.-G., 0000-0002-5354-5261; N.M.-C., 0000-0002-5184-9464; J.6A.G.-M., 0000-0002-8993-5982; J.H.-R., 0000-0003-4550-757X; F.M., 0000-0001-5096-3145; J.L., 0000-0002-2969-3002; M.R.W., 0000-0001-5423-3270; D.E.-S., 0000-0002-2752-5814; P.M.B., 0000-0002-9733-401X; J.P., 0000-0001-7498-3159; P.E.M.P., 0000-0003-3320-3034; S.F.H., 0000-0001-7071-6475; H.P., 0000-0002-9707-8167; H.W., 0000-0003-2618-711X; K.M.I., 0000-0002-0234-139X; M.S., 0000-0002-3365-6562.

Correspondence: Anthony R. Mato, Memorial Sloan Kettering Cancer Center, 1275 York Ave, New York, NY 10065; e-mail: matoa@msckcc.org.

### Footnotes

Submitted 24 March 2021; accepted 7 July 2021; prepublished online on *Blood* First Edition 23 July 2021.

\*L.E.R. and T.A.E. contributed equally to this study.

Original data are available by e-mail request to the corresponding author.

The online version of this article contains a data supplement.

There is a *Blood* Commentary on this article in this issue.

#### REFERENCES

- 1. Asch DA, Sheils NE, Islam MN, et al. Variation in US hospital mortality rates for patients admitted with COVID-19 during the first 6 months of the pandemic. JAMA Intern Med. 2021;181(4):471-478.
- Dennis JM, McGovern AP, Vollmer SJ, Mateen BA. Improving survival of critical care patients with coronavirus disease 2019 in England: a national cohort study, March to June 2020. *Crit Care Med.* 2021;49(2):209-214.
- Horwitz LI, Jones SA, Cerfolio RJ, et al. Trends in COVID-19 risk-adjusted mortality rates. J Hosp Med. 2021;16(2):90-92.
- Mato AR, Roeker LE, Lamanna N, et al. Outcomes of COVID-19 in patients with CLL: a multicenter international experience. *Blood.* 2020; 136(10):1134-1143.
- Scarfò L, Chatzikonstantinou T, Rigolin GM, et al. COVID-19 severity and mortality in patients with chronic lymphocytic leukemia: a joint study by ERIC, the European Research Initiative on CLL, and CLL Campus. *Leukemia*. 2020;34(9):2354-2363.

- Beigel JH, Tomashek KM, Dodd LE, et al; ACTT-1 Study Group Members. Remdesivir for the treatment of Covid-19 - final report. N Engl J Med. 2020;383(19):1813-1826.
- Kalil AC, Patterson TF, Mehta AK, et al; ACTT-2 Study Group Members. Baricitinib plus remdesivir for hospitalized adults with Covid-19. N Engl J Med. 2021;384(9):795-807.
- An EUA for bamlanivimab-A monoclonal antibody for COVID-19. JAMA. 2021;325(9):880-881.
- Weinreich DM, Sivapalasingam S, Norton T, et al; Trial Investigators. REGN-COV2, a neutralizing antibody cocktail, in outpatients with Covid-19. N Engl J Med. 2021;384(3):238-251.
- Recovery\_Collaborative\_Group; Horby P, Lim WS, Emberson JR, et al. Dexamethasone in hospitalized patients with Covid-19. N Engl J Med. 2021;384(8):693-704.
- Villar J, Ferrando C, Martínez D, et al; dexamethasone in ARDS network. Dexamethasone treatment for the acute respiratory distress syndrome: a multicentre, randomised controlled trial. *Lancet Respir Med.* 2020;8(3): 267-276.
- Roeker LE, Knorr DA, Pessin MS, et al. Anti-SARS-CoV-2 antibody response in patients with chronic lymphocytic leukemia. *Leukemia*. 2020; 34(11):3047-3049.
- Sinisalo M, Aittoniemi J, Käyhty H, Vilpo J. Vaccination against infections in chronic lymphocytic leukemia. *Leuk Lymphoma*. 2003;44(4):649-652.
- Pleyer C, Ali MA, Cohen JI, et al. Effect of Bruton tyrosine kinase inhibitor on efficacy of adjuvanted recombinant hepatitis B and zoster vaccines. *Blood.* 2021;137(2):185-189.
- Mauro FR, Giannarelli D, Galluzzo CM, et al. Response to the conjugate pneumococcal vaccine (PCV13) in patients with chronic lymphocytic leukemia (CLL). *Leukemia*. 2021;35(3):737-746.
- Sinisalo M, Aittoniemi J, Oivanen P, Käyhty H, Olander RM, Vilpo J. Response to vaccination against different types of antigens in patients

with chronic lymphocytic leukaemia. *Br J Haematol.* 2001;114(1): 107-110.

- Zent CS, Brady MT, Delage C, et al. Short term results of vaccination with adjuvanted recombinant varicella zoster glycoprotein E during initial BTK inhibitor therapy for CLL or lymphoplasmacytic lymphoma. *Leukemia*. 2021;35(6):1788-1791.
- Hartkamp A, Mulder AH, Rijkers GT, van Velzen-Blad H, Biesma DH. Antibody responses to pneumococcal and haemophilus vaccinations in patients with B-cell chronic lymphocytic leukaemia. Vaccine. 2001;19(13-14):1671-1677.
- Pasiarski M, Rolinski J, Grywalska E, et al. Antibody and plasmablast response to 13-valent pneumococcal conjugate vaccine in chronic lymphocytic leukemia patients-preliminary report. *PLoS One.* 2014;9(12): e114966.
- Bland JM, Altman DG. Survival probabilities (the Kaplan-Meier method). BMJ. 1998;317(7172):1572-1580.
- 21. Stata Statistical Software: Release 16 [computer program]. College Station, TX: StataCorp LLC; 2019.
- Salvi F, Miller MD, Grilli A, et al. A manual of guidelines to score the modified cumulative illness rating scale and its validation in acute hospitalized elderly patients. J Am Geriatr Soc. 2008;56(10):1926-1931.
- Recovery\_Collaborative\_Group, Horby P, Mafham M, et al. Effect of hydroxychloroquine in hospitalized patients with Covid-19. N Engl J Med. 2020;383(21):2030-2040.
- Simonovich VA, Burgos Pratx LD, Scibona P, et al; PlasmAr Study Group. A randomized trial of convalescent plasma in Covid-19 severe pneumonia. N Engl J Med. 2021;384(7):619-629.

DOI 10.1182/blood.2021011841

#### © 2021 by The American Society of Hematology

#### TO THE EDITOR:

# Torque teno mini virus as a cause of childhood acute promyelocytic leukemia lacking PML/RARA fusion

Annalisa Astolfi,<sup>1,\*</sup> Riccardo Masetti,<sup>2,\*</sup> Valentina Indio,<sup>3</sup> Salvatore Nicola Bertuccio,<sup>4</sup> Daria Messelodi,<sup>4</sup> Simone Rampelli,<sup>5</sup> Davide Leardini,<sup>6</sup> Matteo Carella,<sup>7</sup> Salvatore Serravalle,<sup>2</sup> Virginia Libri,<sup>2</sup> Jessica Bandini,<sup>2</sup> Stefano Volinia,<sup>1</sup> Marco Candela,<sup>5</sup> and Andrea Pession<sup>2</sup>

<sup>1</sup>Department of Translational Medicine, University of Ferrara, Ferrara, Italy; <sup>2</sup>Pediatric Unit, IRCCS Azienda Ospedaliero-Universitaria di Bologna, Bologna, Italy; <sup>3</sup>Interdepartmental Center Alma Mater Institute on Healthy Planet, <sup>4</sup>Department of Medical and Surgical Sciences, <sup>5</sup>Unit of Holobiont Microbiome and Microbiome Engineering, Department of Pharmacy and Biotechnology, <sup>6</sup>Residency School of Pediatrics, and <sup>7</sup>Residency School of Hematology, University of Bologna, Bologna, Italy

The treatment of patients with acute promyelocytic leukemia (APL) can serve as a paradigm for cancer therapy.<sup>1</sup> The outcome of this disease, in adults and in children, has significantly improved with the introduction of target-specific agents, such as all-trans retinoic acid (ATRA) and arsenic trioxide (ATO), providing long-term survival for most patients.<sup>2,3</sup> Although morphologic and clinical suspicion is sufficient to immediately initiate ATRA, definitive diagnosis relies on the demonstration of *PML/RARA* translocation, on alternative *RARA* rearrangements represented by *RARA* gene fusion to other partners, cryptic insertion into the *PML* gene, or vice versa that altogether occur in more than 98% of APL cases.<sup>4-6</sup> Here we describe an even more challenging situation where all conventional diagnostic approaches failed to detect an oncogenic event associated with the diagnosis of APL.

A 6-year-old girl was admitted to the emergency department with a 3-day history of shoulder pain and fever. On physical examination, she presented with pallor and ecchymosis of the lower limbs. A full blood count showed a hemoglobin concentration of 9.7 g/dL, and leukocyte and platelet counts of  $2.8 \times 10^3/\mu$ L and  $101 \times 10^3/\mu$ L, respectively. Coagulation tests showed consumptive coagulopathy with slightly prolonged prothrombin and activated partial thromboplastin times, an increased D-dimer concentration, and hypofibrinogenemia. A peripheral blood smear showed atypical promyelocytes packed with numerous azurophilic granules (Figure 1A), whereas analysis of the bone marrow aspirate demonstrated markedly hypercellular marrow containing 85% abnormal promyelocytes with Auer rods (CD33<sup>+</sup>, CD13<sup>+</sup>, CD38<sup>+</sup>, CD99<sup>+</sup>, HLA-DR<sup>low</sup>), with strong and diffuse reactivity to