

TO THE EDITOR:

Hydroxyurea does not affect the spermatogonial pool in prepubertal patients with sickle cell disease

Anne-Sophie Gille,^{1,2} Corinne Pondarré,^{3,4} Jean-Hugues Dalle,^{5,6} Françoise Bernaudin,³ Céline Chalas,⁷ Mony Fahd,⁶ Camille Jean,^{7,8} Harry Lezeau,⁹ Lydia Riou,² Véronique Drouineaud,⁷ Annabel Paye-Jaouen,¹⁰ Annie Kamdem,³ Bénédicte Neven,^{8,11} Cécile Arnaud,³ Saba Azamouh,⁶ Karima Yakouben,⁶ Sabine Samacki,^{8,12} Mariane de Montalembert,¹³ Eva Maria Comperat,^{5,14} Gilles Lenaour,⁸ Mathilde Sibony,^{8,15} Nathalie Dhédin,¹⁶ Daniel Vaiman,¹ Jean-Philippe Wolf,^{1,7,8} Catherine Patrat,^{1,7,8} Pierre Fouchet,² Catherine Poirot,^{5,7,16,*} and Virginie Barraud-Lange^{1,7,8,*}

¹Département de Génétique, Développement et Cancer. Team From Gametes to Birth, Institut Cochin, INSERM U1016, Paris, France; ²UMRE008 Stabilité Génétique, Cellules Souches et Radiations, Laboratoire des Cellules Souches Germinales, Institut de Radiobiologie Cellulaire et Moléculaire, Université de Paris, Université Paris-Saclay, CEA, Fontenay-aux-Roses, France; ³Centre Hospitalier Intercommunal de Créteil, Centre de Référence de la Drépanocytose, Créteil, France; ⁴INSERM Unité 955, Université Paris XII, Créteil, France; ⁵Faculté de Médecine, Sorbonne Université, Paris, France; ⁶Assistance Publique-Hôpitaux de Paris, Hôpital Robert Debré, Service d'Immuno-Hématologie Pédiatrique, Paris, France; ⁷Assistance Publique-Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, CHU Cochin, Laboratoire d'Histologie Embryologie Biologie de la Reproduction CECOS, Paris, France; ⁸UFR Médecine Paris Centre-Université de Paris, Paris, France; ⁹Service de Chirurgie Viscérale, Urologie, Traumatologie, Centre Hospitalier Intercommunal de Créteil, Créteil, France; ¹⁰Assistance Publique-Hôpitaux de Paris, Hôpital Robert Debré, Service de Chirurgie Viscérale et Urologie Pédiatriques, Paris, France; ¹¹Assistance Publique-Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, CHU Necker-Enfant Malades, Service d'Immuno-Hématologie et Rhumatologie Pédiatrique, Hôpital Necker-Enfants Malades, Paris, France; ¹²Assistance Publique-Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, CHU Necker-Enfant Malades, Service de Chirurgie Viscérale et Urologie Pédiatriques, Paris, France; ¹³Assistance Publique-Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, CHU Necker-Enfant Malades, Service de Pédiatrie Générale et Maladies Infectieuses, Centre de Référence des Syndromes Drépanocytaires Majeurs, Labex GR-Ex, Hôpital Necker-Enfants Malades, Paris, France; ¹⁴Assistance Publique-Hôpitaux de Paris, Hôpital Tenon, Service d'Anatomie et Cytologie Pathologiques, Paris, France; ¹⁵Assistance Publique-Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, CHU Cochin, Service de Pathologie, Paris, France; and ¹⁶Assistance Publique-Hôpitaux de Paris, Hôpital Saint-Louis, Service d'Hématologie, Unité Adolescents Jeunes Adultes, Paris, France

Myeloablative conditioning regimens before hematopoietic stem cell transplantation (HSCT) entail a very high risk of infertility.^{1,2} Prepubertal boys who do not yet produce sperm are offered a testicular biopsy along with cryopreservation of the immature testicular tissue (ITT-CP) to preserve the spermatogonial stem cells that give rise to spermatozoa at puberty.³⁻⁶ The use of cryopreserved ITT in nonhuman primates has been shown to lead to full restoration of spermatogenesis and births, thus providing genuine hope for future successes in human fertility.⁷⁻⁹ ITT-CP is an option for prepubertal boys with sickle cell disease (SCD) who are undergoing HSCT to treat severe genotypes (HbSS and HbS β^0 thalassemias). Before HSCT, hydroxyurea (HU) is widely used as a SCD treatment to reduce the frequency of vaso-occlusive and painful crises.¹⁰ However, alterations in sperm parameters have been reported in men with SCD,¹¹ and HU is known to worsen this disorder.^{11,12} There are limited data available regarding the potential effect of SCD and HU on the spermatogonial pool. This is, however, of particular relevance for successful clinical use of ITT in fertility restoration. Several studies to date have reported a reduced spermatogonial pool in immature testicular tissue of prepubertal boys with SCD treated with HU.¹³⁻¹⁵ To analyze the specific effect of HU on immature testis, we compared the spermatogonial quantity in testicular tissue collected for fertility preservation in prepubertal boys with SCD who had or who had not been exposed to HU. We also compared the results with established reference values in healthy boys.¹⁶ This study was approved by the Research Ethics Committee of the Cochin Hospital, Paris, France (AAA-2019-08014) and was conducted in accordance with the Declaration of Helsinki.

Thirty patients with SCD who had undergone ITT retrieval (March 2010 to April 2019) were included. Thirteen patients had not been exposed to HU, and 17 had been administered HU at a median dose of 22.0 mg/kg per day and a median time of exposure of 36.0 months (Table 1; supplemental Table available on the *Blood* Web site). Six of the exposed patients were on-HU at the time of the ITT-CP, whereas the 11 others were off-HU, with a median washout period of 5.2 months. The surgery was bilateral for 18 patients and unilateral for 12 patients (supplemental Table). MAGE-A4 and GATA-4 markers were used to detect the spermatogonia and Sertoli cells, respectively (supplemental Material and Methods; Figure 1A-C).^{17,18} The spermatogonial pool was evaluated as the number of spermatogonia (S) per round cross section of seminiferous tubule (T) (S/T ratio), which has been proposed as a standard.^{13,16} The proportion of seminiferous tubules with Sertoli cells only (SCO), without germ cells, was also quantified.

The HU-exposed and nonexposed groups were comparable in terms of the patient age at ITT-CP and the mean number of tubular cross sections observed (Table 1). Histologic analysis revealed that the spermatogonial pool was not statistically different between the two groups: S/T ratio = 2.5 ± 3.3 vs 1.7 ± 0.6 , respectively, ($P = .61$); SCO = 42 ± 21 vs $38 \pm 16\%$, respectively, ($P = .52$) (Table). The spermatogonial pool was also not statistically different between the off-HU and the on-HU subgroups (S/T ratio = 3.1 ± 4.0 vs 1.5 ± 0.7 , respectively, $P = .84$) (Table). The latter data warrant additional confirmation with more samples, although it is in accordance with the absence of a correlation between the wash-out delay and the spermatogonial count ($r = 0.09$, $P = .73$). The S/T ratio for each of the

Table 1. Histologic analysis of testicular tissue samples retrieved from prepubertal patients with SCD in the context of fertility preservation according to exposure to hydroxyurea

	Exposed to HU			Not exposed to HU	P	
	Off-HU	On-HU	Total		P1	P2
Patients, n	11	6	17	13		
Samples, n	17	8	25	23		
Treatment characteristics						
Age at HU onset, median (range), y	5.2 (3.4-10.9)	4.2 (2.2-9.0)	5.0 (2.2-10.9)	—	—	.33
HU dosing, median (range), mg/kg/d	22.2 ± 4.0 (15-28)	22.5 ± 2.7 (20-25)	22.0 (15-28)	—	—	.91
HU time of exposure, median (range), mo	27.6 (8.0-67.0)	37.5 (9.5-66.0)	36.0 (8.0-67.0)	—	—	.80
Washout period, median (range), mo	5.2 (2.2-36.9)	0.0	3.0 (0.0-36.9)	—	—	—
Transfusion therapy, median (range), mo	24.9 (2.4-71.4)*	13.2 (3.0-36.0)*	15.4 (2.4-71.4)	15.4 (0.9-42.8)†	.70	.62
ITT histologic analysis						
Age at ITT-CP, median (range), y	10.1 (5.8-15)	7.9 (4.2-11.3)	8.8 (4.2-15.0)	8.0 (4.2-11.8)	.39	.42
Cross-sections analyzed, mean ± SD (range), n	77 ± 49 (10-165)	102 ± 33 (60-143)	85 ± 45 (10-165)	87 ± 31 (51-148)	.9	.26
S/T ratio, mean ± SD (range)	3.1 ± 4.0 (0.6-14.0)	1.5 ± 0.7 (0.8-2.4)	2.5 ± 3.3 (0.6-14.0)	1.7 ± 0.6 (0.7-2.7)	.61	.84
SCO tubules, mean ± SD (range), %	44 ± 23 (0-77)	40 ± 17 (18-55)	42 ± 21 (0-77)	38 ± 16	.52	.72

In the comparison of HU-exposed and nonexposed groups, patients with bilateral surgery account for a single value (see supplemental Materials and Methods). P1, comparison of values between HU-exposed and nonexposed patients, Student t test; P2, comparison of values between off-HU and on-HU patients, Mann-Whitney test.

*One missing data point.

†Two missing data points.

48 testicular tissue samples was plotted on the meta-regression fit line of S/T reference values (Figure, panel D).¹⁶ Comparison of the S/T ratio of the patients with SCD to the age-related S/T reference values confirmed that the spermatogonial quantity in SCD patients was lower than in healthy boys, with a highly significant P value (Wilcoxon signed-ranks test: $P < .0001$). Interestingly, the subgroup of the 6 teenagers (≥ 11 years of age) with a mean S/T value of 5.6 ± 4.6 was not different from the healthy boys for this age group ($P = .34$). The latter result nevertheless needs to be interpreted with caution because of the small sample size and the large standard deviation. Finally, there was no correlation between the duration of the transfusion therapy and the spermatogonial count ($r = -.15$, $P = .33$).

To the best of our knowledge, this study is the largest quantitative study of the spermatogonial pool in boys with SCD. Unlike previous reports in which all the boys with SCD were exposed to HU,¹³⁻¹⁵ we compared HU-exposed vs nonexposed patients. As a result, the similar S/T ratio and proportion of SCO between the groups indicates that the spermatogonial depletion observed in prepubertal patients with SCD treated with HU was largely related to the disease itself and not to toxicity of the HU. The less frequent division of the spermatogonial stem cells could explain their relative resistance to HU, specifically to the inhibition of DNA synthesis and the induction of cell cycle arrest and apoptosis. An alternative could be that the daily exposure to HU at pharmacologic doses used in SCD boys is too low to induce a lasting detrimental effect on the spermatogonial pool. We emphasize that we could not address the gonadic effect of HU in infancy, because the median age at HU onset in the studied patients was 5 years, with only 1 patient aged 2 years. We therefore cannot exclude that very early HU introduction may have a specific effect on the testis of very young infants. Of note,

a research team recently reported a nonaffected spermatogonial pool in 3 SCD patients.¹⁹ Other than the very small sample size, this contradictory result could be explained by the fact that the patients were compared with nontreated cancer patients, who may have low spermatogonial counts, unlike healthy boys. SCD is a hemoglobinopathy that affects patients from very early infancy as a result of chronic hemolytic anemia, multiple vaso-occlusive complications, and painful crises.²⁰ The low spermatogonial quantity in patients with SCD may be related to testicular vaso-occlusive subclinical episodes and possible asymptomatic infarction,^{21,22} as well as testicular perfusion deficit as a result of chronic anemia. The establishment of spermatogonial DNA methylation has been shown to be altered in prepubertal patients with SCD.¹⁹ This could be another explanation for the spermatogonial depletion, as correct establishment of the methylation profile is necessary for spermatogonial maintenance. SCD treatments, such as transfusion therapy, did not provide any beneficial effect on the spermatogonial count. This could be because of the complexity of the mechanisms involved in the spermatogonial depletion affecting SCD. We noted, however, that spermatogonial mitotic expansion, physiologically described in peripubertal healthy boys,¹⁶ also occurred for the teenagers of our series. Despite the limitation of the small sample size, this suggests that the spermatogonia in young patients with SCD retain a certain capacity to self-renew and to expand at puberty.

In conclusion, we showed that depletion of the spermatogonial pool in prepubertal patients with severe SCD genotypes is related to the disease itself and not to HU toxicity. In the absence of adverse side effects on the spermatogonial quantity, concerns regarding HU gonadotoxicity should hence not affect treatment decisions in young patients with severe SCD genotypes.

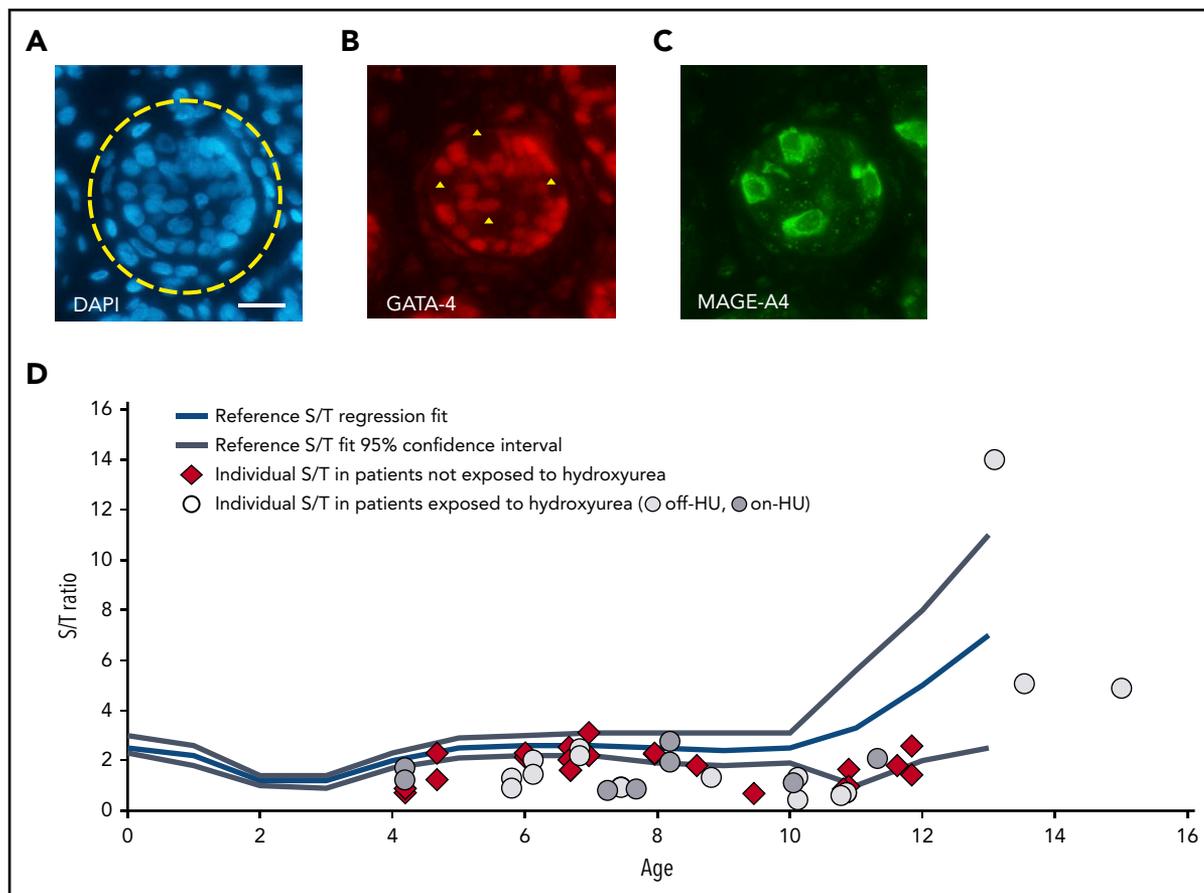


Figure 1. Spermatogonia in the testicular tissue of prepubertal patients with SCD compared with reference values. (A-C) Immunostained section of the testicular tissue of a patient aged 6.1 years who had previously been exposed to hydroxyurea (Off-HU patient 3). (A) Selection of a round tubular cross section (yellow circle). 4',6-Diamidino-2-phenylindole staining (blue). (B) Expression of GATA-4 nuclear protein in Sertoli cells (red staining). (C) Expression of MAGE-A4 cytoplasmic protein in spermatogonial cells (green staining). Negative images of spermatogonial cells in panel C (yellow triangle). Four spermatogonia could be counted in this selected section. Scale bar = 50 μ m. (D) The number of spermatogonia per round tubular cross section (S/T) of patients exposed to hydroxyurea (light gray circle: off-HU; dark gray circle: on-HU) or not exposed to hydroxyurea (red diamond). Individual S/T values plotted on a meta-regression fit line of S/T reference values during testis development.¹⁶ Each mark represents the mean S/T value of a patient. The S/T values of prepubertal patients with SCD were significantly lower than the reference S/T values (Wilcoxon signed-rank test, $P < .0001$).

Acknowledgments

A.-S.G. is supported by a grant from ARC Fondation pour la Recherche contre le Cancer (DOC20180507400). The human Spermatogonial Stem Cell research project (A.-S.G., L.R., J.-P.W., P.F., and V.B.-L.) is funded by grants from the Agence de Biomédecine and Electricité De France. It also benefits from donations of the Laurette Fugain and the Entraide aux Greffés de Moelle Osseuse Associations.

Authorship

Contribution: A.-S.G. and V.B.-L. designed and performed the experiments; A.-S.G., V.B.-L., C. Poirot, and D.V. analyzed and interpreted the data; A.-S.G., V.B.-L., C. Poirot, and P.F. were in charge of the preparation, drafting, and editing of the manuscript; C. Poirot performed fertility preservation counseling for all patients; all other authors contributed to the data collection and to critically reviewing the manuscript; and all authors agreed to the final version of the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: A.-S.G., 0000-0002-2427-1994; C. Pondarré, 0000-0003-4885-8885; J.-H.D., 0000-0002-8406-0793; C.A., 0000-0002-4216-3401; S.S., 0000-0003-4304-5578; M.d.M., 0000-0002-4900-4753; J.-P.W., 0000-0001-6604-9534; V.B.-L., 0000-0001-8507-7303.

Correspondence: Virginie Barraud-Lange, Assistance Publique-Hôpitaux de Paris, Hôpitaux Universitaires Paris Centre, CHU Cochin, Laboratoire d'Histologie Embryologie Biologie de la Reproduction CECOS, 123 Boulevard de Port Royal, 75014 Paris, France; e-mail: virginie.barraud-lange@aphp.fr.

Footnotes

Submitted 13 July 2020; accepted 9 November 2020; prepublished online on *Blood* First Edition 1 December 2020.

*C. Poirot and V.B.-L. contributed equally to this study.

Contact the corresponding author for original data.

The online version of this article contains a data supplement.

There is a *Blood* Commentary on this article in this issue.

REFERENCES

- Lee SJ, Schover LR, Partridge AH, et al. American Society of Clinical Oncology recommendations on fertility preservation in cancer patients. *J Clin Oncol*. 2006;24(18):2917-2931.

2. Lukusa AK, Vermeylen C, Vanabelle B, et al. Bone marrow transplantation or hydroxyurea for sickle cell anemia: long-term effects on semen variables and hormone profiles. *Pediatr Hematol Oncol*. 2009;26(4):186-194.
3. Bahadur G, Chatterjee R, Ralph D. Testicular tissue cryopreservation in boys. Ethical and legal issues: case report. *Hum Reprod*. 2000;15(6):1416-1420.
4. Keros V, Hultenby K, Borgstrom B, Fridstrom M, Jahnukainen K, Hovatta O. Methods of cryopreservation of testicular tissue with viable spermatogonia in pre-pubertal boys undergoing gonadotoxic cancer treatment. *Hum Reprod*. 2007;22(5):1384-1395.
5. Picton HM, Wyns C, Anderson RA, et al. A European perspective on testicular tissue cryopreservation for fertility preservation in prepubertal and adolescent boys. *Hum Reprod*. 2015;30(11):2463-2475.
6. Wyns C, Curaba M, Petit S, et al. Management of fertility preservation in prepubertal patients: 5 years' experience at the Catholic University of Louvain. *Hum Reprod*. 2011;26(4):737-747.
7. Brinster RL, Avarbock MR. Germline transmission of donor haplotype following spermatogonial transplantation. *Proc Natl Acad Sci USA*. 1994; 91(24):11303-11307.
8. Fayomi AP, Peters K, Sukhwani M, et al. Autologous grafting of cryopreserved prepubertal rhesus testis produces sperm and offspring. *Science*. 2019;363(6433):1314-1319.
9. Hermann BP, Sukhwani M, Winkler F, et al. Spermatogonial stem cell transplantation into rhesus testes regenerates spermatogenesis producing functional sperm. *Cell Stem Cell*. 2012;11(5):715-726.
10. Ferster A, Tahriri P, Vermeylen C, et al. Five years of experience with hydroxyurea in children and young adults with sickle cell disease. *Blood*. 2001;97(11):3628-3632.
11. Berthaut I, Bachir D, Kotti S, et al. Adverse effect of hydroxyurea on spermatogenesis in patients with sickle cell anemia after 6 months of treatment. *Blood*. 2017;130(21):2354-2356.
12. Berthaut I, Guignedoux G, Kirsch-Noir F, et al. Influence of sickle cell disease and treatment with hydroxyurea on sperm parameters and fertility of human males. *Haematologica*. 2008;93(7):988-993.
13. Stukenborg JB, Alves-Lopes JP, Kurek M, et al. Spermatogonial quantity in human prepubertal testicular tissue collected for fertility preservation prior to potentially sterilizing therapy. *Hum Reprod*. 2018;33(9):1677-1683.
14. Valli-Pulaski H, Peters KA, Gassei K, et al. Testicular tissue cryopreservation: 8 years of experience from a coordinated network of academic centers. *Hum Reprod*. 2019;34(6):966-977.
15. Van Saen D, Pino Sanchez J, Ferster A, van der Werff ten Bosch J, Tournaye H, Goossens E. Is the protein expression window during testicular development affected in patients at risk for stem cell loss? *Hum Reprod*. 2015;30(12):2859-2870.
16. Masiukaite I, Hagen JM, Jahnukainen K, et al. Establishing reference values for age-related spermatogonial quantity in prepubertal human testes: a systematic review and meta-analysis. *Fertil Steril*. 2016;106(7):1652-1657.
17. Di Persio S, Saracino R, Fera S, et al. Spermatogonial kinetics in humans. *Development*. 2017;144(19):3430-3439.
18. Chui K, Trivedi A, Cheng CY, et al. Characterization and functionality of proliferative human Sertoli cells. *Cell Transplant*. 2011;20(5):619-635.
19. Portela JMD, Heckmann L, Wistuba J, et al. Development and disease-dependent dynamics of spermatogonial subpopulations in human testicular tissues. *J Clin Med*. 2020;9(1):224.
20. Platt OS, Brambilla DJ, Rosse WF, et al. Mortality in sickle cell disease. Life expectancy and risk factors for early death. *N Engl J Med*. 1994;330(23): 1639-1644.
21. Li M, Fogarty J, Whitney KD, Stone P. Repeated testicular infarction in a patient with sickle cell disease: a possible mechanism for testicular failure. *Urology*. 2003;62(3):551.
22. Mueller FE. Segmental testicular infarction in sickle cell anemia. *Urologe A*. 2014;53(5):725-727.

DOI 10.1182/blood.2020008146

© 2021 by The American Society of Hematology