

tissues, including the spleen, lymph nodes, and the kidneys. Accordingly, VQ mice present a more profound impairment of hematopoiesis and ensuing anemia and increased immunoglobulin deposition in the kidneys, when compared with the Vκ*MYC model. Attesting to the aggressive nature of the disease in the VQ mouse, plasma cells are highly proliferative, and, intriguingly, present a gene expression pattern reminiscent of the high-risk myeloma signature (UAMS-70). As expected, the survival of the VQ mice is severely affected, with the animals living only for a few months.

The translational relevance of the VQ model has also been explored by Wen et al and is quite remarkable. For example, as it has been for the Vκ*MYC model,⁴ the VQ mouse was successfully tested as a drug screening in vivo platform, with drugs already used in the clinic to treat myeloma patients or more experimental compounds. In addition, VQ myeloma cells preserve both the PD1 and the TIGIT immune checkpoint pathways, which mediate T-cell suppression in myeloma patients. As such, the VQ mice could be exploited for the preclinical evaluation of immunotherapeutic strategies in an aggressive myeloma setting. Finally, tumor cells derived from this model could be transplanted into sublethally irradiated syngeneic recipients for several passages and, importantly, could be transduced with both retro- and lentiviruses, thus enabling additional genetic manipulations.

Thus, are we there yet? Is this the end of the journey? The model presented herein represents an important step forward, toward the generation of a reliable mouse model of aggressive myeloma. However, in solid tumors, the knowledge of the underlying genetic lesions has led to the generation of increasingly refined mouse models, as a result of the accrual of layers of genetic engineering.² Although MYC and NRAS certainly represent crucial genetic events in the development of the disease,^{4,5} myeloma is a genetically heterogeneous disease, with several other genes and pathways that are engaged in its development, in specific patient subsets.⁵ These lesions have not yet been properly modeled. There is indeed the possibility that the platform described in this study may represent the foundation for models where additional lesions could be engineered, to capture more closely the puzzling

intra- and interpatient heterogeneity of human myeloma.

Conflict-of-interest disclosure: The author declares no competing financial interests. ■

REFERENCES

1. Wen Z, Rajagopalan A, Flietner E, et al. Expression of *Nras*^{Q61R} and MYC transgene in germinal center B cells induces a highly malignant multiple myeloma in mice. *Blood*. 2021;137(1):61-74.
2. Frese KK, Tuveson DA. Maximizing mouse cancer models. *Nat Rev Cancer*. 2007;7(9):645-658.
3. Rossi M, Botta C, Arbitrio M, Grembale RD, Tagliaferri P, Tassone P. Mouse models of multiple myeloma: technologic platforms and perspectives. *Oncotarget*. 2018;9(28):20119-20133.
4. Chesi M, Robbiani DF, Sebag M, et al. AID-dependent activation of a MYC transgene induces multiple myeloma in a conditional mouse model of post-germinal center malignancies. *Cancer Cell*. 2008;13(2):167-180.

5. Kumar SK, Rajkumar V, Kyle RA, et al. Multiple myeloma. *Nat Rev Dis Primers*. 2017;3:17046.
6. Chapman MA, Lawrence MS, Keats JJ, et al. Initial genome sequencing and analysis of multiple myeloma. *Nature*. 2011;471(7339):467-472.
7. Li S, Balmain A, Counter CM. A model for RAS mutation patterns in cancers: finding the sweet spot. *Nat Rev Cancer*. 2018;18(12):767-777.
8. Voice JK, Klemke RL, Le A, Jackson JH. Four human ras homologs differ in their abilities to activate Raf-1, induce transformation, and stimulate cell motility. *J Biol Chem*. 1999;274(24):17164-17170.
9. Burd CE, Liu W, Huynh MV, et al. Mutation-specific RAS oncogenicity explains NRAS codon 61 selection in melanoma. *Cancer Discov*. 2014;4(12):1418-1429.
10. Lin YT, Way GP, Barwick BG, et al. Integrated phosphoproteomics and transcriptional classifiers reveal hidden RAS signaling dynamics in multiple myeloma. *Blood Adv*. 2019;3(21):3214-3227.

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MYELOID NEOPLASIA

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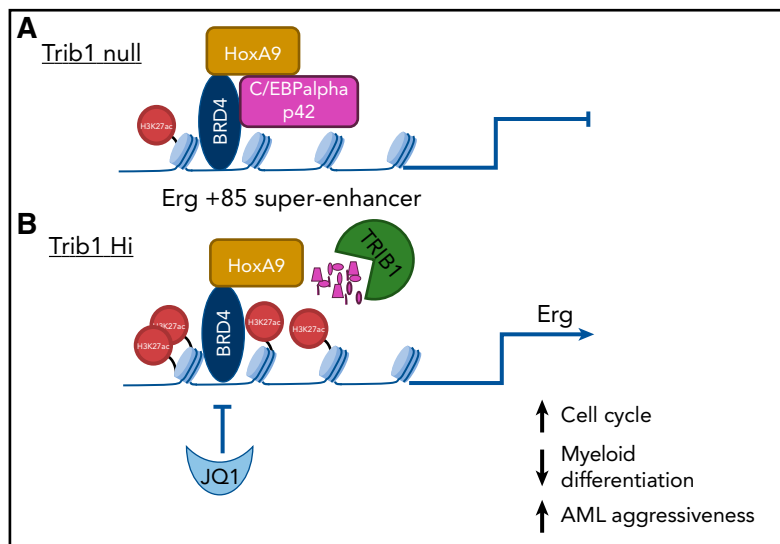
Superenhancing AML with Trib1

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In this issue of *Blood*, Yoshino et al use the genetic Trib1-ROSA26-Cre-knockout murine model and retroviral-mediated HoxA9 overexpression, together or not with Trib1 reexpression (Trib1 Hi and Trib1 null), to investigate the mechanistic basis for HoxA9-mediated acute myeloid leukemia (AML).¹ They have deciphered the mechanism involved in HoxA9-associated AML, linking an Erg-specific superenhancer with the Trib1-C/EBPα axis. It has previously been shown that TRIB proteins, including TRIB1 and TRIB2, are AML oncoproteins capable of driving disease development. Degradation of the p42 isoform of C/EBPα (an important myeloid transcription factor) and increased MAPK/ERK signaling (a proliferation and survival indicator) are key molecular mechanisms for Trib oncogenic activity.² Cooperation with HoxA9 leads to accelerated AML development,^{3,4} which is important because HoxA9 overexpression alone is unable to drive AML in vivo. The role that TRIB proteins may have across the wide heterogeneity of AML phenotypes is still underappreciated; however, given that deregulated Hox signaling is associated with ~70% of all AMLs, it is important to understand the involvement of Trib oncogenic activities in HoxA9-mediated AML. It remained an open question how TRIB proteins cooperate with Hoxa9 at the mechanistic level.

Previous data have shown that HoxA9 is organized into enhanceosomes containing lineage-restricted transcription factors,

including C/EBPα.⁵ Together with what is known about TRIB1 oncoprotein function, this led the investigators to hypothesize that



Schematic depiction of Erg +85 superenhancer in Trib1 null and Trib1 Hi HoxA9-expressing leukemia cells. (A) Trib1 null; (B) Trib1 Hi.

Trib1-mediated degradation of C/EBP α p42 isoform would lead to enhanced remodeling at HoxA9-associated genomic loci and, thus, be a key event in HoxA9-Trib1 cooperation in AML. The investigators tested this using microarray gene expression and chromatin immunoprecipitation sequencing analysis and could show that, in the absence of Trib1 (Trib1 null) in HoxA9-expressing cells, C/EBP α p42 is expressed, and a HoxA9-C/EBP complex and H3K27ac are present at distal enhancers. The HoxA9-C/EBP complex is associated with gene activation and repression and is known to contribute to the aberrant proliferative phenotype in an AML model that required coexpression of MEIS1.⁶ In this article, the investigators now advance our understanding of HoxA9 leukemogenesis (see figure); HoxA9 DNA binding peaks and H3K27ac peaks were not significantly different between TRIB1 hi and TRIB1 null cells, but HoxA9-associated superenhancers specific to TRIB1 hi AML cells were enriched, most notably at the Erg +85 superenhancer, together with H3K27ac. Importantly, it was the removal of C/EBP α p42 isoform mediated by Trib1 that occurred at these specific HoxA9-associated superenhancer genomic loci leading to enhanced H3K27ac and elevated Erg expression. There is a requirement for C/EBP α expression in HoxA9-associated and TRIB1-mediated

AML,^{6,7} which can seem counterintuitive. However, this is explained in the literature by the expression and opposing functions of C/EBP α p42 and p30 isoforms. The degradation of p42 isoform, while p30 expression remains intact, is a feature of AML⁸ and is consistent with the literature; here, it is shown to be a driving force for enhancer modification at the HoxA9-binding specific Erg superenhancer.

The investigators tested the therapeutic targeting of this mechanism with the use of the BRD4 inhibitor JQ1, because BRD4 is abundant at superenhancers. JQ1 suppressed the leukemic superenhancer activity of TRIB1 hi AML cells in vitro and in vivo, consistent with a block in Erg target gene expression. It remains an open question to what degree the expressions of Hox proteins and Trib proteins are linked across AML. Using messenger RNA expression has not revealed strong correlations among HoxA9, Trib1 or Trib2, and Erg; however, as elegantly shown in this article, oncoprotein activity and protein isoform expression are the key factors distinguishing AML with highly aggressive features that can be therapeutically targeted. Erg expression is associated with poor prognosis in

AML,⁹ as well as a more aggressive phenotype¹⁰; therefore, its elevated expression as a result of Trib1 function in HoxA9-associated leukemia has important implications for patient prognostication and, potentially, AML stratification.

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REFERENCES

1. Yoshino S, Yokoyama T, Sunami Y, et al. Trib1 promotes acute myeloid leukemia progression by modulating the transcriptional programs of Hoxa9. *Blood*. 2021;137(1):75-88.
2. Keeshan K, He Y, Wouters BJ, et al. Tribbles homolog 2 inactivates C/EBP α and causes acute myelogenous leukemia. *Cancer Cell*. 2006;10(5):401-411.
3. Jin G, Yamazaki Y, Takuwa M, et al. Trib1 and Evi1 cooperate with Hoxa and Meis1 in myeloid leukemogenesis. *Blood*. 2007;109(9):3998-4005.
4. Keeshan K, Shestova O, Ussin L, Pear WS. Tribbles homolog 2 (Trib2) and HoxA9 cooperate to accelerate acute myelogenous leukemia. *Blood Cells Mol Dis*. 2008;40(1):119-121.
5. Huang Y, Sitwala K, Bronstein J, et al. Identification and characterization of Hoxa9 binding sites in hematopoietic cells. *Blood*. 2012;119(2):388-398.
6. Collins C, Wang J, Miao H, et al. C/EBP is an essential collaborator in Hoxa9/Meis1-mediated leukemogenesis. *Proc Natl Acad Sci USA*. 2014;111(27):9899-9904.
7. O'Connor C, Lohan F, Campos J, et al. The presence of C/EBP α and its degradation are both required for TRIB2-mediated leukaemia. *Oncogene*. 2016;35(40):5272-5281.
8. Pabst T, Mueller BU, Zhang P, et al. Dominant-negative mutations of CEBPA, encoding CCAAT/enhancer binding protein-alpha (C/EBP α), in acute myeloid leukemia. *Nat Genet*. 2001;27(3):263-270.
9. Marcucci G, Baldus CD, Ruppert AS, et al. Overexpression of the ETS-related gene, ERG, predicts a worse outcome in acute myeloid leukemia with normal karyotype: a Cancer and Leukemia Group B study. *J Clin Oncol*. 2005;23(36):9234-9242.
10. Stavropoulou V, Kaspar S, Brault L, et al. MLL-AF9 expression in hematopoietic stem cells drives a highly invasive AML expressing EMT-related genes linked to poor outcome. *Cancer Cell*. 2016;30(1):43-58.

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