

employed in this study, proximity ligation assays. The new opportunities linked to all these studies include unprecedented insight into the architecture of the tumor microenvironment and into spatial relationships of different cell types at single-cell resolution. These relationships have relevance not only to HL but also to other lymphoma entities that rely on a complex cellular ecosystem of malignant cells with reactive immune cells. However, the molecular underpinnings (the “interior design”) in the form of somatic gene mutations, receptor-ligand interactions, and involved soluble factors (cytokines/chemokines) are still only partially understood and are rarely combined with the characteristic morphology features (“architecture”) of HL. Thus, the present study can be seen as a paradigm for an accelerating process deciphering the relatedness of “architecture” and “interior design.”

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REFERENCES

- Veldman J, Visser L, Huberts-Kregel M, et al. Rosetting T cells in Hodgkin lymphoma are activated by immunological synapse components HLA class II and CD58. *Blood*. 2020; 136(21):24372441.
- Scott DW, Gascoyne RD. The tumour micro-environment in B cell lymphomas. *Nat Rev Cancer*. 2014;14(8):517-534.
- Stuart AE, Williams AR, Habeshaw JA. Rosetting and other reactions of the Reed-Sternberg cell. *J Pathol*. 1977;122(2):81-90.
- Diepstra A, van Imhoff GW, Karim-Kos HE, et al. HLA class II expression by Hodgkin Reed-Sternberg cells is an independent prognostic factor in classical Hodgkin's lymphoma. *J Clin Oncol*. 2007;25(21):3101-3108.
- Roemer MGM, Redd RA, Cader FZ, et al. Major histocompatibility complex class II and programmed death ligand 1 expression predict outcome after programmed death 1 blockade in classic Hodgkin lymphoma. *J Clin Oncol*. 2018;36(10):942-950.
- Schneider M, Schneider S, Zühlke-Jenisch R, et al. Alterations of the CD58 gene in classical Hodgkin lymphoma. *Genes Chromosomes Cancer*. 2015;54(10):638-645.
- Steidl C, Shah SP, Woolcock BW, et al. MHC class II transactivator CIITA is a recurrent gene fusion partner in lymphoid cancers. *Nature*. 2011;471(7338):377-381.
- Patel SS, Weirather JL, Lipschitz M, et al. The microenvironmental niche in classic Hodgkin lymphoma is enriched for CTLA-4-positive

T cells that are PD-1-negative. *Blood*. 2019; 134(23):2059-2069.

- Cader FZ, Schackmann RCJ, Hu X, et al. Mass cytometry of Hodgkin lymphoma reveals a CD4⁺ regulatory T-cell-rich and exhausted T-effector microenvironment. *Blood*. 2018; 132(8):825-836.

- Aoki T, Chong LC, Takata K, et al. Single-cell transcriptome analysis reveals disease-defining T-cell subsets in the tumor micro-environment of classic Hodgkin lymphoma. *Cancer Discov*. 2020;10(3):406-421.

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MYELOID NEOPLASIA

Comment on Dzama et al, page 2442

MLL-menin and FLT3 inhibitors team up for AML

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In this issue of *Blood*,¹ Dzama and colleagues demonstrate that inhibiting MLL binding to menin synergizes with FLT3 inhibition in preclinical models of NPM1c and KMT2A (MLL)-rearranged AML, thus identifying a novel, rational combination for clinical development.

Potent, selective *Fms*-like tyrosine kinase 3 (FLT3) inhibitors improve remission rate and survival compared with standard chemotherapy in patients with relapsed or refractory FLT3-mutated acute myeloid leukemia (AML).² However, single-agent FLT3 inhibitors are not curative, prompting studies of drug resistance. These data identified clonal selection for cells bearing new mutations in Ras or related signaling proteins downstream of FLT3 as well as on-target FLT3 kinase domain resistance mutations that impair drug binding or facilitate activation.^{3,4} In addition to combining FLT3 inhibitors with cytotoxic agents and moving these drugs earlier in therapy, there is a need to develop rational combinations of FLT3 inhibitors with other biologically targeted drugs. Recent data suggest nucleophosmin (NPM1) mutations, and KMT2A (MLL) translocations might identify promising populations for this approach, due to shared biologic features and cooperativity with FLT3 in leukemogenesis.

NPM1 encodes a phosphoprotein that shuttles between nucleolus and cytoplasm and contributes to histone chaperoning, ribosome trafficking, DNA damage response, and centrosome duplication. NPM1 is commonly mutated in AML by 4 base-pair insertions in exon 12 that cause frameshift, truncation, and loss of a C-terminal nucleolar localizing segment. Consequently, mutated NPM1 localizes exclusively cytoplasmic (NPM1c), which

perturbs multiple cellular functions and strongly promotes leukemogenesis. Although NPM1 mutations are thought to initiate AML, the full leukemic phenotype requires comutations. FLT3 mutations are found in approximately half of cases and can be associated with high rates of treatment failure and generally poor survival, especially with comutation of DNMT3A and/or high FLT3-internal tandem duplication (ITD) allele burden.⁵

NPM1c is mutually exclusive with other balanced translocations that initiate AML, such as RUNX1-RUNX1T1, CBFB-MYH11, PML-RARA, and lysine methyltransferase 2A (KMT2A) rearrangements (formerly mixed-lineage leukemia, MLL). This suggests overlapping functions of NPM1c and these gene fusions. Gene expression profiling highlights overlapping patterns in NPM1c and KMT2A-rearranged AML, specifically overexpression of homeobox (HOX) A cluster genes, which resemble hematopoietic stem cells' expression pattern.⁶ In particular, HOXA9 and its cofactor MEIS1 overexpression are thought to be critical for enhanced self-renewal in both of these AML subtypes.

KMT2A, the human homolog of the drosophila gene *trithorax*, is essential for both fetal and adult hematopoiesis. KMT2A is a member of a large multiprotein complex that interacts with chromatin and normally acts as a histone H3 lysine 4 (H3K4) methyltransferase, a

function catalyzed by its SET domain. Balanced translocations of *KMT2A* with >60 partner genes occur in ~3% of adult AML. These gene fusions create novel oncoproteins that initiate leukemia. Cases of AML with *KMT2A* rearrangements have among the lowest mutation burden of any described cancer, and these translocations alone may be sufficient to generate the full leukemic phenotype. Through translocation, *KMT2A* always loses its SET domain and associated H3K4 methylation activity. However, fusion oncoproteins frequently gain novel epigenetic activity via interaction with disruptor of telomeric silencing 1-like (DOT1L), an H3K79 histone methyltransferase. In addition, the translocation partner gene, canonically a transcription factor, becomes functionally deregulated to promote a stereotypical pattern of novel target gene expression, including overexpression of *FLT3*, *HOXA9*, and *MEIS1*. Transcription of the latter 2 genes promotes self-renewal and is critically dependent on the *KMT2A*-fusion protein binding to menin.

Small molecules that disrupt the protein-protein interaction between *MLL* and menin show substantial antileukemic efficacy in cellular and murine models of *KMT2A*-rearranged AML.^{7,8} In addition, using a CRISPR/cas9 negative selection screen, Kühn and Armstrong confirmed wild-type *MLL* binding to menin was critical for expression of *HOXA9*, *MEIS1*, and *FLT3* in *NPM1*-mutated AML cells.⁹ Cellular consequences of *MLL*-menin inhibition included growth arrest and induction of hematopoietic differentiation. Subsequently, this class of drugs was studied in murine models of *NPM1/FLT3* mutated AML and shown to reduce leukemic burden, eliminate established leukemia, and improve survival.¹⁰ Two drugs that inhibit *MLL*-menin binding have recently entered clinical testing as single agents for AML (K0539, NCT04067336 and SNDX-5613, NCT04065399); after safety and dose optimization are established, efficacy testing will follow in patients with *MLL*-rearrangements or *NPM1* mutations. Thus, *MLL*-menin inhibitors could emerge as important targeted therapies for *NPM1c* and *KMT2A*-rearranged AML and might open considerable therapeutic options.

Dzama and colleagues now take the logical step to combine *MLL*-menin targeted therapy with *FLT3* inhibitors in preclinical models of *FLT3*-ITD⁺ AML and *NPM1c* or *KMT2A* rearrangements. As noted above, small molecule inhibition of *MLL*-menin interaction alone clearly induces differentiation of the leukemic clone. Here, *MLL*-menin inhibitors or a menin knockdown by short hairpin downregulated and dephosphorylated *FLT3*, potentiating kinase inhibitor antileukemic effects. A marked leftward shift in the 50% inhibitory concentration for combinations over the individual drugs was observed, indicating in vitro synergy. Encouragingly, the combination of *MLL*-menin inhibition and *FLT3* inhibitor appeared to overcome clinically relevant, drug-resistant tyrosine kinase domain mutations in *FLT3*.^{3,4} Adding to these in vitro data, a murine model of *KMT2A*-rearranged, *FLT3*-ITD⁺ AML showed improved survival with the combination therapy.

These data are encouraging for eventual clinical translation, although several important questions remain unanswered. First, given known roles of *FLT3* in regulating stem and progenitor populations, there is potential for significant myelosuppression from overlapping toxicity, which might not become manifest until human testing. Second, resistance to *FLT3* inhibitors can develop by selection for cells whose growth is not dependent upon *FLT3*, including *FLT3*-wild-type clones.³ Further work is needed to characterize whether such cells' growth requires *MLL* binding menin. In addition, because clinical use of *FLT3* inhibitors induces differentiation, functional redundancy with *MLL*-menin inhibitors might fail to recapitulate the in vitro synergy suggested by preclinical studies. *MLL*-menin inhibitors also might more reasonably be combined with drugs that induce apoptosis and eliminate *NPM1* mutated cells, such as BH3 mimetics or traditional cytotoxic chemotherapy. Last, the clinical translation of these results to *KMT2A*-rearranged patients may be limited by the low incidence of *FLT3* comutations.

Overall, however, these are encouraging data with a clear translational application. Results of single-agent *MLL*-menin inhibitor

trials in *NPM1c* and *KMT2A*-rearranged AML are eagerly awaited. With these data, Dzama and colleagues have cleared a path to show the next step for dual-novel targeted agent regimens for these patients.

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REFERENCES

1. Dzama MM, Steiner M, Rausch J, et al. Synergistic targeting of *FLT3* mutations in AML via combined menin-*MLL* and *FLT3* inhibition. *Blood*. 2020;136(21):2442-2456.
2. Perl AE, Martinelli G, Cortes JE, et al. Gilteritinib or chemotherapy for relapsed or refractory *FLT3*-mutated AML. *N Engl J Med*. 2019;381(18):1728-1740.
3. McMahon CM, Ferng T, Canaan J, et al. Clonal selection with RAS pathway activation mediates secondary clinical resistance to selective *FLT3* inhibition in acute myeloid leukemia. *Cancer Discov*. 2019;9(8):1050-1063.
4. Smith CC, Wang Q, Chin CS, et al. Validation of ITD mutations in *FLT3* as a therapeutic target in human acute myeloid leukaemia. *Nature*. 2012;485(7397):260-263.
5. Papaemmanuil E, Gerstung M, Bullinger L, et al. Genomic classification and prognosis in acute myeloid leukemia. *N Engl J Med*. 2016;374(23):2209-2221.
6. Spencer DH, Young MA, Lamprecht TL, et al. Epigenomic analysis of the HOX gene loci reveals mechanisms that may control canonical expression patterns in AML and normal hematopoietic cells. *Leukemia*. 2015;29(6):1279-1289.
7. Grembecka J, He S, Shi A, et al. Menin-*MLL* inhibitors reverse oncogenic activity of *MLL* fusion proteins in leukemia. *Nat Chem Biol*. 2012;8(3):277-284.
8. Krivtsov AV, Evans K, Gadrey JY, et al. A menin-*MLL* inhibitor induces specific chromatin changes and eradicates disease in models of *MLL*-rearranged leukemia. *Cancer Cell*. 2019;36(6):660-673.e611.
9. Kühn MW, Song E, Feng Z, et al. Targeting chromatin regulators inhibits leukemogenic gene expression in *NPM1* mutant leukemia. *Cancer Discov*. 2016;6(10):1166-1181.
10. Uckelmann HJ, Kim SM, Wong EM, et al. Therapeutic targeting of preleukemia cells in a mouse model of *NPM1* mutant acute myeloid leukemia. *Science*. 2020;367(6477):586-590.

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