

REFERENCES

- Charles NJ, Boyer DF. Mixed-phenotype acute leukemia: diagnostic criteria and pitfalls. Arch Pathol Lab Med. 2017;141(11):1462-1468.
- Meyer C, Burmeister T, Gröger D, et al. The MLL recombinome of acute leukemias in 2017. *Leukemia*. 2018;32(2):273-284.
- Krivtsov AV, Armstrong SA. MLL translocations, histone modifications and leukaemia stem-cell development. Nat Rev Cancer. 2007;7(11):823-833.
- Döhner H, Estey E, Grimwade D, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood*. 2017;129(4):424-447.
- Krivtsov AV, Evans K, Gadrey JY, et al. A menin-MLL inhibitor induces specific chromatin changes and eradicates disease in models of MLLrearranged leukemia. *Cancer Cell*. 2019;36(6):660-673.e11.
- Kühn MW, Song E, Feng Z, et al. Targeting chromatin regulators inhibits leukemogenic gene expression in NPM1 mutant leukemia. *Cancer Discov*. 2016;6(10):1166-1181.
- Uckelmann HJ, Kim SM, Wong EM, et al. Therapeutic targeting of preleukemia cells in a mouse model of NPM1 mutant acute myeloid leukemia. Science. 2020;367(6477):586-590.
- Burrows F, Wu T, Kessler L, et al. A novel small molecule menin-MLL inhibitor for potential treatment of MLL-rearranged leukemias and NPM1/DNMT3Amutant AML [abstract]. *Mol Cancer Ther.* 2018;17(suppl). Abstract LB-A27.
- Daigle SR, Olhava EJ, Therkelsen CA, et al. Potent inhibition of DOT1L as treatment of MLL-fusion leukemia. *Blood*. 2013;122(6):1017-1025.

- Stein EM, Garcia-Manero G, Rizzieri DA, et al. The DOT1L inhibitor pinometostat reduces H3K79 methylation and has modest clinical activity in adult acute leukemia. *Blood.* 2018;131(24):2661-2669.
- 11. Chen CW, Koche RP, Sinha AU, et al. DOT1L inhibits SIRT1-mediated epigenetic silencing to maintain leukemic gene expression in MLL-rearranged leukemia. *Nat Med.* 2015;21(4):335-343.
- Chen CW, Armstrong SA. Targeting DOT1L and HOX gene expression in MLL-rearranged leukemia and beyond. Exp Hematol. 2015;43(8):673-684.
- Deshpande AJ, Chen L, Fazio M, et al. Leukemic transformation by the MLL-AF6 fusion oncogene requires the H3K79 methyltransferase Dot1l. *Blood.* 2013;121(13):2533-2541.
- 14. Bernt KM, Armstrong SA. A role for DOT1L in MLL-rearranged leukemias. *Epigenomics.* 2011;3(6):667-670.
- Bernt KM, Zhu N, Sinha AU, et al. MLL-rearranged leukemia is dependent on aberrant H3K79 methylation by DOT1L. Cancer Cell. 2011;20(1):66-78.
- Stauffer F, Weiss A, Scheufler C, et al. New potent DOT1L inhibitors for in vivo evaluation in mouse. ACS Med Chem Lett. 2019;10(12):1655-1660.
- Wang K, Sanchez-Martin M, Wang X, et al. Patient-derived xenotransplants can recapitulate the genetic driver landscape of acute leukemias. *Leukemia*. 2017;31(1):151-158.

DOI 10.1182/blood.2020006113

© 2020 by The American Society of Hematology

TO THE EDITOR:

Sialic acid-bearing paraproteins are implicated in heparin-like coagulopathy in patients with myeloma

Abdulrahman Saadalla,¹ Jansen Seheult,² Paula Ladwig,¹ Julie Tange,¹ Rachel Leger,¹ Mindy Kohlhagen,¹ Aneel Ashrani,^{1,3} and David Murray¹

¹Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN; ²Department of Pathology, University of Pittsburgh, Pittsburgh, PA; and ³Department of Internal Medicine, Mayo Clinic, Rochester, MN

Plasma cell dyscrasias, including multiple myeloma (MM), have been associated with acquired bleeding diathesis.^{1,2} Rarely, this coagulopathy can be due to the presence of circulating heparinlike anticoagulant.³⁻⁵ We present 2 patients with MM with hemorrhagic complications due to the presence of heparin-like anticoagulant that we attribute to increased negatively charged sialic acid (SA)-bearing monoclonal paraproteins.

Patient 1

A 73 year-old man with relapsed immunoglobulin G (IgG) MM on elotuzumab, lenalidomide, and dexamethasone presented with severe epistaxis leading to a hemoglobin drop to 5 g/dL, necessitating packed red blood cell transfusion and local measures to control epistaxis. Platelet count was 35000 per microliter, requiring platelet transfusion. No other hemostatic agents were used. He had prolonged activated partial thromboplastin time (APTT) and prothrombin time (PT), as well as dilute Russell viper venom (dRVV) screening time; these did not correct on 1:1 mixing with pooled normal plasma (PNP), suggesting the presence of an inhibitor. The dRVV confirmatory test provided no evidence for lupus-like anticoagulant activity. Testing of individual coagulation factors for prolonged PT and APTT workup did not reveal any coagulation factor deficiencies. Notably, thrombin time (TT) was prolonged, with normal reptilase time (RT), a pattern attributed to heparin or direct thrombin inhibitor (DTI) effects; however, no source of exposure to exogenous heparin/DTI was identified. His hepatic transaminases, bilirubin, and alkaline phosphatase were within the normal reference range. Measured serum M-spike and λ free light chain (FLC) levels were at their highest since the initial MM diagnosis 5 years earlier (Figure 1A-B). For his aggressive disease, he had received multiple biochemotherapeutic regimens, including autologous stem cell transplant (ASCT) 11 months postdiagnosis. Interestingly, prior to the placement of a central venous catheter in preparation for ASCT, his screening APTT and PT were prolonged (66 seconds and 14.2 seconds, respectively), although they were not evaluated further.

The occurrence of the bleeding and coagulopathy at the time of MM relapse was at the peak of monoclonal paraprotein and FLC concentrations, leading us to hypothesize that the elevated monoclonal protein could be the cause of the heparin-like anticoagulant effects. To investigate further, we retrieved archived sera obtained from the patient at 4 time points, including at MM diagnosis and following the bleeding episode. Paraproteins were purified from the sera using camelid-derived single domain antibody coated beads (CaptureSelect; Thermo Fisher Scientific) specific for κ or λ light chain (LC) or for IgG, IgM, or IgA heavy chains.⁶ Eluted reduced immunoglobulins were analyzed by matrix-assisted laser desorption/ionization time-of-flight

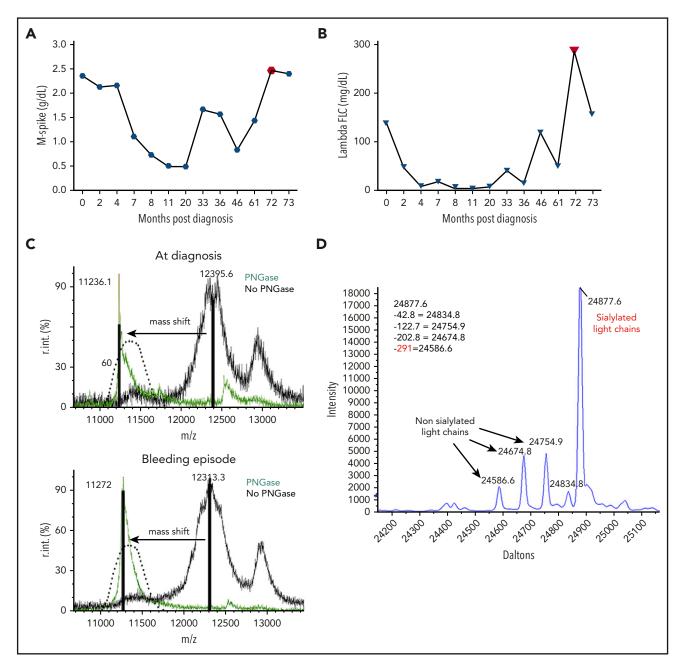


Figure 1. Characterization of monoclonal protein over the course of disease. Line graphs of measured serum M-spike (A) and λ FLC levels (B) in patient 1 at different testing time points. Diagnosis = 0 months; red point at 72 months is the bleeding time point. (C) Overlaid MALDI-TOF/MS LC [M +2H]²⁺ spectra of immunopurified IgG of patient 1 at diagnosis (upper panel) and bleeding (lower panel) time points. λ LCs (vertical black lines) are of a greater mass than predicted normal range (dotted lines). Arrows show mass shifts from untreated IgG (black graph) to PNGase F-treated IgG (green graph), indicating the cleavage of N-linked glycan structures by PNGase F. (D) Deconvoluted λ LC mass spectra generated by high-resolution MS of immunopurified IgG. Arrows point to sialylated and nonsialylated forms; 291 Da corresponds to SA.

(MALDI-TOF) mass spectrometry (MS) (microflex LT; Bruker, MA). The resulting spectra were consistent with an IgG λ paraprotein, and all time points had congruent mass/charge (*m/z*) values, indicating persistence of the same plasma cells clone. Notably, the λ LCs were of higher *m/z* than the expected typical mass range (Figure 1C). We immunopurified IgG from sera collected at diagnosis and bleeding time points and treated it overnight with PNGase F (New England Biolabs, MA) to remove *N*-linked glycans. PNGase F treatment reduced the λ LC mass by 2000 Da, to 2100 Da, indicating the presence of *N*-linked glycans on the LC (Figure 1C). To further characterize

these glycans, purified IgG was analyzed using liquid chromatography coupled to high-resolution electrospray ionization time-of-flight MS (TripleTOF 5600; Sciex).^{7,8} Deconvoluted spectra revealed monoclonal LCs with δ masses of 291 Da, corresponding to SA (Figure 1D). Like heparin, the highly negatively charged SA containing LCs may bind positively charged motifs present on thrombin and antithrombin, mimicking heparin-like anticoagulant in vitro and potentially in vivo. This effect may be concentration dependent and might be most pronounced at high levels of monoclonal paraproteinemia.

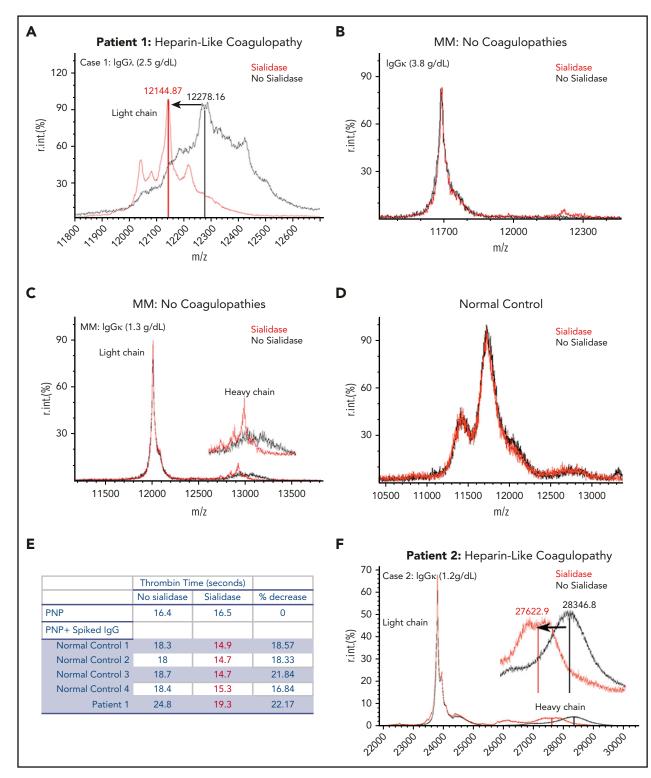


Figure 2. Properties of the monoclonal protein that altered the coagualtion profile. (A) Overlaid MALDI-TOF/MS LC [M +2H]²⁺ spectra of sialidase-treated and untreated immunopurified IgG from patient 1. Arrow indicates mass shift of sialidase-treated IgG. Mass spectra similar to (A) of 2 patients with myeloma with no bleeding or coagulation abnormalities (B-C) and a normal healthy control (D). Note the absence of mass shifts in LC. (C) Smaller heavy chain mass shifts were noted with sialidase treatment (inset). (E) Table of clot-based TT measured in sialidase-treated or untreated PNP and PNP spiked with sialidase-treated or untreated immunopurified IgG from sera of patient 1 and 4 normal controls, at 1:10 ratio. The percentage decrease in TT with sialidase treatment is shown. (F) MALDI-TOF spectra of sialidase-treated or untreated immunopurified IgA from patient 2. Arrow shows a clear mass shifts in sialidase-treated IgA heavy chain (inset). No LC mass shift was noted with sialidase treatment.

To confirm the presence of SA residues on the monoclonal immunoglobulins, immuno-purified IgG was treated with sialidase enzyme (Roche Diagnostics) overnight and analyzed with MALDI-TOF/MS. λ LC in the sialidase-treated samples demonstrated a mass shift, indicating the presence of SA (Figure 2A), whereas treatment of sera collected from 3 patients with MM without coagulation abnormalities, as well as from 4 normal controls, did not display any LC mass shifts (Figure 2B-D). Of note, occasional smaller shifts were noted on the immunoglobulin heavy chain peaks (Figure 2C).

To test the effect of LC SA on the action of thrombin and TT, purified IgG from serum collected at the bleeding time point was split and were either untreated or treated untreated with sialidase, and then spiked at a 1:10 ratio into PNP (Precision BioLogic). TT was consistently shortened in all plasma samples spiked with sialidase-treated IgG but not in controls (Figure 2E). The addition of sialidase enzyme to PNP did not alter the TT (Figure 2E). Based on the above findings, we hypothesize that the increased LC sialylation of the patient's monoclonal paraprotein contributed to the TT prolongation in the setting of a normal RT and the heparin-like activity in vitro and in vivo.

Patient 2

A 75-year-old woman, with relapsed IgAk MM on melphalan, prednisone, and thalidomide, developed hemoptysis and respiratory failure soon after hospitalization for acute-on-chronic renal failure secondary to cast nephropathy. Her serum IgA M-spike was 1.2 g/dL with κ FLC of 924 mg/dL. She had prolonged APTT and dRVV screen not corrected on 1:1 mixing with PNP, suggesting the presence of an inhibitor. The PT was within the normal reference range. Testing of individual coagulation factors for prolonged APTT workup did not reveal any coagulation factor deficiencies. The TT was prolonged, with normal RT, suggesting heparin-like anticoagulant effects; however, she had no heparin/DTI exposure. Heparin anti-Xa activity was at the lowest level of detection (0.05 IU/mL). She was treated with high-dose Solu-Medrol, plasmapheresis, and hemodialysis for cast nephropathy. Because platelet transfusion and protamine sulfate were not effective in controlling her hemoptysis, recombinant factor VIIa (NovoSeven) was administered, with temporary improvement. She expired on day 15 following hospitalization.

Her archived serum samples were retrieved for analysis. Similar analysis of immunopurified paraprotein by MALDI-TOF/MS showed LC mass shift with PNGase F treatment (data not shown), but not with sialidase treatment, indicating the absence of SA on the LC (Figure 2F). However, the heavy chains showed a clear mass shift following sialidase treatment. The 23-aa-long hinge region of the IgA1 heavy chain molecule typically harbors 3 to 6 variably sialylated *O*-linked glycan structures.⁹ We hypothesize that the abundantly sialylated IgA paraproteins with disease progression led to an exaggerated net negative charge and heparin-like activity that may have contributed to bleeding manifestation.¹⁰

The negatively charged SA residues present in the glycan structures linked to the circulating monoclonal immunoglobulins may function similarly to heparin by binding to antithrombin and potentiating its activity; alternatively, they may have direct antithrombin-like activity by binding to and inhibiting thrombin directly, with no effect on RT.¹¹ This is supported by the finding that sialidase-treated monoclonal paraproteins did not prolong the TT, as was noted with the untreated paraprotein. Mártinez-Mártinez et al presented an IgG λ MM patient with prolonged TT and normal RT and demonstrated paraprotein affinity to

antithrombin.¹² In most reported cases, positively charged protamine sulfate has been the mainstay treatment to manage acute bleeding episodes^{1,13}; however, it was not effective in patient 2. Chemotherapy has also been reported as essential in resolving the coagulopathy.^{3,14-16}

In summary, we suggest that the increased net negative charge of heavily sialylated paraproteins may be one of the mechanisms for its heparin-like anticoagulant effects in MM patients.

Authorship

Contribution: A.S. designed and performed experiments and wrote the manuscript; P.L., R.L., J.T., and M.K. performed experiments; and J.S., A.A., and D.M. designed experiments and provided supervision.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: J.S., 0000-0002-6850-7495; P.L., 0000-0002-6176-4501.

Correspondence: David Murray, Department of Laboratory Medicine and Pathology, Mayo Clinic, 200 First St SW, Rochester, MN 55905; e-mail: murray.david@mayo.edu.

Footnotes

Submitted 28 February 2020; accepted 2 June 2020; prepublished online on *Blood* First Edition 26 June 2020.

Data sharing requests should be sent to David Murray (murray.david@ mayo.edu).

REFERENCES

- Eby C. Pathogenesis and management of bleeding and thrombosis in plasma cell dyscrasias. Br J Haematol. 2009;145(2): 151-163.
- Coppola A, Tufano A, Di Capua M, Franchini M. Bleeding and thrombosis in multiple myeloma and related plasma cell disorders. Semin Thromb Hemost. 2011;37(8):929-945.
- Llamas P, Outeiriño J, Espinoza J, Santos AB, Román A, Tomás JF. Report of three cases of circulating heparin-like anticoagulants. Am J Hematol. 2001;67(4):256-258.
- Palmer RN, Rick ME, Rick PD, Zeller JA, Gralnick HR. Circulating heparan sulfate anticoagulant in a patient with a fatal bleeding disorder. N Engl J Med. 1984;310(26):1696-1699.
- Torjemane L, Guermazi S, Ladeb S, et al. Heparin-like anticoagulant associated with multiple myeloma and neutralized with protamine sulfate. Blood Coagul Fibrinolysis. 2007;18(3):279-281.
- Mills JR, Kohlhagen MC, Dasari S, et al. Comprehensive assessment of M-proteins using nanobody enrichment coupled to MALDI-TOF mass spectrometry. *Clin Chem.* 2016;62(10):1334-1344.
- Bondt A, Rombouts Y, Selman MH, et al. Immunoglobulin G (IgG) Fab glycosylation analysis using a new mass spectrometric high-throughput profiling method reveals pregnancy-associated changes. *Mol Cell Proteomics*. 2014;13(11):3029-3039.
- Barnidge DR, Dasari S, Botz CM, et al. Using mass spectrometry to monitor monoclonal immunoglobulins in patients with a monoclonal gammopathy. J Proteome Res. 2014;13(3):1419-1427.
- Förger F, Villiger PM. Immunological adaptations in pregnancy that modulate rheumatoid arthritis disease activity [published correction appears in Nat Rev Rheumatol. 2020 Mar;16(3):184]. Nat Rev Rheumatol. 2020;16(2):113-122.

- 10. Mattu TS, Pleass RJ, Willis AC, et al. The glycosylation and structure of human serum IgA1, Fab, and Fc regions and the role of N-glycosylation on Fc α receptor interactions. *J Biol Chem.* 1998;273(4):2260-2272.
- 11. Carter WJ, Cama E, Huntington JA. Crystal structure of thrombin bound to heparin. J Biol Chem. 2005;280(4):2745-2749.
- Martínez-Martínez I, González-Porras JR, Cebeira MJ, et al. Identification of a new potential mechanism responsible for severe bleeding in myeloma: immunoglobulins bind the heparin binding domain of antithrombin activating this endogenous anticoagulant. *Haematologica*. 2016;101(10):e423-e426.
- Willner CA, Chisti MM. Treatment of bleeding diathesis associated with a heparin-like anticoagulant in plasma cell neoplasia using protamine. Case Rep Hematol. 2018;2018:4342301.
- Chapman GS, George CB, Danley DL. Heparin-like anticoagulant associated with plasma cell myeloma. Am J Clin Pathol. 1985;83(6): 764-766.
- Tefferi A, Nichols WL, Bowie EJ. Circulating heparin-like anticoagulants: report of five consecutive cases and a review. Am J Med. 1990;88(2): 184-188.
- Khoory MS, Nesheim ME, Bowie EJ, Mann KG. Circulating heparan sulfate proteoglycan anticoagulant from a patient with a plasma cell disorder. *J Clin Invest.* 1980;65(3):666-674.

DOI 10.1182/blood.2020005604

© 2020 by The American Society of Hematology