#### RARE SYSTEMIC HEMATOLOGIC DISORDERS

## Langerhans cell histiocytosis

Carlos Rodriguez-Galindo<sup>1,2</sup> and Carl E. Allen<sup>3</sup>

<sup>1</sup>Department of Global Pediatric Medicine and <sup>2</sup>Department of Oncology, St. Jude Children's Research Hospital, Memphis, TN; and <sup>3</sup>Division of Pediatric Hematology-Oncology, Department of Pediatrics, Texas Children's Cancer Center and Baylor College of Medicine, Houston, TX

Langerhans cell histiocytosis (LCH) is caused by clonal expansion of myeloid precursors that differentiate into CD1a<sup>+</sup>/CD207<sup>+</sup> cells in lesions that leads to a spectrum of organ involvement and dysfunction. The pathogenic cells are defined by constitutive activation of the MAPK signaling pathway. Treatment of LCH is risk-adapted: patients with single lesions may respond well to local treatment, whereas patients with multisystem disease require systemic therapy. Although survival rates for patients without organ dysfunction is excellent, mortality rates for patients with organ dysfunction may reach 20%. Despite progress made in the treatment of LCH, disease reactivation rates remain above 30%, and standard second-line treatment is yet to be established. Treatment failure is associated with increased risks for death and long-term morbidity, including LCH-associated neurodegeneration. Early case series report promising clinical responses in patients with relapsed and refractory LCH treated with BRAF or MEK inhibitors, although potential for this strategy to achieve cure remains uncertain. (*Blood.* 2020;135(16):1319-1331)

#### Introduction

Langerhans cell histiocytosis (LCH) is a disease characterized by clonal expansion of myeloid precursors that differentiate into CD1a<sup>+</sup>/CD207<sup>+</sup> in lesions. It presents at all ages with various degrees of systemic involvement, and although cure rates are high, severe long-term neurological or endocrine complications may affect quality of life. Our understanding of the pathogenesis of the disease has evolved from a reactive clonal proliferation of Langerhans cells (LCs) to an inflammatory myeloid neoplasia; this evolution from a disorder of immune dysregulation to a bona fide neoplastic disorder has reclassified the disease and opened the door for the development of targeted therapies.

## Epidemiology

The reported incidence of LCH ranges from 2.6 to 8.9 cases per million children younger than 15 years per year, with a median age at diagnosis of 3 years.<sup>1-4</sup> The exact incidence of LCH in adults is much less defined: the only available data are for disseminated disease, with 0.07 cases per million per year.<sup>5,6</sup>

The causes and risk factors for developing LCH are unclear.<sup>7</sup> Population-based studies have shown differences in the incidence of multisystem LCH by race and ethnicity; in children, higher incidence has been reported for Hispanics, and lower for blacks.<sup>8</sup> The lower incidence in the black population has also been reported in adults.<sup>6</sup> In a population-based, case-control study, Hispanic mothers were more likely to have children who developed LCH compared with non-Hispanic whites; this risk increased when both parents were Hispanic. Non-Hispanic black mothers were less likely to give birth to offspring who developed LCH compared with non-Hispanic whites.<sup>9</sup> The association with Hispanic ancestry has been further documented in a genomewide association study, which identified a novel risk variant within *SMAD6*.<sup>10</sup> The observation of increased incidence in monozygotic twins of affected patients has also suggested the presence of a germline predisposition, at least for a small proportion of cases.<sup>11</sup>

The association between LCH and other malignancies has been described, with frequencies varying from 2.6% in children<sup>12</sup> to 32% in adults.<sup>13</sup> Different solid malignancies have been associated with LCH; lung carcinoma has been consistently reported in adult series, 13,14 and thyroid carcinoma has been noted to occur in conjunction with thyroid infiltration by LCH in both adults and children.<sup>15,16</sup> Hodgkin and non-Hodgkin lymphomas have been described in association with LCH, often occurring concurrently in the same nodes.<sup>14,17</sup> The most common hematologic malignancy reported is acute myeloid leukemia, often occurring years after LCH.<sup>12-14,18</sup> In contrast, LCH in association with acute lymphoblastic leukemia (ALL) commonly occurs during treatment.<sup>18-20</sup> Reports have shown that coincident LCH and ALL share the same oncogenic mutations or have an identical T-cell receptor or immunoglobulin rearrangement, suggesting the presence of a clonal relationship between LCH and ALL.<sup>19,20</sup>

## Pathology

More than 100 different histiocytic disorders have been described, and a classification system consisting of 5 groups of diseases has been proposed: LCH-related, cutaneous and mucocutaneous non-LCH histiocytoses, Rosai-Dorfman disease,

#### Table 1. Classification of histiocytoses

Histiocytosis group	Diseases	
L Group	LCH Indeterminate-cell histiocytosis (ICH) Erdheim-Chester Disease (ECD) Mixed LCH/ECD	
C Group	Cutaneous non-LCH Xanthomatous granuloma (XG) family: JXG, AXG, SRH, BCH, GEH, PNH Non-XG family: cutaneous RDD, NXG, other Cutaneous non-LCH with a major systemic component XG family: XD Non-XG family: MRH	
R Group	Familial RDD Sporadic RDD Classical RDD Extranodal RDD RDD with neoplasia or immune disease Unclassified	
M Group	Primary Malignant Histiocytoses Secondary Malignant Histiocytoses	
H Group	Primary HLH: Monogenic inherited conditions leading to HLH Secondary HLH (non-Mendelian HLH) HLH of unknown/uncertain origin	

Adapted from Emile et al.<sup>21</sup>

AXG, adult xanthogranuloma; BCH, benign cephalic histiocytosis; GEH, generalized eruptive histiocytosis; JXG, juvenile xanthogranuloma; MRH, multicentric reticulohistiocytosis; NXG, necrobiotic xanthogranuloma; PNH, progressive nodular histiocytosis; RDD, Rosai-Dorfman Disease; SRH, solitary reticulohistiocytoma; XD, xanthoma disseminatum.

malignant histiocytoses, and hemophagocytic lymphohistiocytosis and macrophage activation syndrome (Table 1). $^{21}$ 

Diagnosis of LCH requires a clonal neoplastic proliferation with expression of CD1a, CD207 (Langerin), and S100 (Figure 1). The cells are generally large, round to oval in shape, with a coffee-bean nuclear grove, and without the branching that characterizes inflammatory CD1a<sup>+</sup> dendritic cells. On electron microscopy, pentalaminar cytoplasmic rod-shaped inclusions (Birbeck granules) can be identified, although electron microscopy is no longer required for diagnosis in the presence of CD207<sup>+</sup> staining. Because LCH cells activate and recruit other immunologic cells, microscopic examination shows an inflammatory pattern consisting of eosinophils, neutrophils, lymphocytes, and macrophages in addition to the LCs; this appearance is what is described as eosinophilic granuloma.<sup>22</sup>

## **Clinical presentation**

LCH presents in a continuum of systemic involvement, ranging from a solitary eosinophilic granuloma to widespread disseminated disease with organ dysfunction.<sup>23</sup> The current classification system is based on the site of lesions, number of involved sites (single or multisystem/local or multifocal), and whether the disease is involving risk (of mortality) organs (hematopoietic system, liver, or spleen; Tables 2 and 3). In a large cohort review, single-system and multisystem disease accounted for approximately half of the patients each. Among the patients with multisystem disease, approximately 15% of them had involvement of a risk organ.<sup>24</sup> The skeleton is the most commonly affected system, as bone lesions are present in approximately 80% of patients with LCH,<sup>25</sup> and in half of them, lesions are single.<sup>24</sup> The most common site of bone involvement is the skull, followed by spine, limbs, and pelvis.<sup>26</sup>

Most organs can be affected by LCH, and therefore a comprehensive evaluation is indicated.<sup>27</sup> Symptoms and physical and laboratory examination should guide the extent of diagnostic studies; focus should be on assessing the number of systems and sites involved, and on the involvement of risk organs. Bone imaging studies reveal a lytic lesion without marginal sclerosis, with or without periosteal reaction. (Figure 2) Radioisotope imaging is recommended to assess the number of bone lesions; fluorodeoxyglucose-positron emission tomography scans can be useful in defining the extent of the disease and the response to therapy.<sup>28,29</sup> (Figure 2) The skull, including the skull base, is very commonly involved; typical locations include the bones of the orbit or the temporal bone (typically the mastoid). Involvement of the vertebral bodies is also common, and the presence of a vertebra plana is frequent. Pain and tumor formation in a localized area of bone is a very common presentation of LCH.

Skin involvement is also common, particularly in infants, where it presents as seborrheic eczema, and in adults, where it may present as refractory eczema in intertriginous and genital areas (Figure 3). Isolated skin involvement usually carries a good prognosis, with an approximately 60% chance of regression with topical treatments.<sup>30-34</sup> The lesions of congenital self-healing LCH are often present at or shortly after birth; can appear as eroded or ulcerated papules, pustules, or vesicles with hemorrhagic crusting; and may masquerade as diffuse neonatal hemangiomatosis<sup>33</sup> or blueberry muffin rash.<sup>34</sup> Close monitoring is required in infants, as reactivation or progression to multisystem involvement has been observed in up to 40% of cases.<sup>35,36</sup> In adults, cutaneous involvement commonly presents as papules and intertrigo, with significant scaling and crusting, most commonly in the scalp, although mucosal involvement of the genitalia or oral cavity is also common.37-39

In children, lung involvement usually occurs in the context of multisystem disease, where it has been reported to occur in up to 35% of patients.<sup>40</sup> Radiographic findings are typical for the presence of a reticulonodular pattern with bullae formation (Figure 2). In the absence of other risk organ involvement, pulmonary disease is not a predictor of adverse outcome.<sup>41-43</sup> Isolated pulmonary involvement is a rare presentation that is almost exclusive of adults with a smoking habit.<sup>44</sup>

The presence of hematopoietic dysfunction in the form of cytopenias is a poor prognostic sign. It occurs in the context of multisystem involvement, usually in very young children. Its pathophysiology is multifactorial, including direct involvement of the bone marrow as well as peripheral destruction resulting from hypersplenism from LC infiltrates in the spleen.<sup>45</sup>



Figure 1. LCH lesion histology. Images demonstrate typical histology of LCH lesion obtained from a bone biopsy with pathologic histiocytes and inflammatory infiltrate. (A-B) Hematoxylin and eosin stain demonstrates histiocytes with pale cytoplasm and reniform nuclei. (C) Birbeck granules on electron microscopy; immunohistochemistry strongly positive for (D) langerin/CD207, (E) CD1a, and (F) S100a. (Courtesy of M. J. Hicks)

Liver involvement also carries a very poor prognosis. Patients present with hypoalbuminemia, hepatomegaly, or conjugated hyperbilirubinemia. A well-described but rare complication in young children and in adults is the development of sclerosing cholangitis and hepatic fibrosis, which commonly evolve to end-stage liver failure.<sup>46-48</sup>

#### LCH in Adults

Adult LCH usually presents after the fourth decade, and approximately two-thirds of patients have multisystem involvement at diagnosis.<sup>49</sup> Its association with other neoplastic diseases is common, especially other myeloproliferative neoplasms.<sup>13</sup> In a

#### Table 2. Clinical Classification of LCH

significant proportion of patients, LCH and Erdheim-Chester lesions may coexist.<sup>50</sup> In general, the clinical presentation and organs involved are similar to pediatric patients, with the more frequent involvement of the genitalia, particularly in females.<sup>37,49</sup> The rarity of LCH in adults, combined with the nonspecific and varied clinical presentations, typically result in missed and delayed diagnosis.<sup>51,52</sup>

#### **Pulmonary LCH**

Isolated pulmonary LCH (PLCH) is primarily a disease of young adult smokers, with more than 90% of patients endorsing a smoking history.<sup>53-55</sup> PLCH presents with respiratory symptoms,

Clinical group	Description	
<b>Multisystem</b> With risk organ involvement Without risk organ involvement	Two or more systems involved Involvement of liver, spleen or bone marrow Without involvement of liver, spleen or bone marrow	
<b>Single-system</b> Single site Multiple sites Special site	Only 1 system involved Skin, bone, lymph node, other (thyroid, thymus) Multifocal bone disease Skull-base lesion with intracranial extension or vertebral lesion with intraspinal soft tissue extension	
Pulmonary LCH	Isolated lung disease	
CNS LCH	Tumorous lesions Neurodegenerative disease LACI LACS	

#### Table 3. Definition of risk organ involvement in LCH-IV

Hematopoietic involvement (with or without bone marrow involvement*), at least 2 of the following:		
Anemia: hemoglobin <100 g/L (<10 g/dl), infants <90 g/L (<9.0 g/dL), not a result of other causes (eg, iron deficiency) Leukocytopenia: leukocytes <4.0 × 10°/L (4000/μL) Thrombocytopenia: platelets <100 × 10°/L (100 000/μL)		
<b>Spleen involvement enlargement:</b> >2 cm below costal margin in the midclavicular line†		

#### Liver involvement, one or more of the following:

Enlargement >3 cm below costal margin in the midclavicular line† Dysfunction (ie, hypoproteinemia <55 g/L, hypoalbuminemia <25 g/L, not as a result of other causes) Histopathological findings of active disease

\*Bone marrow involvement is defined as presence of CD1a positive cells on marrow slides.

†Enlargement in centimeters below the costal margin as assessed by physical examination.

mainly cough and dyspnea on exertion, in approximately twothirds of the cases. Less frequently, patients may present with spontaneous pneumothorax or with asymptomatic lesions on routine chest X-ray.<sup>56</sup> Extrapulmonary organ involvement occurs in 10% to 15% of patients.<sup>54</sup> High-resolution CT shows a pattern of bilateral reticulonodular and cystic changes, with apical and midlung predominance, sparing the bases and costophrenic angles.<sup>56</sup> Transbronchial lung biopsies may be diagnostic of PLCH in expert centers; however, because of the focal nature of the disease, the diagnostic yield varies between 15% and 40%, and a thoracoscopic lung biopsy is usually recommended.<sup>56</sup>

Pathology of PLCH shows nodular lesions with the typical histology. In late disease, nodules are replaced by advanced bullous and cystic lesions, often in association with hyperinflation and honeycombing.<sup>57</sup> *BRAFV*600E and *MAP2K1* mutations have been reported at similar frequency as in extrapulmonary LCH, although lower mutation rates are identified in the more fibrotic lesions.<sup>58,59</sup>

#### LCH of the central nervous system

Central nervous system (CNS) involvement in LCH (LCH-CNS) represents a spectrum of diseases ranging from active infiltration by LCH to long-term effects. Its prevalence has been noted to range from 3.4% to 57%.<sup>60</sup> LCH-CNS can be divided in focal mass lesions and lesions associated with progressive neurodegeneration.<sup>60,61</sup>

Mass lesions tylically present in meninges, choroid plexus, and brain parenchyma. Characteristic neuroimaging findings include hypothalamic-pituitary involvement, often with diabetes insipidus, infundibular thickening, and absent bright spot in posterior pituitary; enlargement and enhancement of the pineal gland; thickening and enhancement of choroid plexus; or intraparenchymal masses.<sup>61</sup> Among patients with anterior pituitary dysfunction, the most common deficiency is in antidiuretic hormone, followed by growth hormone (which occurs in up to 50% of patients with diabetes insipidus), gonadotropin, and thyrotropin.<sup>62,63</sup> Anterior pituitary dysfunction is more common in childhood-onset patients and in those with multisystem disease.<sup>12,62</sup> Diabetes insipidus, the hallmark of this dysfunction, has been reported to occur in up to 24% of patients with LCH, but in half of patients with multisystem disease<sup>12,64</sup>; in one third of cases, the diabetes insipidus precedes or is concurrent with the diagnosis of LCH, and in the remaining two-thirds of the cases, it is diagnosed later.<sup>12,40,65</sup> With the use of more comprehensive risk-adapted management of LCH, the incidence of endocrinopathies has decreased to 10% to 15% in recent large cohort studies.<sup>24,66</sup>

Neurodegenerative LCH (LCH-ND) is characterized by progressive radiologic and clinical abnormalities. As recently reviewed by Yeh et al,<sup>60</sup> 2 separate clinical forms are identified: LCH-associated abnormal CNS imaging (LACI), which includes asymptomatic patients with radiologic findings, and LCHassociated abnormal CNS symptoms (LACS), which describes patients with abnormal cognitive and psychological findings. LACI and LACS are associated with increased T2-weighted MRI signal in the dentate nucleus of the cerebellum, basal ganglia, and pons (Figure 2; Table 4).



Figure 2. LCH imaging. Images demonstrate typical presentation of LCH lesions including (A) single skull lesion on X-ray, (B) vertebra plana on X-ray, (C) femur lesion on X-ray, (D) vertebral lesion on positron emission tomography/computed tomography scan, and (E) cystic lung disease on computed tomography scan. (F) Brain magnetic resonance imaging demonstrates T2 hyperintensity in cerebellum in LCH-associated neurodegeneration. (Courtesy of P. Campbell) Figure 3. LCH skin disease. A range of LCH skin disease is presented. Rashes and skin lesions are pleotropic in LCH and can mimic other more common pediatric rashes associated with (A) cradle cap, (B) eczema, (C) scarlet fever or scalded skin syndrome, (D) herpes gingivostomatitis, and (E) immune thrombocytopenic purpura. (Courtesy of P. Campbell)



LACS is a neurodegenerative syndrome of variable severity and course. The incidence of long-term neurodegeneration has been estimated to be between 1.9% and 11%,<sup>12,67</sup> and it seems to be higher in patients with multisystem disease, diabetes insipidus, history of involvement of bones of the skull base and orbit,  $^{12,64,67,68}$  or *BRAFV*600E-mutated LCH.<sup>47</sup> Of particular therapeutic relevance are the skull-based lesions (CNS-risk lesions), as this risk association has been considered an indication for the use of systemic therapy, rather than local control measures only.

Lesion type and site	Pathology	MRI characteristics
Tumorous lesions Cerebral white and gray matter	Typical LCH morphology with CD1a/CD207 + histiocytes	Nodular or space-occupying lesions; T2 hyperintensity and T1 iso- or hypointensity; variably contrast enhancing; can present mass effect
LACI		
Dentate nuclei of the cerebellum	Loss of Purkinje cells with gliosis in the cerebellar cortex	Bilateral and symmetrical slight T1-w hyperintensity, followed by development of T1-w hypointensity and/or T2-w hyperintensity
Infratentorial white matter (cerebellum, brainstem)	Neuroaxonal loss with secondary demyelination; pronounced inflammatory process dominated by CD8 <sup>+</sup> T-lymphocytes and microglial activation; BRAFV600E <sup>+</sup> perivascular myeloid cells and increased frequency of <i>BRAFV</i> 600E <sup>+</sup> peripheral blood mononuclear cells (for patients with BRAFV600E <sup>+</sup> systemic LCH)	Bilateral and symmetrical abnormalities (T2-w hyperintensity, T1-w isointensity or hypointensity)
Basal ganglia	_	Bilateral and symmetrical leukoencephalopathy-like abnormalities, or confluent lesions in a vascular pattern, with T2 hyperintensity and T1 hypointensity
Supratentorial white matter	Reactive gliosis and microglial activation decreased <i>BRAFV600E</i> <sup>+</sup> cells compared with cerebellum/brainstem	Bilateral and symmetrical leukoencephalopathy-like abnormalities, or confluent lesions in a vascular pattern, with T2 hyperintensity and T1 hypointensity
<b>Prominent, dilated perivascular spaces</b> Cerebral white matter	_	Bilateral and symmetrical punctate lesions in a vascular pattern. T2-w hyperintensity, and T1 iso- or hypointensity; variable contrast enhancement and mass effect

#### Table 4. Clinical, histological, and imaging characteristics of LCH of the central nervous system

Adapted from Yeh et al<sup>60</sup> with permission.



The appearance of clinical and radiographic signs of LCH-ND can occur with the initial LCH diagnosis, although it commonly occurs years later.<sup>60,61,69</sup> Symptoms may initially include tremors, abnormal reflexes, gait disturbance, motor spasticity, ataxia, dysarthria, dysphagia, behavioral changes, learning disorder, or psychiatric problems. Some patients develop a progressive cerebellar syndrome, with spastic tetraparesis, pseudobulbar palsy, and cognitive deterioration.<sup>60,61</sup> Magnetic resonance imaging shows a characteristic infratentorial predilection, with symmetric abnormalities of the dentate nuclei and of the white matter of the cerebellum and pons (Figure 2; Table 4). Outside the infratentorial compartment, abnormalities of the basal ganglia, optic nerves, and tracts; dilatation of the Virchow-Robin spaces; or diffuse abnormalities of the hemispheric white matter consistent with leukoencephalopathy are also common.<sup>60,61</sup> Serial imaging and neurocognitive evaluations are recommended when the disease is suspected.60,70

Whether CNS involvement with degeneration represents active disease or a radiologic scar remains undefined. Until recently, the only histologic study of LACS reported absence of CD1a<sup>+</sup> histiocytes, an inflammatory collection of CD8<sup>+</sup> lymphocytes with neuronal and axonal degeneration, and extensive myelin loss, supporting the view of a late consequence of an inflammatory phenomenon.<sup>71</sup> However, a recent study supports hematopoietic origin of myeloid cells that share precursors with LCH lesion CD207<sup>+</sup> cells.<sup>72</sup> Clinical and radiological responses to BRAF inhibitors further support this view.<sup>73</sup>

## **Biology**

#### **Ontogeny and function of epidermal LCs**

During development, a wave of LCs arises from yolk sac progenitors and fetal liver-derived monocytes and seeds the epidermis. This population is maintained locally, with tissueresident precursors during steady state. However, when skin is injured or inflamed, monocyte-derived blood cells have the ability to migrate to epidermis and differentiate into LC-like cells.<sup>74-76</sup> Migration of activated LC from epidermis to draining lymph node is dependent on C-C motif chemokine receptor 7 (CCR7).<sup>77</sup> In the lymph node, LCs present antigen and activate T cells (Figure 4A).

#### The evolving identity of LCH

Before LCH was histologically defined, cases of children with unusual constellations of symptoms were described. In the mid-1900s, pathologists noted the common histology of those disease presentations and hypothesized these conditions to represent a common disease entity.<sup>78,79</sup> Lichtenstein proposed the nomenclature of Histiocytosis X, reflecting the uncertain cell of origin. In the 1960s, the Birbeck granules were identified and were thought to be exclusively associated with epidermal LCs (Figure 1). When Nezelof and colleagues identified Birbeck

granules in Histiocytosis X lesion cells, Histiocytosis X was reframed as LC histiocytosis.  $^{\rm 80}$ 

#### Inflammation and LCH

Histologic similarities between LCH cells and epidermal LCs set the stage for a long-standing debate about LCH as a disorder of pathologic activation of epidermal LCs vs neoplastic transformation.<sup>81</sup> The physiologic function of dendritic cells is to interact with and activate T cells, and LCH lesions are characterized by a robust immune infiltrate, although mechanisms driving inflammation are not well understood. The pathologic CD207<sup>+</sup> dendritic cells (LCs) constitute a median of 8% of LCH lesion cells.<sup>82</sup> The remainder of the lesion is composed of inflammatory infiltrate, including a significant population of T cells (enriched for activated CD4<sup>+</sup> regulatory suppressor T cells) and abundant inflammatory cytokines.<sup>83-85</sup> The LCs of an LCH lesion express high levels of the programmed cell death protein.<sup>86</sup>

# Recurrent somatic activating MAPK pathway gene mutations in LCH

In a long-standing cancer vs inflammation debate, nonrandom X-inactivation of CD1a<sup>+</sup> cells supported clonality of LCH lesion LCs.<sup>87,88</sup> However, LCs are not hyperproliferative within the lesion,<sup>83,84,89</sup> and no gross genetic alterations were reported.<sup>90</sup> With improved sequencing technologies, Rollins and colleagues analyzed CD1a<sup>+</sup> cells isolated from LCH lesion biopsies and identified recurrent *BRAFV*600E mutations in more than 50% of the cases.<sup>91</sup> Several groups subsequently found that *BRAFV*600E or alternative activating MAPK pathway gene mutations are nearly universal in LCH, including other *BRAF* mutations and mutations in *MAP2K1* (encoding MEK1; Figure 5).<sup>92-96</sup>

#### **Origins of LCH lesion LCs**

The unique phenotype of CD1a<sup>+</sup>CD207<sup>+</sup> histiocytes made the epidermal LCs a likely culprit in LCH. However, gene expression comparing epidermal LCs with LCH lesion LCs identified a signature consistent with more immature myeloid precursors in the LCH cells.<sup>84</sup> Further, multiple dendritic cell lineages beyond epidermal LCs have the capacity to express Langerin with potential for wide tissue distribution.<sup>76</sup> Together, these observations supported potential for a hematopoietic origin and tissue distribution of precursor cells beyond the epidermis.

The discovery of somatic *BRAFV*600E mutation provided a molecular tag for lineage tracing to test the hypothesis that LCH could arise from hematopoietic precursors. Analysis of peripheral blood mononuclear cells (PBMCs) identified a very small (<1%) but consistent presence of *BRAFV*600E<sup>+</sup> in myeloid cells (CD11c<sup>+</sup> myeloid dendritic cell precursors and CD14<sup>+</sup> monocytes) in patients with high-risk LCH with *BRAFV*600E<sup>+</sup> lesions. Similarly, in bone marrow aspirates, *BRAFV*600E could be identified in CD34<sup>+</sup> hematopoietic stem cells of many patients with high-risk LCH, ~50% of which were reported as histologically normal. In contrast, *BRAFV*600E is typically absent

Figure 4. Models of LCH pathogenesis. (A) Ontogeny of physiologic epidermal LCs and microglia may inform mechanisms of LCH pathogenesis. Physiologic epidermal LCs arise from yolk sac and fetal liver; microglia arise from yolk sac (blue arrows). Subsequently, epidermal LC and microglia may be replace monocytes derived from bone marrow after activation or injury (orange arrows). (B) The variant allele frequency (BRAPV600E or other MAPK mutation) is found in very rare population in myeloid precursors in bone marrow and peripheral blood in patients with HR LCH and some with LR multifocal LCH. The Misguided Model of LCH Pathogenesis proposes that extent of disease is defined by the state of differentiation at which an activating somatic MAPK pathway gene mutation arises (red). (C) Lack of detectable PBMCs with BRAFV600E and self-resolving course support potential fetal liver origin, where identification of PBMCs with BRAFV600E and perivascular BRAF-V600E<sup>+</sup> cells at sites of neurodegeneration support potential for hematopoietic cell.



Figure 5. Activating MAPK pathway mutations in LCH. (A) Physiologic MAPK signaling transduces extracellular signal through receptor tyrosine kinase (RTK), which activates RAS, then RAF, then MEK, then ERK proteins, which in turn regulate cell-specific nuclear targets and gene transcription programs. (B) Activating mutations in LCH such as BRAFV600E drive constitutive ERK activation and downstream transcriptional targets including increased transcription of BCL2L1 and decreased CCR7.

from PBMCs of patients with active single lesion LCH and rarely identified in PBMCs of patients with multifocal low-risk LCH.<sup>97</sup> These observations support the "misguided myeloid differentiation model" of LCH ontogeny, in which extent of disease is defined by differentiation of the cell in which activating somatic *MAPK* gene mutation (eg, *BRAF*V600E) arises (Figure 4B). Beyon The MA

Pattems of skin LCH in infants and LCH-ND may inform pathogenic pathways in LCH. In an institutional series, *BRAFV*600E was frequently identified in PBMCs from children with multisystem LCH and skin lesions; however, *BRAFV*600E<sup>+</sup> PBMCs were not typically detected in infants with skin-limited disease.<sup>98</sup> This pattern could be consistent with skin-limited LCH in infants arising from a yolk sac or liver precursor during the wave of epidermal LC seeding that does not contribute to hematopoiesis. Multisystem LCH with *BRAFV*600E<sup>+</sup> would be more consistent with a hematopoietic precursor (Figure 4C).

As discussed earlier, a recent series identified *BRAFV600E*<sup>+</sup> cells in brain biopsies of patients with LCH-ND. Remarkably, a quantitative analysis of sections from whole-brain autopsy identified regions of brain stem and cerebellum with more than 10% *BRAFV600E*<sup>+</sup> cells. In addition, *BRAFV600E*<sup>+</sup> cells were identified in PBMCs in patients with LCH-ND who had no other systemic findings of active LCH, whereas *BRAFV600E* was not identified in patients cured from systemic LCH without LCH-ND. *BRAFV600E*<sup>+</sup> cells concentrated around blood vessels and lack the phenotype of P2RY12<sup>+</sup> microglia that arise during embryonic development.<sup>72</sup> These observations support a model in which a hematopoietic clone could contribute to microglia-like cells

that drive neurodegeneration (Figure 4C). The possibility of embryonic origin of LCH-ND is supported by a mouse model in which a yolk sac erythromyeloid precursor contributes to an isolated neurodegenerative process.<sup>99</sup>

#### **Beyond BRAF**

The MAPK pathway transduces extracellular signals that requlate transcriptional programs of cell growth, differentiation, and survival. The MAPK pathway is the most common dysregulated pathway in cancer, and BRAFV600E is identified in  $\sim$ 8% of all cancers. It is infrequently associated with hematologic malignancies with the notable exception of hairy cell leukemia, and ERK activation affects myeloid cell differentiation and maturation.<sup>100,101</sup> Cellular context likely plays a major role on the effect of MAPK pathway activation. In mice with enforced expression of BRAFV600E in CD11c cells, MAPK activation abrogates CCR7 expression in skin myeloid dendritic cells, which is required for their activation and migration to draining lymph nodes. Similarly, CCR7 expression is absent in CD207<sup>+</sup> LCH lesion LC, but is rescued with MAPK inhibition. In addition, the BCL21 (encoding BCLXL), which inhibits apoptosis, is upregulated in myeloid dendritic cells with increased MAPK activation. Therefore, MAPK activation contributes to LCH pathogenesis by trapping cells at sites of lesions, where they accumulate and resist cell death.89

Do specific mutations matter? In vitro studies demonstrate differential ERK activation from different MAPK gene mutations, which might explain some differences in clinical presentation and outcomes. Increased rates of resistance to front-line chemotherapy, relapse, and neurodegeneration have been described in patients with *BRAFV600E*.<sup>72,97,67,102</sup> Identifying mutation-specific pathogenic mechanisms may inform opportunities for clinical risk stratification and precision therapy.

## **Treatment of LCH**

The difficulties in developing effective therapies for LCH are linked to the deficiencies in the understanding of its pathogenesis. Patients are now stratified into different risk categories based on the disease extent and the degree of organ dysfunction; patients with single-system disease confined to a single site usually require only local therapy or observation, whereas patients with more extensive disease require systemic therapy.<sup>23,27</sup> These advances in risk-adapted treatment have resulted in better characterization of the natural history of the disease and an overall improvement in outcomes. Populationbased studies have documented a significant increase in survival for patients with disseminated LCH, although these improvements appear to have favored children over adults, with 5-year relative survival rates of 90% vs 70%, respectively.<sup>6,8</sup> Despite these improvements in survival, disease reactivations occur in approximately one-third of the patients, 24,65,103 and their prevention has become one of the major objectives of current clinical trials. The current standard of care for front-line therapy of patients with multifocal LCH or unifocal disease in CNS-risk sites is vinblastine/prednisone for 1 year, with the potential addition of mercaptopurine for high-risk LCH.<sup>66</sup> Robust data to guide treatment after first and subsequent treatment failures are lacking. LCH responds to increasing doses of nucleoside analogs with efficacy in other myeloid malignancies (cytarabine, cladribine, and clofarabine).<sup>104-106</sup> For patients with low-risk disease recurrence, including patients with reactivation of single-system or multifocal bone disease or multisystem disease without risk organ involvement, less toxic regimens have proven to be effective, including oral 6-mercaptopurine and methotrexate,<sup>107</sup> indomethacin,<sup>108</sup> bisphosphonates,<sup>109</sup> and hydroxyurea.<sup>110</sup> The international LCH-IV protocol (ClinicalTrials.gov identifier: NCT02205762) represents a comprehensive effort to address the most relevant clinical and therapeutic challenges, including the management of upfront and relapsed LCH and the treatment of CNS disease.

Early-phase trials in adults with LCH and Erdheim-Chester disease support near-universal responses in patients treated with MAPK pathway inhibitors.<sup>50,111-114</sup> Two recently reported retrospective pediatric series also report high response rates in cohorts including children with high-risk LCH, patients with multiple previous treatment failures, and patients with LCH-ND.<sup>73,115</sup> However, rapid reactivations occur in the majority of patients after discontinuation of therapy, and reintroduction of the BRAF inhibitor is usually effective.<sup>115</sup> Prospective trials will be required to determine optimal duration of therapy and potential for combination with other targeted or cytotoxic therapies.

A comprehensive review of historical and current clinical trials is detailed in the supplemental Materials, available on the *Blood* Web site.

#### Special treatment considerations

**Treatment of adult LCH** Treatment of LCH in adults follows similar guidelines to those recommended for children, with some modifications.<sup>51,116</sup> The more severe skin manifestations

have shown to respond well to phototherapy,<sup>117</sup> low-dose methotrexate,<sup>118</sup> and thalidomide or lenalidomide.<sup>38</sup> For patients requiring systemic therapy, vinblastine-based regimens remain quite effective in the adult population, with similar outcomes to children.<sup>119</sup> However, given the diminished tolerance of adults to corticosteroids and vinblastine, treatment with cytarabine or cladribine is generally preferred,<sup>51,116,120</sup> although BRAF inhibitors are being increasingly used in this population.<sup>50,121,122</sup>

For adults with pulmonary LCH, smoking cessation is critical for stabilization and improvement of symptoms, and a trial of observation after discontinuing smoking is recommended. For patients with severe or progressive disease, and for patients with multisystem disease, systemic therapy is recommended. Corticosteroids, either inhaled or systemic, have not shown to be of benefit. The most recommended treatment is cladribine, although the use of BRAF or MEK inhibitors in eligible patients should also be considered.<sup>56</sup>

**Treatment of LCH CNS disease** There are no standard guidelines for treatment of LCH CNS disease. For tumorous lesions and new-onset diabetes insipidus, treatment with a standard LCH regimen is indicated; vinblastine and prednisone or singleagent cladribine have been shown to be effective.<sup>123,124</sup> Treatment of LCH-ND is less defined. Improvement in the neurological condition has been reported with the use of cytarabine,<sup>125</sup> intravenous immunoglobulins,<sup>126</sup> rituximab,<sup>127</sup> infliximab,<sup>128</sup> and *cis*-retinoic acid.<sup>129</sup> More recently, the documentation of diffuse perivascular infiltration by *BRAFV*600E cells in biopsies of patients with LCH-ND has provided a strong rationale for the use of targeted therapies<sup>72</sup>; responses to BRAF inhibitors have been documented in 12 of 13 patients.<sup>73</sup>

## Late effects

Up to 50% of survivors have at least 1 permanent consequence.<sup>12,24,65</sup> Long-term effects have been reported to be more frequent among patients with multisystem disease and patients with multiple reactivations.<sup>12,65</sup> The most commonly reported late effects are diabetes insipidus and orthopedic abnormalities, which may occur in up to or slightly above 20% of patients, followed by growth retardation, hearing loss, and neurodegeneration in approximately 10% of the patients, and biliary cirrhosis, and respiratory insufficiency in less than 5% of patients.<sup>12,24,65</sup> Of particular relevance is the neurodegenerative syndrome that usually occurs years after the original diagnosis, and which has been discussed here and extensively reviewed elsewhere.<sup>60,61</sup>

### Future perspectives

Further research in the biology of LCH and its correlation with clinical presentation and outcomes will be required for better refinement of treatment of the disease and its complications, such as LCH-ND. Although the universal activation of the MAPK pathway provides a strong rationale for the use of pathway inhibitors, their role, including indications, optimal duration of treatment, and combination with standard chemotherapy, needs to be investigated.

## Acknowledgments

This work has been supported in part by a grant from the St. Baldrick's Foundation for the North American Consortium for Histiocytosis (CRG and CEA), a Translational Research Program grant from the

Leukemia and Lymphoma Society (CEA and CRG), and the American Lebanese Syrian Associated Charities (ALSAC).

## Authorship

Contribution: C.R.-G. and C.E.A. jointly wrote the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: C.R.-G., 0000-0002-2360-8946; C.E.A., 0000-0002-6625-739X.

#### REFERENCES

- Alston RD, Tatevossian RG, McNally RJ, Kelsey A, Birch JM, Eden TO. Incidence and survival of childhood Langerhans cell histiocytosis in Northwest England from 1954 to 1998. Pediatr Blood Cancer. 2007;48(5): 555-560.
- Guyot-Goubin A, Donadieu J, Barkaoui M, Bellec S, Thomas C, Clavel J. Descriptive epidemiology of childhood Langerhans cell histiocytosis in France, 2000-2004. *Pediatr Blood Cancer*. 2008;51(1):71-75.
- Salotti JA, Nanduri V, Pearce MS, Parker L, Lynn R, Windebank KP. Incidence and clinical features of Langerhans cell histiocytosis in the UK and Ireland. Arch Dis Child. 2009; 94(5):376-380.
- Stålemark H, Laurencikas E, Karis J, Gavhed D, Fadeel B, Henter JI. Incidence of Langerhans cell histiocytosis in children: a population-based study. *Pediatr Blood Cancer.* 2008;51(1):76-81.
- Baumgartner I, von Hochstetter A, Baumert B, Luetolf U, Follath F. Langerhans'-cell histiocytosis in adults. *Med Pediatr Oncol.* 1997;28(1):9-14.
- Goyal G, Shah MV, Hook CC, et al. Adult disseminated Langerhans cell histiocytosis: incidence, racial disparities and long-term outcomes. Br J Haematol. 2018;182(4): 579-581.
- Bhatia S, Nesbit ME Jr., Egeler RM, Buckley JD, Mertens A, Robison LL. Epidemiologic study of Langerhans cell histiocytosis in children. J Pediatr. 1997;130(5):774-784.
- Ribeiro KB, Degar B, Antoneli CBG, Rollins B, Rodriguez-Galindo C. Ethnicity, race, and socioeconomic status influence incidence of Langerhans cell histiocytosis. *Pediatr Blood Cancer.* 2015;62(6):982-987.
- Peckham-Gregory EC, McClain KL, Allen CE, Scheurer ME, Lupo PJ. The role of parental and perinatal characteristics on Langerhans cell histiocytosis: characterizing increased risk among Hispanics. Ann Epidemiol. 2018; 28(8):521-528.
- Peckham-Gregory EC, Chakraborty R, Scheurer ME, et al. A genome-wide association study of LCH identifies a variant in SMAD6 associated with susceptibility. Blood. 2017;130(20):2229-2232.
- Arico' M, Scappaticci S, Danesio C. The genetics of Langerhans cell histiocytosis. In: Weitzman S, Egeler RM, eds. Histiocytic disorders of children and adults, Cambridge: Cambridge University Press; 2005:83-94.

- Haupt R, Nanduri V, Calevo MG, et al. Permanent consequences in Langerhans cell histiocytosis patients: a pilot study from the Histiocyte Society-Late Effects Study Group. Pediatr Blood Cancer. 2004;42(5):438-444.
- Ma J, Laird JH, Chau KW, Chelius MR, Lok BH, Yahalom J. Langerhans cell histiocytosis in adults is associated with a high prevalence of hematologic and solid malignancies. *Cancer Med.* 2019;8(1):58-66.
- Egeler RM, Neglia JP, Puccetti DM, Brennan CA, Nesbit ME. Association of Langerhans cell histiocytosis with malignant neoplasms. *Cancer.* 1993;71(3):865-873.
- Wu X, Chen S, Zhang LY, Luo YP, Jiang Y, Feng RE. Langerhans cell histiocytosis of the thyroid complicated by papillary thyroid carcinoma: a case report and brief literature review. *Medicine (Baltimore)*. 2017;96(35):e7954.
- Moschovi M, Adamaki M, Vlahopoulos S, Rodriguez-Galindo C. Synchronous and metachronous thyroid cancer in relation to Langerhans cell histiocytosis; involvement of V600E BRAF-mutation? *Pediatr Blood Can*cer. 2015;62(1):173-174.
- Pina-Oviedo S, Medeiros LJ, Li S, et al. Langerhans cell histiocytosis associated with lymphoma: an incidental finding that is not associated with BRAF or MAP2K1 mutations. *Mod Pathol.* 2017;30:734-744.
- Egeler RM, Neglia JP, Aricò M, et al; The LCH-Malignancy Study Group of the Histiocyte Society. The relation of Langerhans cell histiocytosis to acute leukemia, lymphomas, and other solid tumors. *Hematol Oncol Clin North Am.* 1998;12(2):369-378.
- Yokokawa Y, Taki T, Chinen Y, et al. Unique clonal relationship between T-cell acute lymphoblastic leukemia and subsequent Langerhans cell histiocytosis with TCR rearrangement and NOTCH1 mutation. *Genes Chromosomes Cancer*. 2015;54(7):409-417.
- Kato M, Seki M, Yoshida K, et al. Genomic analysis of clonal origin of Langerhans cell histiocytosis following acute lymphoblastic leukaemia. Br J Haematol. 2016;175(1):169-172.
- Emile J-F, Abla O, Fraitag S, et al; Histiocyte Society. Revised classification of histiocytoses and neoplasms of the macrophagedendritic cell lineages. *Blood*. 2016;127(22): 2672-2681.
- Picarsic J, Jaffe R. Nosology and Pathology of Langerhans Cell Histiocytosis. *Hematol* Oncol Clin North Am. 2015;29(5):799-823.

Correspondence: Carlos Rodriguez-Galindo, Department of Global Pediatric Medicine, St. Jude Children's Research Hospital, 262 Danny Thomas PI MS-721, Memphis TN 38105; e-mail: carlos.rodriguezgalindo@stjude.org.

## Footnotes

Submitted 12 November 2019; accepted 7 January 2020; prepublished online on *Blood* First Edition 27 February 2020. DOI 10.1182/blood. 2019000934.

The online version of this article contains a data supplement.

- 23. Allen CE, Merad M, McClain KL. Langerhanscell histiocytosis. N Engl J Med. 2018;379(9): 856-868.
- Rigaud C, Barkaoui MA, Thomas C, et al. Langerhans cell histiocytosis: therapeutic strategy and outcome in a 30-year nationwide cohort of 1478 patients under 18 years of age. Br J Haematol. 2016;174(6):887-898.
- Donadieu J, Egeler RM, Pritchard J. Langerhans cell histiocytosis: a clinical update. In: Weitzman S, Egeler RM, eds. Histiocytic disorders of children and adults, Cambridge: Cambridge University Press; 2005:95-129.
- 26. Morimoto A, Shioda Y, Imamura T, et al; Japan LCH Study Group. Intensification of induction therapy and prolongation of maintenance therapy did not improve the outcome of pediatric Langerhans cell histiocytosis with single-system multifocal bone lesions: results of the Japan Langerhans Cell Histiocytosis Study Group-02 Protocol Study. Int J Hematol. 2018;108(2):192-198.
- Haupt R, Minkov M, Astigarraga I, et al; Euro Histio Network. Langerhans cell histiocytosis (LCH): guidelines for diagnosis, clinical workup, and treatment for patients till the age of 18 years. *Pediatr Blood Cancer*. 2013;60(2): 175-184.
- Kaste SC, Rodriguez-Galindo C, McCarville ME, Shulkin BL. PET-CT in pediatric Langerhans cell histiocytosis. *Pediatr Radiol.* 2007;37(7):615-622.
- Phillips M, Allen C, Gerson P, McClain K. Comparison of FDG-PET scans to conventional radiography and bone scans in management of Langerhans cell histiocytosis. *Pediatr Blood Cancer*. 2009;52(1):97-101.
- Dennin MH, Roman CJ, Stein SL. Congenital Langerhans cell histiocytosis presenting in a 27-week-gestation neonate. *Pediatr Dermatol.* 2018;35(2):e140-e141.
- Gothwal S, Gupta AK, Choudhary R. Congenital self healing Langerhans cell histiocytosis. *Indian J Pediatr.* 2018;85(4): 316-317.
- Yu J, Rubin AI, Castelo-Soccio L, Perman MJ. Congenital self-healing Langerhans cell histiocytosis. J Pediatr. 2017;184:232-232.e1.
- Kalpana S, Lakshmi V, Seth A, et al. Correspondence: systemic congenital Langerhans cell histiocytosis masquerading as diffuse neonatal hemangiomatosis. *Indian Pediatr.* 2018;55(7):613.
- 34. Schmitt AR, Wetter DA, Camilleri MJ, Khan SP, Tollefson MM. Langerhans cell

histiocytosis presenting as a blueberry muffin rash. *Lancet*. 2017;390(10090):155.

- Lau L, Krafchik B, Trebo MM, Weitzman S. Cutaneous Langerhans cell histiocytosis in children under one year. *Pediatr Blood Cancer*. 2006;46(1):66-71.
- Abla O, Egeler RM, Weitzman S. Langerhans cell histiocytosis: current concepts and treatments. *Cancer Treat Rev.* 2010;36(4): 354-359.
- Crickx E, Bouaziz JD, Lorillon G, et al. Clinical spectrum, quality of life, BRAF mutation status and treatment of skin involvement in adult Langerhans cell histiocytosis. Acta Derm Venereol. 2017;97(7):838-842.
- Ruiz Beguerie J, Fernández J, Stringa MF, Anaya J. Vulvar Langerhans cell histiocytosis and thalidomide: an effective treatment option. Int J Dermatol. 2017;56(3):324-326.
- Jiang W, Li L, He YM, Yang KX. Langerhans cell histiocytosis of the female genital tract: a literature review with additional three case studies in China. Arch Gynecol Obstet. 2012; 285(1):99-103.
- Gao Y-J, Su M, Tang J-Y, Pan C, Chen J. Treatment outcome of children with multisystem Langerhans cell histiocytosis: the experience of a single children's hospital in Shanghai, China. J Pediatr Hematol Oncol. 2018;40(1):e9-e12.
- Braier J, Latella A, Balancini B, et al. Outcome in children with pulmonary Langerhans cell Histiocytosis. *Pediatr Blood Cancer*. 2004;43(7):765-769.
- Ronceray L, Potschger U, Janka G, et al. Pulmonary involvement in pediatric-onset multisystem Langerhans cell histiocytosis: effect on course and outcome. J Pediatr. 2012;161(1):129-133.
- 43. Morimoto A, Shioda Y, Imamura T, et al. Intensified and prolonged therapy comprising cytarabine, vincristine and prednisolone improves outcome in patients with multisystem Langerhans cell histiocytosis: results of the Japan Langerhans Cell Histiocytosis Study Group-02 Protocol Study. Int J Hematol. 2016;104(1):99-109.
- Tazi A. Adult pulmonary Langerhans' cell histiocytosis. Eur Respir J. 2006;27(6): 1272-1285.
- Galluzzo ML, Braier J, Rosenzweig SD, Garcia de Dávila MT, Rosso D. Bone marrow findings at diagnosis in patients with multisystem langerhans cell histiocytosis. *Pediatr Dev Pathol.* 2010;13(2):101-106.
- Hatemi I, Baysal B, Senturk H, Behzatoglu K, Bozkurt ER, Ozbay G. Adult Langerhans cell histiocytosis and sclerosing cholangitis: a case report and review of the literature. *Hepatol Int.* 2010;4(3):653-658.
- Braier J, Ciocca M, Latella A, de Davila MG, Drajer M, Imventarza O. Cholestasis, sclerosing cholangitis, and liver transplantation in Langerhans cell Histiocytosis. *Med Pediatr Oncol.* 2002;38(3):178-182.
- Gadner H, Grois N, Pötschger U, et al; Histiocyte Society. Improved outcome in multisystem Langerhans cell histiocytosis is associated with therapy intensification. *Blood*. 2008;111(5):2556-2562.

- Aricò M, Girschikofsky M, Généreau T, et al. Langerhans cell histiocytosis in adults. Report from the International Registry of the Histiocyte Society. Eur J Cancer. 2003; 39(16):2341-2348.
- Haroche J, Cohen-Aubart F, Emile JF, et al. Dramatic efficacy of vemurafenib in both multisystemic and refractory Erdheim-Chester disease and Langerhans cell histiocytosis harboring the BRAF V600E mutation. *Blood.* 2013;121(9):1495-1500.
- Girschikofsky M, Arico M, Castillo D, et al. Management of adult patients with Langerhans cell histiocytosis: recommendations from an expert panel on behalf of Euro-Histio-Net. Orphanet J Rare Dis. 2013;8(1): 72.
- Pierro J, Vaiselbuh SR. Adult Langerhans cell histiocytosis as a diagnostic pitfall. J Clin Oncol. 2016;34(6):e41-e45.
- Tazi A, de Margerie C, Naccache JM, et al. The natural history of adult pulmonary Langerhans cell histiocytosis: a prospective multicentre study. Orphanet J Rare Dis. 2015;10(1):30.
- Tazi A, de Margerie-Mellon C, Vercellino L, et al. Extrathoracic investigation in adult patients with isolated pulmonary langerhans cell histiocytosis. Orphanet J Rare Dis. 2016; 11(1):11.
- DeMartino E, Go RS, Vassallo R. Langerhans Cell Histiocytosis and Other Histiocytic Diseases of the Lung. *Clin Chest Med.* 2016; 37(3):421-430.
- Lorillon G, Tazi A. How I manage pulmonary Langerhans cell histiocytosis. *Eur Respir Rev.* 2017;26(145):170070.
- Roden AC, Yi ES. Pulmonary Langerhans cell histiocytosis: an update from the pathologists' perspective. Arch Pathol Lab Med. 2016;140(3):230-240.
- Mourah S, How-Kit A, Meignin V, et al. Recurrent NRAS mutations in pulmonary Langerhans cell histiocytosis. Eur Respir J. 2016;47(6):1785-1796.
- Kamionek M, Ahmadi Moghaddam P, Sakhdari A, et al. Mutually exclusive extracellular signal-regulated kinase pathway mutations are present in different stages of multi-focal pulmonary Langerhans cell histiocytosis supporting clonal nature of the disease. *Histopathology*. 2016;69(3): 499-509.
- 60. Yeh EA, Greenberg J, Abla O, et al; North American Consortium for Histiocytosis. Evaluation and treatment of Langerhans cell histiocytosis patients with central nervous system abnormalities: Current views and new vistas. Pediatr Blood Cancer. 2018;65(1):65.
- Grois N, Fahrner B, Arceci RJ, et al. Central nervous system disease in Langerhans cell histiocytosis. J Pediatr. 2010;156(6):873-881.
- Sagna Y, Courtillot C, Drabo JY, et al. Endocrine manifestations in a cohort of 63 adulthood and childhood onset patients with Langerhans cell histiocytosis. Eur J Endocrinol. 2019;181(3):275-285.
- 63. Donadieu J, Rolon M-A, Pion I, et al; French LCH Study Group. Incidence of growth hormone deficiency in pediatric-onset

Langerhans cell histiocytosis: efficacy and safety of growth hormone treatment. *J Clin Endocrinol Metab.* 2004;89(2):604-609.

- Donadieu J, Rolon M-A, Thomas C, et al; French LCH Study Group. Endocrine involvement in pediatric-onset Langerhans' cell histiocytosis: a population-based study. *J Pediatr.* 2004;144(3):344-350.
- Chow TW, Leung WK, Cheng FWT, et al. Late outcomes in children with Langerhans cell histiocytosis. Arch Dis Child. 2017; 102(9):830-835.
- Gadner H, Minkov M, Grois N, et al; Histiocyte Society. Therapy prolongation improves outcome in multisystem Langerhans cell histiocytosis. *Blood.* 2013;121(25): 5006-5014.
- Héritier S, Barkaoui MA, Miron J, et al. Incidence and risk factors for clinical neurodegenerative Langerhans cell histiocytosis: a longitudinal cohort study. Br J Haematol. 2018;183(4):608-617.
- Grois N, Pötschger U, Prosch H, et al; DALHX- and LCH I and II Study Committee. Risk factors for diabetes insipidus in langerhans cell histiocytosis. *Pediatr Blood Cancer*. 2006;46(2):228-233.
- Garg D, Pedapati R, Nakra T, et al. Langerhans cell histiocytosis presenting as a rapidly evolving frontotemporal syndrome. *Neurol Sci.* 2019;40(5):1055-1058.
- Sieni E, Barba C, Mortilla M, et al. Early diagnosis and monitoring of neurodegenerative Langerhans cell histiocytosis. *PLoS One*. 2015;10(7):e0131635.
- Grois N, Prayer D, Prosch H, Lassmann H; CNS LCH Co-operative Group. Neuropathology of CNS disease in Langerhans cell histiocytosis. *Brain*. 2005;128(Pt 4): 829-838.
- McClain KL, Picarsic J, Chakraborty R, et al. CNS Langerhans cell histiocytosis: Common hematopoietic origin for LCH-associated neurodegeneration and mass lesions. *Cancer.* 2018;124(12):2607-2620.
- Eckstein OS, Visser J, Rodriguez-Galindo C, Allen CE; NACHO-LIBRE Study Group. Clinical responses and persistent *BRAF* V600E<sup>+</sup> blood cells in children with LCH treated with MAPK pathway inhibition. *Blood*. 2019;133(15):1691-1694.
- Merad M, Manz MG, Karsunky H, et al. Langerhans cells renew in the skin throughout life under steady-state conditions [published correction appears in Nat Immunol. 2002;3(12):1125-1126]. Nat Immunol. 2002; 3(12):1135-1141.
- Hoeffel G, Wang Y, Greter M, et al. Adult Langerhans cells derive predominantly from embryonic fetal liver monocytes with a minor contribution of yolk sac-derived macrophages. J Exp Med. 2012;209(6):1167-1181.
- Merad M, Ginhoux F, Collin M. Origin, homeostasis and function of Langerhans cells and other langerin-expressing dendritic cells. Nat Rev Immunol. 2008;8(12):935-947.
- Ohl L, Mohaupt M, Czeloth N, et al. CCR7 governs skin dendritic cell migration under inflammatory and steady-state conditions. *Immunity*. 2004;21(2):279-288.

- Farber S. The nature of "solitary or eosinophilic granuloma" of bone. Am J Pathol. 1941;17:625-626.
- Lichtenstein L. Histiocytosis X; integration of eosinophilic granuloma of bone, Letterer-Siwe disease, and Schüller-Christian disease as related manifestations of a single nosologic entity. AMA Arch Pathol. 1953;56(1): 84-102.
- Nezelof C, Basset F, Rousseau MF. Histiocytosis X histogenetic arguments for a Langerhans cell origin. *Biomedicine (Paris)*. 1973;18(5):365-371.
- Arceci RJ, Brenner MK, Pritchard J. Controversies and new approaches to treatment of Langerhans cell histiocytosis. *Hematol Oncol Clin North Am.* 1998;12(2): 339-357.
- Berres ML, Merad M, Allen CE. Progress in understanding the pathogenesis of Langerhans cell histiocytosis: back to Histiocytosis X? Br J Haematol. 2015;169(1):3-13.
- Senechal B, Elain G, Jeziorski E, et al. Expansion of regulatory T cells in patients with Langerhans cell histiocytosis. *PLoS Med.* 2007;4(8):e253.
- Allen CE, Li L, Peters TL, et al. Cell-specific gene expression in Langerhans cell histiocytosis lesions reveals a distinct profile compared with epidermal Langerhans cells. *J Immunol.* 2010;184(8):4557-4567.
- Laman JD, Leenen PJM, Annels NE, Hogendoorn PC, Egeler RM. Langerhans-cell histiocytosis "insight into DC biology". *Trends Immunol.* 2003;24(4):190-196.
- Gatalica Z, Bilalovic N, Palazzo JP, et al. Disseminated histiocytoses biomarkers beyond BRAFV600E: frequent expression of PD-L1. Oncotarget. 2015;6(23): 19819-19825.
- Willman CL. Detaction of clonal histiocytes in Langerhans cell histiocytosis: biology and clinical significance. *Br J Cancer*. 1994;(23): S29-S33.
- Yu RC, Chu C, Buluwela L, Chu AC. Clonal proliferation of Langerhans cells in Langerhans cell histiocytosis. *Lancet*. 1994; 343(8900):767-768.
- Hogstad B, Berres ML, Chakraborty R, et al. RAF/MEK/extracellular signal-related kinase pathway suppresses dendritic cell migration and traps dendritic cells in Langerhans cell histiocytosis lesions. J Exp Med. 2018;215(1): 319-336.
- da Costa CE, Szuhai K, van Eijk R, et al. No genomic aberrations in Langerhans cell histiocytosis as assessed by diverse molecular technologies. *Genes Chromosomes Cancer*. 2009;48(3):239-249.
- Badalian-Very G, Vergilio J-A, Degar BA, et al. Recurrent BRAF mutations in Langerhans cell histiocytosis. *Blood*. 2010;116(11): 1919-1923.
- Nelson DS, van Halteren A, Quispel WT, et al. MAP2K1 and MAP3K1 mutations in Langerhans cell histiocytosis. Genes Chromosomes Cancer. 2015;54(6):361-368.
- 93. Chakraborty R, Burke TM, Hampton OA, et al. Alternative genetic mechanisms of

BRAF activation in Langerhans cell histiocytosis. *Blood.* 2016;128(21):2533-2537.

- Chakraborty R, Hampton OA, Shen X, et al. Mutually exclusive recurrent somatic mutations in MAP2K1 and BRAF support a central role for ERK activation in LCH pathogenesis. *Blood.* 2014;124(19):3007-3015.
- Brown NA, Furtado LV, Betz BL, et al. High prevalence of somatic MAP2K1 mutations in BRAF V600E-negative Langerhans cell histiocytosis. *Blood.* 2014;124(10):1655-1658.
- Héritier S, Hélias-Rodzewicz Z, Chakraborty R, et al. New somatic BRAF splicing mutation in Langerhans cell histiocytosis. *Mol Cancer*. 2017;16(1):115.
- Berres ML, Lim KP, Peters T, et al. BRAF-V600E expression in precursor versus differentiated dendritic cells defines clinically distinct LCH risk groups. J Exp Med. 2014; 211(4):669-683.
- Simko SJ, Garmezy B, Abhyankar H, et al. Differentiating skin-limited and multisystem Langerhans cell histiocytosis. *J Pediatr.* 2014; 165(5):990-996.
- Mass E, Jacome-Galarza CE, Blank T, et al. A somatic mutation in erythro-myeloid progenitors causes neurodegenerative disease. *Nature*. 2017;549(7672):389-393.
- Pratilas CA, Xing F, Solit DB. Targeting oncogenic BRAF in human cancer. Curr Top Microbiol Immunol. 2012;355:83-98.
- Michaloglou C, Vredeveld LC, Mooi WJ, Peeper DS. BRAF(E600) in benign and malignant human tumours. *Oncogene*. 2008; 27(7):877-895.
- 102. Héritier S, Emile JF, Barkaoui MA, et al. BRAF mutation correlates with high-risk Langerhans cell histiocytosis and increased resistance to first-line therapy. J Clin Oncol. 2016;34(25):3023-3030.
- 103. Minkov M, Steiner M, Pötschger U, et al; International LCH Study Group. Reactivations in multisystem Langerhans cell histiocytosis: data of the international LCH registry. J Pediatr. 2008;153(5):700-705.
- 104. Rodriguez-Galindo C, Kelly P, Jeng M, Presbury GG, Rieman M, Wang W. Treatment of children with Langerhans cell histiocytosis with 2-chlorodeoxyadenosine. Am J Hematol. 2002;69(3):179-184.
- 105. Abraham A, Alsultan A, Jeng M, Rodriguez-Galindo C, Campbell PK. Clofarabine salvage therapy for refractory high-risk langerhans cell histiocytosis. *Pediatr Blood Cancer*. 2013;60(6):E19-E22.
- 106. Simko SJ, Tran HD, Jones J, et al. Clofarabine salvage therapy in refractory multifocal histiocytic disorders, including Langerhans cell histiocytosis, juvenile xanthogranuloma and Rosai-Dorfman disease. Pediatr Blood Cancer. 2014;61(3):479-487.
- 107. Womer RB, Anunciato KR, Chehrenama M. Oral methotrexate and alternate-day prednisone for low-risk Langerhans cell histiocytosis. Med Pediatr Oncol. 1995;25(2):70-73.
- 108. Braier J, Rosso D, Pollono D, et al. Symptomatic bone Langerhans cell histiocytosis treated at diagnosis or after

reactivation with indomethacin alone. *J Pediatr Hematol Oncol.* 2014;36(5): e280-e284.

- 109. Morimoto A, Ikushima S, Kinugawa N, et al; Japan Langerhans Cell Histiocytosis Study Group. Improved outcome in the treatment of pediatric multifocal Langerhans cell histiocytosis: Results from the Japan Langerhans Cell Histiocytosis Study Group-96 protocol study. *Cancer.* 2006;107(3): 613-619.
- 110. Zinn DJ, Grimes AB, Lin H, Eckstein O, Allen CE, McClain KL. Hydroxyurea: a new old therapy for Langerhans cell histiocytosis. *Blood*. 2016;128(20):2462-2465.
- Hyman DM, Puzanov I, Subbiah V, et al. Vemurafenib in multiple nonmelanoma cancers with BRAF V600 mutations. N Engl J Med. 2015;373(8):726-736.
- 112. Diamond EL, Subbiah V, Lockhart AC, et al. Vemurafenib for BRAF V600-mutant Erdheim-Chester disease and Langerhans cell histiocytosis: analysis of data from the histology-independent, phase 2, open-label VE-BASKET study. JAMA Oncol. 2018;4(3): 384-388.
- 113. Haroche J, Cohen-Aubart F, Emile JF, et al. Reproducible and sustained efficacy of targeted therapy with vemurafenib in patients with BRAF(V600E)-mutated Erdheim-Chester disease. J Clin Oncol. 2015;33(5): 411-418.
- 114. Diamond EL, Durham BH, Ulaner GA, et al. Efficacy of MEK inhibition in patients with histiocytic neoplasms. *Nature*. 2019; 567(7749):521-524.
- 115. Donadieu J, Larabi IA, Tardieu M, et al. Vemurafenib for refractory multisystem Langerhans Cell Histiocytosis in children: An international observational study. J Clin Oncol. 2019;37(31):2857-2865.
- 116. Cantu MA, Lupo PJ, Bilgi M, Hicks MJ, Allen CE, McClain KL. Optimal therapy for adults with Langerhans cell histiocytosis bone lesions. *PLoS One*. 2012;7(8):e43257.
- 117. Imafuku S, Shibata S, Tashiro A, Furue M. Cutaneous Langerhans cell histiocytosis in an elderly man successfully treated with narrowband ultraviolet B. Br J Dermatol. 2007; 157(6):1277-1279.
- 118. Steen AE, Steen KH, Bauer R, Bieber T. Successful treatment of cutaneous Langerhans cell histiccytosis with low-dose methotrexate. Br J Dermatol. 2001;145(1): 137-140.
- 119. Tazi A, Lorillon G, Haroche J, et al. Vinblastine chemotherapy in adult patients with langerhans cell histiocytosis: a multicenter retrospective study. *Orphanet J Rare Dis.* 2017;12(1):95.
- 120. Minami M, Shima T, Kato K, et al. Successful treatment of adult Langerhans cell histiocytosis with intensified chemotherapy. *Int J Hematol.* 2015;102(2):244-248.
- 121. Gandolfi L, Adamo S, Pileri A, Broccoli A, Argnani L, Zinzani PL. Multisystemic and multiresistant Langerhans cell histiocytosis: a case treated with BRAF inhibitor. J Natl Compr Canc Netw. 2015;13(6):715-718.

- 122. Charles J, Beani J-C, Fiandrino G, Busser B. Major response to vemurafenib in patient with severe cutaneous Langerhans cell histiocytosis harboring BRAF V600E mutation. J Am Acad Dermatol. 2014;71(3):e97-e99.
- 123. Dhall G, Finlay JL, Dunkel IJ, et al. Analysis of outcome for patients with mass lesions of the central nervous system due to Langerhans cell histiocytosis treated with 2-chlorodeoxyadenosine. *Pediatr Blood Cancer*. 2008;50(1):72-79.
- 124. Ng Wing Tin S, Martin-Duverneuil N, Idbaih A, et al; French LCH study group. Efficacy of vinblastine in central nervous system Langerhans cell histiocytosis: a nationwide

retrospective study. *Orphanet J Rare Dis.* 2011;6(1):83.

- 125. Allen CE, Flores R, Rauch R, et al. Neurodegenerative central nervous system Langerhans cell histiocytosis and coincident hydrocephalus treated with vincristine/ cytosine arabinoside. *Pediatr Blood Cancer*. 2010;54(3):416-423.
- 126. Imashuku S, Arceci RJ. Strategies for the prevention of central nervous system complications in patients with Langerhans cell histiocytosis: the problem of neurodegenerative syndrome. *Hematol Oncol Clin North Am.* 2015;29(5): 875-893.
- 127. Eckstein O, McAtee CL, Greenberg J, et al. Rituximab therapy for patients with Langerhans cell histiocytosis-associated neurologic dysfunction. *Pediatr Hematol Oncol.* 2018; 35(7-8):427-433.
- 128. Chohan G, Barnett Y, Gibson J, Reddel SWR, Barnett MH. Langerhans cell histiocytosis with refractory central nervous system involvement responsive to infliximab. J Neurol Neurosurg Psychiatry. 2012;83(5):573-575.
- 129. Idbaih A, Donadieu J, Barthez MA, et al. Retinoic acid therapy in "degenerative-like" neuro-langerhans cell histiocytosis: a prospective pilot study. *Pediatr Blood Cancer*. 2004;43(1):55-58.