



### IRON METABOLISM AND ITS DISORDERS

## Liver iron sensing and body iron homeostasis

Chia-Yu Wang and Jodie L. Babitt

Division of Nephrology, Program in Membrane Biology, Center for Systems Biology, Massachusetts General Hospital, Harvard Medical School, Boston, MA

**The liver orchestrates systemic iron balance by producing and secreting hepcidin. Known as the iron hormone, hepcidin induces degradation of the iron exporter ferroportin to control iron entry into the bloodstream from dietary sources, iron recycling macrophages, and body stores. Under physiologic conditions, hepcidin production is reduced by iron deficiency and erythropoietic drive to increase the iron supply when needed to support red blood cell production and other essential functions. Conversely, hepcidin production is induced by iron loading and inflammation to prevent the toxicity of iron excess and limit its availability to pathogens. The inability to appropriately regulate hepcidin production in response to these physiologic cues underlies genetic disorders of iron overload and deficiency, including hereditary hemochromatosis and iron-refractory iron deficiency anemia.**

**Moreover, excess hepcidin suppression in the setting of ineffective erythropoiesis contributes to iron-loading anemias such as  $\beta$ -thalassemia, whereas excess hepcidin induction contributes to iron-restricted erythropoiesis and anemia in chronic inflammatory diseases. These diseases have provided key insights into understanding the mechanisms by which the liver senses plasma and tissue iron levels, the iron demand of erythrocyte precursors, and the presence of potential pathogens and, importantly, how these various signals are integrated to appropriately regulate hepcidin production. This review will focus on recent insights into how the liver senses body iron levels and coordinates this with other signals to regulate hepcidin production and systemic iron homeostasis. (*Blood*. 2019;133(1):18-29)**

### Roles of iron in biology

Iron is an essential element required for numerous fundamental cellular and organismal processes, including oxygen transport, mitochondrial respiration, nucleic acid replication, intermediary and xenobiotic metabolism, host defense, and cell signaling. These biological functions depend on iron's property as a transition metal to readily donate or accept electrons to participate in oxidation-reduction reactions. Iron performs these essential functions as a constituent of proteins in the form of heme, iron sulfur clusters, or other functional groups, thereby targeting and limiting its reactivity. However, when excess "free" iron is present, this redox cycling can also catalyze the production of free radicals, which can damage DNA, proteins, and lipids. To meet the dual challenge of providing adequate iron to perform essential functions while limiting the toxicity of excess iron, organisms have evolved specialized proteins and homeostatic mechanisms to tightly control iron availability.

### Systemic iron homeostasis

At a systemic level, iron homeostasis is maintained by balancing iron supply with iron utilization and losses (reviewed in Dev and Babitt<sup>1</sup> and Muckenthaler et al<sup>2</sup>). In humans, ~20 mg iron is used per day to generate 200 billion erythrocytes, accounting for 80% of daily iron needs. Erythrocytes and other cells obtain iron from the plasma, which contains only 2 to 4 mg iron and

therefore must be turned over every few hours to meet the daily requirement. Iron is supplied to the plasma by reticuloendothelial macrophages that salvage iron from aged erythrocytes (20-25 mg daily) and duodenal enterocytes that absorb dietary iron (1-2 mg daily), with the remainder provided by body stores. Iron losses (~1-2 mg/day) occur mainly through desquamation and blood loss in an unregulated process. Since iron losses are unregulated, iron homeostasis is mainly controlled by regulating iron supply.

The major suppliers of iron to the plasma are duodenal enterocytes and iron-recycling macrophages (reviewed in Muckenthaler et al<sup>2</sup>). Dietary elemental iron is transported across the enterocyte apical membrane by divalent metal transporter 1 (DMT1) after reduction by ferriductases such as duodenal cytochrome B.<sup>3-6</sup> Alternative mechanisms exist for heme iron uptake, but the specific transport mechanisms are unknown. Reticuloendothelial macrophages phagocytose aged erythrocytes for degradation in phagolysosomes, followed by iron release by heme oxygenase 1. In both enterocytes and macrophages, imported iron can be either used for the cell's own metabolic processes, stored in an inert form in ferritin, or exported into the plasma for use by other tissues. Elemental iron export is coupled to oxidation by the ferroxidases hephaestin and ceruloplasmin for loading onto the plasma iron carrier transferrin for delivery to erythrocyte precursors and other tissues.<sup>7-9</sup>

## Regulation of systemic iron homeostasis by the hepcidin-ferroportin axis

Physiologic studies dating back to the mid-1900s supported the existence of several regulators that modulate the iron supply to maintain stable iron stores (stores regulator), provide adequate iron for erythropoiesis (erythroid regulator), and limit iron availability to pathogenic microorganisms (inflammatory regulator) (reviewed in Finch<sup>10</sup>). In 2000-2001, 2 key proteins were identified, hepcidin<sup>11-13</sup> and ferroportin,<sup>14-16</sup> which began to unravel the puzzle of how communication was coordinated among organs involved in iron utilization, recycling, storage, and acquisition to maintain systemic iron homeostasis. Hepcidin is a 25-amino-acid hormone that is synthesized and secreted predominantly by the liver, circulates in the bloodstream, and is excreted by the kidneys.<sup>11-13</sup> A key role for hepcidin in iron homeostasis regulation was recognized when mutations or knockout of this gene were associated with the iron overload disorder hereditary hemochromatosis (HH), characterized by excessive dietary iron absorption and iron release from macrophage stores leading to serum and tissue iron overload with consequent organ dysfunction.<sup>17,18</sup> Conversely, hepcidin excess in transgenic mice or humans with hepcidin-expressing adenomas resulted in iron deficiency anemia.<sup>19,20</sup> The role of hepcidin as an effector for the stores, erythroid, and inflammatory regulators was suggested by the upregulation of hepcidin expression by iron loading and inflammation<sup>12</sup> and its suppression by erythropoietic drive.<sup>21</sup>

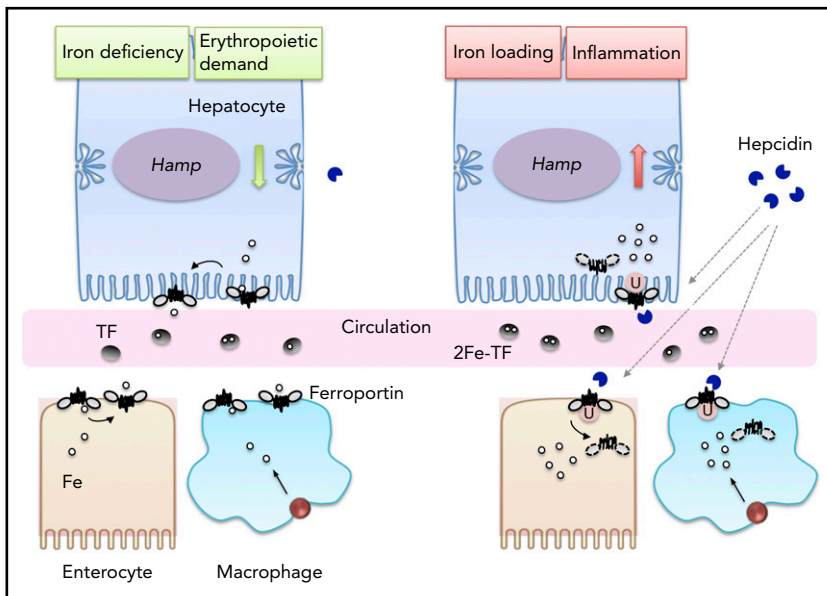
Coincident with the discovery of hepcidin, ferroportin was identified as an iron transport protein expressed on the basolateral surface of duodenal enterocytes, reticuloendothelial macrophages, hepatocytes, and placental syncytiotrophoblasts, where it plays a key role in iron export into the plasma.<sup>14-16</sup> Inactivating ferroportin mutations in zebrafish resulted in iron deficiency anemia due to inadequate maternal iron transport to the fetus.<sup>15</sup> Conditional ferroportin deletion from intestinal enterocytes, macrophages, and hepatocytes in mice cemented the important functional role of ferroportin in dietary iron absorption and iron mobilization to maintain systemic iron homeostasis.<sup>22-24</sup> Recent work has also discovered a local role for ferroportin in other cell types, including cardiac myocytes<sup>25</sup> and erythroid cells,<sup>26</sup> to prevent toxicity from iron excess. Ferroportin mutations in humans lead to 2 distinct phenotypes, one resembling classical HH and the other (so-called ferroportin disease) manifesting in an autosomal-dominant fashion by iron loading predominantly in reticuloendothelial macrophages with a tendency toward reduced plasma iron and anemia.<sup>27-29</sup>

Shortly after their discovery, hepcidin and ferroportin were functionally linked when it was recognized that hepcidin binds ferroportin and induces its internalization and degradation.<sup>30</sup> Data from crystal structure of a putative bacterial ferroportin homolog, computational modeling, patient mutations, and *in vitro* studies have refined our understanding of how ferroportin transports iron and how its activity is regulated by hepcidin.<sup>31-37</sup> A member of the major facilitator superfamily of transporters, ferroportin is a 12-transmembrane domain protein organized into 2 lobes connected by a large cytoplasmic loop.<sup>31,32</sup> In the current model, the 2 lobes form a central cavity containing a metal-binding site, and transport activity occurs by

a conformational change from an inward-facing state open intracellularly to an outward-facing state open extracellularly.<sup>33</sup> The N terminus of hepcidin<sup>34,35</sup> binds to the central cavity of ferroportin, where it interacts with up to 4 helices.<sup>36</sup> Hepcidin binding triggers ferroportin to undergo a conformational change, ubiquitination of multiple lysine residues on the cytoplasmic loop, endocytosis, and degradation.<sup>37,38</sup> Hepcidin binding also occludes ferroportin and interferes with iron export independent of endocytosis activity.<sup>36</sup> These studies provide a molecular explanation for the pathophysiology of HH due to hepcidin mutations and the differing patient phenotypes resulting from ferroportin mutations. Loss of hepcidin function leads to iron overload as a consequence of unregulated ferroportin activity, hyperabsorption of dietary iron, and excess iron release from body stores. Ferroportin mutations that interfere with hepcidin binding or the conformational change required for endocytosis and degradation result in a gain-of-function phenotype resembling classical HH. Ferroportin mutations that interfere with its localization or iron export function result in a loss-of-function phenotype and ferroportin disease. An unresolved question is why loss-of-function mutations act in a dominant-negative fashion to cause autosomal-dominant disease.

The ability of hepcidin to control ferroportin expression and activity also provides a molecular explanation for how hepcidin functions as the key hormone to coordinate signals from iron, erythropoietic drive, and inflammation to control systemic iron homeostasis (Figure 1). Iron loading increases hepcidin expression, thereby inhibiting ferroportin and limiting iron entry into the bloodstream to maintain steady-state iron levels. Hepcidin is regulated not only by the classically described iron "stores regulator" but also by acute increases in plasma iron, even when stores are unchanged.<sup>39,40</sup> This explains why the plasma iron concentration remains relatively stable despite its rapid turnover. Inflammation also induces hepcidin, thereby inhibiting ferroportin to limit iron availability to pathogens. Conversely, iron deficiency and erythropoietic drive suppress hepcidin, thereby increasing ferroportin to increase iron availability for erythropoiesis and other essential functions.

After discovery of the hepcidin-ferroportin axis, it quickly became clear that excessive or insufficient hepcidin production plays a central role in most major disorders of systemic iron homeostasis. Whereas hepcidin suppression by anemia or hypoxia has a physiologic role to increase iron availability for erythropoiesis, excessive hepcidin suppression by ineffective erythropoiesis contributes to pathologic iron overload in thalassemias and other iron-loading anemias. Whereas hepcidin induction may be protective to sequester iron during infection, excessive hepcidin induction contributes to iron-restricted erythropoiesis and anemia in chronic inflammatory diseases. The inability of iron loading to appropriately upregulate hepcidin leads to HH due not only to mutations in hepcidin or ferroportin themselves but also to other genetic causes. The inability of iron deficiency to appropriately suppress hepcidin leads to iron-refractory iron deficiency anemia (IRIDA). Targeting hepcidin and/or ferroportin function has therefore become an attractive new therapeutic strategy for these iron disorders, with efficacy demonstrated in many animal models and numerous candidates now being translated to clinical studies.<sup>41,42</sup> Understanding the genetic and molecular basis of these iron disorders has also yielded important insights into



**Figure 1. Systemic iron homeostasis regulation by hepcidin and ferroportin.** Hepcidin and ferroportin control iron entry into the circulation from dietary sources and body stores to maintain systemic iron homeostasis. Iron deficiency and erythropoietic demand suppress hepcidin production by hepatocytes (left). In the absence of hepcidin, ferroportin is stabilized on the basolateral surface of duodenal enterocytes, iron-recycling macrophages, and hepatocytes, where it transports iron (Fe) from the intracellular space to the plasma for loading onto transferrin (TF) and delivery to red blood cells and other tissues. Iron loading and inflammation stimulate hepatocyte hepcidin production and secretion into the circulation, where it binds to ferroportin to cause its ubiquitination (U), endocytosis, and degradation (right). Hepcidin binding also interferes with ferroportin export activity independent of endocytosis. Hepcidin thereby limits iron overload and sequesters iron from invading pathogens.

how hepcidin is regulated by its various stimuli and has opened up additional therapeutic opportunities that target hepcidin production.

## Hepcidin regulation by iron: lessons from HH and IRIDA

In addition to hepcidin and gain-of-function ferroportin mutations, mutations in several other genes, including *HFE*, transferrin receptor 2 (*TFR2*), and hemojuvelin (*HJV*), lead to HH.<sup>43</sup> All patients and animal models with a global or hepatocyte-specific knockout or mutation in *HFE*, *TFR2*, or *HJV* exhibit hepcidin deficiency and iron overload, suggesting that the key function of all of these proteins is in regulating hepatic hepcidin production in response to iron.<sup>44-52</sup> Mutations in transmembrane serine protease 6 (*TMPRSS6*) lead to an opposite phenotype of IRIDA as a consequence of hepcidin excess, suggesting that *TMPRSS6* plays in an important functional role to suppress hepcidin production in response to iron deficiency.<sup>53-55</sup>

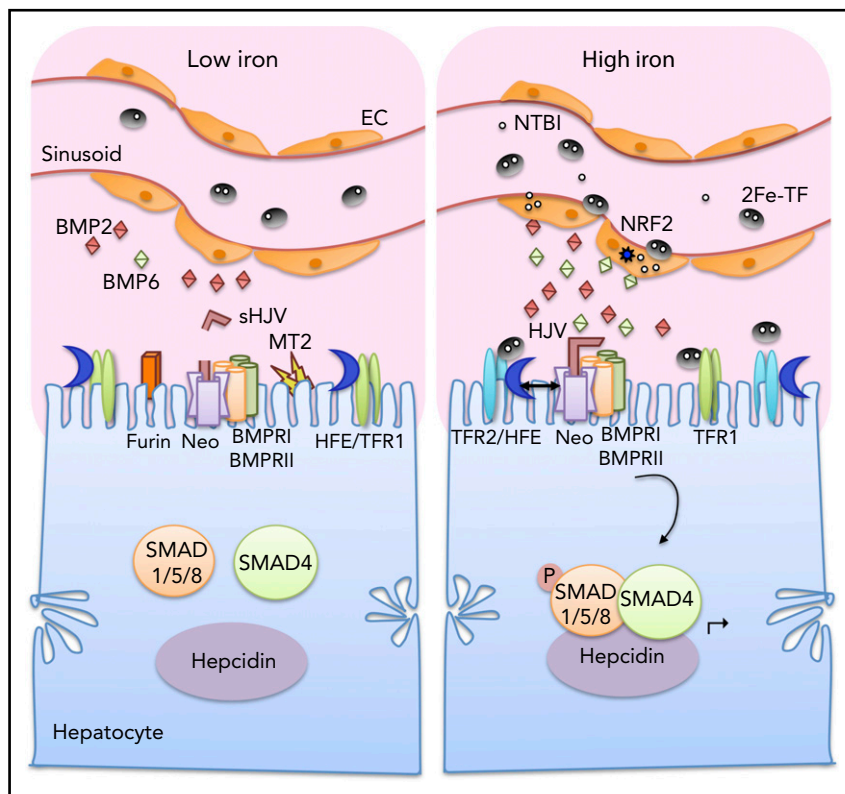
## Hepcidin regulation by HJV and the bone morphogenetic protein (BMP)-SMAD pathway

In 2006, HJV, like its repulsive guidance molecule family homologs RGMA and RGMB,<sup>56,57</sup> was recognized to function as a coreceptor for the BMP-SMAD pathway.<sup>58</sup> This led to the discovery of the BMP-SMAD signaling pathway as a major transcriptional regulator of hepcidin expression in hepatocytes<sup>58</sup> (Figure 2). HJV binds directly to BMP ligands and facilitates the formation of an active signaling complex containing BMP type I and type II receptors.<sup>58-60</sup> This signaling complex phosphorylates receptor-activated SMADs (R-SMADs), which bind to the common mediator SMAD4 and translocate to the nucleus, where they bind 2 specific elements on the hepcidin promoter<sup>61,62</sup> to induce transcription. BMP signaling may also impact hepcidin expression by epigenetic mechanisms.<sup>63</sup>

Mouse genetic studies have identified the key molecular players in the BMP-SMAD pathway that work in concert with HJV to regulate hepcidin and systemic iron homeostasis: BMP6<sup>64,65</sup> and BMP2 ligands,<sup>66,67</sup> ALK2 and ALK3 type I receptors,<sup>68</sup> ACVR2A and BMPR2 type II receptors,<sup>69</sup> R-SMADs 1 and 5,<sup>70</sup> and SMAD4,<sup>63</sup> since global or tissue-specific knockout of these molecules resulted in hepcidin deficiency and HH in mice (Table 1). SMAD7 and SMAD6 are inhibitory SMADs that are induced by BMP-SMAD signaling and feedback to inhibit the pathway by interfering with type I receptor function or SMAD complex formation. Hepatocyte *Smad7* knockout mice exhibited hepcidin excess and mild iron deficiency anemia consistent with a physiologic role in inhibiting hepcidin expression in vivo<sup>71</sup> (Table 1). A similar inhibitory effect of *Smad6* on hepcidin expression was also demonstrated in vitro.<sup>72</sup> Neogenin is another HJV-interacting protein that binds a distinct domain from BMP ligands<sup>60</sup> and may function as a scaffold protein to assist complex formation and localization to modulate signaling.<sup>73-75</sup> Neogenin mutant mice exhibited impaired BMP signaling, hepcidin deficiency, and iron overload consistent with an in vivo role<sup>76</sup> (Table 1). Whereas most of these proteins function to regulate hepcidin directly in hepatocytes, BMP2 and BMP6 ligands originate from liver endothelial cells, where they have paracrine effects on HJV and the BMP receptor complex.<sup>67,77</sup> The relevance of these molecular players in humans is corroborated by reports of higher ferritin in *HFE* hemochromatosis patients with a common *BMP2* polymorphism<sup>78</sup> and iron overload in patients with heterozygous mutations in the *BMP6* prodomain,<sup>79,80</sup> although the causal role of the human *BMP6* mutations has been questioned.<sup>81</sup> Building on these molecular discoveries, pharmacologic BMP-SMAD pathway inhibitors lowered hepcidin and increased serum and tissue iron in normal mice and reversed hepcidin excess, iron-restricted erythropoiesis, and anemia in animal models of anemia of inflammation.<sup>64,66,82-85</sup> Conversely, exogenous BMP2 or BMP6 induced hepcidin and reduced serum iron in normal mice and ameliorated hemochromatosis in *Hfe*<sup>-/-</sup> mice.<sup>58,64,86</sup> These data support the therapeutic possibility of targeting BMP-SMAD signaling in diseases of hepcidin excess or deficiency.

**Figure 2. Current model of iron sensing in the liver to regulate hepcidin production.** Both tissue iron and plasma iron levels are sensed by the liver to regulate hepcidin production.

Iron loading increases transferrin-bound iron (2Fe-TF) and non-transferrin-bound iron (NTBI) in the circulation (right). Iron is taken up by liver endothelial cells (ECs), which play an important role in sensing tissue iron levels. Iron loading in ECs increase BMP6 and, to a lesser extent, BMP2 expression. BMP6 is partially regulated by nuclear factor (erythroid-derived 2)-like 2 (NRF2), which is activated by iron-induced reactive oxygen species. The increased BMP2 and BMP6 bind to HJV on the hepatocyte membrane and facilitate the formation of a signaling complex consisting of BMP type I and type II receptors (BMPRI and BMPRII). Neogenin (Neo), a scaffold protein binding to HJV, assists with the signaling complex formation and localization. The signaling complex phosphorylates SMAD1/5/8, which binds to SMAD4 and translocates to the nucleus to induce hepcidin (*Hamp*) transcription. Plasma iron loading is also sensed directly by hepatocytes, where 2Fe-TF binds to TFR1 and TFR2 on the hepatocyte membrane, favoring the displacement of HFE from TFR1 and the interaction between HFE and TFR2. Both HFE and TFR2 stimulate hepcidin expression via a functional interaction with the SMAD pathway, possibly by forming a complex with HJV and stabilizing BMP type I receptor ALK3. Under low-iron conditions, BMP2 and BMP6 ligand production is reduced in ECs (left). HFE binds to TFR1, whose expression levels increase, whereas TFR2 levels decrease. Iron deficiency also increases TMPRSS6/matriptase-2 (MT2) and furin, which cleave membrane HJV to generate soluble HJV (sHJV). Suppression of BMP expression, HJV cleavage from the membrane, sequestration of HFE with TFR1, and reduced TFR2 expression all diminish BMP-SMAD signaling, thus suppressing hepcidin expression.



BMP ligands and receptors function in a multimeric complex containing a dimeric ligand, 2 type I receptors, and 2 type II receptors. In some contexts, heterodimeric ligands or receptor complexes (containing 2 different ligands or type I receptors) are critical for biological function.<sup>87,88</sup> Although individual components have been identified, the full nature of the BMP-HJV-receptor signaling complex(es) for hepcidin regulation remains unknown. Whereas HJV enhances BMP induction of hepcidin in cell culture systems and animals, HJV is not required for BMP2 or BMP6 signaling, since these ligands can still induce hepcidin, albeit less potently, in *Hjv*<sup>-/-</sup> primary hepatocytes.<sup>58,89</sup> Moreover, mice with a knockout of both *Bmp6* and *Hjv* have a stronger iron overload phenotype than single *Hjv* knockout mice<sup>90</sup> (Table 2). Interestingly, the crystal structure of BMP2 in complex with HJV reveals that HJV binds to the same interface as the type I receptor and even competes for type I receptor binding in cell-free binding assays.<sup>60</sup> Since type I receptors are required for R-SMAD phosphorylation, how HJV facilitates rather than inhibits signaling in cell culture systems and in vivo is unclear. One hypothesis suggests that HJV enhances BMP binding on the cell surface, and after internalization, type I receptors replace HJV in the acidic endosomal compartment that favors BMP type I receptor interactions over BMP-HJV interactions.<sup>60</sup> This hypothesis awaits validation by functional studies. HJV may also interact with other proteins that modulate optimal BMP-SMAD signal transduction in response to iron, such as neogenin<sup>60</sup> (as discussed above) or HFE, TFR2, and TMPRSS6 (as discussed below).<sup>91,92</sup> Another open question involves the relative roles of individual BMP ligands and receptors, whether they are activated under different conditions, have different signaling efficiency, or involve heteromeric ligand or receptor complexes. Type II receptors have completely redundant functions, since

single hepatocyte knockout of either *Acvr2a* or *Bmpr2* did not impact iron homeostasis, whereas double knockout caused HH.<sup>69</sup> In contrast, hepatocyte knockout of either type I receptor caused HH, but *Alk3* knockout had a stronger phenotype than *Alk2* knockout, suggesting at least some nonoverlapping functions.<sup>68</sup> Endothelial knockout of either *Bmp2* or *Bmp6* also caused HH, consistent with nonredundant roles<sup>66,67,77</sup>; however, severity was not directly comparable due to different genetic backgrounds (Tables 1 and 2).

## Hepcidin regulation by HFE and TFR2

Like mutations in *HJV* or hepcidin, mutations in *HFE* or *TFR2* also lead to HH, although with less severe iron overload that presents later in life.<sup>43</sup> HFE is an atypical major histocompatibility complex class I-like molecule.<sup>93</sup> TFR2 is a receptor for the plasma iron carrier transferrin with more limited tissue distribution and a lower affinity than TFR1, the primary receptor for transferrin-bound iron uptake.<sup>94</sup> Both HFE and TFR2 stimulate hepcidin expression through a functional interaction with the SMAD signaling pathway (Figure 2), as demonstrated by blunted SMAD signaling and hepcidin expression in mice and/or patients with mutations in these genes.<sup>95-100</sup> In vitro overexpression systems demonstrated that HFE and TFR2 bind to each other<sup>101</sup> and to HJV,<sup>91</sup> and these protein interactions are thought to be one mechanism by which HFE and TFR2 influence hepcidin production. HFE was also demonstrated to bind ALK3 and increase its stability by interfering with ubiquitination and degradation.<sup>102</sup> Whether these protein interactions occur with endogenously expressed proteins in vivo remains uncertain. Notably, genetic studies suggest that the functions of these proteins are not entirely overlapping and that some proteins have a more

**Table 1. Phenotype of mice with a single knockout of key players in liver iron sensing**

| Single mutant  | Hepcidin          | Liver iron | Extrahepatic iron loading | Anemia | References  |
|--|-------------------|------------|---------------------------|--------|-------------|
| <b>Ligand</b>  |                   |            |                           |        |             |
| <i>Bmp6</i>  | ↓↓↓               | ↑↑↑↑       | Yes (M>F)                 | No     | 64,65,77,90 |
| <i>Bmp2*</i>   | ↓↓↓               | ↑↑↑↑       | Yes (M>F)                 | No     | 66,67       |
| <b>Cell membrane-sensing molecules and receptors</b> |                   |            |                           |        |             |
| <i>Hjv</i>   | ↓↓↓               | ↑↑↑↑       | Yes (M>F)                 | No     | 47,90       |
| <i>Hfe</i> or $\beta 2m$                             | ↓                 | ↑↑         | No                        | No     | 107         |
| <i>Tfr2</i>  | ↓↓                | ↑↑↑        | No                        | No     | 107         |
| <i>Alk2</i> †  | ↓                 | ↑          | n/a                       | No     | 68          |
| <i>Alk3</i> †  | ↓↓↓               | ↑↑↑↑       | n/a                       | No     | 68          |
| <i>Acvr2a</i> or <i>Bmpr2</i> †                      | →                 | →          | n/a                       | No     | 69          |
| <i>Tmprss6</i>                                       | ↑                 | ↓          | No                        | Yes    | 109,112     |
| Neogenin   | ↓                 | ↑↑         | n/a                       | No     | 76          |
| <b>Intracellular molecules</b>                       |                   |            |                           |        |             |
| <i>Smad1</i> or <i>Smad5</i> †                       | 12 d ↓<br>Adult → | →          | n/a                       | No     | 70          |
| <i>Smad4</i> †                                       | ↓↓↓               | ↑↑↑↑       | Yes‡                      | No     | 63          |
| <i>Smad7</i> †                                       | ↑                 | ↓          | n/a                       | Yes    | 71          |

Levels of hepcidin expression and tissue iron loading are subject to variation based on background strains, ages, and analytical variations in different studies.

F, female; M, male; n/a, data not available.

\*Endothelial conditional knockout.

†Hepatocyte conditional knockout.

‡Study did not indicate sex of mice used.

dominant role than others (Tables 1 and 2). For example, combined knockout/mutation of *HFE* and *TFR2* resulted in more severe iron overload than single knockout/mutation of either gene in both mice and humans.<sup>97,103</sup> Moreover, *Tfr2* mutant mice overexpressing an *Hfe* transgene in hepatocytes had hepcidin excess and iron deficiency, suggesting that HFE can induce hepcidin in the absence of TFR2.<sup>104</sup> *Hjv* knockout mice had more severe iron overload than *Hfe* or *Tfr2* knockout mice (similar to the human phenotype<sup>43</sup>) and were not worsened by a combined loss.<sup>105,106</sup> Knockout of *Hjv*, *Tfr2*, or  $\beta 2$  microglobulin (a chaperone required for HFE localization) further dampened SMAD signaling, hepcidin expression, and iron overload in *Bmp6* knockout mice,<sup>107</sup> suggesting that these proteins can modulate hepcidin using BMP2 (or other BMP ligands) in the absence of BMP6. Taken together, the current data suggest that HJV, HFE, and TFR2 all converge on the same pathway to modulate BMP-SMAD signaling and thereby hepcidin. Although all proteins must be present for optimal signaling, multiple types of BMP ligand/receptor/accessory protein complexes appear to be capable of stimulating hepcidin production to some degree, and none of the single protein components are absolutely required, so loss of any one protein results in a modulatory effect rather than a complete abolishment of signaling. HFE and/or TFR2 may also have additional roles outside of modulating the BMP-SMAD pathway to regulate hepcidin production. For example, TFR2 is also expressed in erythroid cells, where it has a functional role to modulate erythropoietin (EPO) signaling and erythropoiesis according to iron availability<sup>108,109</sup> and therefore may also contribute to hepcidin regulation via the erythroid signal.<sup>108,109</sup>

## Hepcidin regulation by TMPRSS6

Mutations or knockout of *TMPRSS6* result in IRIDA, characterized by hepcidin excess and iron deficiency anemia that does not respond to oral iron and only partially responds to parental iron.<sup>53-55</sup> Common variants in *TMPRSS6* are also associated with alterations in iron and red blood cell parameters.<sup>110,111</sup> A serine protease that is highly expressed in the liver, *TMPRSS6* (also known as matriptase-2), also appears to regulate hepcidin by modulating the central BMP-HJV-SMAD signaling pathway (Figure 2). In particular, *Tmprss6* knockout mice exhibited increased SMAD signaling in parallel to hepcidin excess, and ablation of *Tmprss6* in *Hjv* knockout mice did not reverse hepcidin deficiency or iron loading<sup>112</sup> (Table 2). The mechanism of action of *TMPRSS6*, as demonstrated by in vitro overexpression systems, is to bind and cleave HJV, thereby releasing the coreceptor from the membrane surface and blunting BMP-SMAD signaling.<sup>92</sup> Although recombinant soluble HJV fused to immunoglobulin G Fc inhibited BMP-SMAD signaling by sequestering BMP ligands from the receptor signaling apparatus,<sup>82</sup> *TMPRSS6*-cleaved HJV did not inhibit signaling.<sup>113</sup> Interestingly, *Tmprss6* knockout mice exhibited reduced rather than increased liver HJV expression,<sup>114</sup> as might be anticipated based on its proposed mechanism of action. Recently, *TMPRSS6* overexpression in hepatocytes was demonstrated to reduce hepcidin expression in *Hjv* knockout mice, suggesting the existence of another mechanism for *TMPRSS6*-mediated hepcidin suppression independent of HJV.<sup>115</sup> In this study, *TMPRSS6* was also shown to cleave other members of the BMP-SMAD signaling apparatus, including ALK3, ALK2, ACVR2A, BMPR2, HFE, TFR2, and neogenin, in in vitro overexpression systems.<sup>115</sup> Nevertheless, HFE

**Table 2. Phenotype of mice with double vs single knockout of key players in liver iron sensing**

| Double mutant                 | Compared with                   | Hamp levels              | Liver iron | Extrahepatic iron loading | Anemia | References |
|-------------------------------|---------------------------------|--------------------------|------------|---------------------------|--------|------------|
| <i>Bmp6</i> × <i>Hjv</i>      | <i>Bmp6</i> or <i>Hjv</i>       | (F) ↓↓↓<br>(M) ↓↓        | →          | (F) ↑↑<br>(M) →           | No     | 90,107     |
| <i>Bmp6</i> × <i>β2m</i>      | <i>β2m</i><br><i>Bmp6</i>       | ↓↓↓<br>(F) ↓↓↓<br>(M) ↓↓ | ↑↑<br>→    | ↑↑<br>(F) ↑↑<br>(M) →     | No     | 107        |
| <i>Bmp6</i> × <i>Tfr2</i>     | <i>Tfr2</i><br><i>Bmp6</i>      | ↓↓↓<br>(F) ↓↓↓<br>(M) ↓↓ | ↑<br>→     | ↑↑<br>(F) ↑<br>(M) →      |        | 107        |
| <i>Hjv</i> × <i>Hfe</i>       | <i>Hfe</i><br><i>Hjv</i>        | ↓↓<br>→                  | ↑↑↑<br>→   | n/a<br>n/a                | No     | 10,106     |
| <i>Hjv</i> × <i>Tfr2</i>      | <i>Tfr2</i><br><i>Hjv</i>       | →<br>→                   | ↑<br>→     | n/a<br>n/a                | No     | 105,106    |
| <i>Hfe</i> × <i>Tfr2</i>      | <i>Hfe</i><br><i>Tfr2</i>       | ↓↓<br>↓                  | ↑↑<br>↑    | n/a<br>n/a                | No     | 97         |
| <i>Acvr2a</i> × <i>Bmpr2*</i> | <i>Acvr2a*</i> or <i>Bmpr2*</i> | ↓↓↓                      | ↑↑↑↑       | n/a                       | No     | 69         |
| <i>Smad1</i> × <i>Smad5*</i>  | <i>Smad1*</i> or <i>Smad5*</i>  | ↓↓↓                      | ↑↑↑↑       | n/a                       | No     | 70         |
| <i>Tmprss6</i> × <i>Hfe</i>   | <i>Tmprss6</i><br><i>Hfe</i>    | →<br>↑                   | →<br>↓     | →<br>↓                    | Yes    | 116        |
| <i>Tmprss6</i> × <i>Hjv</i>   | <i>Tmprss6</i><br><i>Hjv</i>    | ↓↓↓<br>→                 | ↑↑↑↑<br>→  | ↑↑<br>→                   | No     | 109        |
| <i>Tmprss6</i> × <i>Tfr2</i>  | <i>Tmprss6</i>                  | →                        | →          | n/a                       | Yes    | 109,117    |
| <i>Tmprss6</i> × <i>Bmp6</i>  | <i>Tmprss6</i><br><i>Bmp6</i>   | ↓<br>↑↑                  | ↑<br>↓↓    | →<br>↓                    | No     | 149        |

Levels of hepcidin expression and tissue iron loading are subject to variation based on background strains ages, and analytical variations in different studies.

\*Hepatocyte conditional knockout.

and TFR2 do not appear critical to TMPRSS6 function *in vivo*, since mice with a combined deletion of *Tfr2* and/or *Hfe* and *Tmprss6* still exhibited a similar degree of hepcidin excess and iron deficiency anemia as single *Tmprss6* knockout mice<sup>109,116,117</sup> (Table 2). A limitation for most of these studies is the use of an overexpression system to ascertain the molecular targets of TMPRSS6. The identification of TMPRSS6 as an endogenous hepcidin suppressor has resulted in the development of novel TMPRSS6-inhibiting therapies that have shown promise for increasing hepcidin to limit iron overload in mouse models of  $\beta$ -thalassemia and HH.<sup>118,119</sup>

## Iron sensing in the liver

There are 2 distinct iron signals that regulate hepatocyte hepcidin production: acute changes in plasma iron and changes in tissue iron.<sup>39,40</sup> Both of these iron signals stimulate hepcidin production by activating the BMP-SMAD signaling cascade, but through different mechanisms (Figure 2). Tissue iron is sensed by liver endothelial cells to regulate production of BMP ligands, which have paracrine effects to induce hepcidin production in hepatocytes. Indeed, chronic dietary iron loading or deficiency that alters liver iron levels concordantly regulated *Bmp6* and

*Bmp2* messenger RNA (mRNA) expression in mouse liver endothelial cells,<sup>66,120</sup> although *Bmp6* expression was modulated more than *Bmp2*.<sup>66</sup> BMP6 regulation by iron appears to be a cell-autonomous event in liver endothelial cells, since *Bmp6* was induced more strongly by dietary iron in *Zip14/Slc39a14* knockout mice that fail to load iron in hepatocytes but load more iron in liver nonparenchymal cells,<sup>121</sup> and importantly, *Bmp6* was induced by iron loading in isolated liver endothelial cell primary cultures.<sup>122</sup> One mechanism by which iron induces BMP6 in liver endothelial cells is through transcriptional regulation by NRF2, which is activated by iron-induced reactive oxygen species.<sup>123</sup> NRF2 activation in this context has the dual benefit of inducing other antioxidant protective pathways in addition to limiting further iron loading as a consequence of BMP6-mediated hepcidin upregulation.<sup>121</sup>

Acute increases in plasma iron in the absence of liver iron loading also induce hepcidin expression, but without inducing liver *Bmp6* mRNA expression.<sup>39</sup> This is associated with an induction of the downstream SMAD1/5/8 signaling cascade.<sup>39</sup> Whether SMAD1/5/8 activation in this context occurs downstream of BMP6 or in response to BMP2 or other BMP ligands is still not known. Hepatocyte HFE and TFR2 are both proposed to play a critical role in

sensing plasma iron levels in the form of iron-bound transferrin (holo-transferrin). TFR2 binds directly to holo-transferrin.<sup>94</sup> HFE binds to transferrin receptor 1 (TFR1) at the transferrin binding site and is displaced by increased holo-transferrin.<sup>124,125</sup> Interestingly, mice expressing mutant TFR1 that does not bind HFE exhibited high hepcidin.<sup>126</sup> Thus, displacement of HFE by holo-transferrin may be one mechanism by which hepatocytes sense plasma iron levels to upregulate hepcidin.<sup>126</sup> Cell culture models suggest holo-transferrin favors the interaction between HFE and TFR2 and this may also play an important role in hepcidin induction by iron.<sup>101</sup> Notably, iron reduces TFR1 and increases TFR2 levels, which may also help favor the release of HFE from TFR1 and/or interaction with TFR2. For TFR1, mRNA stability is reduced by intracellular iron loading through the iron regulatory protein (IRP)/iron response element (IRE) system, which is key mediator of cellular iron homeostasis (discussed below).<sup>2</sup> In contrast, TFR2 is stabilized by holo-transferrin,<sup>127,128</sup> which redirects it from a degradation to a recycling pathway.<sup>129</sup> Holo-transferrin also reduces release of TFR2 from the membrane surface.<sup>130</sup>

Additional mechanisms may also be involved in sensing iron deficiency (Figure 2). A low-iron diet and cellular iron deficiency increase TMPRSS6 by inhibiting its degradation.<sup>131-133</sup> Iron deficiency also increases HJV cleavage by the furin family of pro-protein convertases,<sup>134-136</sup> generating a distinct form of soluble HJV (sHJV) compared with TMPRSS6 cleavage. Circulating sHJV was increased in mice on a low-iron diet, consistent with increased HJV cleavage by one or both of these mechanisms.<sup>137</sup> Thus, iron deficiency may suppress hepcidin production by release of membrane-bound HJV from hepatocytes. Furin-cleaved sHJV has also been demonstrated to act as a decoy to inhibit BMP-mediated hepcidin induction, although this property is not shared by TMPRSS6-cleaved sHJV.<sup>113</sup> However, circulating sHJV levels in vivo are relatively low<sup>137</sup> and may not be sufficient to inhibit BMP signaling.

## Hepcidin regulation by inflammation, erythropoietic demand, and other stimuli

Hepcidin induction by inflammation plays a protective role against some infections by lowering available circulating iron<sup>138,139</sup>; however, excessive hepcidin induction in chronic inflammatory diseases contributes to iron restricted erythropoiesis and anemia.<sup>41</sup> Mechanistically, inflammation upregulates hepcidin predominantly by interleukin-6,<sup>140</sup> which induces phosphorylation of signal transducer and activator of transcription 3 that binds to the hepcidin promoter adjacent to the proximal BMP response element.<sup>141-143</sup> Other inflammatory cytokines such as interleukin-1 $\beta$  and activin B may also contribute, although their functional roles in vivo are less clearly defined.<sup>41</sup>

Erythropoietic drive due to hypoxia, anemia, or pathologic conditions of ineffective erythropoiesis induce the kidney to produce EPO, which not only increases proliferation and terminal differentiation of red blood cells but also induces erythroblasts to secrete erythroferrone (ERFE) and possibly other "erythroid regulators" that function to suppress hepcidin.<sup>2</sup> This not only ensures the adequate delivery of iron to support erythropoiesis but also contributes to pathologic iron loading in conditions of ineffective erythropoiesis, such as in  $\beta$ -thalassemia. A key role for ERFE was demonstrated by the failure of *Erfe* knockout mice to

suppress hepcidin in response to hemorrhage or EPO injection<sup>144</sup> and the amelioration of hepcidin deficiency and iron loading in a *Erfe* knockout thalassemia model.<sup>145</sup> Other proposed erythroid regulators include platelet-derived growth factor BB, which is induced by hypoxia in humans and mice and suppresses hepcidin when injected into mice.<sup>146</sup> Growth and differentiation factor 15 is induced in the blood of patients with thalassemia,<sup>147</sup> but its functional role in hepcidin suppression is less certain.<sup>2</sup>

Hepcidin regulation by erythropoietic drive and inflammation depend on a functional SMAD signaling pathway, suggesting important cross talk with the iron-sensing pathway. EPO and ERFE suppressed liver SMAD1/5 phosphorylation in wild-type mice and primary hepatocytes, respectively, and failed to suppress hepcidin in hepatocyte *Smad1/5* conditional knockout mice.<sup>70</sup> EPO likely suppresses the SMAD1/5 pathway at least in part by increasing iron consumption for erythropoiesis, reducing circulating holo-transferrin, and triggering the liver iron sensing pathway.<sup>148,149</sup> The mechanism of action of ERFE to suppress SMAD1/5 signaling and hepcidin directly is less certain. Its effects do not require BMP6 or HJV, since EPO still suppressed hepcidin in *Bmp6* and *Hjv* knockout mice.<sup>149,150</sup> Its direct effects are also independent of TMPRSS6, since ERFE efficiently suppressed hepcidin in *Tmprss6* knockout primary hepatocytes.<sup>151</sup> However, *Tmprss6* knockout mice failed to suppress hepcidin in response to EPO despite ERFE induction, similar to mice on a high-iron diet, suggesting that ERFE's actions in vivo are impaired when SMAD signaling is high.<sup>149</sup> Notably, *Tmprss6* inhibition ameliorated hepcidin deficiency and iron overload in a thalassemia mouse model, demonstrating the overall importance of TMPRSS6 function for erythroid-mediated hepcidin suppression in conditions of ineffective erythropoiesis with potential therapeutic implications.<sup>118,119,152</sup>

The cross talk between inflammation and BMP-SMAD signaling in hepcidin regulation is demonstrated by blunted hepcidin induction by inflammation in hepatocyte conditional *Smad4* knockout mice,<sup>63</sup> when the hepcidin promoter proximal BMP response element was mutated,<sup>153</sup> or in the presence of BMP inhibitors.<sup>84,85</sup> This cross talk is postulated to occur at the level of the hepcidin promoter or by stimulating production of the BMP/transforming growth factor- $\beta$  superfamily member activin B, which can use BMP type I receptors to induce SMAD1/5/8 phosphorylation and hepcidin expression in hepatocytes.<sup>89,154</sup> However, a functional role for activin B in vivo is uncertain given preserved hepcidin induction by inflammation in activin B knockout mice.<sup>155</sup>

In addition to inflammation and erythropoietic drive, a variety of other stimuli have been demonstrated to regulate hepcidin production, including endoplasmic reticulum stress, hormones (testosterone, estrogen, and progesterone), growth factors (hepatocyte growth factor and epidermal growth factor), and other signaling pathways (Ras/RAF and mTOR).<sup>156,157</sup> Many of these mediators also cross talk with the central BMP-SMAD pathway.<sup>156,158</sup>

## Cross talk between systemic and cellular iron homeostasis

Iron sensing at the cellular level in nonhepatic tissues involved in systemic iron transport and erythropoiesis can also contribute to

systemic iron homeostasis. A key mediator of intracellular iron homeostasis is the IRP/IRE system.<sup>2</sup> In iron-deficient conditions, IRPs bind to IREs in the 3' untranslated region of transcripts such as TFR1 and DMT1 to increase mRNA stability or in the 5' untranslated region of transcripts such as ferritin or ferroportin to inhibit translation.<sup>2</sup> IRPs are inactivated in iron-replete conditions either by binding to an iron sulfur cluster or by being targeted for proteosomal degradation by the iron-binding hemerythrin and E3-ubiquitin ligase F-box/leucine-rich repeat protein 5.<sup>2</sup> Another sensor of cellular iron status is the hypoxia-inducible factor (HIF) family of transcription factors that are stabilized to regulate gene transcription by iron deficiency or hypoxia. HIFs are inactivated in iron-replete conditions by the oxygen- and iron-dependent prolyl hydroxylases that target regulatory  $\alpha$  subunits for degradation.<sup>1</sup> A prime example where cellular iron homeostasis can influence systemic iron homeostasis is duodenal enterocytes, where HIF-2 $\alpha$  mediates upregulation of DMT1, duodenal cytochrome B, and ferroportin in response to iron deficiency to influence duodenal iron uptake.<sup>159-161</sup> Indeed, genetic disruption of intestinal HIF-2 $\alpha$  signaling reduced serum and tissue iron levels and exacerbated iron deficiency and anemia by a low-iron diet despite appropriate hepcidin suppression.<sup>159,160</sup> IRP1 appears to have more of a counterregulatory effect in this system by downregulating HIF-2 $\alpha$ .<sup>162</sup>

## Conclusions

The hepcidin-ferroportin axis is a key mediator of systemic iron homeostasis by controlling iron availability from dietary sources and body stores to support erythropoiesis and other essential functions but prevent the toxicity of iron excess. Insufficient or excess activity of this pathway contributes to most disorders of iron metabolism, including HH, iron-loading anemias, anemia of inflammation, and IRIDA. Identifying the genetic basis of iron disorders has provided important advances in understanding

how the liver senses plasma and tissue iron levels and integrates these signals with inflammation and erythropoietic drive to control hepcidin production. Future research is needed to fully comprehend the molecular mechanisms of iron sensing and the complex interplay among the proteins involved in hepcidin regulation. These studies are already contributing to the development of novel therapies for iron disorders.

## Acknowledgments

This work was supported by the National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases (grant RO1-DK087727) (J.L.B.).

## Authorship

Contribution: J.L.B. wrote the manuscript and edited the figures and tables; and C.-Y.W. created the figures and tables and edited the manuscript.

Conflict-of-interest disclosure: J.L.B. has ownership interest in Ferrumax Pharmaceuticals and has received consulting fees from Keryx Biopharmaceuticals and Disc Medicine. C.-Y.W. declares no competing financial interests.

ORCID profiles: CY.W., 0000-0003-3679-6453; J.L.B., 0000-0003-1228-1551.

Correspondence: Jodie L. Babitt, Division of Nephrology, Program in Membrane Biology, Center for Systems Biology, Massachusetts General Hospital, 185 Cambridge St, CPZN-8208, Boston, MA 02114; e-mail: babitt.jodie@mgh.harvard.edu.

## Footnote

Submitted 14 June 2018; accepted 21 July 2018. Prepublished online as Blood First Edition paper, 6 November 2018; DOI 10.1182/blood-2018-06-815894.

## REFERENCES

- Dev S, Babitt JL. Overview of iron metabolism in health and disease. *Hemodial Int*. 2017;21(Suppl 1):S6-S20.
- Muckenthaler MU, Rivella S, Hentze MW, Galy B. A red carpet for iron metabolism. *Cell*. 2017;168(3):344-361.
- Fleming MD, Trenor CC III, Su MA, et al. Microcytic anaemia mice have a mutation in *Nramp2*, a candidate iron transporter gene. *Nat Genet*. 1997;16(4):383-386.
- Gunshin H, Mackenzie B, Berger UV, et al. Cloning and characterization of a mammalian proton-coupled metal-ion transporter. *Nature*. 1997;388(6641):482-488.
- Gunshin H, Fujiwara Y, Custodio AO, Drenzo C, Robine S, Andrews NC. *Slc11a2* is required for intestinal iron absorption and erythropoiesis but dispensable in placenta and liver. *J Clin Invest*. 2005;115(5):1258-1266.
- McKie AT, Barrow D, Latunde-Dada GO, et al. An iron-regulated ferric reductase associated with the absorption of dietary iron. *Science*. 2001;291(5509):1755-1759.
- Vulpe CD, Kuo YM, Murphy TL, et al. Hephaestin, a ceruloplasmin homologue implicated in intestinal iron transport, is defective in the *sla* mouse. *Nat Genet*. 1999; 21(2):195-199.
- Harris ZL, Durley AP, Man TK, Gitlin JD. Targeted gene disruption reveals an essential role for ceruloplasmin in cellular iron efflux. *Proc Natl Acad Sci USA*. 1999;96(19): 10812-10817.
- Cherukuri S, Potla R, Sarkar J, Nurko S, Harris ZL, Fox PL. Unexpected role of ceruloplasmin in intestinal iron absorption. *Cell Metab*. 2005;2(5):309-319.
- Finch C. Regulators of iron balance in humans. *Blood*. 1994;84(6):1697-1702.
- Park CH, Valore EV, Waring AJ, Ganz T. Hepcidin, a urinary antimicrobial peptide synthesized in the liver. *J Biol Chem*. 2001; 276(11):7806-7810.
- Pigeon C, Ilyin G, Courselaud B, et al. A new mouse liver-specific gene, encoding a protein homologous to human antimicrobial peptide hepcidin, is overexpressed during iron overload. *J Biol Chem*. 2001;276(11): 7811-7819.
- Krause A, Neitz S, Mägert HJ, et al. LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity. *FEBS Lett*. 2000;480(2-3):147-150.
- McKie AT, Marciani P, Rolfs A, et al. A novel duodenal iron-regulated transporter, IREG1, implicated in the basolateral transfer of iron to the circulation. *Mol Cell*. 2000;5(2): 299-309.
- Donovan A, Brownlie A, Zhou Y, et al. Positional cloning of zebrafish ferroportin1 identifies a conserved vertebrate iron exporter. *Nature*. 2000;403(6771):776-781.
- Abboud S, Haile DJ. A novel mammalian iron-regulated protein involved in intracellular iron metabolism. *J Biol Chem*. 2000;275(26):19906-19912.
- Nicolas G, Bennoun M, Devaux I, et al. Lack of hepcidin gene expression and severe tissue iron overload in upstream stimulatory factor 2 (USF2) knockout mice. *Proc Natl Acad Sci USA*. 2001;98(15):8780-8785.
- Roetto A, Papanikolaou G, Politou M, et al. Mutant antimicrobial peptide hepcidin is associated with severe juvenile hemochromatosis. *Nat Genet*. 2003;33(1):21-22.
- Nicolas G, Bennoun M, Porteu A, et al. Severe iron deficiency anemia in transgenic mice expressing liver hepcidin. *Proc Natl Acad Sci USA*. 2002;99(7):4596-4601.
- Weinstein DA, Roy CN, Fleming MD, Loda MF, Wolfsdorf JI, Andrews NC.



- Inappropriate expression of hepcidin is associated with iron refractory anemia: implications for the anemia of chronic disease. *Blood*. 2002;100(10):3776-3781.
21. Nicolas G, Chauvet C, Viatte L, et al. The gene encoding the iron regulatory peptide hepcidin is regulated by anemia, hypoxia, and inflammation. *J Clin Invest*. 2002;110(7):1037-1044.
  22. Donovan A, Lima CA, Pinkus JL, et al. The iron exporter ferroportin/Slc40a1 is essential for iron homeostasis. *Cell Metab*. 2005;1(3):191-200.
  23. Zhang DL, Senecal T, Ghosh MC, Ollivierre-Wilson H, Tu T, Rouault TA. Hepcidin regulates ferroportin expression and intracellular iron homeostasis of erythroblasts. *Blood*. 2011;118(10):2868-2877.
  24. Zhang Z, Zhang F, Guo X, An P, Tao Y, Wang F. Ferroportin1 in hepatocytes and macrophages is required for the efficient mobilization of body iron stores in mice. *Hepatology*. 2012;56(3):961-971.
  25. Lakhal-Littleton S, Wolna M, Carr CA, et al. Cardiac ferroportin regulates cellular iron homeostasis and is important for cardiac function. *Proc Natl Acad Sci USA*. 2015;112(10):3164-3169.
  26. Zhang DL, Wu J, Shah BN, et al. Erythrocytic ferroportin reduces intracellular iron accumulation, hemolysis, and malaria risk. *Science*. 2018;359(6383):1520-1523.
  27. Montosi G, Donovan A, Totaro A, et al. Autosomal-dominant hemochromatosis is associated with a mutation in the ferroportin (SLC11A3) gene. *J Clin Invest*. 2001;108(4):619-623.
  28. Njajou OT, Vaessen N, Joosse M, et al. A mutation in SLC11A3 is associated with autosomal dominant hemochromatosis. *Nat Genet*. 2001;28(3):213-214.
  29. Pietrangelo A. Ferroportin disease: pathogenesis, diagnosis and treatment. *Haematologica*. 2017;102(12):1972-1984.
  30. Nemeth E, Tuttle MS, Powelson J, et al. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science*. 2004;306(5704):2090-2093.
  31. Liu XB, Yang F, Haile DJ. Functional consequences of ferroportin 1 mutations. *Blood Cells Mol Dis*. 2005;35(1):33-46.
  32. Rice AE, Mendez MJ, Hokanson CA, Rees DC, Björkman PJ. Investigation of the biophysical and cell biological properties of ferroportin, a multipass integral membrane protein iron exporter. *J Mol Biol*. 2009;386(3):717-732.
  33. Taniguchi R, Kato HE, Font J, et al. Outward- and inward-facing structures of a putative bacterial transition-metal transporter with homology to ferroportin. *Nat Commun*. 2015;6:8545.
  34. Nemeth E, Preza GC, Jung CL, Kaplan J, Waring AJ, Ganz T. The N-terminus of hepcidin is essential for its interaction with ferroportin: structure-function study. *Blood*. 2006;107(1):328-333.
  35. Preza GC, Ruchala P, Pinon R, et al. Minihepcidins are rationally designed small peptides that mimic hepcidin activity in mice and may be useful for the treatment of iron overload. *J Clin Invest*. 2011;121(12):4880-4888.
  36. Aschemeyer S, Qiao B, Stefanova D, et al. Structure-function analysis of ferroportin defines the binding site and an alternative mechanism of action of hepcidin. *Blood*. 2018;131(8):899-910.
  37. Qiao B, Sugianto P, Fung E, et al. Hepcidin-induced endocytosis of ferroportin is dependent on ferroportin ubiquitination. *Cell Metab*. 2012;15(6):918-924.
  38. Ross SL, Tran L, Winters A, et al. Molecular mechanism of hepcidin-mediated ferroportin internalization requires ferroportin lysines, not tyrosines or JAK-STAT. *Cell Metab*. 2012;15(6):905-917.
  39. Corradini E, Meynard D, Wu Q, et al. Serum and liver iron differently regulate the bone morphogenetic protein 6 (BMP6)-SMAD signaling pathway in mice. *Hepatology*. 2011;54(1):273-284.
  40. Ramos E, Kautz L, Rodríguez R, et al. Evidence for distinct pathways of hepcidin regulation by acute and chronic iron loading in mice. *Hepatology*. 2011;53(4):1333-1341.
  41. Wang CY, Babitt JL. Hepcidin regulation in the anemia of inflammation. *Curr Opin Hematol*. 2016;23(3):189-197.
  42. Casu C, Nemeth E, Rivella S. Hepcidin agonists as therapeutic tools. *Blood*. 2018;131(16):1790-1794.
  43. Pietrangelo A. Genetics, genetic testing, and management of hemochromatosis: 15 years since hepcidin. *Gastroenterology*. 2015;149(5):1240-1251.e1244.
  44. Papanikolaou G, Samuels ME, Ludwig EH, et al. Mutations in HFE2 cause iron overload in chromosome 1q-linked juvenile hemochromatosis. *Nat Genet*. 2004;36(1):77-82.
  45. Bridle KR, Frazer DM, Wilkins SJ, et al. Disrupted hepcidin regulation in HFE-associated hemochromatosis and the liver as a regulator of body iron homeostasis. *Lancet*. 2003;361(9358):669-673.
  46. Nemeth E, Roetto A, Garozzo G, Ganz T, Camaschella C. Hepcidin is decreased in TFR2 hemochromatosis. *Blood*. 2005;105(4):1803-1806.
  47. Huang FW, Pinkus JL, Pinkus GS, Fleming MD, Andrews NC. A mouse model of juvenile hemochromatosis. *J Clin Invest*. 2005;115(8):2187-2191.
  48. Niederkofler V, Salie R, Arber S. Hemojuvelin is essential for dietary iron sensing, and its mutation leads to severe iron overload. *J Clin Invest*. 2005;115(8):2180-2186.
  49. Kawabata H, Fleming RE, Gui D, et al. Expression of hepcidin is down-regulated in Tfr2 mutant mice manifesting a phenotype of hereditary hemochromatosis. *Blood*. 2005;105(1):376-381.
  50. Muckenthaler M, Roy CN, Custodio AO, et al. Regulatory defects in liver and intestine implicate abnormal hepcidin and Cybrd1 expression in mouse hemochromatosis. *Nat Genet*. 2003;34(1):102-107.
  51. Nicolas G, Viatte L, Lou DQ, et al. Constitutive hepcidin expression prevents iron overload in a mouse model of hemochromatosis. *Nat Genet*. 2003;34(1):97-101.
  52. Wallace DF, Summerville L, Lusby PE, Subramaniam VN. First phenotypic description of transferrin receptor 2 knockout mouse, and the role of hepcidin. *Gut*. 2005;54(7):980-986.
  53. Du X, She E, Gelbart T, et al. The serine protease TMPRSS6 is required to sense iron deficiency. *Science*. 2008;320(5879):1088-1092.
  54. Finberg KE, Heeney MM, Campagna DR, et al. Mutations in TMPRSS6 cause iron-refractory iron deficiency anemia (IRIDA). *Nat Genet*. 2008;40(5):569-571.
  55. Folgueras AR, de Lara FM, Pendás AM, et al. Membrane-bound serine protease matrilysin-2 (Tmprss6) is an essential regulator of iron homeostasis. *Blood*. 2008;112(6):2539-2545.
  56. Babitt JL, Zhang Y, Samad TA, et al. Repulsive guidance molecule (RGMa), a DRAGON homologue, is a bone morphogenetic protein co-receptor. *J Biol Chem*. 2005;280(33):29820-29827.
  57. Samad TA, Rebbapragada A, Bell E, et al. DRAGON, a bone morphogenetic protein co-receptor. *J Biol Chem*. 2005;280(14):14122-14129.
  58. Babitt JL, Huang FW, Wrighting DM, et al. Bone morphogenetic protein signaling by hemojuvelin regulates hepcidin expression. *Nat Genet*. 2006;38(5):531-539.
  59. Wu Q, Sun CC, Lin HY, Babitt JL. Repulsive guidance molecule (RGM) family proteins exhibit differential binding kinetics for bone morphogenetic proteins (BMPs). *PLoS One*. 2012;7(9):e46307.
  60. Healey EG, Bishop B, Elegheert J, Bell CH, Padilla-Parra S, Siebold C. Repulsive guidance molecule is a structural bridge between neogenin and bone morphogenetic protein. *Nat Struct Mol Biol*. 2015;22(6):458-465.
  61. Truksa J, Lee P, Beutler E. Two BMP responsive elements, STAT, and bZIP/HNF4/COUP motifs of the hepcidin promoter are critical for BMP, SMAD1, and HJV responsiveness. *Blood*. 2009;113(3):688-695.
  62. Casanovas G, Mleczo-Sanecka K, Altamura S, Hentze MW, Muckenthaler MU. Bone morphogenetic protein (BMP)-responsive elements located in the proximal and distal hepcidin promoter are critical for its response to HJV/BMP/SMAD. *J Mol Med (Berl)*. 2009;87(5):471-480.
  63. Wang RH, Li C, Xu X, et al. A role of SMAD4 in iron metabolism through the positive regulation of hepcidin expression. *Cell Metab*. 2005;2(6):399-409.
  64. Andriopoulos B Jr, Corradini E, Xia Y, et al. BMP6 is a key endogenous regulator of hepcidin expression and iron metabolism. *Nat Genet*. 2009;41(4):482-487.
  65. Meynard D, Kautz L, Darnaud V, Canonne-Hergaux F, Coppin H, Roth MP. Lack of the

- bone morphogenetic protein BMP6 induces massive iron overload. *Nat Genet.* 2009; 41(4):478-481.
66. Canali S, Wang CY, Zumbrennen-Bullough KB, Bayer A, Babitt JL. Bone morphogenetic protein 2 controls iron homeostasis in mice independent of Bmp6. *Am J Hematol.* 2017; 92(11):1204-1213.
  67. Koch PS, Olsavszky V, Ulbrich F, et al. Angiocrine Bmp2 signaling in murine liver controls normal iron homeostasis. *Blood.* 2017;129(4):415-419.
  68. Steinbicker AU, Bartnikas TB, Lohmeyer LK, et al. Perturbation of hepcidin expression by BMP type I receptor deletion induces iron overload in mice. *Blood.* 2011;118(15): 4224-4230.
  69. Mayeur C, Leyton PA, Kolodziej SA, Yu B, Bloch KD. BMP type II receptors have redundant roles in the regulation of hepatic hepcidin gene expression and iron metabolism. *Blood.* 2014;124(13):2116-2123.
  70. Wang CY, Core AB, Canali S, et al. Smad1/5 is required for erythropoietin-mediated suppression of hepcidin in mice. *Blood.* 2017;130(1):73-83.
  71. An P, Wang H, Wu Q, et al. Smad7 deficiency decreases iron and haemoglobin through hepcidin up-regulation by multilayer compensatory mechanisms. *J Cell Mol Med.* 2018;22(6):3035-3044.
  72. Vujčić Spasić M, Sparla R, Mlecško-Sanecka K, et al. Smad6 and Smad7 are co-regulated with hepcidin in mouse models of iron overload. *Biochim Biophys Acta.* 2013; 1832(1):76-84.
  73. Zhao N, Maxson JE, Zhang RH, Wahedi M, Enns CA, Zhang AS. Neogenin facilitates the induction of hepcidin expression by hemojuelin in the liver. *J Biol Chem.* 2016; 291(23):12322-12335.
  74. Tian C, Shi H, Xiong S, Hu F, Xiong WC, Liu J. The neogenin/DCC homolog UNC-40 promotes BMP signaling via the RGM protein DRAG-1 in *C. elegans*. *Development.* 2013; 140(19):4070-4080.
  75. Hagihara M, Endo M, Hata K, et al. Neogenin, a receptor for bone morphogenetic proteins. *J Biol Chem.* 2011;286(7): 5157-5165.
  76. Lee DH, Zhou LJ, Zhou Z, et al. Neogenin inhibits HJV secretion and regulates BMP-induced hepcidin expression and iron homeostasis. *Blood.* 2010;115(15):3136-3145.
  77. Canali S, Zumbrennen-Bullough KB, Core AB, et al. Endothelial cells produce bone morphogenetic protein 6 required for iron homeostasis in mice. *Blood.* 2017;129(4): 405-414.
  78. Milet J, Dehais V, Bourgain C, et al. Common variants in the BMP2, BMP4, and HJV genes of the hepcidin regulation pathway modulate HFE hemochromatosis penetrance. *Am J Hum Genet.* 2007;81(4):799-807.
  79. Daher R, Kannengiesser C, Houamel D, et al. Heterozygous mutations in BMP6 pro-peptide lead to inappropriate hepcidin synthesis and moderate iron overload in humans. *Gastroenterology.* 2016;150(3): 672-683.e674.
  80. Piubelli C, Castagna A, Marchi G, et al. Identification of new BMP6 pro-peptide mutations in patients with iron overload. *Am J Hematol.* 2017;92(6):562-568.
  81. McDonald CJ, Rishi G, Wallace DF, Subramaniam VN. Genetic variants in the BMP6 pro-peptide may not cause iron loading and should be interpreted with caution. *Gastroenterology.* 2016;151(4): 770-771.
  82. Babitt JL, Huang FW, Xia Y, Sidis Y, Andrews NC, Lin HY. Modulation of bone morphogenetic protein signaling in vivo regulates systemic iron balance. *J Clin Invest.* 2007; 117(7):1933-1939.
  83. Yu PB, Hong CC, Sachidanandan C, et al. Dorsomorphin inhibits BMP signals required for embryogenesis and iron metabolism. *Nat Chem Biol.* 2008;4(1):33-41.
  84. Theurl I, Schroll A, Sonnweber T, et al. Pharmacologic inhibition of hepcidin expression reverses anemia of chronic inflammation in rats. *Blood.* 2011;118(18): 4977-4984.
  85. Steinbicker AU, Sachidanandan C, Vonner AJ, et al. Inhibition of bone morphogenetic protein signaling attenuates anemia associated with inflammation. *Blood.* 2011; 117(18):4915-4923.
  86. Corradini E, Schmidt PJ, Meynard D, et al. BMP6 treatment compensates for the molecular defect and ameliorates hemochromatosis in Hfe knockout mice. *Gastroenterology.* 2010;139(5):1721-1729.
  87. Little SC, Mullins MC. Bone morphogenetic protein heterodimers assemble heteromeric type I receptor complexes to pattern the dorsoventral axis. *Nat Cell Biol.* 2009;11(5): 637-643.
  88. Isaacs MJ, Kawakami Y, Allendorph GP, Yoon BH, Izpisua Belmonte JC, Choe S. Bone morphogenetic protein-2 and -6 heterodimer illustrates the nature of ligand-receptor assembly. *Mol Endocrinol.* 2010; 24(7):1469-1477.
  89. Canali S, Core AB, Zumbrennen-Bullough KB, et al. Activin B induces noncanonical SMAD1/5/8 signaling via BMP type I receptors in hepatocytes: evidence for a role in hepcidin induction by inflammation in male mice. *Endocrinology.* 2016;157(3): 1146-1162.
  90. Latour C, Besson-Fournier C, Gourbeyre O, Meynard D, Roth MP, Coppin H. Deletion of BMP6 worsens the phenotype of HJV-deficient mice and attenuates hepcidin levels reached after LPS challenge. *Blood.* 2017; 130(21):2339-2343.
  91. D'Alessio F, Hentze MW, Muckenthaler MU. The hemochromatosis proteins HFE, TfR2, and HJV form a membrane-associated protein complex for hepcidin regulation. *J Hepatol.* 2012;57(5):1052-1060.
  92. Silvestri L, Pagani A, Nai A, De Domenico I, Kaplan J, Camaschella C. The serine protease matriptase-2 (TMPRSS6) inhibits hepcidin activation by cleaving membrane hemojuelin. *Cell Metab.* 2008;8(6):502-511.
  93. Feder JN, Gnirke A, Thomas W, et al. A novel MHC class I-like gene is mutated in patients with hereditary haemochromatosis. *Nat Genet.* 1996;13(4):399-408.
  94. Kawabata H, Yang R, Hiramata T, et al. Molecular cloning of transferrin receptor 2. A new member of the transferrin receptor-like family. *J Biol Chem.* 1999;274(30): 20826-20832.
  95. Corradini E, Garuti C, Montosi G, et al. Bone morphogenetic protein signaling is impaired in an HFE knockout mouse model of hemochromatosis. *Gastroenterology.* 2009; 137(4):1489-1497.
  96. Kautz L, Meynard D, Besson-Fournier C, et al. BMP/Smad signaling is not enhanced in Hfe-deficient mice despite increased Bmp6 expression. *Blood.* 2009;114(12):2515-2520.
  97. Wallace DF, Summerville L, Crampton EM, Frazer DM, Anderson GJ, Subramaniam VN. Combined deletion of Hfe and transferrin receptor 2 in mice leads to marked dysregulation of hepcidin and iron overload. *Hepatology.* 2009;50(6):1992-2000.
  98. Bolondi G, Garuti C, Corradini E, et al. Altered hepatic BMP signaling pathway in human HFE hemochromatosis. *Blood Cells Mol Dis.* 2010;45(4):308-312.
  99. Ryan JD, Ryan E, Fabre A, Lawless MW, Crowe J. Defective bone morphogenetic protein signaling underlies hepcidin deficiency in HFE hereditary hemochromatosis. *Hepatology.* 2010;52(4):1266-1273.
  100. Corradini E, Rozier M, Meynard D, et al. Iron regulation of hepcidin despite attenuated Smad1,5,8 signaling in mice without transferrin receptor 2 or Hfe. *Gastroenterology.* 2011;141(5):1907-1914.
  101. Gao J, Chen J, Kramer M, Tsukamoto H, Zhang AS, Enns CA. Interaction of the hereditary hemochromatosis protein HFE with transferrin receptor 2 is required for transferrin-induced hepcidin expression. *Cell Metab.* 2009;9(3):217-227.
  102. Wu XG, Wang Y, Wu Q, et al. HFE interacts with the BMP type I receptor ALK3 to regulate hepcidin expression. *Blood.* 2014; 124(8):1335-1343.
  103. Pietrangolo A, Caleffi A, Henrion J, et al. Juvenile hemochromatosis associated with pathogenic mutations of adult hemochromatosis genes. *Gastroenterology.* 2005; 128(2):470-479.
  104. Schmidt PJ, Fleming MD. Transgenic HFE-dependent induction of hepcidin in mice does not require transferrin receptor-2. *Am J Hematol.* 2012;87(6):588-595.
  105. Wu Q, Wang H, An P, et al. HJV and HFE play distinct roles in regulating hepcidin. *Antioxid Redox Signal.* 2015;22(15):1325-1336.
  106. Gutschow P, Schmidt PJ, Han H, et al. A competitive enzyme-linked immunosorbent assay specific for murine hepcidin-1: correlation with hepatic mRNA expression in established and novel models of dysregulated iron homeostasis. *Haematologica.* 2015;100(2):167-177.
  107. Latour C, Besson-Fournier C, Meynard D, et al. Differing impact of the deletion of hemochromatosis-associated molecules HFE and transferrin receptor-2 on the iron phenotype of mice lacking bone morphogenetic

- protein 6 or hemojuvelin. *Hepatology*. 2016; 63(1):126-137.
108. Nai A, Lidonnici MR, Rausa M, et al. The second transferrin receptor regulates red blood cell production in mice. *Blood*. 2015; 125(7):1170-1179.
  109. Wallace DF, Secondes ES, Rishi G, et al. A critical role for murine transferrin receptor 2 in erythropoiesis during iron restriction. *Br J Haematol*. 2015;168(6):891-901.
  110. Benyamin B, Ferreira MA, Willemsen G, et al. Common variants in *Tmprss6* are associated with iron status and erythrocyte volume. *Nat Genet*. 2009;41(11):1173-1175.
  111. Chambers JC, Zhang W, Li Y, et al. Genome-wide association study identifies variants in *Tmprss6* associated with hemoglobin levels. *Nat Genet*. 2009;41(11):1170-1172.
  112. Finberg KE, Whittlesey RL, Fleming MD, Andrews NC. Down-regulation of *Bmp/Smad* signaling by *Tmprss6* is required for maintenance of systemic iron homeostasis. *Blood*. 2010;115(18):3817-3826.
  113. Maxson JE, Chen J, Enns CA, Zhang AS. Matriptase-2- and proprotein convertase-cleaved forms of hemojuvelin have different roles in the down-regulation of hepcidin expression. *J Biol Chem*. 2010;285(50):39021-39028.
  114. Krijt J, Fujikura Y, Ramsay AJ, Velasco G, Nečas E. Liver hemojuvelin protein levels in mice deficient in matriptase-2 (*Tmprss6*). *Blood Cells Mol Dis*. 2011;47(2):133-137.
  115. Wahedi M, Wortham AM, Kleven MD, et al. Matriptase-2 suppresses hepcidin expression by cleaving multiple components of the hepcidin induction pathway. *J Biol Chem*. 2017;292(44):18354-18371.
  116. Finberg KE, Whittlesey RL, Andrews NC. *Tmprss6* is a genetic modifier of the Hfe-hemochromatosis phenotype in mice. *Blood*. 2011;117(17):4590-4599.
  117. Lee P, Hsu MH, Welser-Alves J, Peng H. Severe microcytic anemia but increased erythropoiesis in mice lacking Hfe or Tfr2 and *Tmprss6*. *Blood Cells Mol Dis*. 2012;48(3):173-178.
  118. Guo S, Casu C, Gardenghi S, et al. Reducing *Tmprss6* ameliorates hemochromatosis and  $\beta$ -thalassemia in mice. *J Clin Invest*. 2013; 123(4):1531-1541.
  119. Schmidt PJ, Toudjarska I, Sendamarai AK, et al. An RNAi therapeutic targeting *Tmprss6* decreases iron overload in *Hfe(-/-)* mice and ameliorates anemia and iron overload in murine  $\beta$ -thalassemia intermedia. *Blood*. 2013;121(7):1200-1208.
  120. Kautz L, Meynard D, Monnier A, et al. Iron regulates phosphorylation of *Smad1/5/8* and gene expression of *Bmp6*, *Smad7*, *Id1*, and *Atoh8* in the mouse liver. *Blood*. 2008;112(4):1503-1509.
  121. Jenkitkasemwong S, Wang CY, Coffey R, et al. *SLC39A14* is required for the development of hepatocellular iron overload in murine models of hereditary hemochromatosis. *Cell Metab*. 2015;22(1):138-150.
  122. Canali S, Wang CY, Zumbrennan-Bullough KB, et al. Endothelial cells produce BMP6 required for iron homeostasis in mice and provide a model for studying iron-mediated regulation of the BMP6-hepcidin axis [abstract]. *Am J Hematol*. 2017;92(8):e191.
  123. Lim PJ, Duare TL, Pasricha SR, et al. NRF2 controls systemic iron homeostasis via BMP6 and hepcidin and alleviates hemochromatosis [abstract]. *Am J Hematol*. 2017;92(8):e189.
  124. Bennett MJ, Lebrón JA, Bjorkman PJ. Crystal structure of the hereditary haemochromatosis protein HFE complexed with transferrin receptor. *Nature*. 2000;403(6765):46-53.
  125. Giannetti AM, Bjorkman PJ. HFE and transferrin directly compete for transferrin receptor in solution and at the cell surface. *J Biol Chem*. 2004;279(24):25866-25875.
  126. Schmidt PJ, Toran PT, Giannetti AM, Bjorkman PJ, Andrews NC. The transferrin receptor modulates Hfe-dependent regulation of hepcidin expression. *Cell Metab*. 2008;7(3):205-214.
  127. Johnson MB, Enns CA. Diferric transferrin regulates transferrin receptor 2 protein stability. *Blood*. 2004;104(13):4287-4293.
  128. Robb A, Wessling-Resnick M. Regulation of transferrin receptor 2 protein levels by transferrin. *Blood*. 2004;104(13):4294-4299.
  129. Johnson MB, Chen J, Murchison N, Green FA, Enns CA. Transferrin receptor 2: evidence for ligand-induced stabilization and redirection to a recycling pathway. *Mol Biol Cell*. 2007;18(3):743-754.
  130. Pagani A, Vieillevoeye M, Nai A, et al. Regulation of cell surface transferrin receptor-2 by iron-dependent cleavage and release of a soluble form. *Haematologica*. 2015;100(4):458-465.
  131. Zhang AS, Anderson SA, Wang J, et al. Suppression of hepatic hepcidin expression in response to acute iron deprivation is associated with an increase of matriptase-2 protein. *Blood*. 2011;117(5):1687-1699.
  132. Zhao N, Nizzi CP, Anderson SA, et al. Low intracellular iron increases the stability of matriptase-2. *J Biol Chem*. 2015;290(7):4432-4446.
  133. Frýdlová J, Prikryl P, Truksa J, et al. Effect of erythropoietin, iron deficiency and iron overload on liver matriptase-2 (*Tmprss6*) protein content in mice and rats. *PLoS One*. 2016;11(2):e0148540.
  134. Lin L, Nemeth E, Goodnough JB, Thapa DR, Gabayan V, Ganz T. Soluble hemojuvelin is released by proprotein convertase-mediated cleavage at a conserved polybasic RNRR site. *Blood Cells Mol Dis*. 2008;40(1):122-131.
  135. Silvestri L, Pagani A, Camaschella C. Furin-mediated release of soluble hemojuvelin: a new link between hypoxia and iron homeostasis. *Blood*. 2008;111(2):924-931.
  136. Kuninger D, Kuns-Hashimoto R, Nili M, Rotwein P. Pro-protein convertases control the maturation and processing of the iron-regulatory protein, *RGMc/hemojuvelin*. *BMC Biochem*. 2008;9:9.
  137. Chen W, Sun CC, Chen S, Meynard D, Babitt JL, Lin HY. A novel validated enzyme-linked immunosorbent assay to quantify soluble hemojuvelin in mouse serum. *Haematologica*. 2013;98(2):296-304.
  138. Ganz T. Iron and infection. *Int J Hematol*. 2018;107(1):7-15.
  139. Stefanova D, Raychev A, Deville J, et al. Hepcidin protects against lethal *Escherichia coli* sepsis in mice inoculated with isolates from septic patients. *Infect Immun*. 2018; 86(7):e00253-18.
  140. Nemeth E, Rivera S, Gabayan V, et al. IL-6 mediates hypoferrremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J Clin Invest*. 2004; 113(9):1271-1276.
  141. Wrighting DM, Andrews NC. Interleukin-6 induces hepcidin expression through *STAT3*. *Blood*. 2006;108(9):3204-3209.
  142. Verga Falzacappa MV, Vujic Spasic M, Kessler R, Stolte J, Hentze MW, Muckenthaler MU. *STAT3* mediates hepatic hepcidin expression and its inflammatory stimulation. *Blood*. 2007; 109(1):353-358.
  143. Pietrangelo A, Dierssen U, Valli L, et al. *STAT3* is required for IL-6-gp130-dependent activation of hepcidin in vivo. *Gastroenterology*. 2007;132(1):294-300.
  144. Kautz L, Jung G, Valore EV, Rivella S, Nemeth E, Ganz T. Identification of erythroferrone as an erythroid regulator of iron metabolism. *Nat Genet*. 2014;46(7):678-684.
  145. Kautz L, Jung G, Du X, et al. Erythroferrone contributes to hepcidin suppression and iron overload in a mouse model of  $\beta$ -thalassemia. *Blood*. 2015;126(17):2031-2037.
  146. Sonnweber T, Nachbaur D, Schroll A, et al. Hypoxia induced downregulation of hepcidin is mediated by platelet derived growth factor *BB*. *Gut*. 2014;63(12):1951-1959.
  147. Tanno T, Bhanu NV, Oneal PA, et al. High levels of *GDF15* in thalassemia suppress expression of the iron regulatory protein hepcidin. *Nat Med*. 2007;13(9):1096-1101.
  148. Mirciov CSG, Wilkins SJ, Hung GCC, Helman SL, Anderson GJ, Frazer DM. Circulating iron levels influence the regulation of hepcidin following stimulated erythropoiesis. *Haematologica*. 2018;103(10):1616-1626.
  149. Nai A, Rubio A, Campanella A, et al. Limiting hepatic *Bmp-Smad* signaling by matriptase-2 is required for erythropoietin-mediated hepcidin suppression in mice. *Blood*. 2016; 127(19):2327-2336.
  150. Krijt J, Jonášová A, Neuwirtová R, Necas E. Effect of erythropoietin on hepcidin expression in hemojuvelin-mutant mice. *Blood Cells Mol Dis*. 2010;44(4):257-261.
  151. Aschemeyer S, Gabayan V, Ganz T, Nemeth E, Kautz L. Erythroferrone and matriptase-2 independently regulate hepcidin expression. *Am J Hematol*. 2017;92(5):E61-E63.
  152. Nai A, Pagani A, Mandelli G, et al. Deletion of *Tmprss6* attenuates the phenotype in a mouse model of  $\beta$ -thalassemia. *Blood*. 2012;119(21):5021-5029.

153. Verga Falzacappa MV, Casanovas G, Hentze MW, Muckenthaler MU. A bone morphogenetic protein (BMP)-responsive element in the hepcidin promoter controls HFE2-mediated hepatic hepcidin expression and its response to IL-6 in cultured cells. *J Mol Med (Berl)*. 2008;86(5):531-540.
154. Besson-Fournier C, Latour C, Kautz L, et al. Induction of activin B by inflammatory stimuli up-regulates expression of the iron-regulatory peptide hepcidin through Smad1/5/8 signaling. *Blood*. 2012;120(2):431-439.
155. Besson-Fournier C, Gineste A, Latour C, et al. Hepcidin upregulation by inflammation is independent of Smad1/5/8 signaling by activin B. *Blood*. 2017;129(4):533-536.
156. Core AB, Canali S, Babitt JL. Hemojuvelin and bone morphogenetic protein (BMP) signaling in iron homeostasis. *Front Pharmacol*. 2014;5:104.
157. Li X, Rhee DK, Malhotra R, et al. Progesterone receptor membrane component-1 regulates hepcidin biosynthesis. *J Clin Invest*. 2016;126(1):389-401.
158. Canali S, Vecchi C, Garuti C, Montosi G, Babitt JL, Pietrangelo A. The SMAD pathway is required for hepcidin response during endoplasmic reticulum stress. *Endocrinology*. 2016;157(10):3935-3945.
159. Mastrogiannaki M, Matak P, Keith B, Simon MC, Vaulont S, Peyssonnaud C. HIF-2alpha, but not HIF-1alpha, promotes iron absorption in mice. *J Clin Invest*. 2009;119(5):1159-1166.
160. Shah YM, Matsubara T, Ito S, Yim SH, Gonzalez FJ. Intestinal hypoxia-inducible transcription factors are essential for iron absorption following iron deficiency. *Cell Metab*. 2009;9(2):152-164.
161. Taylor M, Qu A, Anderson ER, et al. Hypoxia-inducible factor-2alpha mediates the adaptive increase of intestinal ferroportin during iron deficiency in mice. *Gastroenterology*. 2011;140(7):2044-2055.
162. Anderson SA, Nizzi CP, Chang YI, et al. The IRP1-HIF-2alpha axis coordinates iron and oxygen sensing with erythropoiesis and iron absorption. *Cell Metab*. 2013;17(2):282-290.