STRATEGIES TO IMPROVE GRAFT-VERSUS-LEUKEMIA EFFECTS

The potential role of $\gamma\delta$ T cells after allogeneic HCT for leukemia

Rupert Handgretinger and Karin Schilbach

Department of Hematology/Oncology, Children's University Hospital, Tuebingen, Germany

Allogeneic hematopoetic stem cell transplantation (HCT) offers an option for patients with hematologic malignancies, in whom conventional standard therapies failed or are not effective enough to cure the disease. Successful HCT can restore functional hematopoiesis and immune function, and the new donor-derived immune system can exert a graft-versus-leukemia (GVL) effect. However, allogenic HCT can also be associated with serious risks for transplantation-related morbidities or mortalities such as graft-versus-host disease (GVHD) or life-threatening infectious complications. GVHD is caused by alloreactive T lymphocytes, which express the $\alpha\beta$ T-cell receptor, whereas lymphocytes expressing the $\gamma\delta$ T-cell receptor are not alloreactive and do not induce GVHD but can exhibit potent antileukemia and antiinfectious activities. Therefore, $\gamma\delta$ T cells are becoming

Introduction

 $\gamma\delta$ T cells are a unique and conserved population of lymphocytes that, as a first line of defense, mediate innate immunity against a wide variety of infections and play unique roles in immune surveillance and tissue homeostasis. Because of their ability to kill tumor cells of various origins, including leukemic blasts, and the fact that allogeneic T-cell receptor (TCR) $\gamma\delta$ T cells, in contrast to allogeneic TCR $\alpha\beta$ T cells, are not alloreactive and do not cause graft-versus-host disease (GVHD), $\gamma\delta$ T cells are increasingly the focus of research for adoptive immunotherapy in the context of allogeneic hematopoetic stem cell transplantation (HCT). $\gamma\delta$ T cells are part of the innate lymphocyte family and differ from conventional $\alpha\beta$ T cells by the absence of CD4 and CD8 coreceptors. Their TCR heterodimer consists of a γ chain and a δ chain, each encoded with a restricted repertoire of V, D, and J gene segments, rearranged with the help of recombination activating genes. According to their TCR δ chain usage, human $\gamma\delta$ T cells are generally divided into 2 major subsets, namely V&1 and V&2.1 Although V δ 1⁺ T cells are predominantly associated with a γ chain of the Vg I gene family comprising V γ 2/3/4/5/8, the majority of V δ 2⁺ T cells express an invariant TCR harboring Vy9.² A small subset of $V\delta 3^+$ T cells has been described, which might play a role in immunity against cytomegalovirus (CMV).³ In humans, $\gamma\delta$ T cells account for 1% to 20% of all CD3⁺ circulating T lymphocytes but increasingly interesting in allogeneic HCT, and clinical strategies to exploit the full function of these lymphocytes have been and are being developed. Such strategies comprise the in vivo activation of $\gamma\delta$ T cells or subsets after HCT by certain drugs or antibodies or the ex vivo expansion and manipulation of either patientderived or donor-derived $\gamma\delta$ T cells and their subsets and the adoptive transfer of the ex vivo-activated lymphocytes. On the basis of the absence of dysregulated alloreactivity, such approaches could induce potent GVL effects in the absence of GVHD. The introduction of large-scale clinical methods to enrich, isolate, expand, and manipulate $\gamma\delta$ T cells will facilitate future clinical studies that aim to exploit the full function of these beneficial nonalloreactive lymphocytes. (Blood. 2018; 131(10):1063-1072)

constitute the major subset of resident T cells in skin and mucosa.^{4,5} Whereas the V_Y982 TCR is expressed by 50% to 95% of the peripheral $\gamma\delta$ T cells, TCR including other V δ elements are predominantly found in the epithelia and the skin. On activation, $\gamma\delta$ T cells can respond to signals from microbes or tumors with the production of Th1, Th2, Th17, and other (inflammatory) cyto- and chemokines and interact with dendritic cells (DCs). V_Y9V δ 2 T cells exhibit a potent antigen-presenting capacity and can prime CD4⁺ and CD8⁺ T cells more efficiently than do DCs.⁶⁻⁸ In this review, we describe physiological and manipulated antileukemic cytotoxic mechanisms of $\gamma\delta$ T cells and how their antileukemic properties can be exploited posttransplant in patients with leukemia.

Antitumor cytotoxicity of $\gamma\delta$ T cells Physiological antigen recognition of $\gamma\delta$ T cells

 $\gamma\delta$ T cells can exert effective antitumor activity against various solid tumors⁹ and hematologic malignancies, such as lymphoma¹⁰ and multiple myeloma.¹¹ In contrast to $\alpha\beta$ T cells, $\gamma\delta$ T cells are not HLA-restricted for antigen recognition, and their activation does not require antigen processing and presentation by antigen-presenting cells (APCs). Therefore, $\gamma\delta$ T cells can be rapidly activated during the early phase of an immune response. Very similar to natural killer (NK) cells, they can respond to stress-induced self-ligands such as major histocompatibility complex

class I-related chain A and B (MICA/B) and UL16-binding proteins (ULBPs) via activation of the NKG2D receptor expressed on $\gamma\delta$ T cells and via the V δ 1 receptor.¹²⁻¹⁵ Some of the NKGD2 ligands can directly engage the $\gamma\delta$ TCR in V δ 2 T cells.¹⁶ In addition, $\gamma\delta$ T cells also express pattern-recognition receptors, such as toll-like receptors, which augment their antitumor activity.¹⁷ It has also been shown that $\gamma\delta$ T cells express natural cytotoxicity receptors NKp30 and NKp44 and that $\gamma\delta$ T cells can kill lymphoid leukemia cell lines and leukemic blasts from patients with chronic myeloid leukemia through NKp30.¹⁸ Finally, with the activating receptor DNAM-1, γδ T cells efficiently target nectin-2 (CD112) and poliovirus receptor (CD155) positive acute myeloid leukemia and multiple myeloma cells.^{19,20} Upregulation of Fas ligand and TRAIL through TCR activation also leads to enhanced tumor killing.²¹ yo T cells recognize a ligand repertoir heterogeneous in size, composition, and molecular structure, mainly including non-major histocompatibility complex (MHC) cell surface molecules, soluble proteins, and smaller peptides, phospholipids, prenyl pyrophosphates, and sulfatides that are bound to or expressed on the cell surface.²² Through their TCR, peripheral V₈2 T cells recognize nonpeptidic phosphorylated antigens, so-called phosphoantigens (PAgs). Some natural phosphoantigens such as (E)-4-hydroxy-3methyl-but-2-enylpyrophosphate---intermediates of the non-mevalonate isoprenoid pathway in bacteria-are strong activators of Vy9V82 T cells^{23,24}; others, such as isopentenyl pyrophophate (IPP), are produced during metabolic processes in eukaryotic cells via the mevalonate isoprenoid synthesis pathway. Although physiologic levels of IPP do not activate $\gamma\delta$ T cells, the increased metabolism of tumor cells does, by producing increased levels of PAgs, which can be recognized by $\gamma\delta$ T cells as tumor-associated antigens.²⁵

Pharmacological ways of stimulating, activating, and modifying $\gamma\delta$ T cell responses

In vivo stimulation of V δ 2 $\gamma\delta$ T cells can be achieved by using aminobisphosphonates (eg, zoledronate, pamidronate, risedronate), which as inhibitors of the farnesyl pyrophosphate synthetase (FPPS) in the mevalonate pathway provoke the accumulation of IPPs and thus activation of $\gamma\delta$ T cells in vivo. A direct correlation between bisphosphonate-induced FPPS inhibition, and thus accumulation of PAqs and tumor recognition by V γ 9V δ 2 T cells, has been shown for more than 50 tumor cell lines for zoledronate²⁶ as well as for risedronate.²⁷ A comprehensive analysis of 19 primary acute myeloid leukemia (AML) samples by Gundermann et al correlated genetic, metabolic, and immunophenotypic features with the recognition of the blasts by allogeneic $V\gamma 9V\delta 2$ T cells and found that an intrinsic susceptibility for $\gamma\delta$ T cell recognition was associated with ULBP1 expression, whereas pretreatment of nonsusceptible blasts with bisphosphonates increased their susceptibility for lysis by $\gamma\delta$ T cells.²⁸ Noteworthy, in the absence of accessory cells, nucleotide phosphoantigens as precursors of pyrophosphate antigens can deliver strong agonists intracellularly when transformed into strong pyrophosphate agonists by nucleotide pyrophosphatase activity present in serum, resulting in prolonged and strengthened activity.²⁹ Gerner-Dardenne et al showed high cytolytic potential toward AML blasts via TCR- and DNAM-1 activation for in vitro stimulated V γ 9V δ 2 T cells.¹⁹ In vitro cytotoxicity of $\gamma\delta$ T cells against acute lymphoblastic leukemia (ALL) blasts was demonstrated by Lamb et al.³⁰ It was shown in vitro in an allogeneic transplant setting that donor-derived $\gamma\delta$ T cells were activated and proliferated in response to recipient ALL blasts, whereas the $\gamma\delta$ T cells did not proliferate in a mixed lymphocyte culture with HLA-mismatched stimulator cells. In addition, the proliferating $\gamma\delta$ T cells were highly cytotoxic against myeloid and lymphoid target cells.

More recently, an emerging role of butyrophilins (BTNs) in $\gamma\delta$ T cell activation has been described³¹. BTNs are a large family of proteins and members of the extended B7 family of costimulatory molecules.³² CD277/butyrophilin 3 (BTN3A1) was identified as a crucial molecule indispensable for activation of V γ 9 T cells by PAgs.³³ In a mechanism termed "inside-out signaling," V γ 9V δ 2 TCR activation is modulated by the guanosine triphospatase activity of RhoB and its redistribution to BTN3A1. This stabilizes BTN3A1 in the membrane, and IPP accumulation induces a conformational change of BTN3A1, which leads to recognition of the extracellular domains by V γ 9V δ 2 TCRs³⁴. This mechanism is reported to be active on leukemic stem cells but not healthy CD34⁺ stem cells.³⁴

Accordingly, anti-BTN3A1 antibodies can mimic phosphoantigen stimulation and selectively activate Vy9 T cells, whereas inhibitory anti-BNT3A1 antibodies can neutralize PAg stimulation. Incubation of AML blasts with anti-BNT3A1 triggered BTN3A1 on the blasts and resulted in an enhanced Vy9V δ 2 T cell–mediated killing and could also specifically sensitize resistant blasts to Vy9V δ 2 T cell lysis. Furthermore, in a preclinical xenotransplantation model of AML, application of the agonistic anti-CD277/BTN3A1 antibody mAb 20.1 enhanced the therapeutic efficacy of adoptively transferred Vy9V δ 2 T cells.³⁵

Another key mode of antitumor activity is the activation of $\gamma\delta$ T cells via the Fc receptor FcRyIII (CD16). This Fc receptor, by binding to the Fc region of immunoglobulins (IgG), can induce tumor cell lysis via antibody-dependent cellular cytotoxity (ADCC) in the presence of antibodies that target and bind antigens expressed by the tumor.³⁶ Stimulation of $\gamma\delta$ T cells with phosphoantigens leads to upregulation of CD16 and increased ADCC,³⁷ and antibodies against CD20 or HER2/neu have been shown to enhance the cytotoxicity of $\gamma\delta$ T cells against tumor cells expressing these target antigens.³⁸ More recently, Seidel et al presented data on ADCC by freshly isolated as well as in vitro expanded $\gamma\delta$ T cells against ALL blasts in the presence of an Fc-optimized anti-CD19 antibody.³⁹ Similar results were reported by Schiller et al, who generated triple bodies ds(19-16-19).40 These triple bodies have 2 binding sites for CD19 and 1 for CD16 and are able to mediate a very rapid lysis of CD19-positive leukemic blasts mediated by primary and in vitro expanded $\gamma\delta$ T cells.

The role of $\gamma\delta$ T cells in allogeneic HCT

On the basis of the observations that $\gamma\delta$ T cells are not HLArestricted for antigen recognition, these cells might offer the opportunity for graft-versus-leukemia (GVL) effects in the absence of GVHD. Therefore, the detailed analysis of the reconstitution of $\gamma\delta$ T cells and their subsets, their in vivo activation, and the adoptive transfer of $\gamma\delta$ T cells after allogeneic transplantation are areas of increasing interest.⁴¹ Whereas earlier reconstitution studies showed increased numbers of $\gamma\delta$ T cells in patients who developed acute GVHD up to 3 months after allogeneic HCT,⁴² later studies could not corroborate these findings, and no significant correlation between $\gamma\delta$ T cell recovery and the incidence of GVHD in the first 12 months posttransplant was found by Cela et al.⁴³ In another study, a lower number of $\gamma\delta$ T cells was seen in In a larger study in patients with leukemia, who received a T-cell depleted bone marrow graft from partially HLA-mismatched donors, a significantly better disease-free survival (DSF) at 30 months posttransplant was observed in those patients in whom the percentage of $\gamma\delta$ T cells exceeded 10% of the total lymphocytes.⁴⁵ Most interestingly, there was no significant difference in the incidence of acute or chronic GVHD, suggesting an enhanced GVL effect in the absence of GVHD. In an extended 42-month follow-up study, these data were confirmed⁴⁶ and a further 8-year follow-up study with additional patients (n = 153) showed a significantly better 5-year leukemia-free survival and overall survival for patients who recovered with an increased proportion of $\gamma\delta$ T cells.⁴⁷

A more recent study on the reconstitution of $\gamma\delta$ T cells was performed with 102 pediatric patients with leukemia.48 In this single-center study at St. Jude Children's Research Hospital, the number of $\gamma\delta$ T cells was analyzed after allogeneic HCT of unmanipulated grafts from matched sibling, matched unrelated, or unrelated cord blood donor or of CD3⁺ T-cell depleted mobilized peripheral stem cell grafts from haploidentical donors. A significantly better event-free survival (EFS) and overall survival (OS) were observed in patients with elevated $\gamma\delta$ T cells within a median follow-up of 2.7 years. Interestingly, a significantly lower incidence of infections, with no bacterial infection, was seen in the patients with high levels of $\gamma\delta$ T cells, suggesting an important antibacterial⁴⁹ and antiviral function of these cells.³ The anti-CMV response of $V\delta 2^{neg} \gamma \delta T$ cells after allo-HCT especially seems of interest, because this subset expands upon CMV reactivation, and these CMV-induced $\gamma\delta$ T cells are able to recognize and also lyse primary leukemic blasts.⁵⁰ This may at least in part explain the favorable effect of CMV reactivation after HCT on the risk of relapse.⁵¹

In a manner similar to NK cells, $\gamma\delta$ T cells express inhibitory HLA class I receptors. This might play a role in tumor immunity and autoimmunity, ^{52,53} and it has been shown that V γ 9 δ 1 T cells isolated from the peripheral blood of an AML patient after HCT effectively lysed freshly isolated AML blasts and AML cell lines and that the inhibitory killer immunoglobuline-like receptor (KIR) CD158b negatively regulated the cytotoxic function of the $\gamma\delta$ T cells.⁵⁴ It is currently not clear, however, whether alloreactive and nonalloreactive $\gamma\delta$ T cells based on their KIR expression exist and whether the donor KIR phenotype or genotype plays a role in allogeneic HCT in the context of $\gamma\delta$ T cells.⁵⁵

Adoptive transfer of autologous and allogenic $\gamma\delta$ T cells

Protocols for ex vivo–manipulations of $\gamma\delta$ T cells

Various methods for the ex vivo expansion of $\gamma \delta$ T cells for autologous and allogeneic adoptive transfer have been established. Combinations of interleukin 2 (IL-2) and IPPs or aminobisphosphonates (eg, zoledronate) are established protocols for their expansion from peripheral blood mononuclear cells (PBMCs).⁵⁶ Incubation of PBMCs with zoledronate via the inhibition of the farnesyl pyrophosphate synthase leads to the accumulation of mevalonate metabolites, such as IPPs selectively in monocytes, which become APCs and stimulate V γ 9V δ 2 T cells. Therefore monocytes are required for $\gamma\delta$ T cell expansion.⁵⁷ In the absence of monocytes, IPPs or related phophoantigens are necessary. Besides IL-2, other cytokines that share the common cytokine receptor γ chain γ c are investigated, and it has been shown that IL-15 combined with IPPs enhanced the proliferation of purified $\gamma\delta$ T cells.⁵⁸ In addition, the expanded $\gamma\delta$ T cells had an activated phenotype, exerted increased cytotoxicity toward lymphoma and multiple myeloma target cells, and secreted high amounts of proinflammatory cytokines, comprising interferon- γ (IFN- γ) and tumor necrosis factor– α (TNF- α). Adding IL-15 to IL-2/zoledronate also boosted the expansion of $\gamma\delta$ T cells from PBMCs.

Although most clinical trials of adoptive transfer of ex vivo expanded $\gamma\delta$ T cells have been performed with autologous T cells and in solid tumors,⁵⁹ only 1 study so far has been performed with patients with hematologic malignancies by using allogeneic $\gamma\delta$ T cells. The successful adoptive transfer and in vivo expansion of haploidentical $\gamma\delta$ T cells was reported by Wilhelm et al in a nontransplant setting.⁶⁰ In this study, unstimulated leukapheresis products from haploidentical family donors were CD4- and CD8-depleted and the remaining $\gamma\delta$ T cells infused into patients after lymphopenia-induced chemotherapy comprising fludarabine and cyclophosphamide. The median of infused $\gamma\delta$ T cells was 2.17 \times 10⁶/kg (range, 0.9-3.84). After infusion, patients received zoledronate at day 0 and 1 \times 10 6 IU/m 2 IL-2 on day +1 through +6. A marked expansion of $\gamma\delta$ T cells (mean, 68-fold) was observed, with a peak at day 8 and persistence up to 28 days.

Despite moderate results in hitherto performed clinical studies in solid malignancies, 61 intratumoral $\gamma\delta$ T-cell signatures emerged as the most significant favorable cancer-wide prognostic populations in 25 human cancers, including lymphoma and multiple myeloma, thus pointing to a key role of these cells in cancer immune surveillance. 62

Adoptive transfer of $\gamma\delta$ T cells after allo-HCT

Another method for the adoptive transfer of allogenic $\gamma\delta$ T cells in transplant settings with a stem cell graft is the use of mobilized peripheral stem cells (PBSCs) depleted from T-cell receptor– $\alpha\beta$ (TCRαβ) positive T lymphocytes using a clinical-grade technology first described by Chaleff et al.63 The purpose of this strategy is to conserve mature NK and $\gamma\delta$ T cells, both of these lymphocyte subsets presumably being active against leukemia and viral infections but having a limited role in inducing GVHD. It was shown by Otto et al that stem cell mobilization of the donors with granulocyte colony-stimulating factor (G-CSF) does not impair the function of $\gamma\delta$ T cells and that they retain strong tumoricidal activity and produce immunomodulatory cytokines, such as IFN- γ and TNF- α .⁶⁴ Interestingly, in some of the G-CSFmobilized donors, a high proportion of $V_{\gamma}9V\delta1$ T cells was seen. This subset was associated with effective killing of fresh AML blasts after allogeneic HCT,⁵⁴ and more research is necessary to investigate the influence of G-CSF mobilization on $\gamma\delta$ T cells and their subsets.

The TCR $\alpha\beta$ depletion method was further extended by Schumm et al⁶⁵ and was performed in 102 mobilized apheresis products. The median number of $\gamma\delta$ T cells in the mobilized grafts prior to depletion was 345 \times 10⁶ (range, 16-2852) and 272 \times 10⁶ (range,

20-2339) after TCR $\alpha\beta$ depletion, with a median recovery of $\gamma\delta$ T cells of 83%. An improved immune recovery after TCR $\alpha\beta$ depleted stem cells from haploidentical donors in pediatric patients was reported by Lang et al.⁶⁶ The median number of cotransplanted $\gamma\delta$ T cells was 11 \times 10⁶/kg (range, 0.9-89.7). Recovery of donor-derived $\gamma\delta$ T cells already started at day +7 posttransplant and preceded the reconstitution of $\alpha\beta$ T cells. In the early posttransplant period, $\gamma\delta$ T cells represented the majority of CD3⁺ lymphocytes. Similar results were reported in children with AML who received TCRaβ-depleted grafts from matched unrelated donors.⁶⁷ Airoldi et al report a more detailed analysis of the $\gamma\delta$ T-cell reconstitution after haploidentical transplantation of PBSCs that were depleted of $\alpha\beta$ T cells.⁶⁸ These authors analyzed the $\gamma\delta$ T-cell reconstitution and their $V\gamma 9V\delta 2$ and $V\gamma 9V\delta 1$ subsets in 27 pediatric patients. Fifteen of the patients had acute leukemia. The median number of $\gamma\delta$ T cells coinfused with the graft was 7.85 \times 106/kg (range, 1.5-94.5). Phenotypic analysis of the reconstituting lymphocytes collected 2 to 4 weeks posttransplant also showed that the majority of the CD3⁺ T lymphocytes were of the $\gamma\delta$ lineage (mean, 91.5% of the gated CD3⁺ T cells) ranging from 70% to 100%. As has been reported already by Lang et al,⁶⁶ the $\gamma\delta$ T- cell population decreased over time, and the percentage of the $\alpha\beta$ T cells exceeded that of $\gamma\delta$ T cells in most patients after 2 months posttransplant. Interestingly, a higher number of infused $\gamma\delta$ T cells was significantly correlated with a later increase of $\alpha\beta$ T cells. A more detailed analysis of the reconstituting $\gamma\delta$ T-cell subsets mainly showed 3 subsets, namely V δ 1⁺, V δ 2⁺, and $V\delta 1^{neg}V\delta 2^{neg} \gamma \delta T$ cells. One month posttransplant, $\gamma \delta T$ cells were V δ 2⁺ (mean, 65%) and V γ 1⁺ (mean, 25%) and V δ 1^{neg}V δ 2^{neg} (9.6%). This composition was very similar to that of the $\gamma\delta$ subsets present in the mobilized stem cell donors as well as in healthy individuals. The authors then compared the $\gamma\delta$ T-cell recovery with pediatric patients who had received a CD34+-enriched haploidentical graft and found a significant difference. Three months posttransplant, the percentage of $\gamma\delta$ T cells in the CD34⁺ cohort was significantly lower, and most cells were of the V δ 1 phenotype. In those patients who experienced CMV reactivation, a significant increase of the V δ 1 subset was observed. Incubation of the $\gamma\delta$ T cells collected at various time points posttransplant with zoledronate and IL-2 led to an expansion of the $V\gamma 9V\delta 2$ T cells. Although the expanded cells showed only a low cytotoxicity toward primary leukemia cells, the lysis capability was strongly augmented by sensitizing the leukemic target cells with zoledronate. This was shown for primary myeloid blasts, T-ALL, and B-cell precursor ALL blasts.

The same group then presented very interesting data on the clinical application of zoledronate to 43 pediatric patients with ALL (n = 30) and AML (n = 13), all of whom received a haploidentical TCR $\alpha\beta$ -depleted PBSC graft.⁶⁹ Zoledronate was given every 28 days, and most patients were treated at least twice. Treatment with zoledronate was associated with an increase of V δ 2 subsets and an increase of the cytotoxicity against primary leukemic blasts. Patients given 3 or more infusions of zoledronate had a better probability of survival than did those who received only 1 or 2 treatments (86% vs 54%, respectively; P = .008). It is noteworthy that all the reported studies on haploidentical transplantation of TCR $\alpha\beta$ -depleted PBSC grafts were performed with no or only short-term pharmacological GVHD prophylaxes, which might favor a better immune recovery than those with transplantation of T-replete grafts with long-term posttransplant immunosuppression for GVHD prophylaxis.⁷⁰ Whether the rapid expansion of $\gamma\delta$ T cells early after transplantation can exclusively be explained by the expansion of the coinfused donor $\gamma\delta$ T cells is yet unclear, because Ziegler et al have shown that a small population of CD4⁺V δ 1⁺ can transdifferentiate extrathymically directly into $\alpha\beta$ T cells under inflammatory conditions.⁷¹ This small subset also expresses CD34 and is present in donor PBSCs and in TCR $\alpha\beta$ -depleted grafts. Further research is necessary to investigate whether this subset contributes to the improved $\alpha\beta$ T-cell recovery after transplantation of TCR $\alpha\beta$ -depleted grafts.

Future perspectives

The fact that $\gamma\delta$ T cells do not cause GVHD but can exert strong GVL effects and can mediate strong antiviral and antibacterial activity predisposes this cell population for cellular immunotherapeutic strategies in the context of allogeneic HCT.⁷² In vivo activation of $\gamma\delta$ T cells posttransplant by the repetitive infusion of zoledronate⁶⁹ has shown promising results in pediatric leukemia, and further clinical studies are warranted, also in combination with cytokines (eg, IL-2 and IL-15).

Because $\gamma\delta$ T cells can be obtained and expanded in vivo and ex vivo from stem cell donors as well as from patients after lymphocyte recovery,⁶⁰ various strategies for posttransplant adoptive transfer can be envisioned, depending on the purpose of the adoptive transfer. $\gamma\delta$ T cell–enriched TCR $\alpha\beta$ -depleted PBSC grafts have been used as an immunological booster to treat graft failure in 5 patients after transplantation of HLA-matched related and unrelated grafts.⁷³ Besides NK cells, such donor lymphocyte infusions (DLIs) mainly comprise $V\gamma 9V\delta 2$ T cells and lead to increased levels of white blood cells, platelets, granulocytes, or a combination of these 30 days after infusion. The number of infused $\gamma\delta$ T cells ranged from 0.2 \times 10⁶/kg to 83.5 \times 10⁶/kg, and most important, no signs of GVHD were seen. A positive effect on the infectious complications was seen in 4 of the 5 patients. Therefore, future clinical studies using $\gamma\delta$ T-cell DLIs in patients with graft failure, decreasing donor chimerism, or posttransplant infectious complications are warranted. It is currently not clear whether different subsets of $\gamma\delta$ T cells would have different effects.

It has already been reported that CMV reactivations after allogeneic HCT led to an in vivo expansion of V $\delta^{2^{neg}}\gamma\delta$ T cells, which then also cross-recognized leukemic blasts.⁵⁰ Therefore, the adoptive transfer of V $\delta^{2^{neg}}$ -donor $\gamma\delta$ T cells could be an alternative to the use of CMV-specific $\alpha\beta$ T cells,⁷⁴ because this subset is also present in CMV-naïve cord bloods⁵⁰ and in CMV negative donors. On the basis of the cross-recognition of CMV-induced V $\delta^{2^{neg}}$ T cells of leukemic blasts, this subset would also be of interest as DLIs to prevent or treat leukemic relapses. In this context, a V $\delta^{2^{neg}}$ V δ^{1+} subset is of increasing interest because of its antimalignancies activity.⁷⁵

Circulating V δ 1⁺ T cells have been associated with nonprogression of low-risk B–chronic lymphocytic leukemia (B-CLL)⁷⁶ and showed effective antitumor responses against neuroblastoma target cell lines.⁷⁷ Although some activating ligands are known, such as stress-induced self-antigens, other ligands are still to be identified. CD1d is widely recognized by V δ 1⁺ T cells by germ line residues spanning the complementarity-determining region



Figure 1. The various physiologic and manipulated antileukemic cytotoxic mechanisms of $\gamma\delta$ T cells. (1) V γ 9V δ 2 T cells recognize blasts via the CD277/IPP/rhoB axis,³⁴ and (2) CD1 ligands are recognized by V δ 1 $\gamma\delta$ T cells.⁷⁸ (3) Stress-induced self-ligands (MICA/B, ULBPs) are recognized by NKG2D¹⁵; (4) human leukocyte B-associated transcript 3 is recognized by natural cytotoxicity receptors NKp30¹⁸; (5) nectin-2 (CD112) and poliovirus receptor (CD155) are the ligands for DNAM.1^{19,20}; and (6) therapeutic monoclonal antibodies can induce antibody-dependent cellular cytotoxicity via the Fc receptor III.³⁹ (7) Bispecific antibodies can activate $\gamma\delta$ T cells via anti-V γ 9 single chain Fv.⁹⁸ and (8) $\gamma\delta$ T cells can be redirected using CARs⁴⁶ or (9) through the transduction of an exogenous $\alpha\beta$ TCR directed against a tumor-associated antigen.⁹⁹ (10) The transfer of a V γ 9V δ 2 TCR receptor III; TAA, tumor-associated antigen.

(CDR) loop of V $\delta1$, and CD1d-bound α GalCer⁷⁸ and sulfatide are separately recognized by non–germ line CDR3 δ -encoded residues. These findings provide structural demonstration of MHC-like recognition of a self-lipid by $\gamma\delta$ T cells and reveal the prevalence of lipid recognition by innate-like T-cell populations⁷⁹; they may also be useful for manipulating the number of $\gamma\delta$ T cells. CD1-bound α GalCer has already been investigated in clinical trials in patients with asymptomatic myeloma in context with invariant NKT cells⁸⁰ and might also be considered in context with $\gamma\delta$ T cells. Upon activation with IL-2, the small V δ 3 subset can also be expanded, recognize CD1d and lyse CD1d⁺ target cells, and release cytokines such as IL-17 and IFN- γ .⁸¹ Such strategies could also be envisioned posttransplant to exploit the antileukemia activity of V δ subsets.

Although most clinical ex vivo expansion strategies expand V γ 9V δ 2 T cells, various small-grade procedures, but only 1 clinical-grade method for the selective ex vivo expansion of V δ 1 cells, have been described. The latter expands V δ 1⁺ cells up to



Figure 2. Possible scenarios for how antitumor function of nonalloreactive $\gamma\delta$ T cells could be exploited after HCT. Aminobisphosphonates in combination with IL-2 or the application of therapeutic monoclonal antibodies can activate $\gamma\delta$ T cells in vivo. By using TCR $\alpha\beta$ -depleted grafts, large numbers of $\gamma\delta$ T cells are coinfused. $\gamma\delta$ T cells and their subsets either from patients or from their donors can be expanded ex vivo and manipulated prior to adoptive transfer into the patients.

2000-fold and generates Th1 cytokine but not IL-17-producing, long-lived functional V δ 1⁺ T cells, which inhibited tumor growth and prevented dissemination of CLL xenografts.82 Siegers used mitogen-stimulated $\gamma\delta$ T cells and IL-2 and IL-4.83 Upon injection into leukemia-bearing mice, Vô1⁺ cells after having been expanded ex vivo for 16 to 21 days were still viable 5 weeks after injection. A 24 000-fold expansion was achieved with PBMCs that were stimulated with Concanavalin A and then depleted for $\alpha\beta$ T cells. Lamb et al reported a 1200-fold expansion of V δ 1⁺ T cells from PBMC after depletion of $\alpha\beta$ T cells and culture with irradiated leukemia feeder cells and low levels of IL-2.³⁰ An additional approach to expand V δ 1⁺ and V δ 2⁺ T cells is the use of an anti- $\gamma\delta$ TCR antibody.⁸⁴ These expanded $\gamma\delta$ T cells showed an effective antilymphoma cytotoxicity and might be more suitable for adoptive immunotherapy of lymphoid malignancies.

Besides ex vivo expansion, $\gamma\delta$ T cells can be redirected against tumor targets using chimeric antigen receptors (CARs),⁸⁵ and peripheral blood-derived V γ 9V δ 2 T cells were transduced with retroviral vectors encoding either disialoganglioside GD2- or CD19-specific CARs.⁸⁶ These $\gamma\delta$ CAR T cells could be expanded and showed antigen-specific IFN- γ secretion and cytotoxcity against GD2⁺ neuroblastoma cells and CD19⁺ leukemic blasts. Deniger et al electroporated PBMC for the transfer of sleepingbeauty transposase and a synthetic CD19-specific T-cell receptor encoding transposon to enforce expression of CD19-specific CARs. The CARpos $\gamma\delta$ T cells were expanded on CD19⁺ artificial APCs and showed a polyclonal repertoire of endogenous $\gamma\delta$ T-cell receptors and an enhanced killing of CD19⁺ leukemic blasts in comparison with CAR^{neg} $\gamma\delta$ T cells. After adoptive transfer of the CARpos $\gamma\delta$ T cells to CD19⁺ leukemia-bearing NSG mice, the tumor burden was significantly reduced in comparison with untreated mice.87 Another approach comprises the ex vivo expansion of cytokine-induced killer cells and Vy9V82 T cells for CAR T-cell therapy.⁸⁸ PBMCs were expanded in vitro by using zoledronate, IFN- γ , IL-2, anti-CD3 antibody, and engineered K562 feeder cells. A nearly 20 000-fold expansion was achieved, with over 20% of the expanded cells being $\gamma\delta$ T cells. The expanded cells were then modified to express CD19 CARs and showed enhanced cytotoxicity in vitro as well as in vivo in a xenograft model.⁸⁸ $\gamma\delta$ T cells can also be redirected against leukemia by introducing an exogenous $\alpha\beta$ TCR of known antitumor specificity in $\gamma\delta$ T cells. 89,90 These cells retain the functionality of their original TCR and respond to stimuli via $\alpha\beta$ and $\gamma\delta$ TCR with rapid, $\gamma\delta$ -like kinetics.⁹¹ Another approach is the transduction of PAg-reactive V γ 9V δ 2 TCR into $\alpha\beta$ T cells to redirect them to cancer cells. The concomitant downregulation of the endogenous $\alpha\beta$ TCRs might abrogate alloreactive responses, and such effector cells might also play a role in the allogeneic HCT setting.92 This strategy not only overcomes diversity of different affinities of $\gamma\delta$ TCRs but also generates appropriate help through CD4-engineered cells.⁹²

Another interesting approach is the use of bispecific T-cell engager (BiTE) antibodies, which are engineered by combining 2 single chain (sc) Fv domains of 2 different antibodies. One scFv binds to effector cells, and the other scFv binds to a tumor-associated target. In ALL, the CD3-CD19 BiTE blinatumomab has shown impressive clinical efficacy in the treatment of ALL with manifest disease in children^{93,94} as well as in adults.⁹⁵ It also showed impressive efficacy in the treatment of ALL patients with

minimal residual disease.⁹⁶ Moreover, BiTEs are currently also developed for the treatment of AML and solid tumors.⁹⁷ A similar strategy has been described for $\gamma\delta$ T cells,⁹⁸ and a Her2 × V γ 9 BiTE has been generated. The construct triggered target-cell lysis by $\gamma\delta$ T cells with freshly isolated PBMC or after short-term activation. While this bispecific antibody did not induce $\gamma\delta$ T-cell proliferation and expansion, it might be more suitable for tumor cell killing than for $\gamma\delta$ T cells activated with PAgs, mainly because in contrast to PAgs activation, the Her2 × V γ 9 BiTE did not trigger activation-induced cell death, which might hamper the antitumor efficacy of PAgs-activated $\gamma\delta$ T cells. On the basis of the lack of alloreactivity of $\gamma\delta$ T cells, such bispecific antibodies (eg, anti-CD19/anti-V γ 9) could also be envisioned for the treatment of CD19⁺ ALL or other leukemias in the context of allogeneic HCT.

The various possible approaches to exploiting $\gamma\delta$ T cells as effector cells against leukemic blasts are depicted in Figure 1. In Figure 2, possible scenarios show how $\gamma\delta$ T cells can be activated directly in vivo and how patient- or donor-derived $\gamma\delta$ T cells can be expanded, manipulated, and adoptively transferred into the patient after allogeneic HCT.

Conclusions

Over the years, $\gamma\delta$ T cells have been established as important effector cells for immunotherapeutic strategies in hematologic malignancies in the context of allogeneic stem cell transplantation (allo-SCT). Many clinical trials exploiting in vivo activation or ex vivo expansion of $\gamma\delta$ T cells have been or are currently being performed,⁹⁹ and a meta-analysis of $\gamma\delta$ T-cell-based tumor immunotherapy in various tumor entities showed their efficacy and safety.¹⁰⁰ Preliminary data on the antileukemic efficacy in the allogeneic HCT setting are promising, and more clinical studies in hematological malignancies are warranted. The introduction of the clinical-grade TCR $\alpha\beta$ depletion mainly in the haploidentical setting and the coinfusion of large numbers of $\gamma\delta$ T cells together with the stem cell grafts provided valuable insights into the antileukemic and anti-infectious effects of $\gamma\delta$ T cells in allogeneic HCT.

However, it is still not clear which subset of $\gamma\delta$ T cells is more effective, and it might well be that different subsets are

REFERENCES

- Bottino C, Tambussi G, Ferrini S, et al. Two subsets of human T lymphocytes expressing gamma/delta antigen receptor are identifiable by monoclonal antibodies directed to two distinct molecular forms of the receptor. *J Exp Med.* 1988;168(2):491-505.
- Wesch D, Hinz T, Kabelitz D. Analysis of the TCR Vgamma repertoire in healthy donors and HIV-1-infected individuals. *Int Immunol.* 1998;10(8):1067-1075.
- Knight A, Madrigal AJ, Grace S, et al. The role of Vδ2-negative γδ T cells during cytomegalovirus reactivation in recipients of allogeneic stem cell transplantation. *Blood*. 2010;116(12):2164-2172.
- Vantourout P, Hayday A. Six-of-the-best: unique contributions of γδ T cells to immunology. Nat Rev Immunol. 2013;13(2): 88-100.

- Ravens S, Schultze-Florey C, Raha S, et al. Human γδ T cells are quickly reconstituted after stem-cell transplantation and show adaptive clonal expansion in response to viral infection. Nat Immunol. 2017;18(4): 393-401.
- Brandes M, Willimann K, Moser B. Professional antigen-presentation function by human gammadelta T cells. Science. 2005;309(5732):264-268.
- Moser B, Eberl M. γδ T-APCs: a novel tool for immunotherapy? *Cell Mol Life Sci.* 2011; 68(14):2443-2452.
- Brandes M, Willimann K, Bioley G, et al. Cross-presenting human gammadelta T cells induce robust CD8+ alphabeta T cell responses. Proc Natl Acad Sci USA. 2009; 106(7):2307-2312.
- 9. Legut M, Cole DK, Sewell AK. The promise of $\gamma \delta$ T cells and the $\gamma \delta$ T cell receptor for cancer

necessary for different purposes. Accumulating evidence suggests that the V δ 1 subset will play an important role in the prevention or treatment of posttransplant relapses of ALL and AML and that it might become feasible in the future to separate a potent GVL effect from GVHD.¹⁰¹

Because $\gamma\delta$ T cells do not exert dysregulated alloreactivity (but rather dampen excessive, unwanted $\alpha\beta$ T-cell responses) while simultaneously showing potent antileukemic activity, approaches to in vivo activate $\gamma\delta$ T cells directly or expand them ex vivo, either from the stem cell donor or from the patient posttransplant, might offer promising therapeutic strategies for preventing or treating relapse after HCT. However, more basic and clinical research will be necessary to fully exploit the antileukemic effect in the setting of allogeneic transplantation.

Acknowledgments

The authors thank Peter Weber for the graphic art.

This work was supported by grants from the Jürgen Manchot Foundation (K.S.), and by grants from the Stiftung für Krebskranke Kinder Tübingen e.V., the Jose-Carreras Leukemia Foundation, and the Stefan-Morsch-Stiftung (R.H.).

Authorship

Contribution: R.H. and K.S. wrote the review.

Conflict-of-interest disclosure: R.H. is a copatent holder (Germany) of the T-cell receptor $\alpha\beta$ depletion technology, and has received speaker honoraria from Miltenyibiotec. K.S. declares no competing financial interests.

Correspondence: Rupert Handgretinger, Department of Pediatric Hematology and Oncology, University Children's Hospital, Hoppe-Seyler Str 1, D-72076 Tuebingen, Germany; e-mail: rupert.handgretinger@ med.uni-tuebingen.de.

Footnote

Submitted 3 August 2017; accepted 19 October 2017. Prepublished online as *Blood* First Edition paper, 22 January 2018; DOI 10.1182/ blood-2017-08-752162.

immunotherapy. *Cell Mol Immunol*. 2015; 12(6):656-668.

- Kunzmann V, Wilhelm M. Anti-lymphoma effect of gammadelta T cells. *Leuk Lymphoma*. 2005;46(5):671-680.
- Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP, Wilhelm M. Stimulation of gammadelta T cells by aminobisphosphonates and induction of antiplasma cell activity in multiple myeloma. *Blood*. 2000;96(2):384-392.
- Groh V, Steinle A, Bauer S, Spies T. Recognition of stress-induced MHC molecules by intestinal epithelial gammadelta T cells. *Science*. 1998;279(5357):1737-1740.
- Groh V, Wu J, Yee C, Spies T. Tumourderived soluble MIC ligands impair expression of NKG2D and T-cell activation. *Nature*. 2002;419(6908):734-738.

- Wrobel P, Shojaei H, Schittek B, et al. Lysis of a broad range of epithelial tumour cells by human gamma delta T cells: involvement of NKG2D ligands and T-cell receptor- versus NKG2D-dependent recognition. Scand J Immunol. 2007;66(2-3):320-328.
- Rincon-Orozco B, Kunzmann V, Wrobel P, Kabelitz D, Steinle A, Herrmann T. Activation of V gamma 9V delta 2 T cells by NKG2D. *J Immunol.* 2005;175(4):2144-2151.
- Kong Y, Cao W, Xi X, Ma C, Cui L, He W. The NKG2D ligand ULBP4 binds to TCRgamma9/ delta2 and induces cytotoxicity to tumor cells through both TCRgammadelta and NKG2D. *Blood.* 2009;114(2):310-317.
- Shojaei H, Oberg HH, Juricke M, et al. Tolllike receptors 3 and 7 agonists enhance tumor cell lysis by human gammadelta T cells. *Cancer Res.* 2009;69(22):8710-8717.
- Correia DV, Fogli M, Hudspeth K, da Silva MG, Mavilio D, Silva-Santos B. Differentiation of human peripheral blood Vô1+ T cells expressing the natural cytotoxicity receptor NKp30 for recognition of lymphoid leukemia cells. *Blood*. 2011;118(4): 992-1001.
- Gertner-Dardenne J, Castellano R, Mamessier E, et al. Human Vγ9Vδ2 T cells specifically recognize and kill acute myeloid leukemic blasts. J Immunol. 2012;188(9): 4701-4708.
- Knight A, Mackinnon S, Lowdell MW. Human Vdelta1 gamma-delta T cells exert potent specific cytotoxicity against primary multiple myeloma cells. Cytotherapy. 2012;14(9): 1110-1118.
- Braza MS, Klein B. Anti-tumour immunotherapy with Vγ9Vδ2 T lymphocytes: from the bench to the bedside. Br J Haematol. 2013;160(2):123-132.
- Born WK, Kemal Aydintug M, O'Brien RL. Diversity of γδ T-cell antigens. Cell Mol Immunol. 2013;10(1):13-20.
- Hintz M, Reichenberg A, Altincicek B, et al. Identification of (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human gammadelta T cells in Escherichia coli. FEBS Lett. 2001;509(2):317-322.
- Altincicek B, Moll J, Campos N, et al. Cutting edge: human gamma delta T cells are activated by intermediates of the 2-Cmethyl-D-erythritol 4-phosphate pathway of isoprenoid biosynthesis. J Immunol. 2001; 166(6):3655-3658.
- Gober HJ, Kistowska M, Angman L, Jenö P, Mori L, De Libero G. Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells. J Exp Med. 2003;197(2):163-168.
- 26. Idrees AS, Sugie T, Inoue C, et al. Comparison of $\gamma\delta$ T cell responses and farnesyl diphosphate synthase inhibition in tumor cells pretreated with zoledronic acid. *Cancer Sci.* 2013;104(5):536-542.
- Benzaïd I, Mönkkönen H, Bonnelye E, Mönkkönen J, Clézardin P. In vivo phosphoantigen levels in bisphosphonatetreated human breast tumors trigger Vγ9Vδ2 T-cell antitumor cytotoxicity through ICAM-1

engagement. Clin Cancer Res. 2012;18(22): 6249-6259.

- Gundermann S, Klinker E, Kimmel B, et al. A comprehensive analysis of primary acute myeloid leukemia identifies biomarkers predicting susceptibility to human allogeneic Vγ9Vδ2 T cells. J Immunother. 2014;37(6): 321-330.
- Moulin M, Alguacil J, Gu S, et al. Vγ9Vδ2 T cell activation by strongly agonistic nucleotidic phosphoantigens. *Cell Mol Life Sci.* 2017;74(23):4353-4367.
- Lamb LS Jr, Musk P, Ye Z, et al. Human gammadelta(+) T lymphocytes have in vitro graft vs leukemia activity in the absence of an allogeneic response. *Bone Marrow Transplant*. 2001;27(6):601-606.
- Kabelitz D, Lettau M, Janssen O. Immunosurveillance by human γδ T lymphocytes: the emerging role of butyrophilins. F1000Research. 2017;6:F1000 Faculty Rev-782.
- Rhodes DA, Reith W, Trowsdale J. Regulation of immunity by butyrophilins. Annu Rev Immunol. 2016;34(1):151-172.
- Harly C, Guillaume Y, Nedellec S, et al. Key implication of CD277/butyrophilin-3 (BTN3A) in cellular stress sensing by a major human γδ T-cell subset. *Blood*. 2012;120(11): 2269-2279.
- Sebestyen Z, Scheper W, Vyborova A, et al. RhoB mediates phosphoantigen recognition by Vγ9Vδ2 T cell receptor. *Cell Reports*. 2016;15(9):1973-1985.
- Benyamine A, Le Roy A, Mamessier E, et al. BTN3A molecules considerably improve Vγ9Vδ2T cells-based immunotherapy in acute myeloid leukemia. Oncolmmunology. 2016;5(10):e1146843.
- Lafont V, Liautard J, Liautard JP, Favero J. Production of TNF-alpha by human V gamma 9V delta 2 T cells via engagement of Fc gamma RIIIA, the low affinity type 3 receptor for the Fc portion of IgG, expressed upon TCR activation by nonpeptidic antigen. J Immunol. 2001;166(12):7190-7199.
- Gertner-Dardenne J, Bonnafous C, Bezombes C, et al. Bromohydrin pyrophosphate enhances antibody-dependent cell-mediated cytotoxicity induced by therapeutic antibodies. *Blood*. 2009;113(20): 4875-4884.
- Tokuyama H, Hagi T, Mattarollo SR, et al. V gamma 9 V delta 2 T cell cytotoxicity against tumor cells is enhanced by monoclonal antibody drugs—rituximab and trastuzumab. *Int J Cancer.* 2008;122(11):2526-2534.
- Seidel UJ, Vogt F, Grosse-Hovest L, Jung G, Handgretinger R, Lang P. γδ T cell-mediated antibody-dependent cellular cytotoxicity with CD19 antibodies assessed by an impedance-based label-free real-time cytotoxicity assay. Front Immunol. 2014;5:618.
- Schiller CB, Braciak TA, Fenn NC, et al. CD19-specific triplebody SPM-1 engages NK and γδ T cells for rapid and efficient lysis of malignant B-lymphoid cells. *Oncotarget*. 2016;7(50):83392-83408.
- 41. Minculescu L, Sengeløv H. The role of gamma delta T cells in haematopoietic stem

cell transplantation. *Scand J Immunol*. 2015; 81(6):459-468.

- Viale M, Ferrini S, Bacigalupo A. TCR gamma/delta positive lymphocytes after allogeneic bone marrow transplantation. Bone Marrow Transplant. 1992;10(3):249-253.
- Cela ME, Holladay MS, Rooney CM, et al. Gamma delta T lymphocyte regeneration after T lymphocyte-depleted bone marrow transplantation from mismatched family members or matched unrelated donors. Bone Marrow Transplant. 1996;17(2): 243-247.
- Yabe M, Yabe H, Hattori K, et al. Transition of T cell receptor gamma/delta expressing double negative (CD4–/CD8–) lymphocytes after allogeneic bone marrow transplantation. Bone Marrow Transplant. 1994; 14(5):741-746.
- Lamb LS Jr, Henslee-Downey PJ, Parrish RS, et al. Increased frequency of TCR gamma delta + T cells in disease-free survivors following T cell-depleted, partially mismatched, related donor bone marrow transplantation for leukemia. J Hematother. 1996;5(5):503-509.
- Lamb LS Jr, Gee AP, Hazlett LJ, et al. Influence of T cell depletion method on circulating gammadelta T cell reconstitution and potential role in the graft-versus-leukemia effect. *Cytotherapy*. 1999;1(1):7-19.
- Godder KT, Henslee-Downey PJ, Mehta J, et al. Long term disease-free survival in acute leukemia patients recovering with increased gammadelta T cells after partially mismatched related donor bone marrow transplantation. *Bone Marrow Transplant.* 2007; 39(12):751-757.
- Perko R, Kang G, Sunkara A, Leung W, Thomas PG, Dallas MH. Gamma delta T cell reconstitution is associated with fewer infections and improved event-free survival after hematopoietic stem cell transplantation for pediatric leukemia. *Biol Blood Marrow Transplant.* 2015;21(1):130-136.
- Zheng J, Liu Y, Lau YL, Tu W. γδ-T cells: an unpolished sword in human anti-infection immunity. Cell Mol Immunol. 2013;10(1):50-57.
- 50. Scheper W, van Dorp S, Kersting S, et al. $\gamma\delta T$ cells elicited by CMV reactivation after allo-SCT cross-recognize CMV and leukemia. *Leukemia.* 2013;27(6):1328-1338.
- Elmaagacli AH, Steckel NK, Koldehoff M, et al. Early human cytomegalovirus replication after transplantation is associated with a decreased relapse risk: evidence for a putative virus-versus-leukemia effect in acute myeloid leukemia patients. *Blood.* 2011; 118(5):1402-1412.
- Handgretinger R, Geiselhart A, Moris A, et al. Pure red-cell aplasia associated with clonal expansion of granular lymphocytes expressing killer-cell inhibitory receptors. N Engl J Med. 1999;340(4):278-284.
- Fisch P, Moris A, Rammensee HG, Handgretinger R. Inhibitory MHC class I receptors on gammadelta T cells in tumour immunity and autoimmunity. *Immunol Today*. 2000;21(4):187-191.

- 54. Dolstra H, Fredrix H, van der Meer A, de Witte T, Figdor C, van de Wiel-van Kemenade E. TCR gamma delta cytotoxic T lymphocytes expressing the killer cell-inhibitory receptor p58.2 (CD158b) selectively lyse acute myeloid leukemia cells. Bone Marrow Transplant. 2001;27(10):1087-1093.
- Handgretinger R, Lang P, André MC. Exploitation of natural killer cells for the treatment of acute leukemia. *Blood.* 2016; 127(26):3341-3349.
- Nicol AJ, Tokuyama H, Mattarollo SR, et al. Clinical evaluation of autologous gamma delta T cell-based immunotherapy for metastatic solid tumours. *Br J Cancer.* 2011; 105(6):778-786.
- Roelofs AJ, Jauhiainen M, Mönkkönen H, Rogers MJ, Mönkkönen J, Thompson K. Peripheral blood monocytes are responsible for gammadelta T cell activation induced by zoledronic acid through accumulation of IPP/ DMAPP. Br J Haematol. 2009;144(2): 245-250.
- Van Acker HH, Anguille S, Willemen Y, et al. Interleukin-15 enhances the proliferation, stimulatory phenotype, and antitumor effector functions of human gamma delta T cells. J Hematol Oncol. 2016;9(1):101.
- Zou C, Zhao P, Xiao Z, Han X, Fu F, Fu L. γδ T cells in cancer immunotherapy. Oncotarget. 2017;8(5):8900-8909.
- Wilhelm M, Smetak M, Schaefer-Eckart K, et al. Successful adoptive transfer and in vivo expansion of haploidentical γδ T cells. J Transl Med. 2014;12(1):45.
- Deniger DC, Moyes JS, Cooper LJ. Clinical applications of gamma delta T cells with multivalent immunity. *Front Immunol.* 2014; 5:636.
- 62. Gentles AJ, Newman AM, Liu CL, et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat Med.* 2015;21(8):938-945.
- Chaleff S, Otto M, Barfield RC, et al. A largescale method for the selective depletion of alphabeta T lymphocytes from PBSC for allogeneic transplantation. *Cytotherapy*. 2007; 9(8):746-754.
- Otto M, Barfield RC, Iyengar R, et al. Human gammadelta T cells from G-CSF-mobilized donors retain strong tumoricidal activity and produce immunomodulatory cytokines after clinical-scale isolation. *J Immunother*. 2005; 28(1):73-78.
- Schumm M, Lang P, Bethge W, et al. Depletion of T-cell receptor alpha/beta and CD19 positive cells from apheresis products with the CliniMACS device. *Cytotherapy*. 2013;15(10):1253-1258.
- Lang P, Feuchtinger T, Teltschik HM, et al. Improved immune recovery after transplantation of TCRαβ/CD19-depleted allografts from haploidentical donors in pediatric patients. Bone Marrow Transplant. 2015; 50(Suppl 2):S6-S10.
- 67. Maschan M, Shelikhova L, Ilushina M, et al. TCR-alpha/beta and CD19 depletion and treosulfan-based conditioning regimen in unrelated and haploidentical transplantation in children with acute myeloid leukemia.

Bone Marrow Transplant. 2016;51(5): 668-674.

- Airoldi I, Bertaina A, Prigione I, et al. γδ T-cell reconstitution after HLA-haploidentical hematopoietic transplantation depleted of TCR-αβ+/CD19+ lymphocytes. *Blood*. 2015;125(15):2349-2358.
- 69. Bertaina A, Zorzoli A, Petretto A, et al. Zoledronic acid boosts $\gamma\delta$ T-cell activity in children receiving $\alpha\beta$ + T and CD19+ celldepleted grafts from an HLA-haplo-identical donor. *Oncolmmunology*. 2016;6(2): e1216291.
- Lowe EJ, Turner V, Handgretinger R, et al. T-cell alloreactivity dominates natural killer cell alloreactivity in minimally T-celldepleted HLA-non-identical paediatric bone marrow transplantation. Br J Haematol. 2003;123(2):323-326.
- 71. Ziegler H, Welker C, Sterk M, et al. Human peripheral CD4(+) V δ 1(+) $\gamma\delta$ T cells can develop into $\alpha\beta$ T cells. Front Immunol. 2014;5: 645.
- Hu Y, Cui Q, Luo C, Luo Y, Shi J, Huang H. A promising sword of tomorrow: human γδ T cell strategies reconcile allo-HSCT complications. *Blood Rev.* 2016;30(3):179-188.
- Radestad E, Wikell H, Engstrom M, et al. Alpha/beta T-cell depleted grafts as an immunological booster to treat graft failure after hematopoietic stem cell transplantation with HLA-matched related and unrelated donors. J Immunol Res. 2014;2014:578741.
- Feuchtinger T, Opherk K, Bethge WA, et al. Adoptive transfer of pp65-specific T cells for the treatment of chemorefractory cytomegalovirus disease or reactivation after haploidentical and matched unrelated stem cell transplantation. *Blood.* 2010;116(20): 4360-4367.
- 75. Siegers GM, Lamb LS Jr. Cytotoxic and regulatory properties of circulating $V\delta 1 + \gamma \delta$ T cells: a new player on the cell therapy field? *Mol Ther.* 2014;22(8):1416-1422.
- Poggi A, Venturino C, Catellani S, et al. Vdelta1 T lymphocytes from B-CLL patients recognize ULBP3 expressed on leukemic B cells and up-regulated by trans-retinoic acid. *Cancer Res.* 2004;64(24):9172-9179.
- Schilbach K, Frommer K, Meier S, Handgretinger R, Eyrich M. Immune response of human propagated gammadelta-T-cells to neuroblastoma recommend the Vdelta1+ subset for gammadelta-T-cellbased immunotherapy. *J Immunother*. 2008; 31(9):896-905.
- Uldrich AP, Le Nours J, Pellicci DG, et al. CD1d-lipid antigen recognition by the γδ TCR. Nat Immunol. 2013;14(11):1137-1145.
- Luoma AM, Castro CD, Adams EJ. γδ T cell surveillance via CD1 molecules. Trends Immunol. 2014;35(12):613-621.
- Richter J, Neparidze N, Zhang L, et al. Clinical regressions and broad immune activation following combination therapy targeting human NKT cells in myeloma. *Blood*. 2013;121(3):423-430.
- 81. Mangan BA, Dunne MR, O'Reilly VP, et al. Cutting edge: CD1d restriction and Th1/

Th2/Th17 cytokine secretion by human Vδ3 T cells. *J Immunol*. 2013;191(1):30-34.

- Almeida AR, Correia DV, Fernandes-Platzgummer A, et al. Delta one T cells for immunotherapy of chronic lymphocytic leukemia: clinical-grade expansion/ differentiation and preclinical proof of concept. *Clin Cancer Res.* 2016;22(23): 5795-5804.
- Siegers GM, Dhamko H, Wang XH, et al. Human Vδ1 γδ T cells expanded from peripheral blood exhibit specific cytotoxicity against B-cell chronic lymphocytic leukemiaderived cells. Cytotherapy. 2011;13(6): 753-764.
- Zhou J, Kang N, Cui L, Ba D, He W. Anti-γδ TCR antibody-expanded γδ T cells: a better choice for the adoptive immunotherapy of lymphoid malignancies. *Cell Mol Immunol.* 2012;9(1):34-44.
- Mirzaei HR, Mirzaei H, Lee SY, Hadjati J, Till BG. Prospects for chimeric antigen receptor (CAR) γδ T cells: a potential game changer for adoptive T cell cancer immunotherapy. *Cancer Lett.* 2016;380(2):413-423.
- Rischer M, Pscherer S, Duwe S, Vormoor J, Jürgens H, Rossig C. Human gammadelta T cells as mediators of chimaeric-receptor redirected anti-tumour immunity. *Br J Haematol.* 2004;126(4):583-592.
- Deniger DC, Switzer K, Mi T, et al. Bispecific T-cells expressing polyclonal repertoire of endogenous γδ T-cell receptors and introduced CD19-specific chimeric antigen receptor. *Mol Ther.* 2013;21(3):638-647.
- Du SH, Li Z, Chen C, et al. Co-expansion of cytokine-induced killer cells and Vγ9Vδ2 T cells for CAR T-cell therapy. *PLoS One*. 2016;11(9):e0161820.
- van der Veken LT, Hagedoorn RS, van Loenen MM, Willemze R, Falkenburg JH, Heemskerk MH. Alphabeta T-cell receptor engineered gammadelta T cells mediate effective antileukemic reactivity. *Cancer Res.* 2006;66(6):3331-3337.
- van der Veken LT, Coccoris M, Swart E, Falkenburg JH, Schumacher TN, Heemskerk MH. Alpha beta T cell receptor transfer to gamma delta T cells generates functional effector cells without mixed TCR dimers in vivo. J Immunol. 2009;182(1):164-170.
- Hiasa A, Nishikawa H, Hirayama M, et al. Rapid alphabeta TCR-mediated responses in gammadelta T cells transduced with cancerspecific TCR genes. *Gene Ther.* 2009;16(5): 620-628.
- Marcu-Malina V, Heijhuurs S, van Buuren M, et al. Redirecting αβ T cells against cancer cells by transfer of a broadly tumor-reactive γδT-cell receptor. *Blood.* 2011;118(1):50-59.
- Handgretinger R, Zugmaier G, Henze G, Kreyenberg H, Lang P, von Stackelberg A. Complete remission after blinatumomabinduced donor T-cell activation in three pediatric patients with post-transplant relapsed acute lymphoblastic leukemia. *Leukemia*. 2011;25(1):181-184.
- 94. von Stackelberg A, Locatelli F, Zugmaier G, et al. Phase I/phase II study of blinatumomab in pediatric patients with relapsed/refractory

acute lymphoblastic leukemia. *J Clin Oncol.* 2016;34(36):4381-4389.

- 95. Topp MS, Gökbuget N, Zugmaier G, et al. Phase II trial of the anti-CD19 bispecific T cell-engager blinatumomab shows hematologic and molecular remissions in patients with relapsed or refractory B-precursor acute lymphoblastic leukemia. J Clin Oncol. 2014;32(36): 4134-4140.
- Topp MS, Kufer P, Gökbuget N, et al. Targeted therapy with the T-cellengaging antibody blinatumomab of chemotherapy-refractory minimal

residual disease in B-lineage acute lymphoblastic leukemia patients results in high response rate and prolonged leukemia-free survival. *J Clin Oncol.* 2011;29(18):2493-2498.

- Klinger M, Benjamin J, Kischel R, Stienen S, Zugmaier G. Harnessing T cells to fight cancer with BiTE® antibody constructs-past developments and future directions. *Immunol Rev.* 2016;270(1): 193-208.
- Oberg HH, Kellner C, Gonnermann D, et al. γδ T cell activation by bispecific antibodies. *Cell Immunol.* 2015;296(1):41-49.
- Fisher JP, Heuijerjans J, Yan M, Gustafsson K, Anderson J. γδ T cells for cancer immunotherapy: a systematic review of clinical trials. Oncolmmunology. 2014;3(1):e27572.
- 100. Buccheri S, Guggino G, Caccamo N, Li Donni P, Dieli F. Efficacy and safety of γδT cell-based tumor immunotherapy: a meta-analysis. J Biol Regul Homeost Agents. 2014;28(1):81-90.
- 101. Fisher J, Kramer AM, Gustafsson K, Anderson J. Non-V delta 2 gamma delta T lymphocytes as effectors of cancer immunotherapy. Oncolmmunology. 2014; 4(3):e973808.