### Letters to Blood



Check for updates

### To the editor:

# Genome-wide association study to identify variants associated with acute severe vaso-occlusive pain in sickle cell anemia

Shruti Chaturvedi,<sup>1</sup> Pallav Bhatnagar,<sup>2</sup> Christopher J. Bean,<sup>3</sup> Martin H. Steinberg,<sup>4-6</sup> Jaqueline N. Milton,<sup>7</sup> James F. Casella,<sup>8</sup> Emily Barron-Casella,<sup>8</sup> Dan E. Arking,<sup>2</sup> and Michael R. DeBaun<sup>9</sup>

<sup>1</sup>Division of Hematology and Oncology, Department of Medicine, Vanderbilt University Medical Center, Nashville, TN; <sup>2</sup>McKusick-Nathans Institute of Genetic Medicine, School of Medicine, Johns Hopkins University, Baltimore, MD; <sup>3</sup>Division of Blood Disorders, National Center on Birth Defects and Developmental Disabilities, Centers for Disease Control and Prevention, Atlanta, GA; <sup>4</sup>Department of Medicine, <sup>5</sup>Department of Pediatrics, and <sup>6</sup>Department of Pathology and Laboratory Medicine, Boston University School of Medicine, Boston, MA; <sup>7</sup>Department of Biostatistics, Boston University School of Public Health, Boston, MA; <sup>8</sup>Division of Pediatric Hematology, Department of Pediatrics, School of Medicine, Johns Hopkins University, Baltimore, MD; and <sup>9</sup>Department of Pediatrics, Vanderbilt University School of Medicine, Nashville, TN

Acute vaso-occlusive pain episodes are a hallmark of sickle cell anemia (SCA), one of the most common Mendelian disorders worldwide with an estimated >300 000 births annually. 1-3 Although SCA is a monogenic disorder, manifestations and disease severity are highly variable, suggesting additional phenotypic modifiers. The few genetic factors known to act as phenotypic modifiers do not completely explain the clinical heterogeneity in SCA. Previous genetic association studies identified that variants at 3 distinct loci (BCL11A, HBS1L-MYB, and HBB) are strong determinants of fetal hemoglobin level, and the singlenucleotide polymorphism (SNP) variant rs6141803 located upstream of COMMD7 is associated with acute chest syndrome.<sup>4</sup> Also, heme oxygenase-1 gene promoter polymorphisms influence heme oxygenase (HO-1) activity and the incidence of acute chest syndrome in children with sickle cell disease (SCD).<sup>5,6</sup> We conducted this genome-wide association study (GWAS) to identify the variants associated with acute, severe vaso-occlusive pain in children with SCA enrolled in the Cooperative Study for Sickle Cell Disease (CSSCD) and Silent Infarct Transfusion (SIT) trial.

The CSSCD, a multi-institutional prospective cohort, natural history study of SCD, enrolled 3538 individuals with SCD between 1979 and 1981. The SIT trial, a multicenter international trial, screened 1210 children with SCA to test the hypothesis that regular blood transfusions attenuate progression of cerebral infarcts in children with preexisting silent strokes. Both studies were approved by the Institutional Review Boards at Boston University School of Medicine and Vanderbilt University Medical Center.

We included participants identified as being of African descent from both cohorts with available genotype data, and who were diagnosed with SCA. We excluded participants if essential clinical or demographic data (necessary for phenotypic assignment or previously reported to impact the pain phenotype) were missing, or if there was discordance between genetically defined and self-identified sex. We excluded all self-reported first-degree relatives, and cryptic relatedness (including full siblings, parents, and offspring) determined by examining pairwise identity-by-descent in the combined cohort. To harmonize pain phenotypes in the CSSCD and SIT trial cohorts, the age inclusion criterion of 2 to 18 years was used to match the age and length of follow-up in both cohorts. SIT trial participants were between 5 and 15 years of age at the time of registration and the trial included a 3-year retrospective collection of all acute, severe vaso-occlusive pain based on hospitalization and treatment with opioid medication.<sup>8,9</sup> Unlike previous CSSCD pain analyses, where the definition of a pain episode included an acute vaso-occlusive event that lasted at least 2 hours and resulted in a physician visit, we restricted the definition of a pain episode to include only episodes requiring hospitalization to match the SIT trial definition.

CSSCD cohort DNA samples were genotyped at Boston University School of Medicine by using Illumina Human610-Quad arrays (n = 610 000 SNPs) (Illumina, San Diego, CA) and BeadStudio was used to call genotypes. SIT trial samples were genotyped at the Center for Inherited Disease Research at Johns Hopkins University School of Medicine (N = 573) by using the Illumina HumanHap650Y array  $(n = 661\ 000\ SNPs)$  (Illumina) or at the Centers for Disease Control and Prevention (Atlanta, GA) (N = 509) by using the Illumina Infinium HumanOmni1-Quad array (n = 1 134 514 SNPs) (Illumina). After detailed quality control procedures were completed (and excluding CSSCD samples outside the SIT trial age inclusion criteria [≥2 and ≤12 years of age]), 359 and 934 samples from the CSSCD and SIT cohorts, respectively, were included in the analysis (Table 1; supplemental Figure 1A-B; and supplemental Methods, available on the Blood Web site). To infer ungenotyped SNPs and fill in missing data across genotyping platforms in the SIT trial and CSSCD cohort, we merged HumanHap650Y, HumanOmni1-Quad, and Human610-Quad array data sets and performed imputation for autosomal markers by using a Hidden Markov model as implemented in Markov Chain Haplotyping algorithm (MaCH), version 1.16, 10 with 50 rounds and 200 states. Quality control was performed both before and after imputation and poorly imputed SNPs (squared correlation between imputed and true genotypes < 0.3) were excluded; a total of 1 098 907 SNPs remained for analysis. Due to the observed overdispersion of pain episodes in both cohorts, a multivariate quasi-Poisson regression model, with correction for estimated overdispersion, was used to evaluate the possible associations between SNPs and the acute severe vaso-occlusive pain rate, which was treated as a quantitative trait. The model was adjusted for age at enrollment, sex, hematocrit, and the top 10 principal components from the genetic data (to account for population substructure and genetic heterogeneity), assuming additive effects of allele dosage on the acute vaso-occlusive pain rate.

Participant characteristics for the SIT trial and CSSCD cohort are shown in Table 1. Statistically significant, but not clinically relevant, differences were identified between the 2 cohorts in age, percentage of fetal hemoglobin, reticulocyte percentage, pain rate, and follow-up time. The Manhattan plot summarizing the results of the GWAS for acute vaso-occlusive pain in the SIT trial and CSSCD cohort for

Table 1. Summary of SIT and CSSCD cohort demographics and clinical characteristics

Characteristic	CSSCD (n = 349)*	SIT trial (n = 934)†	<b>P</b> ‡
Male sex, n (%)	195 (54.3)	489 (52.3)	.53
Age, mean ± SD, y	6.92 ± 2.78	8.96 ± 2.44	<.001
Follow-up, mean ± SD, y	2.8 ± 0.67	$3.00 \pm 0.0$	<.001
Acute, severe vaso-occlusive pain rate (events per patient year)§	0.58 ± 1.02	0.61 ± 0.83	.01
ACS rate (events per patient year)	0.18 ± 0.41	0.13 ± 0.26	.14
Hematocrit, mean ± SD, %	23.10 ± 2.81	$23.35 \pm 3.43$	.47
Hemoglobin, mean ± SD, g/dL	7.98 ± 0.88	8.12 ± 1.08	.15
Fetal hemoglobin, mean ± SD, %	7.59 ± 4.87	8.94 ± 5.75	.001
Reticulocytes, mean ± SD, %	13.99 ± 5.62	12.02 ± 5.48	<.001
White blood cells, $\times 10^9 \text{/L}$	12.38 ± 2.71	12.58 ± 5.26	.68

ACS, acute chest syndrome.

§Acute, severe vaso-occlusive pain was defined as an acute episode of pain requiring hospitalization and treatment with opioids.

the additive model is shown in Figure 1. The genomic inflation  $\lambda$  coefficient was 1.079, suggesting minimal test-statistic inflation by potential population stratification, cryptic relatedness, or other technical factors. Although none of the SNPs were significant at  $P < 5.0 \times 10^{-8}$ , 1 novel locus approached genome-wide significance: SNP rs3115229 ( $P = 5.63 \times 10^{-8}$ ). This SNP is located 63.7 kb 5' upstream of the *KIAA1109* gene on chromosome 4 (4q27).

The suggested locus includes the *KIAA1109-TENR-IL2-IL21* linkage disequilibrium block, containing 3 known protein-coding genes, *TENR*, *IL2*, and *IL21*, and a predicted gene of unknown function, *KIAA1109*. This locus has been associated with autoinflammatory disorders, such as celiac disease, <sup>11,12</sup> ulcerative colitis, <sup>13,14</sup> and rheumatoid arthritis. <sup>15,16</sup>

Given the nature of GWAS studies, namely associations between a SNP and a phenotype, we can only postulate as to the potential role of this locus in the pathogenesis of acute vaso-occlusive pain, a complex phenomenon involving tissue ischemia, hypoxia-reperfusion injury, immune responses and inflammation, <sup>17,18</sup> and interactions between red blood cells, the endothelium, and leukocytes regulated by T-cell cytokines and adhesion molecules. <sup>19,20</sup> Interleukin 2 (IL-2) and IL-21 may modulate acute pain in SCD through their effects on inflammation

and immune responses. IL-2 is a key cytokine for T-cell activation and proliferation.<sup>21</sup> IL-21 enhances B, T, and natural killer cell proliferation and interferon- $\gamma$  production; inhibiting IL-21 has been shown to dampen inflammatory responses.<sup>22,23</sup> T lymphocytes have also been implicated as mediators of pain hypersensitivity.<sup>24</sup> *KIAA1109* is moderately expressed in all adult and fetal tissues and encodes a protein of unknown function.<sup>25</sup> *TENR* encodes testis nuclear RNA-binding protein, expressed primarily in the testis.

Strengths of the study include the consistent definition of acute, severe vaso-occlusive pain requiring hospitalization, the relatively large non–hydroxyurea treated population (a potential confounder) from 2 independent cohorts of children not on disease modifying therapy with hydroxyurea, with fewer comorbidities, and lower rates of chronic pain than adults with SCA. Pooling the cohorts improved power for our discovery analysis, but precluded validation in a separate cohort.

In summary, we present preliminary evidence of an association between variant rs3115229 and acute, severe vaso-occlusive pain in children with SCA. Our results will require additional validation and functional studies to understand the biology and reveal mechanisms by which candidate SNPs/genes might have their effects.

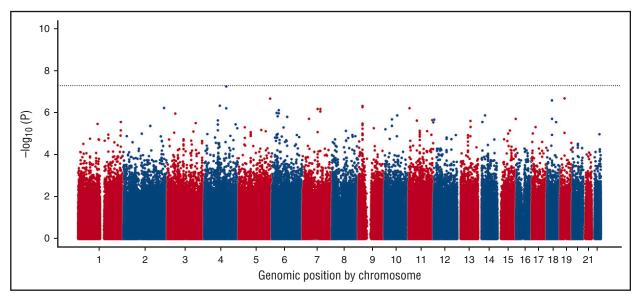


Figure 1. Manhattan plot showing the genome-wide  $-\log_{10} P$  values for association of SNPs with vaso-occlusive pain. Only 1 SNP on chromosome 4 (rs3115229) approached genome-wide significance ( $P = 5.63 \times 10^{-8}$ ).

<sup>\*</sup>Age range at enrollment was 2 to 12 years; pain/ACS events were defined as those occurring 3 years prospectively from date of enrollment; therefore, the age range in which they developed pain is 2 to 15 years.

<sup>†</sup>Age range was 2 to 15 years at enrollment; pain/ACS events were defined as those occurring within 3 years retrospectively (from the date of enrollement); the youngest patient included was 5 years of age.

 $<sup>\</sup>ddagger P$  values for continuous and categorical variables are based on Wilcoxon rank-sum test (with continuity correction) and Pearson's  $\chi^2$  test, respectively.

The online version of this article contains a data supplement.

**Acknowledgments:** The authors thank the families and children with SCD who were participants in the SIT trial and CSSCD.

This work was supported by the National Institutes of Health (NIH) National Heart, Lung, and Blood Institute (U54HL090515, 4U01HL117721, 5R01HL091759, R01 HL87681 [M.H.S.], R01 HL 068970 [M.H.S.], and T32 HL007501 [J.N.M.]), and the NIH National Institute of Neurological Disorders and Stroke (5U01-NS042804-03); the Burroughs Wellcome Foundation (M.R.D.); an American Society of Hematology Research Training Award for Fellows (S.C.); and the Jim and Carol O'Hare Fellowship (S.C.).

The funders had no role in study design, data collection and analysis, preparation of the manuscript, or decision to publish. The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the Centers for Disease Control and Prevention.

Contribution: M.R.D. designed the research and wrote portions of the manuscript; P.B. performed analysis and interpreted the data; S.C. interpreted the data and wrote the manuscript; C.J.B., J.N.M., D.E.A., E.B.-C., J.F.C., and M.H.S. designed the study, collected data, interpreted the analysis, and wrote the manuscript; and all authors read and approved the final draft of the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests

Correspondence: Michael R. DeBaun, Division of Hematology/Oncology, Vanderbilt-Meharry Center in Sickle Cell Disease, 2200 Children's Way, Suite 11206, Nashville, TN 37232; e-mail: m.debaun@vanderbilt.edu.

#### References

- Weatherall DJ. The inherited diseases of hemoglobin are an emerging global health burden. Blood. 2010;115(22):4331-4336.
- Rees DC, Williams TN, Gladwin MT. Sickle-cell disease. Lancet. 2010; 376(9757):2018-2031.
- Vichinsky EP, Styles LA, Colangelo LH, Wright EC, Castro O, Nickerson B; Cooperative Study of Sickle Cell Disease. Acute chest syndrome in sickle cell disease: clinical presentation and course. *Blood*. 1997;89(5):1787-1792.
- Galarneau G, Coady S, Garrett ME, et al. Gene-centric association study of acute chest syndrome and painful crisis in sickle cell disease patients. *Blood*. 2013;122(3):434-442.
- Exner M, Minar E, Wagner O, Schillinger M. The role of heme oxygenase-1 promoter polymorphisms in human disease. Free Radic Biol Med. 2004;37(8):1097-1104.
- Bean CJ, Boulet SL, Ellingsen D, et al. Heme oxygenase-1 gene promoter polymorphism is associated with reduced incidence of acute chest syndrome among children with sickle cell disease. *Blood*. 2012;120(18):3822-3828.
- Gaston M, Rosse WF. The cooperative study of sickle cell disease: review of study design and objectives. Am J Pediatr Hematol Oncol. 1982;4(2):197-201.
- DeBaun MR, Gordon M, McKinstry RC, et al. Controlled trial of transfusions for silent cerebral infarcts in sickle cell anemia. N Engl J Med. 2014;371(8):699-710.

- Platt OS, Thorington BD, Brambilla DJ, et al. Pain in sickle cell disease. Rates and risk factors. N Engl J Med. 1991;325(1):11-16.
- Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR. MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genet Epidemiol*. 2010;34(8):816-834.
- van Heel DA, Franke L, Hunt KA, et al. A genome-wide association study for celiac disease identifies risk variants in the region harboring IL2 and IL21. Nat Genet. 2007;39(7):827-829.
- Adamovic S, Amundsen SS, Lie BA, et al. Association study of IL2/IL21 and FcgRlla: significant association with the IL2/IL21 region in Scandinavian coeliac disease families. Genes Immun. 2008;9(4):364-367.
- Glas J, Stallhofer J, Ripke S, et al. Novel genetic risk markers for ulcerative colitis in the IL2/IL21 region are in epistasis with IL23R and suggest a common genetic background for ulcerative colitis and celiac disease. Am J Gastroenterol. 2009;104(7):1737-1744.
- Festen EA, Goyette P, Scott R, et al. Genetic variants in the region harbouring IL2/IL21 associated with ulcerative colitis. Gut. 2009;58(6):799-804.
- Hollis-Moffatt JE, Chen-Xu M, Topless R, et al. Only one independent genetic association with rheumatoid arthritis within the KIAA1109-TENR-IL2-IL21 locus in Caucasian sample sets: confirmation of association of rs6822844 with rheumatoid arthritis at a genome-wide level of significance. Arthritis Res Ther. 2010;12(3):R116.
- Teixeira VH, Pierlot C, Migliorini P, et al; European Consortium on Rheumatoid Arthritis Families. Testing for the association of the KIAA1109/Tenr/IL2/IL21 gene region with rheumatoid arthritis in a European family-based study. Arthritis Res Ther. 2009;11(2):R45.
- Kaul DK, Hebbel RP. Hypoxia/reoxygenation causes inflammatory response in transgenic sickle mice but not in normal mice. J Clin Invest. 2000;106(3):411-420.
- Cain DM, Vang D, Simone DA, Hebbel RP, Gupta K. Mouse models for studying pain in sickle disease: effects of strain, age, and acuteness. Br J Haematol. 2012;156(4):535-544.
- Duits AJ, Schnog JB, Lard LR, Saleh AW, Rojer RA. Elevated IL-8 levels during sickle cell crisis. Eur J Haematol. 1998;61(5):302-305.
- Graido-Gonzalez E, Doherty JC, Bergreen EW, Organ G, Telfer M, McMillen MA. Plasma endothelin-1, cytokine, and prostaglandin E2 levels in sickle cell disease and acute vaso-occlusive sickle crisis. *Blood*. 1998;92(7):2551-2555.
- 21. Hoyer KK, Dooms H, Barron L, Abbas AK. Interleukin-2 in the development and control of inflammatory disease. *Immunol Rev.* 2008;226:19-28.
- Young DA, Hegen M, Ma HL, et al. Blockade of the interleukin-21/interleukin-21
  receptor pathway ameliorates disease in animal models of rheumatoid arthritis.
   Arthritis Rheum. 2007;56(4):1152-1163.
- Di Fusco D, Izzo R, Figliuzzi MM, Pallone F, Monteleone G. IL-21 as a therapeutic target in inflammatory disorders. Expert Opin Ther Targets. 2014; 18(11):1329-1338.
- Sorge RE, Mapplebeck JC, Rosen S, et al. Different immune cells mediate mechanical pain hypersensitivity in male and female mice. *Nat Neurosci.* 2015; 18(8):1081-1083.
- Kikuno R, Nagase T, Ishikawa K, et al. Prediction of the coding sequences of unidentified human genes. XIV. The complete sequences of 100 new cDNA clones from brain which code for large proteins in vitro. DNA Res. 1999;6(3):197-205.

DOI 10.1182/blood-2017-02-769661

#### To the editor:

## Failure to replicate thrombomodulin genetic variant predictors of venous thromboembolism in African Americans

Aaron R. Folsom, Nicholas S. Roetker, Spencer T. Kelley, Weihong Tang, and Nathan Pankratz

<sup>1</sup>Division of Epidemiology & Community Health, School of Public Health, and <sup>2</sup>Department of Laboratory Medicine and Pathology, School of Medicine, University of Minnesota, Minneapolis, MN

African Americans have a higher risk of venous thromboembolism (VTE) than most other ethnic groups. A case-control, genome-wide association study of VTE in African Americans by Hernandez et al<sup>1</sup>

recently reported that having minor alleles of 3 intergenic singlenucleotide polymorphisms (SNPs) on chromosome 20 (rs2144940, rs2567617, and rs1998081) was associated with more than a doubling