

Comment on Artz et al, page 529

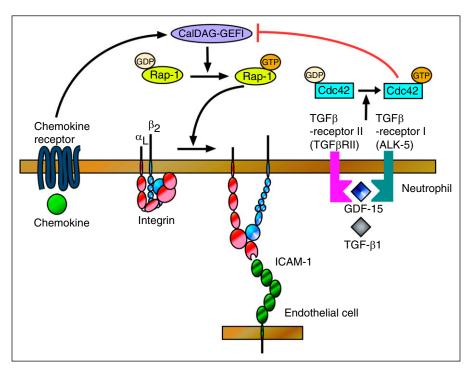
Dampening neutrophil integrins

Rodger P. McEver Oklahoma medical research foundation

In this issue of *Blood*, Artz et al show that growth differentiation factor 15 (GDF-15), a distant relative of transforming growth factor β (TGF- β), inhibits neutrophil integrin activation through canonical TGF- β receptor heterodimers.¹

uring inflammation, neutrophils roll along venules, arrest, and crawl into perivascular tissues. Interactions of neutrophil ligands with endothelial P- and E-selectin mediate rolling. Interactions of neutrophil

integrins with ICAM-1 slow rolling and mediate arrest. Signaling through selectin ligands activates integrin $\alpha L\beta 2$ to an extended, intermediate-affinity conformation that slows rolling. Signaling through chemokine



Signals transmitted through chemokine receptors on rolling neutrophils activate the GTPase Rap-1, in part through CalDAG-GEFI. GTP-bound Rap-1 acts through effectors to convert integrin $\alpha L\beta 2$ to an extended, high-affinity conformation, which mediates neutrophil arrest by binding ICAM-1 on endothelial cells. Artz et al demonstrate that TGF- $\beta 1$ and the distantly related GDF-15 transmit signals through TGF- β -receptor heterodimers, which activate the GTPase Cdc42. GTP-bound Cdc42 inhibits activation of Rap-1, likely by interfering with CalDAG-GEFI. Cdc42, cell division protein 42; GDP, guanosine diphosphate. Illustration by Tadayuki Yago.

receptors activates integrin $\alpha L\beta 2$ to an extended, high-affinity conformation that mediates arrest.² In both signaling cascades, a key downstream step is activation of the guanosine triphosphatase (GTPase) Rasrelated protein 1 (Rap-1), in part through the guanine nucleotide exchange factor (GEF) CalDAG-GEFI (see figure). GTP-bound Rap-1 acts through effectors to recruit talin-1 to the plasma membrane.³ Binding of talin-1 to the $\beta 2$ cytoplasmic tail initiates integrin activation.⁴ The signaling steps that activate integrins have been intensively investigated. Whether signals can inhibit integrin activation has received less attention.

Earlier studies revealed that another GTPase, Cdc42, in its active GTP-bound form, decreases chemokine-induced activation of αLβ2 in lymphocytes.⁵ GDF-15 was then shown to inhibit chemokine-mediated activation of $\alpha L\beta 2$ in neutrophils by activating Cdc42, which blocks activation of Rap-1.6 Inflammatory mediators induce expression of GDF-15 in many tissues. In GDF-15deficient mice, more neutrophils arrest and migrate outside venules after inflammatory challenge.⁶ Thus, GDF-15 is an antiinflammatory cytokine that dampens neutrophil integrin activation. Its weak sequence similarity to other TGF-B family members suggested that it signals by distinct mechanisms. But Artz et al demonstrate that GDF-15, and TGF-β1 itself, inhibits chemokine-triggered integrin activation in neutrophils through heterodimers of TGF-B receptor I (TGF-BRI), also known as activin receptor-like kinase 5, and TGF-BRII. Like GDF-15-deficient mice, mice lacking TGF-BRI or TGF-BRII in myeloid cells have enhanced neutrophil arrest and migration after challenge. Both GDF-15 and TGF-B1 use TGF-B receptor heterodimers to activate Cdc42, which is required to inhibit Rap-1 in neutrophils. Strikingly, neither GDF-15 nor TGF-B1 inhibits chemokine-induced integrin activation in CalDAG-GEFI-deficient neutrophils. The

authors did not test whether the lack of CalDAG-GEFI prevents activation of Cdc42, although the latter has other known GEFs. Instead, they suggest that activated Cdc42 somehow prevents CalDAG-GEFI from activating Rap-1. How this might occur is an intriguing topic for future study.

Although Artz et al do not address this possibility, signaling through TGF-B receptors likely also dampens selectintriggered integrin activation by inhibiting Rap-1. Engaging TGF-β receptors may have other effects. For example, phosphatidylinositol-4phosphate 5-kinase γ (PIP5KI γ) recruits talin-1 to membranes by direct binding and by generating phosphatidylinositol 4,5 bisphosphate (PIP2).³ In lymphocytes, constitutively active Cdc42 decreases chemokine-induced generation of PIP2, implying that it negatively regulates PIP5KIy.5 Thus, TGF-B-receptor activation of Cdc42 could block talin-1 recruitment by inhibiting both Rap-1 and PIP5KIy. Artz et al also show that engaging TGF-B receptors on monocytic cells inhibits chemokine activation of $\alpha 4$ integrins, at least in vitro. Therefore, TGF-B receptors may modulate different integrins in different leukocyte subsets.

Artz et al demonstrate that TGF-B1 inhibits neutrophil integrin activation in vitro. Whether it exerts this function in vivo requires further investigation. Most cells express TGF-β1, which exerts pleiotropic effects depending on cellular context. These effects typically regulate gene expression, whereas Artz et al describe a rapid, transcription-independent signaling pathway. To become active, TGF-B1 must dissociate from a latency complex. Activated platelets release large amounts of latent TGF- β 1 that can be activated by shear stress under some conditions.7 Activated platelets bind to neutrophils arrested in venules and transmit signals that regulate integrin αMβ2-dependent crawling.⁸ Could plateletreleased TGF-B1 contribute to signaling?

Other molecules influence neutrophil integrins. Signaling through neutrophil adenosine A2A receptors hinders selectinand chemokine-induced β 2 integrin activation, as well as β 2 integrin outside-in signaling.⁹ On the other hand, rolling neutrophils secrete myeloid-related proteins 8 and 14 from the cytosol.¹⁰ The secreted proteins bind in an autocrine manner to tolllike receptor 4 on neutrophils, triggering Rap-1–dependent activation of $\beta 2$ integrins. The demonstration that TGF- β receptors negatively regulate $\beta 2$ integrins further emphasizes the complexity of neutrophil responses during inflammation. Determining how diverse signaling pathways cooperate or compete in different contexts may suggest new therapies for inflammatory diseases.

Conflict-of-interest disclosure: The author holds equity in Selexys and Tetherex Pharmaceuticals.

REFERENCES

 Artz A, Butz S, Vestweber D. GDF-15 inhibits integrin activation and mouse neutrophil recruitment through the ALK-5/TGF-βRII heterodimer. *Blood.* 2016;128(4):529-541.

2. Lefort CT, Ley K. Neutrophil arrest by LFA-1 activation. *Front Immunol.* 2012;3:157.

3. Calderwood DA, Campbell ID, Critchley DR. Talins and kindlins: partners in integrin-mediated adhesion. *Nat Rev Mol Cell Biol.* 2013;14(8):503-517.

4. Yago T, Petrich BG, Zhang N, et al. Blocking neutrophil integrin activation prevents ischemia-reperfusion injury. *J Exp Med.* 2015;212(8):1267-1281.

• • • LYMPHOID NEOPLASIA

Comment on Guo et al, page 553

IL-4 regulates B-cell receptor signaling in CLL

Dimitar G. Efremov INTERNATIONAL CENTRE FOR GENETIC ENGINEERING AND BIOTECHNOLOGY

In this issue of *Blood*, Guo et al show that interleukin-4 (IL-4) upregulates surface immunoglobulin M (IgM) expression and B-cell receptor (BCR) signaling capacity of chronic lymphocytic leukemia (CLL) cells, providing an explanation of how anergic CLL cells from the peripheral blood become signal competent when they enter the lymph nodes.¹

he recent approval of the Bruton tyrosine kinase (BTK) inhibitor ibrutinib and the phosphatidylinositol 3-kinase δ (PI3Kδ) inhibitor idelalisib represent major breakthroughs in the treatment of CLL. Although these drugs can inhibit various signaling pathways, they are believed to function primarily by inhibiting signals that the leukemic cells receive through their BCRs.² This mechanism of action is supported by numerous lines of evidence supporting a major role for the BCR pathway in the development and progression of CLL, including the expression of BCRs with particular structural and antigen-binding properties, the expression of high levels of

5. Bolomini-Vittori M, Montresor A, Giagulli C, et al. Regulation of conformer-specific activation of the integrin LFA-1 by a chemokine-triggered Rho signaling module. *Nat Immunol.* 2009;10(2):185-194.

6. Kempf T, Zarbock A, Widera C, et al. GDF-15 is an inhibitor of leukocyte integrin activation required for survival after myocardial infarction in mice. *Nat Med.* 2011;17(5):581-588.

7. Ahamed J, Burg N, Yoshinaga K, Janczak CA, Rifkin DB, Coller BS. In vitro and in vivo evidence for shear-induced activation of latent transforming growth factor- β 1. *Blood.* 2008;112(9):3650-3660.

8. Sreeramkumar V, Adrover JM, Ballesteros I, et al. Neutrophils scan for activated platelets to initiate inflammation. *Science*. 2014;346(6214):1234–1238.

9. Yago T, Tsukamoto H, Liu Z, Wang Y, Thompson LF, McEver RP. Multi-inhibitory effects of A2A adenosine receptor signaling on neutrophil adhesion under flow. *J Immunol.* 2015;195(8):3880-3889.

10. Pruenster M, Kurz AR, Chung KJ, et al. Extracellular MRP8/14 is a regulator of β 2 integrin-dependent neutrophil slow rolling and adhesion. *Nat Commun.* 2015;6:6915.

DOI 10.1182/blood-2016-06-719849

© 2016 by The American Society of Hematology

BCR-target genes in lymph node-derived CLL cells, and the significant association between clinical course and certain BCR-related features, such as the mutational status of the immunoglobulin heavy chain variable (IGHV) region genes.³ This evidence, however, is difficult to reconcile with the relatively impaired capacity of CLL cells in the peripheral blood to respond to BCR engagement. Reduced BCR signaling capacity is a general feature of CLL, but is particularly prominent in the CLL subset with IGHV-mutated genes (M-CLL), where the malignant B cells are often completely anergic.⁴

In the article published in this issue of *Blood*, Guo et al investigate the mechanism(s)