

TRANSPLANTATION

Replication of associations between genetic polymorphisms and chronic graft-versus-host disease

Paul J. Martin,^{1,2} Wenhong Fan,³ Barry E. Storer,^{1,4} David M. Levine,⁴ Lue Ping Zhao,³ Edus H. Warren,^{1,2} Mary E. D. Flowers,^{1,2} Stephanie J. Lee,^{1,2} Paul A. Carpenter,^{1,2} Michael Boeckh,^{2,5} Sangeeta Hingorani,⁶ Li Yan,⁷ Qiang Hu,⁷ Leah Preus,^{8,9} Song Liu,⁷ Stephen Spellman,¹⁰ Xiaochun Zhu,¹¹ Marcelo Pasquini,¹¹ Philip McCarthy,¹² Daniel Stram,¹³ Xin Sheng,¹³ Loreall Pooler,¹³ Christopher A. Haiman,¹³ Lara Sucheston-Campbell,^{8,9} Theresa Hahn,⁷ and John A. Hansen^{1,2}

¹Division of Clinical Research, Fred Hutchinson Cancer Research Center, Seattle, WA; ²Department of Medicine, University of Washington, Seattle, WA; ³Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, WA; ⁴Department of Biostatistics, University of Washington, Seattle, WA; ⁵Division of Vaccines and Infectious Diseases, Fred Hutchinson Cancer Research Center, Seattle, WA; ⁶Department of Pediatrics, University of Washington, Seattle, WA; ⁷Department of Biostatistics & Bioinformatics, Roswell Park Cancer Institute, Buffalo, NY; ⁸College of Pharmacy and ⁹College of Veterinary Medicine, The Ohio State University, Columbus, OH; ¹⁰Center for International Blood and Marrow Transplant Research (CIBMTR), National Marrow Donor Program/Be The Match, Minneapolis, MN; ¹¹CIBMTR, Medical College of Wisconsin, Milwaukee, WI; ¹²Department of Medicine, Roswell Park Cancer Institute, Buffalo, NY; and ¹³Department of Preventive Medicine, University of Southern California, Los Angeles, CA

Key Points

- Most published SNP associations with chronic GVHD are likely to represent false-positive findings.
- HRs for any true-positive SNP associations are likely to be much smaller than reported previously.

Previous studies have identified single-nucleotide polymorphisms (SNPs) associated with the risk of chronic graft-versus-host disease (GVHD) after allogeneic hematopoietic cell transplantation. The current study determined whether these associations could be replicated in large cohorts of donors and recipients. Each SNP was tested with cohorts of patients having the same donor type (HLA-matched related, unrelated, or both) reported in the original publication, and testing was limited to the same genome (recipient or donor) and genetic model (dominant, recessive, or allelic) reported in the original study. The 21 SNPs reported in this study represent 19 genes, and the analysis encompassed 22 SNP association tests. The hazard ratio (HR) point estimates and risk ratio point estimates corresponding to odds ratios in previous studies consistently fall outside the 95% confidence intervals of HR estimates in the current study. Despite the large size of the cohorts available for the current study, the 95% confidence intervals for most HRs did not

exclude 1.0. Three SNPs representing *CTLA4*, *HPSE*, and *IL1R1* showed evidence of association with the risk of chronic GVHD in unrelated donor-recipient pairs from 1 cohort, but none of these associations was replicated when tested in unrelated donor-recipient pairs from an independent cohort. Two SNPs representing *CCR6* and *FGFR1OP* showed possible associations with the risk of chronic GVHD in related donor-recipient pairs but not in unrelated donor-recipient pairs. These results remain to be tested for replication in other cohorts of related donor-recipient pairs. (*Blood*. 2016;128(20):2450-2456)

Introduction

Previous studies have identified genetic variants such as single-nucleotide polymorphisms (SNPs) and other polymorphisms that influence the risk of chronic graft-versus-host disease (GVHD) after allogeneic hematopoietic cell transplantation (HCT).¹⁻¹⁹ Many of the reported variants regulate the function of immune cells, their receptors, effector molecules, cytokines, or chemokines. Some results have suggested that the assessment of these variants before HCT might help to assess the risk of adverse outcomes for each patient, guide the clinical management of patients who are at high risk, and ultimately serve as potential biologic targets for novel therapeutics.

Previous case-control or cohort studies of SNP associations with chronic GVHD evaluated 50 to 353 individuals. No previous study has comprehensively evaluated the association of these variants in the same cohort simultaneously. In this study, we used genotyped or imputed

SNP data to determine how many of the previously published associations we could replicate with substantially larger numbers of individuals from 2 cohorts.

Methods

Literature search

We performed a comprehensive PubMed search using the terms “chronic GVHD” and “polymorphism” to identify all studies published by August 2014 reporting an association of a genetic polymorphism with the odds ratio (OR) or relative risk of chronic GVHD at an α level <0.05 . Studies that did not meet this threshold were not included. Reported associations of genetic deletions,

Submitted 14 July 2016; accepted 20 September 2016. Prepublished online as *Blood* First Edition paper, 6 October 2016; DOI 10.1182/blood-2016-07-728063.

The online version of this article contains a data supplement.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked “advertisement” in accordance with 18 USC section 1734.

© 2016 by The American Society of Hematology

Table 1. Characteristics of the FHCRC study cohort (n = 3918)

Characteristic	No. (%)
Recipient age at transplantation, years	
Median	43
Range	0-78
Diagnosis	
Acute leukemia	1605 (41)
Chronic myeloid leukemia	971 (25)
Myelodysplastic syndromes or myeloproliferative neoplasms	638 (16)
Chronic lymphocytic leukemia	110 (3)
Malignant lymphoma or multiple myeloma	593 (15)
Histiocytic sarcoma	1
Donor-recipient gender combination	
Male to male	1337 (34)
Male to female	873 (22)
Female to male	946 (24)
Female to female	762 (19)
HLA and donor type	
HLA-matched related	1819 (46)
Unrelated HLA-matched, confirmed by high-resolution typing	1213 (31)
Unrelated HLA-matched, unconfirmed by high-resolution typing	239 (6)
HLA-mismatched unrelated	666 (17)
Graft source	
Bone marrow	2179 (56)
Mobilized blood cells	1739 (44)
Conditioning regimen	
Myeloablative with <1000 cGy total body irradiation	1340 (34)
Myeloablative with ≥1000 cGy total body irradiation	1887 (18)
Nonmyeloablative	691 (18)
Prior grade 2-4 acute GVHD*	2848 (73)

*Grades were not assigned for 23 recipients.

insertions, microsatellites, or variable number tandem repeats with chronic GVHD were excluded because the genotyping arrays used for our study do not detect these variants.

Study cohort: FHCRC

Description. The Fred Hutchinson Cancer Research Center (FHCRC) cohort included 3918 donor-recipient pairs of European ancestry who received allogeneic HCT from 1990 through 2011 (Table 1). European ancestry was determined by analysis of principal components. For purposes of this study, the numbers of available non-European pairs in this cohort were not sufficient for meaningful analysis. The cohort included recipients with either HLA-matched related donors (MRDs) (N = 1819) or unrelated donors (URDs) (N = 2019). The number of recipients with HLA-mismatched related donors in our cohort is not sufficient for meaningful stratified analysis. Indications for HCT included hematologic malignancy or myelodysplasia. Patients treated with either myeloablative or nonmyeloablative conditioning regimens were included in the analysis, whereas patients who received T cell–depleted grafts or were treated with rabbit antithymocyte globulin as part of the conditioning regimen were excluded from the analysis, because these interventions decrease the risk of chronic GVHD.

Baseline recipient and donor information was collected during the evaluation before HCT. URD and recipient matching for HLA-A, HLA-B, and HLA-C; DRB1; and DQB1 was confirmed by high-resolution methods in most cases. Grade 2-4 acute GVHD was diagnosed according to previously described criteria.²⁰ Information regarding chronic GVHD and follow-up outcomes was captured prospectively by the Long-Term Follow-Up program through medical records from our outpatient clinic and from referring physicians who provided the primary care for patients. Clinical extensive chronic GVHD was diagnosed by historical criteria.²¹ Patients with clinical limited chronic GVHD were not considered as having chronic GVHD unless and until they developed clinical extensive disease. By National Institutes of Health (NIH) consensus criteria, this definition used in this study includes classical and overlap chronic GVHD and late acute GVHD. In the FHCRC cohort, the cumulative incidence of chronic

GVHD at 2 years after HCT was 44% (95% confidence interval [CI], 42-47) among patients with HLA-MRDs and 48% (95% CI, 45-50) among patients with URDs. Methods for collection of biospecimens, genotyping, and imputation for the FHCRC cohort are summarized in the supplemental Appendix, available on the *Blood* Web site.

Statistical analysis of results in the FHCRC cohort. For purposes of replicating previous results, each SNP was tested with subsets of the FHCRC cohort of patients having the same donor type (MRD, URD, or both) reported in the original publication; initial testing was limited to the same genome (recipient or donor) reported in the original study using genotypes only from the microarray platforms that passed quality control for each SNP (supplemental Table 1). The primary analysis used Cox regression models with no adjustments, treating death and recurrent malignancy as competing risks. A secondary analysis used multivariate Cox regression models with additional covariates for baseline clinical risk factors that have strong effect on the risk of chronic GVHD (HLA-matched URD vs HLA-mismatched URD vs HLA-MRD; female donor for male recipient vs other combinations; mobilized blood cell graft-versus-marrow graft; diagnosis of chronic myeloid leukemia vs other diseases; conditioning with total body irradiation vs without; and patient age).²² HLA-matching of URDs was coded according to categories shown in Table 1. The secondary analysis also included the first 4 principal components to control for population stratification.²³

Each SNP was evaluated for allelic or genotypic (recessive and dominant) association as reported in the original study. For a SNP with a major allele “A” and a minor allele “a,” the recessive model tests the hypothesis that the genotype “aa” is associated with a higher or lower risk compared with the collective genotypes “AA” and “Aa” used as the reference. The dominant model tests the hypothesis that the collective genotypes “Aa” and “aa” are associated with a higher or lower risk compared with the genotype “AA” used as the reference. The allelic model tests the hypothesis that the minor allele “a” is associated with a higher or lower risk compared with the major allele “A,” and the number of copies of the minor allele is modeled as an additive effect. Because the main goal of this analysis was to replicate previously reported associations, a 2-sided $P \leq .05$ was selected as the threshold of significance, despite the multiple comparisons.

To compare results of the current study with those of previous case-control studies, ORs were converted to the corresponding risk ratios after accounting for the minor allele frequency and genetic model, with the incidence of chronic GVHD set at 45%. Where necessary, risk alleles, genetic models, and ORs reported in previous studies were inverted to match the analysis in the current study. With these adjustments, the risk ratio from a previous case-control study is used to approximate the corresponding hazard ratio (HR) that might be expected in a cohort study.

Study cohort: DISCOVeRY-BMT

Description. Recipients and donors in the Determining the Influence of Susceptibility Conveying Variants Related to One Year mortality after Unrelated Donor Allogeneic Blood or Marrow Transplant (DISCOVeRY-BMT) cohort originated from the Center for International Blood and Marrow Transplant Research (CIBMTR) and were selected for the DISCOVeRY-BMT Genome-wide Association Study.^{24,25} CIBMTR is collaborative research program organized by the National Marrow Donor Program/Be The Match Registry and the Medical College of Wisconsin and collects data from a voluntary working group of more than 450 transplant centers throughout the world. Participating centers contribute comprehensive baseline and longitudinal follow-up data as well as pretransplant biospecimens.

DISCOVeRY-BMT cohort 1 included 2609 patients who received a first HLA-A, HLA-B, and HLA-C, DRB1, DQB1-matched unrelated HCT transplant for treatment of acute lymphoblastic leukemia, acute myeloid leukemia, or myelodysplastic syndrome between 2000 and 2008. Patients were excluded if they had received T cell–depleted grafts or cord blood grafts, or if biorepository samples were not available from both the donor and recipient. DISCOVeRY-BMT cohort 2 included 572 patients who were selected according to the same criteria but had HCT between 2009 and 2011, together with 351 patients who received a first HLA-A, HLA-B, and HLA-C, DRB1-matched unrelated HCT for the same indications between 2000 and 2011 without assessment of HLA-DQB1 matching. For the purposes of this replication analysis, we included only individuals of European ancestry and excluded those who were treated with antithymocyte globulin as part of the conditioning regimen, yielding 1656 and

Table 2. SNPs previously associated with risk of chronic GVHD

Gene	Reference	Population	Genome	Graft	N	SNP	Alleles	MAF*	Model	Statistic	95% CI
<i>FAS</i>	11	EUR	R	MRD	107	rs1800682	A/G	0.48	Dominant	OR 3.6	1.1-11
<i>IL10</i>	11	EUR	R	MRD	106	rs1800871	C/T	0.19	Dominant	OR 3.9	1.5-10
<i>IL10</i>	11	EUR	R	MRD	106	rs1800896	G/A	0.48	Dominant	OR 2.3	1.1-5.3
<i>IL10</i>	12	EUR	R	MRD	95	rs1800896	A/G	0.45	Recessive	HR 2.8	1.1-7.0
<i>IL10</i>	8	Korean	R	MRD	53	rs1800871	T/C	0.34	Dominant	OR 0.07	0.01-0.61
<i>IL10</i>	8	Korean	R	MRD	53	rs1800871	T/C	0.34	Recessive	OR 0.16	0.03-0.84
<i>IL10RB</i>	15	EUR	R	MRD	184	rs2834167	A/G	0.31	Dominant†	OR 2.3	1.0-5.0
<i>IL6</i>	16	EUR	R	MRD	99	rs1800795	G/C	0.40	Recessive	HR 0.24	0.07-0.77
<i>CCR6</i>	4	EUR	D	MRD	153	rs3093023	G/A	0.42	Dominant	OR 4.2	1.5-11
<i>FGFR10P</i>	4	EUR	D	MRD	156	rs2301436	G/A	0.45	Dominant	OR 6.3	2.0-20
<i>CTLA4</i>	3	EUR	D	MRD	225	rs231775	A/G	0.43	Recessive	HR 1.8	1.0-3.0
<i>CTLA4</i>	18	EUR	D	URD	147	rs3087243	G/A	0.37	Recessive	HR 1.8	1.1-3.2
<i>GSTP1</i>	12	EUR	D	MRD	95	rs1695	A/G	0.28	Dominant	HR 2.3	1.3-3.8
<i>CD14</i>	18	EUR	R	URD	147	rs2569190	G/A	0.47	Recessive	HR 1.9	1.3-2.9
<i>MADCAM1</i>	1	EUR	R	MRD+URD	70	rs2302217	A/G	0.50	Dominant	OR 3.6	1.1-11
<i>HPSE</i>	17	EUR	R	MRD+URD	225	rs4693608	A/G	0.45	Dominant	HR 0.38	0.15-0.91
<i>PARP1</i>	2	EUR	R	MRD+URD	352	rs1805410	A/G	0.15	Allelic	HR 1.8	1.3-2.5
<i>TNFSF13B</i>	5	EUR	R	MRD+URD	156	rs16972217	C/T	0.26	Allelic	OR 2.7	1.4-5.4
<i>IL1R1</i>	10	EUR	D	MRD+URD	302	rs3917225	A/G	0.44	Dominant	HR 1.3	1.1-1.6
<i>TNFA</i>	19	EUR-AF	R	MRD	82	rs361525	G/A	0.10	Dominant	HR 2.6	1.3-5.2
<i>FCRL3</i>	14	Japanese	R	MRD	112	rs7528684	T/C	0.43	Recessive	OR 0.21	0.06-0.60
<i>IL2</i>	6	Japanese	R	URD	326	rs2069762	T/G	0.33	Dominant	OR 2.3‡	1.4-3.6
<i>CCL5</i>	9	Korean	R	MRD+URD	50	rs1800825	C/G	0.15	Dominant	HR 2.9	1.2-7.0
<i>GZMB</i>	7	Japanese	D	URD	353	rs8192917	A/G	0.20	Dominant§	HR 0.61	0.37-0.99

AF, African; D, donor; EUR, European; MAF, minor allele frequency; R, recipient.

*In some cases, minor allele frequencies reflect 1000 genomes results because data were not reported by the authors. Alleles are shown as major/minor, and, in some cases, strand designations were adjusted for consistency with the current analysis. Results were identical for rs1800872 and rs1800871 because these SNPs are in perfect linkage disequilibrium with each other. Results for rs16972217 were similar for 3 other *TNFSF13B* SNPs in strong linkage disequilibrium.

†Results reflect cases where patients and donors had the same interleukin-10 production levels based on *IL10* genotypes.

‡Results for GT vs others; GG noted to have highest risk.

§Only in patients with acute myeloid leukemia or myelodysplastic syndrome in the multivariate model; not statistically significant in the univariate model.

527 recipients and 1601 and 514 donors in cohorts 1 and 2, respectively. The cumulative incidence of chronic GVHD at 2 years after HCT was 43% (95% CI, 41-45) in the combined cohorts. Methods for collection of biospecimens, genotyping, and imputation for the DISCOVeRY-BMT cohort are summarized in the supplemental Appendix.

Statistical analysis of results in the DISCOVeRY-BMT cohort.

Cox proportional hazards models evaluated time to extensive chronic GVHD. Deaths from any cause and progression or recurrence of malignant disease were treated as competing risks. Multivariate models were unadjusted, including only the SNP of interest and then adjusted for the following covariates: donor sex mismatch (female donor to male recipient or not), graft type (blood, marrow), total body irradiation exposure (>900 cGy, ≤900 cGy, or none), and recipient age (continuous). Dosage data accounting for the probability of each genotype were used in all analyses of imputed data. To combine data from DISCOVeRY-BMT cohorts 1 and 2, the inverse variance weighting method was used as implemented in the R package METAL. For adjusted analyses, we report the METAL HRs, CIs, and *P* values under a fixed effects model because heterogeneity between cohorts was low ($0 \leq I^2 < 25$).²⁶

Results

Testing of SNPs in the FHCR cohort

The primary goal of this study was to determine whether we could replicate previously reported SNP associations with risk of chronic GVHD (Table 2). Previous studies have identified 29 informative SNPs associated with the risk of chronic GVHD according to allelic, dominant, or recessive genetic models. Three of these could not be typed or imputed from results on any of the platforms used for our study. In the current study, results are reported for only 1 of the 4

TNFSF13B SNPs reported by Clark et al⁵ because they have nearly perfect linkage disequilibrium, and results did not differ among them. Likewise, the *IL10* SNPs rs1800872 and rs1800871 are in strong linkage disequilibrium. In the current study, results are reported only for rs1800871.

The 21 SNPs reported in this study represent 19 genes. Seven were donor SNPs representing 6 genes, and 13 were recipient SNPs representing 11 genes. The *TNFA* SNP is located in the major histocompatibility complex and therefore represents both the donor and recipient in HLA-identical related pairs. The analysis encompassed 22 SNP association tests because previous studies of rs1800896 showed both dominant and recessive associations.^{11,12}

In an unadjusted analysis, results of this screen replicated results for 3 of the 22 SNP associations (Table 3). Related donor rs3093023 (*CCR6*) and rs2301436 (*FGFR10P*) genotypes showed statistically significant dominant genetic associations with the risk of chronic GVHD (HR, 1.19; 95% CI, 1.02-1.38; *P* = .02; and HR, 1.23; 95% CI, 1.06-1.44; *P* = .01, respectively). Results of the adjusted analysis were similar. Kochi et al²⁷ previously identified rs968334 as a functional SNP regulating the expression of *CCR6*, and this SNP has strong linkage disequilibrium with rs3093023. Related donor rs968334 genotypes showed a statistically significant dominant genetic association with the risk of chronic GVHD (*N* = 1757; HR, 1.19; 95% CI, 1.03-1.39; *P* = .02). Results of the adjusted analysis were similar (HR, 1.16; 95% CI 0.99-1.35; *P* = .06).

For exploratory purposes, donor *CCR6* rs3093023 and rs968334 genotypes and donor *FGFR10P* rs2301436 genotypes were tested for dominant genetic associations with the risk of chronic GVHD in unrelated recipients (*N* = 2002, 1998, and 2011, respectively). Results showed no statistically significant association in the unadjusted

Table 3. Replication testing of candidate SNPs for association with the risk of chronic GVHD in the FHCRC cohort*

Gene	Genome	Graft	SNP	Alleles	MAF	Model	N†	Unadjusted analysis			Adjusted analysis		
								HR	95% CI	P	HR	95% CI	P
<i>FAS</i>	R	MRD	rs1800682	A/G	0.46	Dominant	1640	0.99	0.85-1.16	.94	0.99	0.84-1.16	.85
<i>IL10</i>	R	MRD	rs1800871	G/A	0.24	Dominant	1622	1.07	0.92-1.24	.39	1.07	0.93-1.25	.34
<i>IL10</i>	R	MRD	rs1800871	G/A	0.24	Recessive	1622	0.96	0.69-1.34	.80	0.97	0.69-1.36	.86
<i>IL10</i>	R	MRD	rs1800896	T/C	0.48	Recessive	1196	1.01	0.83-1.24	.90	1.00	0.81-1.22	.97
<i>IL10RB</i>	R	MRD	rs2834167	A/G	0.26	Dominant‡	674	1.13	0.90-1.42	.30	1.10	0.88-1.39	.40
<i>IL6</i>	R	MRD	rs1800795	G/C	0.40	Recessive	1663	0.94	0.76-1.15	.52	0.93	0.76-1.15	.50
<i>CCR6</i>	D	MRD	rs3093023	G/A	0.44	Dominant	1758	1.19	1.02-1.38	.02	1.15	0.99-1.34	.08
<i>FGFR1OP</i>	D	MRD	rs2301436	C/T	0.47	Dominant	1772	1.23	1.06-1.44	.008	1.18	1.01-1.38	.04
<i>CTLA4</i>	D	MRD	rs231775	A/G	0.38	Recessive	1761	0.92	0.76-1.12	.39	0.92	0.76-1.13	.43
<i>CTLA4</i>	D	URD	rs3087243	G/A	0.45	Recessive	2011	0.84	0.72-0.99	.04	0.84	0.72-0.99	.04
<i>GSTP1</i>	D	MRD	rs1695	A/G	0.35	Dominant	1291	1.01	0.86-1.19	.89	0.99	0.84-1.17	.91
<i>CD14</i>	R	URD	rs2569190	G/A	0.48	Recessive	1958	0.97	0.84-1.13	.69	0.96	0.83-1.11	.58
<i>MADCAM1</i>	R	ALL	rs2302217	A/G	0.47	Dominant	671	1.10	0.85-1.43	.47	1.16	0.89-1.51	.28
<i>HPSE</i>	R	ALL	rs4693608	A/G	0.48	Dominant	2574	1.12	0.98-1.28	.09	1.15	1.01-1.31	.04
<i>PARP1</i>	R	ALL	rs1805410	T/C	0.15	Allelic	3637	1.06	0.97-1.17	.20	1.05	0.96-1.16	.28
<i>TNFSF13B</i>	R	ALL	rs16972217	C/T	0.23	Allelic	2589	0.93	0.84-1.02	.13	0.93	0.84-1.02	.13
<i>IL1R1</i>	D	ALL	rs3917225	A/G	0.44	Dominant	3737	1.06	0.96-1.18	.24	1.07	0.96-1.18	.22
<i>TNF</i>	R	MRD	rs361525	G/A	0.05	Dominant	1662	0.97	0.76-1.24	.82	0.94	0.73-1.20	.62
<i>FCRL3</i>	R	MRD	rs7528684	A/G	0.45	Recessive	1652	1.02	0.86-1.21	.81	1.01	0.85-1.20	.93
<i>IL2</i>	R	URD	rs2069762	A/C	0.29	Dominant	1966	1.08	0.96-1.23	.21	1.06	0.94-1.21	.35
<i>CCL5</i>	R	ALL	rs1800825	A/G	0.02	Dominant	3582	0.89	0.69-1.16	.39	0.89	0.68-1.15	.37
<i>GZMB</i>	D	URD	rs8192917	T/C	0.23	Dominant§	927	1.04	0.85-1.26	.72	0.99	0.82-1.21	.95

*Alleles are shown as major/minor. Results for rs16972217 were similar for 3 other *TNFSF13B* SNPs in near perfect linkage disequilibrium. Statistically significant HRs are highlighted in bold.

†Numbers reflect samples that passed quality control.

‡Comparisons were limited to patients with the same interleukin-10 production level.

§Testing was limited to patients with acute myeloid leukemia or myelodysplastic syndromes.

analyses (HR, 1.07; 95% CI, 0.93-1.22; $P = .36$; HR, 1.07; 95% CI, 0.94-1.23; $P = .30$; and HR 1.04; 95% CI 0.90-1.19; $P = .62$, respectively). Results of the adjusted analyses were similar (data not shown). Tests for homogeneity showed no significant differences between the respective HRs for related and unrelated HCT.

URD rs3087243 (*CTLA4*) genotypes showed statistically significant recessive genetic association with the risk of chronic GVHD (HR, 0.84; 95% CI, 0.72-0.99; $P = .04$). Results of the adjusted analysis were similar. Related donor rs3087243 genotypes, however, did not show a statistically significant recessive genetic association (HR, 0.95; 95% CI, 0.80-1.13; $P = .57$). A test for homogeneity showed no significant difference between the respective HRs for related and unrelated HCT.

Previous studies evaluated 6 SNP associations with chronic GVHD in a combined cohort of related and unrelated graft recipients. Because the balance between related and unrelated grafts varied considerably in these studies, we tested these SNPs with stratification for type of graft (supplemental Table 3). Unrelated patient *HSPE* rs4693608 genotypes showed a statistically significant dominant genetic association with risk of chronic GVHD (HR, 1.26; 95% CI, 1.06-1.51; $P = .01$). Results of the adjusted analysis were similar. Related patient *HSPE* rs4693608 genotypes, however, did not show an association with the risk of chronic GVHD (HR, 0.98; 95% CI, 0.81-1.19; $P = .86$), and a test for homogeneity suggested a significant difference between the respective HRs for related and unrelated HCT ($P = .05$). URD *ILIR1* rs3917225 genotypes showed a statistically significant dominant genetic association with the risk of chronic GVHD (HR, 1.17; 95% CI, 1.02-1.35; $P = .03$). Results of the adjusted analysis were similar. Related donor *ILIR1* rs3917225 genotypes, however, did not show an association with the risk of chronic GVHD (HR, 0.96; 95% CI, 0.82-1.11; $P = .55$). The other 4 SNPs previously associated with chronic GVHD in mixed related and unrelated HCT cohorts showed no statistically significant

associations with chronic GVHD in separate analyses of related and unrelated HCT (supplemental Table 3).

Comparison of current FHCRC results with previously published results

As shown in Figure 1, the HR point estimates and risk ratio point estimates corresponding to ORs in previous studies consistently fall outside the 95% CIs for HRs in the current study of the FHCRC cohort. HRs for SNPs showing a possible association with chronic GVHD in the current study are much closer to 1.0 than those in previous studies.

Further testing of selected SNPs in the DISCOVERy-BMT cohorts

In the FHCRC cohort, SNPs representing *CTLA4*, *HPSE*, and *ILIR1* showed evidence of association in URD HCT but not in MRD HCT, whereas SNPs representing *CCR6* and *FGFR1OP* showed evidence of association with the risk of chronic GVHD in MRD HCT but not in URD HCT. These 5 SNPs were tested for association with chronic GVHD in a cohort of 2183 URD-recipient pairs from the combined DISCOVERy-BMT cohorts, according to the same recipient or donor genome and dominant, recessive, or allelic genetic model reported in the original study. No statistically significant associations were observed in this analysis (supplemental Table 4). No large cohort of related donor-recipient pairs with genotyping data is available for replication of current results from testing SNPs representing *CCR6* and *FGFR1OP* in the FHCRC cohort.

Global survey of candidate SNPs

We also screened the set of 21 SNPs for association with chronic GVHD in both the entire FHCRC cohort and in the related and

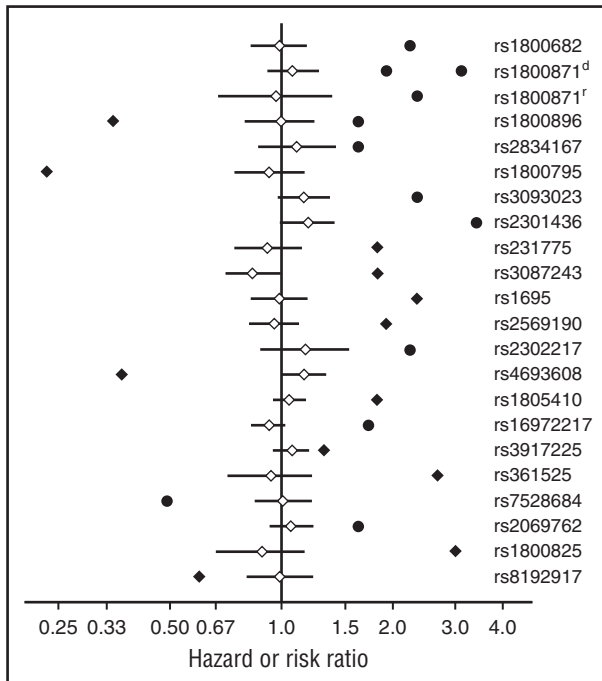


Figure 1. HR point estimates and risk ratio point estimates corresponding to ORs in previous studies consistently fall outside the 95% CIs of HR estimates in the current study. Adjusted HR point estimates (◊) and 95% CIs (—) from the FHCRC cohort are shown together with HR point estimates (◆) or risk ratio estimates corresponding to ORs (●) in previous studies. Results for rs1800871^d and rs1800871^r, respectively, represent dominant and recessive genetic models.

unrelated subsets, testing donor and recipient genotypes with allelic, dominant, and recessive models, using the first 4 principal components as covariates. For this purpose, we set a threshold of interest at $P < .001$ to account for multiple comparisons. No SNP association met this level of statistical significance.

Discussion

Only 2 of the candidate SNPs tested in this study (*CCR6*, *FGFR10P*) remain as possibly associated with the risk of chronic GVHD. These associations were replicated in related donor-recipient pairs but not in URD-recipient pairs from the FHCRC, and the results have yet to be tested in other cohorts of related donor-recipient pairs. An overriding effect of HLA-DP mismatching or more extensive mismatching for minor antigens could explain why genetic associations observed with MRD in our study could not be replicated with URD, but such a mechanism could not explain the similarly frequent cases where genetic associations observed in URD (eg, *CTLA4*, *HPSE*, *IL1R1*) were not observed with MRD. Because we have no demonstrable biological explanation for the lack of correspondence between results with MRD and URD, the evidence for a true association of these SNPs with chronic GVHD remains indeterminate. The HRs for all tested associations were much closer to 1.0 than reported in previous studies. Similar reductions in HRs or ORs toward 1.0 across several studies have been observed in the analysis of SNP associations with other diseases and may have a variety of explanations.²⁸⁻³⁰ A global survey of candidate SNPs did not yield any new findings that would meet the threshold of statistical significance adjusted for multiple comparisons.

The inability to replicate previous results in the current study is reminiscent of a previous study that replicated only 1 of 16 SNPs

previously reported to be associated with acute GVHD.³¹ At least 2 explanations should be considered in accounting for our inability to replicate previous results in the current study. Four SNPs, respectively representing *FCRL3*, *IL2*, *CCL5*, and *GZMB*, were previously tested in Asian cohorts only. Population stratification could account for the lack of association of these SNPs with chronic GVHD in the FHCRC European ancestry cohort. The T allele of rs1800871 in *IL10*, however, was previously associated with an increased risk of chronic GVHD both in a European ancestry cohort¹¹ and in an Asian cohort.⁸ Therefore, population stratification cannot explain why this SNP was not associated with the risk of chronic GVHD in the current study.

As a second explanation, it is possible that unknown heterogeneity in the pathogenesis of chronic GVHD and stratification of genetic risk factors might mask associations that would be apparent in specific subsets of patients.³² Although our conclusions are valid for the FHCRC and DISCOVeRY-BMT cohorts, they might not reflect results in cohorts of patients selected or treated in ways that differ from our cohort. Finally, changes in the criteria for the diagnosis or lack of precision in making the diagnosis of chronic GVHD during the past decade could have contributed to our inability to replicate previous results.³³ Criteria for the diagnosis of chronic GVHD were not defined in 4 of the previous studies.^{1-3,15} Two studies used historical criteria for the diagnosis of extensive chronic GVHD,^{9,18} and only 1 used the 2004 NIH criteria for chronic GVHD.⁵ The remaining studies used historical or modified criteria for the diagnosis of limited or extensive chronic GVHD.²¹ In the current study, we used historical or modified criteria for the diagnosis of extensive chronic GVHD, which include classical and overlap chronic GVHD together with late acute GVHD according to NIH criteria. The cumulative incidence frequencies of chronic GVHD were closely similar between the FHCRC and DISCOVeRY-BMT cohorts. Criteria for the diagnosis in the current study did not include patients with limited chronic GVHD. In the FHCRC cohort, however, <5% of the patients had limited chronic GVHD that never met criteria for extensive chronic GVHD. Therefore, it is unlikely that the use of clinical extensive chronic GVHD as the diagnostic criterion accounts for our inability to replicate previous results.

Previous studies evaluated between 50 and 353 individuals, whereas 18 of the 22 SNPs evaluated in the FHCRC cohort were tested in 1200 to 3737 individuals, and the 5 SNPs evaluated in the DISCOVeRY-BMT cohorts were tested in 2115 to 2183 individuals. HRs with nominal statistical significance ($P < .05$) in the FHCRC cohort were in the range from 1.15 to 1.25. These HRs are consistent with those reported in other studies of immune-mediated diseases.³⁴⁻³⁷ Point estimates for hazard and risk ratios from previous studies consistently fell outside the 95% CIs for HR estimates in the current study, demonstrating that the inability to replicate previous results in the current study cannot be explained by insufficient statistical power.

We were able to test SNPs representing *CTLA4*, *HPSE*, and *IL1R1* for association with chronic GVHD in a large cohort of unrelated pairs from the DISCOVeRY-BMT cohorts. In this cohort, any GVHD persisting after day 100 is considered chronic GVHD, similar to the definition used for the FHCRC cohort in this study. The results did not support any association of these SNPs with the risk of chronic GVHD. In the current study, the minor allele frequencies of SNPs representing *CCR6* and *FGFR10P* spanned a narrow and highly favorable range from 0.42 to 0.47. If the incidence of chronic GVHD incidence is 0.45, a cohort of 1000 to 2500 patients would be needed to provide at least 80% power to verify HRs within the 1.15 to 1.23 range observed for these SNPs in our study. We conclude that a definitive analysis of the chronic GVHD associations of the *CCR6* and *FGFR10P* SNPs reported here will depend on further studies in sufficiently powered large cohorts with chronic GVHD diagnosed by criteria similar to those used in the current study.

Despite our best efforts, we were unable to replicate the results of previous studies and, as such, our results offer no support for biologically plausible inferences resulting from those studies. In the absence of an obvious explanation for the inability to replicate previous results, we would conclude that most, but not necessarily all, of the previous results were false positives. We acknowledge that our study could have false-negative results, but, if so, it is likely that many true effects are too small to be detected, despite the large size of our cohort.

Our results have important implications for future genetic association studies of chronic GVHD. The current study used the historical definition of chronic GVHD to match the diagnostic criteria used in most previous studies. Future studies could benefit by using the more refined criteria for diagnosing chronic GVHD developed by the NIH Consensus Development Project. Previous studies evaluated associations for only a small numbers of SNPs. Future studies would benefit from expanding the number of SNPs to be evaluated. Genomewide surveys offer broad opportunity for discovery but incur a very heavy statistical penalty for multiple comparisons. A curated set of candidate SNPs that are known to alter immune function could be used to elucidate specific biological mechanisms, as suggested by a recent study of SNPs associated with sclerosis in patients with NIH-defined chronic GVHD.³² In designing such studies, careful consideration must be given to the number of SNPs and genetic models that can be tested while accounting appropriately for multiple comparisons, the minimum minor allele frequency, the anticipated effect size, the available numbers of patients and donors, and the need to replicate any new discoveries in an independent cohort.

Infectious Diseases (Public Health Service grant/cooperative agreement 5U24-CA076518); the NHLBI and NCI (grant/cooperative agreement 5U10HL069294); the Health Resources and Services Administration (contract HHS250201200016C); the Office of Naval Research (grants N00014-14-1-0028 and N00014-15-1-0848); grants from Alexion and Amgen, Inc.; an anonymous donation to the Medical College of Wisconsin; and by the Be the Match Foundation, Bristol Myers Squibb Oncology, Celgene Corporation, Chimerix, Inc., Fred Hutchinson Cancer Research Center, Gamida Cell Ltd, Genentech Inc, Genzyme Corporation, Gilead Sciences Inc, Health Research Inc, Roswell Park Cancer Institute, HistoGenetics Inc, Incyte Corporation, Jazz Pharmaceuticals Inc, Jeff Gordon Children's Foundation, The Leukemia & Lymphoma Society, The Medical College of Wisconsin, Merck & Co Inc, Mesoblast, Millennium, The Takeda Oncology Co, Miltenyi Biotec Inc, National Marrow Donor Program, Neovii Biotech NA Inc, Novartis Pharmaceuticals Corporation, Onyx Pharmaceuticals, Optum Healthcare Solutions Inc, Otsuka America Pharmaceutical Inc, Otsuka Pharmaceutical Co Ltd-Japan, Oxford Immunotec, Perkin Elmer Inc, Pharmacyclics, Sanofi US, Seattle Genetics, Sigma- τ Pharmaceuticals, Spectrum Pharmaceuticals Inc, St. Baldrick's Foundation, Sunesis Pharmaceuticals Inc, Swedish Orphan Biovitrum Inc, Telomere Diagnostics Inc, TerumoBCT, Therakos Inc, University of Minnesota, and 7Wellpoint, Inc.

The views expressed in this article do not reflect the official policy or position of the National Institutes of Health, the Department of the Navy, the Department of Defense, Health Resources and Services Administration, or any other agency of the US Government.

Acknowledgments

The authors thank Jenna Gravley for outstanding assistance in maintaining the biospecimen repository at the Fred Hutchinson Cancer Research Center (FHCRC) and verifying the identity of samples used in this study, and the FHCRC Genomics Shared Resource Laboratory (Jeff Delrow, Director) for the genotyping of FHCRC cohort II.

Work at the FHCRC was supported by the National Institutes of Health, National Heart, Lung, Blood Institute (NHLBI; grants R01-HL105914 and R01-HL87690) and the National Cancer Institute, Department of Health and Human Services (grant P30-CA015704). The DISCOVeRY-BMT study is funded by the National Institutes of Health, NHLBI (grant R01 HL102278). The RPCI Biostatistics and Bioinformatics Shared Resource is partially funded by the National Institutes of Health/National Cancer Institute (NCI) (grant P30CA016056). The Center for International Blood and Marrow Transplant Research is supported by the NCI, NHLBI, and the National Institute of Allergy and

Authorship

Contribution: P.J.M., J.A.H., L.P.Z., B.E.S., E.H.W., M.B., S.H., L.Y., L.S.-C., and T.H. designed the study. E.H.W., M.E.D.F., S.J.L., P.A.C., S.S., X.Z., and M.P. provided data and samples. W.F., L.P.Z., D.S., X.S., L. Pooler, and C.A.H. provided genotyping and imputation. W.F., D.M.L., L. Preus, X.Z., L.S.-C., and T.H. provided data quality control and coding. B.E.S., L.Y., Q.H., S.L., and L.S.-C. provided statistical analysis. P.J.M., J.A.H., L.Y., S.J.L., P.M., L.S.-C., and T.H. interpreted results. P.J.M., J.A.H., T.H., and L.S.-C. wrote the manuscript. Funding was awarded to J.A.H., P.J.M., B.E.S., L.S.-C., and T.H. All authors critically revised the manuscript for important intellectual content and approved the manuscript for publication.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

ORCID profiles: P.J.M., 0000-0001-9051-1215.

Correspondence: Paul J. Martin, Fred Hutchinson Cancer Research Center, P.O. Box 19024, Seattle, WA, 98109-1024; e-mail: pmartin@fredhutch.org.

References

- Ambruzova Z, Mrazek F, Raida L, et al. Possible impact of MADCAM1 gene single nucleotide polymorphisms to the outcome of allogeneic hematopoietic stem cell transplantation. *Hum Immunol*. 2009;70(6):457-460.
- Arora M, Lindgren B, Basu S, et al. Polymorphisms in the base excision repair pathway and graft-versus-host disease. *Leukemia*. 2010;24(8):1470-1475.
- Azarian M, Busson M, Lepage V, et al. Donor CTLA-4 +49 A/G*GG genotype is associated with chronic GVHD after HLA-identical haematopoietic stem-cell transplantations. *Blood*. 2007;110(13):4623-4624.
- Broen K, van der Waart AB, Grepink-Draaisma A, et al. Polymorphisms in CCR6 are associated with chronic graft-versus-host disease and invasive fungal disease in matched-related hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant*. 2011;17(10):1443-1449.
- Clark WB, Brown-Gentry KD, Crawford DC, et al. Genetic variation in recipient B-cell activating factor modulates phenotype of GVHD. *Blood*. 2011;118(4):1140-1144.
- Harkensee C, Oka A, Onizuka M, et al; Japan Marrow Donor Program. Single nucleotide polymorphisms and outcome risk in unrelated mismatched hematopoietic stem cell transplantation: an exploration study. *Blood*. 2012;119(26):6365-6372.
- Espinoza LJ, Takami A, Nakata K, et al; Japan Marrow Donor Program. Genetic variants of human granzyme B predict transplant outcomes after HLA matched unrelated bone

- marrow transplantation for myeloid malignancies. *PLoS One*. 2011; 6(8):e23827.
8. Kim DH, Lee NY, Sohn SK, et al. IL-10 promoter gene polymorphism associated with the occurrence of chronic GVHD and its clinical course during systemic immunosuppressive treatment for chronic GVHD after allogeneic peripheral blood stem cell transplantation. *Transplantation*. 2005;79(11):1615-1622.
 9. Kim DH, Jung HD, Lee NY, Sohn SK. Single nucleotide polymorphism of CC chemokine ligand 5 promoter gene in recipients may predict the risk of chronic graft-versus-host disease and its severity after allogeneic transplantation. *Transplantation*. 2007;84(7):917-925.
 10. Kim DD, Yun J, Won HH, et al. Multiple single-nucleotide polymorphism-based risk model for clinical outcomes after allogeneic stem-cell transplantation, especially for acute graft-versus-host disease. *Transplantation*. 2012;94(12):1250-1257.
 11. Mullighan C, Heatley S, Doherty K, et al. Non-HLA immunogenetic polymorphisms and the risk of complications after allogeneic hemopoietic stem-cell transplantation. *Transplantation*. 2004;77(4):587-596.
 12. Rocha V, Franco RF, Porcher R, et al. Host defense and inflammatory gene polymorphisms are associated with outcomes after HLA-identical sibling bone marrow transplantation. *Blood*. 2002; 100(12):3908-3918.
 13. Rocha V, Porcher R, Fernandes JF, et al. Association of drug metabolism gene polymorphisms with toxicities, graft-versus-host disease and survival after HLA-identical sibling hematopoietic stem cell transplantation for patients with leukemia. *Leukemia*. 2009;23(3):545-556.
 14. Shimada M, Onizuka M, Machida S, et al. Association of autoimmune disease-related gene polymorphisms with chronic graft-versus-host disease. *Br J Haematol*. 2007;139(3):458-463.
 15. Sivula J, Turpeinen H, Volin L, Partanen J. Association of IL-10 and IL-10Rbeta gene polymorphisms with graft-versus-host disease after haematopoietic stem cell transplantation from an HLA-identical sibling donor. *BMC Immunol*. 2009;10:24.
 16. Socié G, Loiseau P, Tamouza R, et al. Both genetic and clinical factors predict the development of graft-versus-host disease after allogeneic hematopoietic stem cell transplantation. *Transplantation*. 2001;72(4):699-706.
 17. Ostrovsky O, Shimoni A, Rand A, Vlodavsky I, Nagler A. Genetic variations in the heparanase gene (HPSE) associate with increased risk of GVHD following allogeneic stem cell transplantation: effect of discrepancy between recipients and donors. *Blood*. 2010;115(11):2319-2328.
 18. Vannucchi AM, Guidi S, Guglielmelli P, et al. Significance of CTLA-4 and CD14 genetic polymorphisms in clinical outcome after allogeneic stem cell transplantation. *Bone Marrow Transplant*. 2007;40(10):1001-1002.
 19. Viel DO, Tsuneto LT, Sossai CR, et al. IL2 and TNFA gene polymorphisms and the risk of graft-versus-host disease after allogeneic haematopoietic stem cell transplantation. *Scand J Immunol*. 2007;66(6):703-710.
 20. Leisenring WM, Martin PJ, Petersdorf EW, et al. An acute graft-versus-host disease activity index to predict survival after hematopoietic cell transplantation with myeloablative conditioning regimens. *Blood*. 2006;108(2):749-755.
 21. Lee SJ, Vogelsang G, Flowers ME. Chronic graft-versus-host disease. *Biol Blood Marrow Transplant*. 2003;9(4):215-233.
 22. Flowers ME, Inamoto Y, Carpenter PA, et al. Comparative analysis of risk factors for acute graft-versus-host disease and for chronic graft-versus-host disease according to National Institutes of Health consensus criteria. *Blood*. 2011;117(11):3214-3219.
 23. Patterson N, Price AL, Reich D. Population structure and eigenanalysis. *PLoS Genet*. 2006; 2(12):e190.
 24. Hahn T, Sucheston-Campbell LE, Preus L, et al. Establishment of definitions and review process for consistent adjudication of cause-specific mortality after allogeneic unrelated-donor hematopoietic cell transplantation. *Biol Blood Marrow Transplant*. 2015;21(9):1679-1686.
 25. Sucheston-Campbell LE, Clay A, McCarthy PL, et al. Identification and utilization of donor and recipient genetic variants to predict survival after HCT: are we ready for primetime? *Curr Hematol Malign Rep*. 2015;10(1):45-58.
 26. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics*. 2010;26(17):2190-2191.
 27. Kochi Y, Okada Y, Suzuki A, et al. A regulatory variant in CCR6 is associated with rheumatoid arthritis susceptibility. *Nat Genet*. 2010;42(6):515-519.
 28. Bush WS, Moore JH. Chapter 11: Genome-wide association studies. *PLOS Comput Biol*. 2012; 8(12):e1002822.
 29. Zollner S, Pritchard JK. Overcoming the winner's curse: estimating penetrance parameters from case-control data. *Am J Hum Genet*. 2007;80(4):605-615.
 30. Ioannidis JP. Why most published research findings are false. *PLoS Med*. 2005;2(8):e124.
 31. Chien JW, Zhang XC, Fan W, et al. Evaluation of published single nucleotide polymorphisms associated with acute GVHD. *Blood*. 2012; 119(22):5311-5319.
 32. Inamoto Y, Martin PJ, Flowers MED, et al. Genetic risk factors for sclerotic graft-versus-host disease. *Blood*. 2016;128(11):1516-1524.
 33. Jagasia MH, Greinix HT, Arora M, et al. National Institutes of Health Consensus Development Project on Criteria for Clinical Trials in Chronic Graft-versus-Host Disease: I. The 2014 Diagnosis and Staging Working Group report. *Biol Blood Marrow Transplant*. 2015;21(3):389-401.
 34. Jostins L, Ripke S, Weersma RK, et al; International IBD Genetics Consortium (IIBDGC). Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature*. 2012;491(7422):119-124.
 35. Barrett JC, Clayton DG, Concannon P, et al; Type 1 Diabetes Genetics Consortium. Genome-wide association study and meta-analysis find that over 40 loci affect risk of type 1 diabetes. *Nat Genet*. 2009;41(6):703-707.
 36. Qu HQ, Bradfield JP, Li Q, et al. In silico replication of the genome-wide association results of the Type 1 Diabetes Genetics Consortium. *Hum Mol Genet*. 2010;19(12):2534-2538.
 37. Stahl EA, Raychaudhuri S, Remmers EF, et al; BIRAC Consortium; YEAR Consortium. Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. *Nat Genet*. 2010;42(6):508-514.