

the discovery cohort of COG white patients, back of the envelope calculations indicate an osteonecrosis incidence of 21.5% for affected patients with G/A or A/A and 10.7% for G/G patients (see Table 1 in Karol et al). The polymorphism accounts for ~17% of cases of osteonecrosis, ~1 case in 6. White patients with the G/G genotype have 109 cases in place of the expected 132 cases, a reduction in incidence from 12.9% to 10.7%, and remain at substantial risk for symptomatic osteonecrosis.

Karol et al are impressive for their sample size and scientific rigor. They present compelling arguments for biological plausibility. However, no prior genome-wide association study investigation has linked glutamate receptor genetic variations and osteonecrosis. A variety of other plausible polymorphisms have been inconsistently implicated with similar hazard ratios, involving PAI-1, glucocorticoid metabolism, antifolate pharmacodynamics, fibrinolysis, and lipid and albumin homeostasis.⁵⁻⁸ Few candidate polymorphisms appear in >1 report. As with all retrospective studies, prospective confirmation is needed.⁹

Conflict-of-interest disclosure: P.S.G. serves as a consultant and is on the speakers' bureau for Jazz and Sigma Tau Pharmaceuticals and is on a Data and Safety Monitoring Committee for Bristol Meyers Squibb. ■

REFERENCES

1. Karol SE, Yang W, Van Driest SL, et al. Genetics of glucocorticoid-associated osteonecrosis in children with acute lymphoblastic leukemia. *Blood* 2015;126(15):1770-1776.
2. Mattano LA Jr, Sather HN, Trigg ME, Nachman JB. Osteonecrosis as a complication of treating acute lymphoblastic leukemia in children: a report from the Children's Cancer Group. *J Clin Oncol*. 2000;18(18):3262-3272.
3. Mattano LA Jr, Devidas M, Nachman JB, et al; Children's Oncology Group. Effect of alternate-week versus continuous dexamethasone scheduling on the risk of osteonecrosis in paediatric patients with acute lymphoblastic leukaemia: results from the CCG-1961 randomised cohort trial. *Lancet Oncol*. 2012;13(9):906-915.
4. Te Winkel ML, Pieters R, Wind EJ, Bessems JH, van den Heuvel-Eibrink MM. Management and treatment of osteonecrosis in children and adolescents with acute lymphoblastic leukemia. *Haematologica*. 2014;99(3):430-436.
5. French D, Hamilton LH, Mattano LA Jr, et al; Children's Oncology Group. A PAI-1 (SERPINE1) polymorphism predicts osteonecrosis in children with acute lymphoblastic leukemia: a report from the Children's Oncology Group. *Blood*. 2008;111(9):4496-4499.
6. Bond J, Adams S, Richards S, Vora A, Mitchell C, Goulden N. Polymorphism in the PAI-1 (SERPINE1)

gene and the risk of osteonecrosis in children with acute lymphoblastic leukemia. *Blood*. 2011;118(9):2632-2633.

7. Kawedia JD, Kaste SC, Pei D, et al. Pharmacokinetic, pharmacodynamic, and pharmacogenetic determinants of osteonecrosis in children with acute lymphoblastic leukemia. *Blood*. 2011;117(8):2340-2347.

8. Relling MV, Yang W, Das S, et al. Pharmacogenetic risk factors for osteonecrosis of the hip among

children with leukemia. *J Clin Oncol*. 2004;22(19):3930-3936.

9. Ioannidis JPA. How to make more published research true. *PLoS Med*. 2014;11(10):e1001747.

DOI 10.1182/blood-2015-08-665067

© 2015 by The American Society of Hematology

● ● ● LYMPHOID NEOPLASIA

Comment on Song et al, page 1813

Restoring Ikaros's wings to solve a leukemia maze

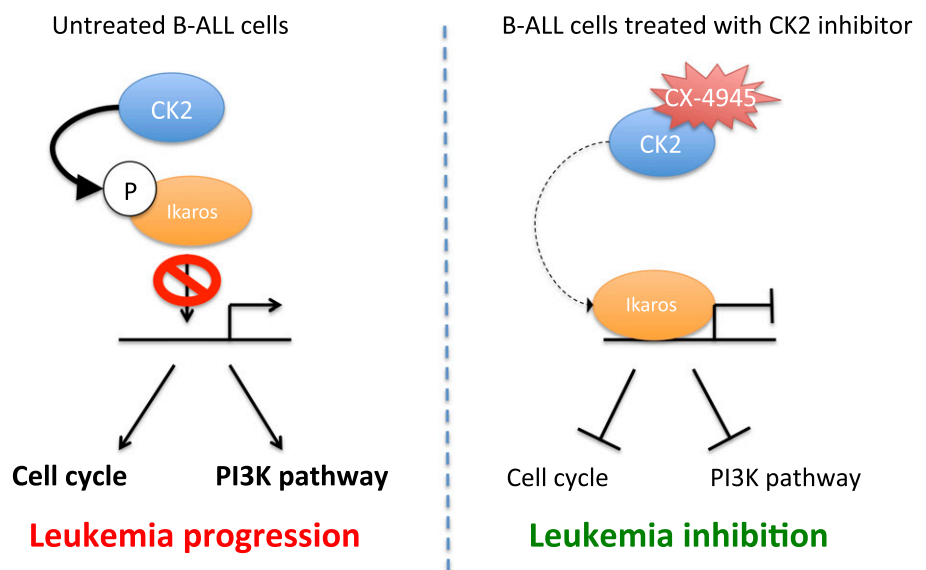
Camille Lobry INSTITUT GUSTAVE ROUSSY

In this issue of *Blood*, Song et al show that tumor suppressor activity of Ikaros is achieved through repression of cell cycle and phosphatidylinositol-3 (PI3) kinase pathway genes and can be reactivated through pharmacologic inhibition of casein kinase 2 (CK2) to eradicate disease in high-risk B-cell acute lymphoblastic leukemia (B-ALL).¹

B-ALL is the most common leukemia diagnosed during childhood. Although frontline risk-adapted chemotherapies have improved overall survival, nearly 20% of children and >50% of adults relapse.² Therefore, this disease remains one of the leading causes of leukemia death. In recent years, extensive genomic profiling has led

to better classification and understanding of high-risk B-ALL.³ Elucidation of the mechanisms involving these genetic alterations in the pathogenesis of B-ALL could lead to the design of specific therapies for the most refractory subgroups.

Among these genetic alterations, the *IKZF1* gene encoding the Ikaros transcription factor



The two "wings" of Ikaros in B-ALL suppression. (Right) In B-ALL cells, CK2 activity is increased and results in Ikaros phosphorylation that prevents its binding to DNA and regulation of cell cycle and PI3K pathway genes. (Left) On treatment with the specific CK2 inhibitor CX-4945, Ikaros is no longer phosphorylated and can actively repress its target genes, leading to leukemia inhibition.

was found to be affected in ~15% of all B-ALL patients. Strikingly, *IKZF1* mutations were found in 70% to 80% of Philadelphia chromosome-positive B-ALL, where it is associated with poor outcome.⁴ Ikaros is a zinc finger transcription factor characterized in the early 1990s that plays an essential role in the development of lymphoid lineages.⁵ Ikaros was identified as a tumor suppressor in lymphoid malignancies, as deletion of 1 *Ikaros* allele in mice resulted in T-cell leukemia.⁶ *IKZF1* alterations include deletions or point mutations of a single allele, resulting in loss of function, haplo-insufficiency, or expression of a dominant negative form. Previous attempts to identify direct transcriptional targets of Ikaros focused on normal lymphoid development in mice⁷ and not in human leukemia. To determine genome-wide occupancy of Ikaros in human B-ALL, Song et al¹ performed chromatin immunoprecipitation followed by massively parallel sequencing (ChIP-seq) in a B-ALL cell line and primary B-ALL samples. They identified Ikaros binding on promoters of genes involved in cell cycle regulation such as cyclins (*CCND3*, *CCNE2*) or cyclin-dependent kinases (*CDK2*, *CDK6*) and genes involved in the phosphatidylinositol pathway such as *PIK3CD* or *PIK3C2B*, among others. Using gain-of-function experiments overexpressing Ikaros or loss-of-function experiments with short hairpin RNA (shRNA) directed against Ikaros mRNA, they demonstrated that Ikaros directly represses expression of these genes. These 2 pathways are essential for proliferation of leukemic cells, and these experiments may define the tumor suppressor activity of Ikaros in B-ALL (see figure). However, progression and development of B-ALL occur even in the presence of 2 normal alleles of Ikaros, suggesting that other posttranscriptional or posttranslational regulatory mechanisms are involved.

Previous studies have shown that phosphorylation of Ikaros by CK2 inhibits its DNA binding⁸ and that CK2 activity is commonly upregulated in B-ALL.⁹ Therefore, the authors tested whether CK2 could directly affect Ikaros activity in B-ALL. They first showed that CK2 activity is indeed increased in primary B-ALL cells compared with normal precursor B cells. Inhibition of CK2 with anti-CK2 shRNA or a pharmacologic inhibitor

of CK2, CX-4945, resulted in downregulation of Ikaros target genes. Moreover, they demonstrated that this transcriptional repression required Ikaros activity because knockdown of Ikaros reverted CK2 inhibitor-induced gene repression. Using a NALM6 cell line xenograft model, they found that treatment with CX-4945 induced repression of Ikaros target genes in vivo and significantly increased survival of the animals. These findings paralleled earlier studies addressing the efficiency of this inhibitor in B-ALL.⁹ The most novel and striking finding of the work by Song et al is the efficacy of CX-4945 in high-risk B-ALL where 1 *IKZF1* allele is deleted. In primary patient samples, treatment with the CK2 inhibitor in vitro resulted in increased Ikaros binding to and repression of its target genes. The authors went on to validate these findings in patient-derived xenograft (PDX) models of high-risk B-ALL using 3 different patient samples presenting with different characteristics: CRLF2 overexpression, high blood count with 95% blasts, or *IKZF1* deletion. After disease establishment, animals were treated for 22 days with CX-4945 inhibitor or vehicle. Cohorts treated with CK2 inhibitor had increased leukemic cell death, decreased infiltration of the bone marrow and spleen, and significantly increased overall survival. Taken together, the experiments by Song et al reveal that the tumor suppressor activity of Ikaros is reduced in high-risk B-ALL due to phosphorylation by CK2 and highlight the therapeutic potential of CK2 inhibitors in this lethal disease.

Recently, CK2 inhibitors have been extended from solid tumors to hematologic malignancies, including chronic lymphocytic leukemia, Hodgkin and non-Hodgkin lymphomas, ALL, and acute myeloid leukemia.¹⁰ This work suggests that CK2 inhibition may be an effective strategy in the treatment of high-risk B-ALL, particularly subsets presenting with *IKZF1* deletion. However, several questions remain. Although the authors find that CX-4945 treatment is well tolerated in their preclinical model, it was only administered for a limited time with a modest gain in overall survival. As CK2 plays numerous essential functions in other tissues, the tolerability and consequences of extended treatment with this inhibitor remain to be seen. Using PDX models, it would be interesting to

examine a possible synergy between moderate doses of CX-4945 and conventional chemotherapies no longer effective on high-risk B-ALL. Finally, as the loss of one *IKZF1* allele is encountered in treatment-refractory high-risk B-ALL, would forced reactivation of the remaining *IKZF1* allele trigger either loss of this allele or selection of minor clones already deleted for both alleles? Nonetheless, results of these studies by Song et al suggest that CK2 inhibition represents a promising therapeutic strategy for the management of high-risk B-ALL and warrants further evaluation of its clinical potential.

Conflict-of-interest disclosure: The author declares no competing financial interests. ■

REFERENCES

1. Song C, Gowda C, Pan X, et al. Targeting casein kinase II restores Ikaros tumor suppressor activity and demonstrates therapeutic efficacy in high-risk leukemia. *Blood*. 2015;126(15):1813-1822.
2. Hunger SP, Lu X, Devidas M, et al. Improved survival for children and adolescents with acute lymphoblastic leukemia between 1990 and 2005: a report from the Children's Oncology Group. *J Clin Oncol*. 2012;30(14):1663-1669.
3. Hunger SP, Mullighan CG. Redefining ALL classification: toward detecting high-risk ALL and implementing precision medicine. *Blood*. 2015;125(26):3977-3987.
4. Mullighan CG, Su X, Zhang J, et al; Children's Oncology Group. Deletion of *IKZF1* and prognosis in acute lymphoblastic leukemia. *N Engl J Med*. 2009;360(5):470-480.
5. Georgopoulos K, Bigby M, Wang JH, et al. The Ikaros gene is required for the development of all lymphoid lineages. *Cell*. 1994;79(1):143-156.
6. Winandy S, Wu P, Georgopoulos K. A dominant mutation in the Ikaros gene leads to rapid development of leukemia and lymphoma. *Cell*. 1995;83(2):289-299.
7. Ferreirós-Vidal I, Carroll T, Taylor B, et al. Genome-wide identification of Ikaros targets elucidates its contribution to mouse B-cell lineage specification and pre-B-cell differentiation. *Blood*. 2013;121(10):1769-1782.
8. Gómez-del Arco P, Maki K, Georgopoulos K. Phosphorylation controls Ikaros's ability to negatively regulate the G₁-S transition. *Mol Cell Biol*. 2004;24(7):2797-2807.
9. Gomes AM, Soares MV, Ribeiro P, et al. Adult B-cell acute lymphoblastic leukemia cells display decreased PTEN activity and constitutive hyperactivation of PI3K/Akt pathway despite high PTEN protein levels. *Haematologica*. 2014;99(6):1062-1068.
10. Chon HJ, Bae KJ, Lee Y, Kim J. The casein kinase 2 inhibitor, CX-4945, as an anti-cancer drug in treatment of human hematological malignancies. *Front Pharmacol*. 2015;6:70.

DOI 10.1182/blood-2015-08-662544

© 2015 by The American Society of Hematology