LYMPHOID NEOPLASIA

A genome-wide association study of susceptibility to acute lymphoblastic leukemia in adolescents and young adults

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Key Points

- In this first ALL GWAS in AYAs, we determined that inherited GATA3 variants strongly influence ALL susceptibility in this age group.
- These findings revealed similarities and differences in the genetic basis of ALL susceptibility between young children and AYAs.

Acute lymphoblastic leukemia (ALL) in adolescents and young adults (AYA) is characterized by distinct presenting features and inferior prognosis compared with pediatric ALL. We performed a genome-wide association study (GWAS) to comprehensively identify inherited genetic variants associated with susceptibility to AYA ALL. In the discovery GWAS, we compared genotype frequency at 635 297 single nucleotide polymorphisms (SNPs) in 308 AYA ALL cases and 6,661 non-ALL controls by using a logistic regression model with genetic ancestry as a covariate. SNPs that reached $P \le 5 \times 10^{-8}$ in GWAS were tested in an independent cohort of 162 AYA ALL cases and 5,755 non-ALL controls. We identified a single genome-wide significant susceptibility locus in *GATA3*: rs3824662, odds ratio (OR), 1.77 ($P = 2.8 \times 10^{-10}$) and rs3781093, OR, 1.73 ($P = 3.2 \times 10^{-9}$). These findings were validated in the replication cohort. The risk allele at rs3824662 was most frequent in Philadelphia chromosome (Ph)-like ALL but also conferred susceptibility to non-Ph-like ALL in AYAs. In 1,827 non-selected ALL cases, the risk allele frequency at this SNP was positively correlated with age at diagnosis

 $(P = 6.29 \times 10^{-11})$. Our results from this first GWAS of AYA ALL susceptibility point to unique biology underlying leukemogenesis and potentially distinct disease etiology by age group. (*Blood.* 2015;125(4):680-686)

Introduction

Cancer survival rates have been steadily increasing in the United States across age groups except for in adolescents and young adults (AYA; age 16 to 39 years), partly because of the persisting inferior treatment response in hematologic malignancies.¹ Particularly with acute lymphoblastic leukemia (ALL), age as a continuous variable is negatively

correlated with prognosis in spite of risk-adapted combination chemotherapy.² In an analysis of 21 626 ALL cases diagnosed between 1990 and 2005 and treated on Children's Oncology Group (COG) frontline protocols, survival rates decreased significantly with increasing age at diagnosis regardless of treatment era (eg, 94.1% for

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Figure 1. GWAS study design. ALL susceptibility variants were identified by comparing SNP genotype frequency in AYA ALL cases compared with non-ALL controls in the discovery GWAS, followed by replication. CALGB: The Alliance-Cancer and Leukemia Group B; COG: Children's Oncology Group; ECOG: Eastern Cooperative Oncology Group; MDACC: MD Anderson Cancer Center; SJ: St. Jude Children's Research Hospital.



age 1 to 10, 84.7% for age 10 to 15, and 75.9% for age 15 to 22 years in the 2000-2005 cohort).³ Although pediatric-based treatment regimens have been tested in AYA populations and have resulted in improved survival, the gap in treatment outcome between age groups persists, and ALL remains one of the leading causes of cancer-related deaths in the AYA population.^{4,5}

The inferior prognosis of AYA is likely to be multifactorial and includes socioeconomic factors, medication adherence, clinical trial enrollment and, importantly, age-related differences in ALL tumor and host biology.⁶ For example, as age increases, there is a progressive rise in prevalence of ALL genetic subtypes with poor prognosis such as Philadelphia chromosome–positive (Ph+),⁷ Ph-like,⁸ or intrachromosomal amplification of chromosome 21,⁹ whereas subtypes with favorable outcome (high hyperdiploidy¹⁰ and *ETV6-RUNX1*¹¹) become less common. These comparisons are informative but are also limited because they are primarily driven by ALL features discovered in children and/or older adults. As a result, differences in tumor biology between AYA and childhood ALL may have been underestimated, and genomic profiling studies focusing on the AYAs are likely to reveal novel molecular features unique to this population.

Inherited genetic variations can strongly influence both the susceptibility to ALL¹²⁻¹⁵ and treatment outcomes.¹⁶⁻²⁰ For example, genome-wide association studies (GWASs) have identified germline genetic variants at ARID5B, IKZF1, CEBPE, PIP4K2A, and CDKN2A/ CDKN2B loci with substantial cumulative effects on ALL disease risk in children. These ALL susceptibility genes are involved in lymphoid cell development, cell cycle control, and tumor suppression, collectively affecting leukemogenesis. Although one's inherited genetic variants remain unchanged over a lifetime, it is possible that the effects of these susceptibility variants vary by age, thus contributing to the agerelated differences in ALL incidence and subtype. In fact, when we examined the risk of ALL conferred by the ARID5B variant, there was a clear trend of diminishing effects with increasing age (ie, allelic odds ratio of 2.01, 1.8, and 1.48 in children younger than age 5, 5 to 10, and older than 10 years, respectively).¹⁵ However, germline variants related to ALL risk in the AYA population have not been comprehensively examined.

To better understand the potential unique leukemia etiology in AYAs, we conducted the first GWAS to systemically interrogate

germline single nucleotide polymorphisms (SNPs) for their contribution to ALL risk in this age group.

Methods

Study design and patients

In the discovery GWAS, the ALL cases consisted of 209 adolescents (median age, 17.4 years; range, 16 to 21 years) and 99 young adults (median age, 24.3 years; range, 21 to 39 years) with newly diagnosed B-cell ALL who were treated on the Children's Oncology Group (COG; N = 202),²¹ the Alliance-Cancer and Leukemia Group B (N = 56),²² Eastern Cooperative Oncology Group E2993 (N = 29),²³ MD Anderson Cancer Center (N = 11)²⁴⁻²⁷, and St. Jude Children's Research Hospital (N = 10) trials.²⁸ The subjects were chosen on the basis of the availability of germline DNA, which was extracted from peripheral blood samples during clinical remission (<5% blasts cells in bone marrow). A total of 6,661 unrelated subjects from the Multi-Ethnic Study of Atherosclerosis (MESA) cohort (dbGaP phs000209.v9) were considered as non-ALL control subjects because the prevalence of adult survivors of childhood ALL is extremely low.¹⁴

For the replication analyses, 162 children with ALL age 16 to 21 years from the COG P9900 protocols (COG P9905 [NCT00005596], COG P9904 [NCT00005585²⁹], COG P9906 [NCT00005603³⁰], and COG AALL0232 [NCT00075725²¹]) were included. A set of 5755 unrelated non-ALL controls were included in the replication analysis: 1228 African Americans (AAs) from the AIDS Linked to Intravenous Experience (ALIVE) cohort,³¹ 880 Hispanic Americans from the Genetics of Asthma in Latino Americans (GALA) study,³² and 3647 European Americans (EAs) from the Genetic Association Informative Network (GAIN) schizophrenia cohort (dbGAP phs000021.v3.p2)³³ and GAIN bipolar cohort (phs000017.v3.p1³⁴; Figure 1).

The clinical trials were approved by local institutional review boards, and informed consent for trial enrollment and banking of specimens for future research was obtained from parents, guardians, or patients, as appropriate. This study was approved by the St. Jude Children's Research Hospital institutional review board.

Genotyping and quality control

Genome-wide SNP genotyping was performed by using the Affymetrix Human SNP Array 6.0 for ALL cases in the discovery GWAS, those in the COG P9905, P9904, and AALL0232 cohorts, and for all non-ALL controls (dbGaP MESA, ALIVE, GAIN, and GALA). Genotype calls (coded as 0, 1, and 2 for AA, AB,

Table 1. Association of GATA3 SNPs with AYA ALL susceptibility in the o	discover	y GWAS an	d replication	cohort
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					BAFS in	Genotype count in cases‡			RAF§ in	Genotype count in controls‡			AYA ALL vs non-ALL		
Chr	Position*	SNP	Alleles†	Cohort	AYA cases (%)	RR	RW	ww	controls (%)	RR	RW	ww	<i>P</i> II	ORII	95% CI
10	8144214	rs3824662	A/C	Discovery	35	36	145	125	20	341	2078	4242	$2.84 imes10^{-10}$	1.77	1.48-2.12
				Replication	39	24	78	59	18	226	1651	3876	$1.52 imes10^{-10}$	2.21	1.72-2.83
10	8141933	rs3781093	C/T	Discovery	33	34	137	136	22	380	2254	4025	$3.20 imes10^{-9}$	1.73	1.44-2.08
				Replication	35	18	75	64	18	236	1702	3809	$1.00 imes 10^{-7}$	1.96	1.52-2.54

Association of SNP genotype and ALL was evaluated by comparing allele frequency between ALL and non-ALL, after adjusting for genetic ancestry. Chr, chromosome.

*Chromosomal locations are based on hg18.

†Bold indicates risk allele for ALL.

‡Genotype is denoted by RR (homozygous for the risk allele), RW (heterozygous), or WW (homozygous for the wild-type allele).

§RAF, risk allele frequency (allele A at rs38246623 and allele C at rs3781093).

IIP values were estimated by the logistic regression test and OR represents the increase in risk of developing ALL for each copy of the risk allele compared with participants who don't carry the risk allele.

and BB genotypes) were determined by the Birdseed v2 (Affymetrix SNP 6.0) algorithm.³⁵ Samples for which genotypes were ascertained for less than 95% of SNPs on the array were deemed to have failed and were excluded from the analyses. For the ALL cases in the COG P9906 trial, genome-wide SNP genotyping was performed by using Affymetrix Human SNP Array 500K, and *GATA3* SNPs were genotyped by polymerase chain reaction and Sanger sequencing, as described previously.³⁶ We did not observe evidence of potential genotyping errors in the germline DNA because of tumor cell contamination (data not shown).

Prior to GWAS, SNPs were subjected to a series of quality control steps (supplemental Figure 1, available on the *Blood* Web site). First, we filtered SNPs on the basis of minor allele frequency (MAF) and SNP call rate: for SNPs with an MAF of 1% to 3%, we excluded those with a call rate <99%; for SNPs with an MAF of 5%, we excluded those with a call rate <98%; for SNPs with an MAF of >5%, we excluded those with a call rate <95%. An additional filtering step was applied in the GWAS involving non-ALL controls: we removed SNPs for which genotype frequencies differed significantly among control groups (ie, dbGaP MESA vs HapMap unrelated CEU or dbGaP MESA vs the GAIN bipolar cohort [dbGaP phs000017.v3]³³; $P < 10^{-6}$ by χ^2 test), and the comparison was restricted to EAs. Finally, those SNPs deviating from Hardy-Weinberg equilibrium (P < .01 in EA cases or controls) were also excluded from the analysis. After quality control filters were applied, 635 297 SNPs were included in the GWAS.

Genetic ancestry and population structure

Genetic ancestry was determined by using STRUCTURE (version 2.2.3),³⁷ based on genotypes at 30 000 SNPs randomly selected from the Affymetrix SNP arrays. HapMap samples from descendants of Northern Europeans (CEU; N = 60), West Africans (YRI; N = 60), East Asians (CHB/JPT; N = 90), and Native Americans (NAs; N = 105)³⁸ references were used to represent European, African, Asian, and Native American ancestries, respectively. We assumed that these four ancestries summed to 100% in each genotyped individual. EAs, AAs, NAs, and Asians were defined as having >95% European genetic ancestry, >70% African ancestry, >90% NA ancestry, and >90% Asian ancestry, respectively. Hispanics were individuals for whom NA ancestry was >10% and greater than African ancestry (including genetically defined NAs). The rest of the subjects were grouped as "Others."

We also performed principal component analysis of ALL cases and controls in the discovery GWAS cohort, including all SNPs that passed the quality control and observed comparable population structure between cases and controls (supplemental Figure 2). In addition, we exhaustively examined potential relatedness within ALL cases and within controls included in the discovery GWAS by computing pairwise identity by descent probabilities. No evidence of first or second-degree relationships was identified.

ALL somatic genomic lesions

In the discovery cohort, the ALL genetic subtypes included high hyperdiploid (>50 chromosomes), *ETV6-RUNX1*, *TCF3-PBX1*, *MLL*-rearranged, *BCR-ABL1* (Ph⁺), and Ph-like (with or without *CRLF2* rearrangements). Ph-like and *ERG*-deregulated ALL were defined by Predictive Analysis of Microarrays.^{21,39} In the COG P9900 series, ALL subtypes included *ETV6-RUNX1*, *TCF3-PBX1*, hyperdiploid, and *MLL*-rearranged, with the remainder of cases considered as B-other. *GATA3* expression was quantified in 237 ALL blasts in 237 AYA cases, using Affymetrix U133A array.⁸

Statistical analysis

In the discovery GWAS, the association test between genotypes at each of the 635 297 SNPs and ALL susceptibility was tested by comparing genotype frequency between AYA ALL cases and non-ALL controls using a logistic regression test under an additive model, including European, African, and NA ancestry (as continuous variables) as covariates using PLINK (v1.07).⁴⁰ Population stratification was assessed by the construction of a quantile-quantile plot (supplemental Figure 3), and there was only a minimal inflation at the upper tail of the distribution ($\lambda = 1.02$). SNPs that reached the association $P \le 5 \times 10^{-8}$ in the discovery GWAS were evaluated in the independent replication series (1-tailed test). In both discovery and replication groups, we also tested *GATA3* SNPs separately in EAs, AAs, and Hispanic Americans.

R (version 2.15.1) statistical software was used for the rest of the analyses unless indicated otherwise. Statistical tests were chosen as appropriate and according to the phenotype distribution (eg, normally or binomially distributed for continuous or categorical variables, respectively). Associations of SNP genotype with somatic lesions and age were estimated by logistic regression and linear regression test, respectively, after adjusting for genetic ancestry. Associations of *GATA3* SNP genotype with *GATA3* gene expression was assessed by linear regression model, adjusting for genetic ancestry.

Results

AYA ALL GWAS

In the discovery GWAS, we compared genotype frequency at 635 297 SNPs between 308 AYA ALL cases and 6,661 non-ALL controls (Figure 1). After adjusting for genetic ancestry, only two SNPs at 10p14 within the *GATA3* gene reached genome-wide significance: rs3824662 (odds ratio [OR], 1.77; 95% confidence interval [CI], 1.48 to 2.12; $P = 2.84 \times 10^{-10}$) and rs3781093 (OR, 1.73; 95% CI, 1.44 to 2.08; $P = 3.20 \times 10^{-9}$; Table 1 and Figure 2). These two SNPs were in strong linkage disequilibrium ($r^2 = 0.94$; D' = 1 in HapMap CEU; supplemental Figure 4), representing a single susceptibility locus. The A allele at rs3824662 was significantly overrepresented in ALL cases compared with controls (35% vs 20%) and was consistent across race/ ethnicity (ie, EAs, 30% vs 17% [$P = 1.09 \times 10^{-5}$]; Hispanics, 50% vs 33% [P = .0008]; and AAs, 20% vs 10% [P = .07]; Figure 3A). rs3781093 was significantly associated with ALL risk in EAs and Hispanics, but not in individuals of African descent in whom it was

Figure 2. Genome-wide association of SNP genotype with ALL susceptibility in AYAs. The association between genotype and ALL susceptibility was evaluated by using a logistic regression model for 635 297 SNPs in 308 AYA ALL cases and 6661 non-ALL controls. *P* values (y-axis) were plotted against respective chromosomal position of each SNP (x-axis). Points above the blue horizontal line indicate SNPs achieving the genome-wide significant threshold ($P < 5 \times 10^{-8}$). Gene symbol was indicated for the *GATA3* locus at 10p14.



no longer in linkage disequilibrium ($r^2 = 0.006$; D' = 0.16) with rs3824662 (supplemental Figure 5A).

To validate the association signals at these *GATA3* SNPs, we tested an independent set of 162 AYA ALL cases enrolled in COG P9900 and ALL0232 protocols and an additional 5,755 non-ALL controls. In the replication analysis, risk alleles at both *GATA3* SNPs were consistently overrepresented in AYA ALL cases compared with non-ALL controls: rs3824662 (OR, 2.21; 95% CI, 1.72 to 2.83; $P = 1.52 \times 10^{-10}$) and rs3781093 (OR, 1.96; 95% CI, 1.52 to 2.54; $P = 1.0 \times 10^{-7}$; Table 1, Figure 3B, and supplemental Figure 5B). rs3824662 was validated across race/ethnicity in the replication group (ie, EAs, 35% vs 18% [$P = 2.0 \times 10^{-7}$]; Hispanics, 55% vs 39% [P = .005]; and AAs, 13% vs 9% [P = .035]; Figure 3B). In contrast, rs3781093 was significant in EAs and Hispanics but not in AAs (supplemental Figure 5B).

We also examined the association signals in AYAs for susceptibility loci previously identified in pediatric populations (supplemental Table 1). *ARID5B*, *IKZF1*, and *PIP4K2A* variants were nominally significant in AYAs in the discovery GWAS and/or in the replication analyses. In contrast, *CEBPE* and *CDKN2A/CDKN2B* were not associated with ALL risk in AYAs in either discovery or replication cohorts. These results imply both similarities and differences in genetic predisposition to ALL between children and AYAs.¹⁵

GATA3 SNP rs3824662 and ALL subtypes in AYAs

We further analyzed the association of the *GATA3* SNP rs3824662 with somatic ALL genomic abnormalities. Among the AYA ALL cases in the discovery cohort, the risk allele at rs3824662 was underrepresented among hyperdiploid ALL cases (22% vs 37%; P = .03; Figure 4 and supplemental Table 2), with a similar trend for *TCF3-PBX1* and *ETV6-RUNX1* ALL albeit not statistically significant. In contrast, the ALL risk allele frequency of rs3824662 was higher in AYA ALL cases with the Ph-like gene expression profile than in those without this signature (48% vs 32%; P = .02; Figure 4 and supplemental Table 2). This was consistent with our prior reports of *GATA3* as a susceptibility gene for Ph-like ALL,³⁶ although there was no overlap in cases included in the current AYA ALL GWAS and those in our previous Ph-like ALL GWAS.³⁶ Within Ph-like ALL, there was a trend with A allele further enriched in cases involving *CRLF2* rearrangements (P = .06; Figure 4).



Figure 3. Association of *GATA3* SNP rs3824662 with ALL in AYAs by race/ethnicity. In the discovery group (A) the A allele at rs3824662 was overrepresented in AYA ALL cases relative to non-ALL controls. This association was true within the European Americans (>95% European genetic ancestry), African Americans (>70% African ancestry), or Hispanic Americans (>10% Native American genetic ancestry, and Native American ancestry > African genetic ancestry). (B) Similar association was confirmed in the replication group (1-tailed test). Genetic ancestry was determined by using STRUCTURE (version 2.2.3) with HapMap CEU, YRI, CHB/JPT, and indigenous Native Americans as reference populations.



Figure 4. *GATA3* SNP genotype and ALL genetic subtypes in AYAs. The allele frequency of rs3824662 varied substantially by ALL somatic genomic abnormalities, with the ALL risk allele underrepresented in hyperdiploid cases and more common in the Ph-like subtype. Numbers are based on the ALL cases included in the discovery GWAS (N = 308). The frequency of A allele at rs3824662 was 20% among unrelated non-ALL controls (MESA).

Importantly, even after excluding Ph-like ALL cases, the risk allele at rs3824664 was still more common in AYA ALL cases compared with non-ALL controls (rs3824662: OR, 1.56 [95% CI, 1.25 to 1.96; $P = 8.13 \times 10^{-5}$]; rs3781093: OR, 1.53 [95% CI, 1.21 to 1.92; P = .0002]; supplemental Figure 6). This suggested that the influence of the *GATA3* variant on ALL susceptibility in AYAs extends beyond the predisposition to Ph-like subtype.

GATA3 SNP rs3824662 and age at ALL diagnosis

Finally, we examined the distribution of the *GATA3* SNP genotype by age at diagnosis in a cohort of largely unselected patients enrolled on the COG P9900 protocols (N = 1,827, age 0.1 to 21 years). When we divided patients into four consecutive age groups (<5, 5 to 10, 10 to 15, and >15 years), we observed a clear progressive increase in the risk allele frequency at rs3824662 ($P = 6.29 \times 10^{-11}$; Figure 5) with increasing allelic ORs (ie, relative risk of ALL conferred by each copy of the A allele at rs3824662; Figure 5, inset plot): 0.96 (95% CI, 0.85 to 1.09), 1.26 (95% CI, 1.08 to 1.48), 1.48 (95% CI, 1.19 to 1.84), and 2.40 (95% CI, 1.81 to 3.19). Similar correlation between *GATA3* genotype frequency and age was evident irrespective of genetic ancestry, but the *GATA3* risk allele was markedly more common in Hispanics (ie, individuals with high NA genetic ancestry; Figure 5). To examine

whether the association with age is confounded by ALL genetic subtype, we compared rs3824662 allele frequency by age in the COG P9900 protocols after stratifying ALL cases into *TCF3-PBX1*, *ETV6-RUNX1*, high hyperdiploid, *MLL*-rearranged, and B-other. There was a trend for the risk allele at this SNP to be more frequent in patients older than age 16 years relative to those younger than age 16 years in 5 subtypes examined, although with a limited sample size (supplemental Figure 7). This suggests that *GATA3* germline variants confer a general ALL disease risk in AYAs. In contrast, the frequency of ALL risk variant in *ARID5B* (rs10821936) decreased progressively with increasing age at diagnosis in the COG P9900 cohort (P = .006), whereas *PIP4K2A*, *CDKN2A/2B*, *IKZF1*, and *CEBPE* variants were not related to age (P > .05; data not shown).

Discussion

Because ALL is the most common cancer in children, previous susceptibility GWAS studies understandably focused on pediatric populations. We hypothesized that ALL in AYAs has distinct tumor biology and genetic etiology, which potentially contribute to the disparities in treatment





Figure 5. *GATA3* SNP rs3824662 and age at ALL diagnosis. In a largely unselected cohort of ALL cases enrolled on the Children's Oncology Group (COG) P9900 trials (N = 1,827), the frequency of ALL risk allele at rs3824662 was positively correlated with patient age at diagnosis consistently across race/ethnicity. Inset figure: the relative risk of ALL (odds ratio) conferred by each copy of the A allele at rs3824662 increased progressively with age, as estimated by logistic regression after adjusting for genetic ancestry. Horizontal dotted line is odds ratio of 1. Unrelated participants from the Multi-ethnic Study of Atherosclerosis (MESA) were considered as non-ALL controls.

outcomes by age. To this end, we performed the first GWAS of ALL susceptibility specifically in the AYA population and identified a single genome-wide significant risk locus within the *GATA3* gene on 10p14.

The susceptibility to ALL varies substantially by age. ALL risk first peaks between 2 and 5 years after birth, followed by gradual decrease into adulthood, but rises again in older individuals (older than age 70 years), suggesting that differential combinations of environmental and genetic factors contribute to leukemogenesis at different ages. For example, it has been hypothesized that infection (and supposedly acquired immunity) may ameliorate susceptibility to ALL in young children, 41,42 which may not be important in ALL that occurs later in life. Similarly, the in utero occurrence of genomic lesions is characteristic in many (if not most) pediatric ALL cases,^{43,44} whereas such early origin of presumed initiating events may not be evident in AYAs ALL. Agedependent differences in lymphocyte development and function are well documented in human and mouse systems,⁴⁵ and rapid growth of hematopoietic cells may render them particularly susceptible to oncogenic assaults.⁴⁶ Thus, it can be postulated that specific ALL susceptibility genes are required during a particular stage of hematopoietic development and preferentially influence ALL risk within a certain age range. For example, loss of Arid5b in mice resulted in reduction of lymphoid cells in bone marrow within 3 weeks after birth, but the effect became blunted by 6 weeks.⁴⁷ In fact, germline ARID5B variants also exhibited increasing influence on ALL predisposition in children as age decreases.¹⁵

GATA3 encodes for a transcription factor critical for lymphoid cell lineage commitment and early T-cell differentiation,48 and loss-offunction somatic mutations have been discovered in early T-cell precursor ALL.⁴⁹ Germline polymorphisms in GATA3, however, appear more important for B-cell malignancies.⁵⁰ We recently reported that rs3824662 was significantly associated with susceptibility to Ph-like ALL in children and risk of relapse.³⁶ A contemporaneous study by Migliorini et al¹⁷ reported the same ALL susceptibility variant in GATA3 in children of European descent and associated it with relapse. Particularly of note, GATA3 risk variants also appeared enriched in older children, even within their predominantly pediatric cohort. The association of rs3824662 with ALL relapse^{17,36} is in line with the negative prognosis by age and higher frequency of the GATA3 variant in AYAs with ALL. Nevertheless, it is unclear whether poor prognosis conferred by a GATA3 variant was driven by its association with a highrisk subtype (ie, Ph-like ALL), novel somatic genomic aberrations specific to AYA, and/or host biology related to antileukemic drug response. Interestingly, in AYA cases included in the discovery GWAS, the number of the risk allele at rs3824662 was significantly associated with GATA3 expression in ALL blasts (P = .02; supplemental Figure 8), consistent with our previous report in pediatric ALL cases of this variant functioning as a *cis*-acting regulatory element of GATA3 transcription.³⁶

The overrepresentation of the GATA3 variant in AYAs is consistent with its association with Ph-like ALL³⁶ for which the frequency increases with age.⁸ However, the risk variant at rs3824662 remained significantly associated with susceptibility to AYA ALL cases without Ph-like expression pattern, suggesting the link to Ph-like ALL contributed only partly to the genome-wide significant association signal at rs3824662. In fact, the GATA3 risk allele tended to be more common in ALL patients age 16 years or older than in those age younger than 16 years consistently across different genetic subtypes, plausibly conferring a general ALL risk in AYAs. It remains unknown how the GATA3 variants influence the risk of developing ALL in older adults, including the elderly (>60 years). Future studies including this age group may provide insights on molecular etiology of ALL across the age spectrum. It is also noteworthy that MLLrearranged cases had the second highest GATA3 risk variant frequency (Figure 4), although the number of patients was relatively small and the difference did not reach statistical significance. Future studies are warranted to comprehensively characterize potential interactions of germline *GATA3* variants with somatic genomic lesions in ALL.

In conclusion, our GWAS identified inherited *GATA3* genetic variants that strongly influence ALL susceptibility in adolescents and young adults, shedding new light on potential age-related differences in ALL biology and treatment outcome.

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Authorship

Contribution: V.P.-A., K.G.R., H.X., C.G.M., and J.J.Y. conceived of and designed the study; R.C.H., D.P.-T., I-M.C., W.L.C., N.A.H., A.J.C., E.A.R., J.M.G.-F., G.M., C.D.B., K.M., J.K., W.S., S.M.K., M.K., E.P., J.M.R., S.M.L., M.S.T., M.D., E.G.B., D.G.T., F.Y., Y.W., C.-H.P., S.J., M.V.R., W.E.E., D.S.G., M.L.L., S.P.H., and C.L.W. provided study materials or patients; V.P.-A., K.G.R., H.X., M.D., I-M.C., C.L.W., R.C.H., M.V.R., and W.E.E. collected and assembled data; V.P.-A., H.X., C.S., W.Y., H.Z., M.D., R.C.H., I-M.C., and J.J.Y. analyzed and interpreted data; V.P.-A., H.X., C.G.M., and J.Y. wrote the manuscript; and all authors gave final approval for the manuscript.

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