Ng S-B, Yan J, Huang G, et al. Dysregulated microRNAs affect pathways and targets of biologic relevance in nasal-type natural killer/T-cell lymphoma. *Blood*. 2011;118(18):4919-4929.

On page 4922 in the 3 November 2011 issue, there is a typographical error regarding the precursor vector, which reads, "miR-186." The correct precursor vector is "miR-30b." In "Methods," the fifth sentence under the heading "Luciferase reporter assay" reads, "PRDM1 reporter construct and miR-186 precursor vector were similarly transfected at a ratio of 1:200 for 72 hours before harvest." The sentence should have read, "PRDM1 reporter construct and miR-30b precursor vector were similarly transfected at a ratio of 1:200 for 72 hours before harvest."

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Munoz J, Guo Y. Basophilic stippling: a lead to the diagnosis. *Blood*. 2011;118(20):5370.

On page 5370 in the 17 November 2011 issue, there is an error in the unit of hemoglobin concentration, which reads, "(hemoglobin 6.6 mg/dL...)." The correct unit is "hemoglobin 6.6 g/dL." The fourth sentence of the first paragraph reads, "Laboratory tests showed microcytic anemia (hemoglobin 6.6 mg/dL, mean corpuscular volume 66.8 fL) with normal iron studies and basophilic stippling on the peripheral smear (panel A)." The sentence should have read, "Laboratory tests showed microcytic anemia (hemoglobin 6.6 g/dL, mean corpuscular volume 66.8 fL) with normal iron studies and basophilic stippling on the peripheral smear (panel A)."

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Zhang W, Dang S, Hong T, et al. A humanized single-chain antibody against beta 3 integrin inhibits pulmonary metastasis by preferentially fragmenting activated platelets in the tumor microenvironment. *Blood*. 2012;120(14):2889-2898.

On page 2895 in the 4 October 2012 issue, Figure 6C is an inadvertent replication of Figure 6B. The corrected Figure 6 is shown.

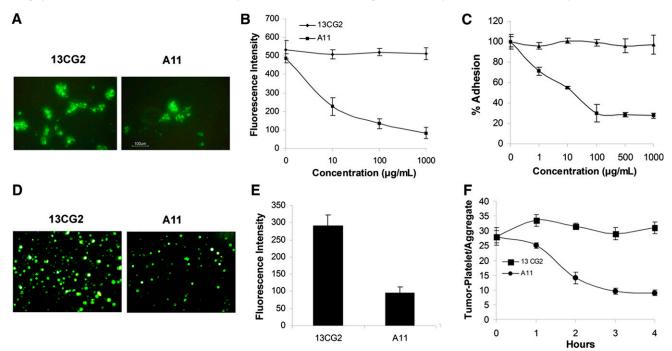


Figure 6. Molecular mechanisms by which A11 inhibits tumor metastasis. (A-B) Effect of A11 on the adhesion of tumor cells to platelets in vitro. (A) The adhesion of LLCs to activated platelets as observed under fluorescence microscope. (B) The quantitative analysis of adhesion of LLC with activated platelet in the presence of various concentration of A11, as measurements of fluorescent intensity under a fluorescence plate reader. The experiment was repeated 3 times and each concentration had 4 wells. (C) Effect of A11 on the adhesion of platelets to HUVECs in vitro. The extent of adhesion was expressed as the percentage of control platelets adhering without preincubation with A11 or control scFv (13CG2). The experiment was repeated 3 times and each concentration had 4 wells. (D-E) Effect of A11 on platelets adhering without preincubation with A11 or control scFv (13CG2). The experiment was repeated 3 times and each concentration had 4 wells. (D-E) Effect of A11 on platelets adhering without preincubation endothelial cell in vitro. B16 melanoma cells adhesion to HUVECs was performed as described in "Assay of tumor cells adhesion to endothelial cells." (D) The adhesion efficiency of B16 tumor cell was observed under a fluorescence microscope. (E) Quantitative result of adhesion of B16 melanoma cells with HUVECs. (F) Effect of A11 on the destruction of already formed tumor platelet aggregates. Data and SD are given for 3 separate experiments at 0.5 µM reagent in which each time point represents 5 measurements.