Platelets in tissue repair: control of apoptosis and interactions with regenerative cells

Meinrad Gawaz and Sebastian Vogel

Department of Cardiology and Cardiovascular Diseases, Eberhard Karls Universität Tübingen, Germany

Besides mediating primary hemostasis and thrombosis, platelets play a critical role in tissue repair and regeneration. They regulate fundamental mechanisms involved in the healing process including cellular migration, proliferation, and angiogenesis. Control of apoptosis/cell survival and interaction with progenitor cells, which are clinically relevant but poorly understood aspects of platelets in tissue repair, will be highlighted in this review. Gaining deeper insight into the less well-characterized molecular mechanisms is necessary to develop new therapeutic platelet-based options. (*Blood.* 2013;122(15):2550-2554)

Platelet: the healer of damaged tissue

For a long time, platelets have been used to treat patients with thrombocytopenia or bleeding events to restore hemostasis. However, platelets also function as circulating cellular sensors that provide a unique link to immune responses and tissue repair.¹ Wound repair indeed is inseparably associated with inflammation and requires a finely tuned interplay of mechanisms regulating cellular migration, extracellular matrix organization/remodeling, cell proliferation, differentiation, and angiogenesis/neovascularization.² Platelets have been recognized to be majorly involved in all these cellular events, which is reviewed elsewhere.³⁻⁶ This review gives an update on the intensively investigated role of platelets in tissue regeneration and highlights clinically relevant but still poorly characterized mechanisms, namely interactions with progenitors and control of apoptosis/ cell survival.

Besides the fact that platelet-rich plasma has increasingly gained attention to seal wounds and enhance wound healing,⁷ experimental and clinical data clearly indicate that platelets are fundamentally involved in repair and regeneration of damaged tissues and preservation of organ function. During tissue injury, for example caused by trauma or local ischemia as seen with myocardial infarction or stroke, the coagulation system and immune responses become activated very early, initiating the process of wound healing. Platelets are the first cells that accumulate at sites of the lesion and, on activation, release a multitude of biologically active mediators into their microenvironment.⁵ Various cytokines, chemokines, and growth factors, including CXCL12 (stromal-derived growth factor 1, SDF-1)^{8,9} and hepatocyte growth factor (HGF),^{10,11} have been identified to be secreted from platelets. Platelet-derived mediators induce and modulate activation of fibroblasts and recruitment of leukocytes, first neutrophils, followed by macrophages, resulting in elimination of dead cells and cellular debris.² Moreover, platelet-released factors induce and control proliferation and migration of other cell types that are critically involved in tissue repair such as smooth muscle cells (SMCs)12 and mesenchymal stem cells (MSCs).¹³ Angiogenesis in damaged tissue, another pivotal mechanism for recovery of tissue function, is also substantially regulated by platelets due to release of a multitude of pro- and antiangiogenic mediators upon platelet activation⁵ (Figure 1).

Nowadays, platelets and their secretory products may successfully be used as feasible therapeutic tools, facilitating repair of injured tissues and organs. For instance, autologous platelet releasate¹⁴ as well as recombinant platelet-derived growth factors¹⁵ may enhance healing of chronic lower extremity diabetic ulcers. Moreover, regeneration of cutaneous wounds,¹⁶ retina,¹⁷ and peri-implant bone¹⁸ by platelets has been reported. However, treatment of surgical lesions with plateletrich plasma has also generated controversial results in clinical trials.¹⁹ In 10 patients with chronic liver disease, platelet transfusion improved distinct parameters of liver function, although adverse events related to platelet transfusion could be seen as well.²⁰

One major achievement in the understanding of platelets and their defects in terms of tissue repair has been made in the field of liver pathophysiology. In a mouse model, Lesurtel et al²¹ identified platelet-derived serotonin as the key player for hepatic regeneration. Interestingly, thrombocytopenia as well as impaired platelet activity in mice substantially abrogated cellular proliferation in the liver. Conversely, thrombopoietin-induced thrombocytosis resulted in strong accumulation of platelets in the sinusoids of liver and induction of hepatocyte proliferation shortly after hepatectomy in mice.²² Moreover, platelets have been shown to be involved in postnatal occlusion of the ductus arteriosus and vessel remodeling.²³ Malfunctioning platelet adhesion/aggregation and defective platelet biogenesis was associated with impaired postnatal occlusion of the ductus in neonatal mice. Moreover, preterm human newborns with thrombocytopenia showed increased risk of persistent open ductus. The impact of abnormal platelet function on tissue repair has also been investigated in atypical hemolytic uremic syndrome with dysfunctional plateletderived complement factor H.²⁴ Besides its function as a complement regulatory protein, factor H exerts antiinflammatory activity and its mutations contribute substantially to hemolytic uremic syndrome and glomerular membrane damage resulting in membranoproliferative glomerulonephritis type II.

Thus, growing evidence indicates that platelets or platelet-derived factors play a pivotal role in determining the balance between tissue repair and tissue damage and may therefore successfully be used for regenerative care. However, the underlying molecular mechanisms involved in platelet-mediated tissue repair are less well characterized,

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Figure 1. Mechanisms governing platelet-mediated tissue repair. Platelets are cellular mediators that orchestrate clinically relevant but still poorly understood mechanisms of tissue repair. They release cytokines, chemokines, and growth factors such as SDF-1 and HGF that control recruitment, proliferation, and activation of fibroblasts, neutrophils, monocytes, SMCs, MSCs, and other cell types critically involved in wound healing. Platelets also regulate angiogenesis in damaged tissue, which is another important mechanism for recovery of tissue function. Recruitment of progenitor cells, including MSCs, SMCs, endothelial progenitors, and CD34-positive progenitors, is influenced by platelets as well, promoting wound repair at least partially due to paracrine mechanisms. Moreover, platelets are capable of modulating the balance between apoptosis and cell survival, which determines the pathophysiology of damaged tissues. They can release proapoptotic (Fas-L, CD40L, TRAIL, TWEAK, and LIGHT) as well as antiapoptotic (HGF, SDF-1, servionin, adenosine diphosphate, and sphingosine-1-phosphate) mediators. Moreover, microparticles derived from platelets can regulate apoptosis in endothelial cells and SMCs as well as provide survival signals to monocytic, endothelial, and neural stem cells. Granzyme B is a mediator of platelet-induced apoptosis in spleen and lung. HMGB1, a danger signal that is exported to the cell surface by platelets upon activation, regulates apoptosis as well as autophagy in tumor cells depending on its redox status. Therefore, platelets control complex mechanisms of tissue repair. ADP, adenosine diphosphate; CD62P, P-selectin; CM, cardiomyocyte; EC, endothelial cell; MØ, macrophage; MP, microparticle; NSC, neural stem cell; pAkt, phosphorylated Akt; PC, progenitor cell; ROS, reactive oxygen species; Ser, serotonin; SP-1, sphingosine-1-phosphate; TC, tumor cell.

and further experimental and clinical studies are needed to define specific targets for future therapeutic interventions.

Platelet/progenitor cell interaction and regeneration

Activated platelets release a whole range of chemokines²⁵ and promote recruitment, adhesion, and proliferation of adult stem cells, including CD34-positive progenitor cells, MSCs, SMC progenitors, and endothelial progenitors^{13,26-28} (Figure 1). The multipotency of these stem cell types and their ability to augment vascular and tissue repair due to paracrine mechanisms²⁹ make them promising therapeutic vehicles in regenerative medicine. Moreover, tissue damage itself generates strong chemo-attractive signals for stem cells, providing the basis for their regenerative activity. Platelet-regulated recruitment of adult stem cells toward injured cells may therefore be a substantial mechanism in exerting regenerative cellular responses.

After myocardial infarction and intramyocardial,³⁰ intracoronary,³¹ or intravenous³² transplantation of MSCs, the cells have been shown to migrate toward the injured heart, prevent ventricular remodeling, and significantly restore cardiac function.³³ Various clinical studies could confirm beneficial effects of MSCs after myocardial damage even though the clinical benefit occasionally turned out to be moderate.^{34,35} Recently, we demonstrated that only apoptotic, but not necrotic or vital, cardiomyocytes induced recruitment of MSCs via HGF/MET-receptor interaction, providing a link between apoptotic cell death and the recruitment of cells with regenerative potential.³⁶ HGF is a growth factor known to be produced after myocardial ischemia, as investigated in animals³⁷ as well as in

humans.³⁸ With its antiapoptotic,³⁹ proangiogenetic,⁴⁰ and immunosuppressive⁴¹ activity, it exerts cardioprotection.⁴² Platelets are also known to release, upon activation, HGF¹¹ and have been described to promote recruitment of MSCs to human arterial endothelial cells.¹³ Modulation of HGF-mediated migration of MSCs to apoptotic tissue cells by platelets is therefore likely and may become a potent therapeutic tool to improve cardiac function after myocardial infarction.

As with HGF, SDF-1, another important mediator involved in stem cell trafficking, is also up-regulated after myocardial ischemia. SDF-1/CXCR4 has been shown to induce recruitment of bone marrow-derived progenitors to the left ventricle after intravenous administration of the cells in a mouse model.⁴³ In clinical trials, CD34-positive progenitor cells have been reported to be critically involved in myocardial repair and regeneration, contributing to preserved cardiac function.44 Moreover, injection of recombinant SDF-1 into the left ventricular cavity of mice before coronary occlusion significantly decreased infarct size compared with control groups.45 To improve efficacy of SDF-1-mediated cardio-protection, we have established a bifunctional protein consisting of an SDF-1 domain and a glycoprotein (GP)VI domain with high binding affinity to CXCR4 as well as to extracellular matrix proteins that become exposed after tissue injury.⁴⁶ After experimental myocardial infarction, administration of SDF1-GPVI had significant cardioprotective effects, promoting migration of CXCR4-positive bone marrowderived progenitors, enhancing endothelial differentiation of the latter, preserving cell survival, and revealing proangiogenic effects. Platelets have an influence on these SDF-1-mediated progenitor cell activities. SDF-1 secreted by activated platelets supported CD34positive progenitor cell recruitment to arterial thrombi and differentiation of the cells to endothelial progenitor cells in vivo.^{8,9} In patients with myocardial infarction, platelet-derived SDF-1 correlated with the number of circulating progenitor cells and was associated with restoration of left ventricular function and improved prognosis.^{47,48} Moreover, formation of circulating platelet/CD34positive progenitor cell coaggregates has been described in patients with acute coronary syndromes, which was associated with a significantly decreased myocardial infarct size and better left ventricular function, as seen with cardiac magnetic resonance imaging at a 3-month follow-up.49 However, platelet-induced differentiation of CD34-positive progenitors into mature foam cells and endothelial cells has been described in an in vitro co-culture system,50 which may be of particular relevance for development of atherosclerotic vascular lesions.

Platelets regulate apoptosis and survival of cells: mechanisms for regeneration

Apoptosis is a precisely executed mode of cell death that sets off processes to limit further tissue damage and is generally associated with immunological tolerance.⁵¹ Increasing evidence indicates that regulation of the balance between apoptosis and cell survival, which determines fate of the injured tissues, is a process that is controlled by platelets⁵²⁻⁵⁶ (Figure 1). Induction of apoptosis is regulated by a diverse range of cell signals, which may originate either extracellularly (extrinsic) or intracellularly (intrinsic).⁵¹ One prominent extrinsic apoptotic pathway involves death receptors that are members of the tumor necrosis factor (TNF) receptor gene superfamily.⁵⁷ TNF- α is a major cytokine regulating apoptosis.⁵⁸ Although the presence of TNF- α in platelets is debatable, they store and secrete a variety of TNF- α -related ligands such as CD95 (Fas-L),⁵⁹ CD154 (CD40L),⁶⁰ Apo2-L (TRAIL),⁶¹ Apo3-L (TWEAK),⁶² and LIGHT,⁶³ which have the potential of regulating apoptosis through paracrine signaling.

In the field of sepsis pathophysiology, pivotal insights could be gained about the significance of platelet-induced apoptosis.⁵²⁻⁵⁵ Incubation of endothelial cells and SMCs with platelet-derived microparticles from septic patients resulted in strong induction of apoptosis in the cells due to production of reactive oxygen species, suggesting a central mechanism in the pathogenesis of septic vascular dysfunction.^{53,54} However, platelet microparticles have also been shown to phosphorylate and activate Akt, a serine-threonine kinase that inactivates the proapoptotic B-cell lymphoma 2 family member BAD (B-cell lymphoma 2-associated death promoter),⁶⁴ and exert antiapoptotic activity in THP-1 cells, a human monocytic leukemia cell line, in a P-selectin-dependent manner.⁵⁶ Interestingly, distinct microparticle types induced differential monocyte responses in terms of intracellular calcium fluxes and release of complement factor C5a as well as TNF- α . Another group demonstrated that platelets from septic mice induced apoptosis in mouse CD4positive splenocytes via a microparticle-independent mechanism.⁵² In this study, apoptosis was mediated by the serine protease granzyme B, which was upregulated in megakaryocytes from the septic mice. Later, the same group demonstrated that platelet granzyme B-mediated apoptosis occurs in spleen and lung depending on direct cell-cell contacts and proper GPIIb/IIIa-function⁵⁵ (Figure 1).

On the other hand, platelets are capable of executing antiapoptotic mechanisms, shifting the balance toward cell survival and tissue repair (Figure 1). In neural stem cells, platelet-derived microparticles induced phosphorylation of Akt, which was associated with neuronal cell proliferation, survival, and differentiation.⁶⁵ Platelet microparticlemediated phosphorylation of Akt has also been observed in endothelial cells, and improved endothelial regeneration took place after injection of microparticle-treated angiogenic early outgrowth cells in a mouse carotid artery wire denudation injury model.⁶⁶ Moreover, platelets secrete, upon activation, mediators with antiapoptotic activity, such as HGF,¹¹ SDF-1,⁶⁷ serotonin,^{12,68} adenosine diphosphate,¹² and sphingosine-1-phosphate,69 promoting survival signals for vascular endothelial cells and SMCs at sites of vascular injury. High mobility group box 1 (HMGB1), a nuclear protein passively released by necrotic cells during tissue injury⁷⁰ or actively secreted by innate immune cells, has been identified as a danger signal that activates immune responses⁷¹ and regulates cell death and survival, as it has been shown for tumor cells, depending on HMGB1-redox status⁷² or formation of complexes with p53-protein.⁷³ Platelets contain endogenous HMGB1, which is exported to the cell surface upon activation,⁷⁴ making it another candidate for platelet-mediated regulation of cell death and survival.

The target cell type as well as regional distribution and intensity of surface expression of the respective death/survival receptors may define the ultimate outcome of pro- and antiapoptotic function of platelets. Further experimental and clinical studies have to be carried out to offer a better understanding of the crosstalk between platelets and mechanisms that control tissue repair, including less wellcharacterized processes such as recruitment of cells with regenerative potential and regulation of apoptosis/cell survival. Such new insights will help us find better therapeutic platelet-based options to facilitate repair and regeneration of injured tissues and organs.

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Authorship

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References

- Nurden AT. Platelets, inflammation and tissue regeneration. *Thromb Haemost.* 2011;105(Suppl 1): S13-S33.
- Gurtner GC, Werner S, Barrandon Y, Longaker MT. Wound repair and regeneration. *Nature*. 2008;453(7193):314-321.
- Gawaz M, Langer H, May AE. Platelets in inflammation and atherogenesis. *J Clin Invest.* 2005;115(12):3378-3384.
- May AE, Seizer P, Gawaz M. Platelets: inflammatory firebugs of vascular walls. *Arterioscler Thromb Vasc Biol.* 2008;28(3):s5-s10.
- Stellos K, Kopf S, Paul A, Marquardt JU, Gawaz M, Huard J, et al. Platelets in regeneration. *Semin Thromb Hemost.* 2010;36(2):175-184.
- Weber C. Platelets and chemokines in atherosclerosis: partners in crime. *Circ Res.* 2005; 96(6):612-616.
- Lacci KM, Dardik A. Platelet-rich plasma: support for its use in wound healing. *Yale J Biol Med.* 2010;83(1):1-9.
- Massberg S, Konrad I, Schürzinger K, et al. Platelets secrete stromal cell-derived factor 1alpha and recruit bone marrow-derived progenitor cells to arterial thrombi in vivo. J Exp Med. 2006;203(5):1221-1233.
- Stellos K, Langer H, Daub K, et al. Plateletderived stromal cell-derived factor-1 regulates adhesion and promotes differentiation of human CD34+ cells to endothelial progenitor cells. *Circulation*. 2008;117(2):206-215.
- 10. Miyazono K, Takaku F. Platelet-derived growth factors. *Blood Rev.* 1989;3(4):269-276.
- Nakamura T, Teramoto H, Ichihara A. Purification and characterization of a growth factor from rat platelets for mature parenchymal hepatocytes in primary cultures. *Proc Natl Acad Sci USA*. 1986; 83(17):6489-6493.
- Crowley ST, Dempsey EC, Horwitz KB, Horwitz LD. Platelet-induced vascular smooth muscle cell proliferation is modulated by the growth amplification factors serotonin and adenosine diphosphate. *Circulation*. 1994;90(4):1908-1918.
- Langer HF, Stellos K, Steingen C, et al. Platelet derived bFGF mediates vascular integrative mechanisms of mesenchymal stem cells in vitro. *J Mol Cell Cardiol*. 2009;47(2):315-325.
- Margolis DJ, Kantor J, Santanna J, Strom BL, Berlin JA. Effectiveness of platelet releasate for the treatment of diabetic neuropathic foot ulcers. *Diabetes Care*. 2001;24(3):483-488.
- Embil JM, Papp K, Sibbald G, Tousignant J, Smiell JM, Wong B, et al. Recombinant human platelet-derived growth factor-BB (becaplermin) for healing chronic lower extremity diabetic ulcers: an open-label clinical evaluation of efficacy. Wound Repair Regen. 2000;8(3):162-168.
- Carter MJ, Fylling CP, Parnell LK. Use of platelet rich plasma gel on wound healing: a systematic review and meta-analysis. *Eplasty*. 2011;11:e38.
- Cullinane AB, O'Callaghan P, McDermott K, Keohane C, Cleary PE. Effects of autologous platelet concentrate and serum on retinal wound healing in an animal model. *Graefes Arch Clin Exp Ophthalmol.* 2002;240(1):35-41.

- Weibrich G, Hansen T, Kleis W, Buch R, Hitzler WE. Effect of platelet concentration in platelet-rich plasma on peri-implant bone regeneration. *Bone*. 2004;34(4):665-671.
- Carreon LY, Glassman SD, Anekstein Y, Puno RM. Platelet gel (AGF) fails to increase fusion rates in instrumented posterolateral fusions. *Spine (Phila Pa 1976)*. 2005;30(9):E243-246; discussion E247.
- Maruyama T, Murata S, Takahashi K, et al. Platelet transfusion improves liver function in patients with chronic liver disease and cirrhosis. *Tohoku J Exp Med.* 2013;229(3):213-220.
- Lesurtel M, Graf R, Aleil B, et al. Platelet-derived serotonin mediates liver regeneration. *Science*. 2006;312(5770):104-107.
- Murata S, Ohkohchi N, Matsuo R, Ikeda O, Myronovych A, Hoshi R. Platelets promote liver regeneration in early period after hepatectomy in mice. *World J Surg.* 2007;31(4):808-816.
- Echtler K, Stark K, Lorenz M, et al. Platelets contribute to postnatal occlusion of the ductus arteriosus. *Nat Med.* 2010;16(1):75-82.
- Zipfel PF. Hemolytic uremic syndrome: how do factor H mutants mediate endothelial damage? *Trends Immunol.* 2001;22(7):345-348.
- von Hundelshausen P, Petersen F, Brandt E. Platelet-derived chemokines in vascular biology. *Thromb Haemost*. 2007;97(5):704-713.
- de Boer HC, Verseyden C, Ulfman LH, et al. Fibrin and activated platelets cooperatively guide stem cells to a vascular injury and promote differentiation towards an endothelial cell phenotype. *Arterioscler Thromb Vasc Biol.* 2006; 26(7):1653-1659.
- Lev EI, Estrov Z, Aboulfatova K, et al. Potential role of activated platelets in homing of human endothelial progenitor cells to subendothelial matrix. *Thromb Haemost*. 2006;96(4):498-504.
- Zernecke A, Schober A, Bot I, et al. SDF-1 alpha/CXCR4 axis is instrumental in neointimal hyperplasia and recruitment of smooth muscle progenitor cells. *Circ Res.* 2005;96(7):784-791.
- Gnecchi M, Zhang Z, Ni A, Dzau VJ. Paracrine mechanisms in adult stem cell signaling and therapy. *Circ Res.* 2008;103(11):1204-1219.
- Amado LC, Saliaris AP, Schuleri KH, et al. Cardiac repair with intramyocardial injection of allogeneic mesenchymal stem cells after myocardial infarction. *Proc Natl Acad Sci USA*. 2005;102(32):11474-11479.
- Chen SL, Fang WW, Ye F, et al. Effect on left ventricular function of intracoronary transplantation of autologous bone marrow mesenchymal stem cell in patients with acute myocardial infarction. *Am J Cardiol.* 2004;94(1): 92-95.
- Nagaya N, Fujii T, Iwase T, et al. Intravenous administration of mesenchymal stem cells improves cardiac function in rats with acute myocardial infarction through angiogenesis and myogenesis. Am J Physiol Heart Circ Physiol. 2004;287(6):H2670-H2676.
- Gnecchi M, He H, Liang OD, et al. Paracrine action accounts for marked protection of ischemic heart by Akt-modified mesenchymal stem cells. *Nat Med.* 2005;11(4):367-368.

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Correspondence: Meinrad Gawaz, Medizinische Klinik III, Kardiologie und Universität Tübingen, Otfried-Müller-Strasse 10, 72076 Tübingen, Germany; e-mail: meinrad.gawaz@med. uni-tuebingen.de.

- Assmus B, Honold J, Schächinger V, et al. Transcoronary transplantation of progenitor cells after myocardial infarction. *N Engl J Med.* 2006; 355(12):1222-1232.
- Wollert KC, Meyer GP, Lotz J, et al. Intracoronary autologous bone-marrow cell transfer after myocardial infarction: the BOOST randomised controlled clinical trial. *Lancet.* 2004;364(9429): 141-148.
- Vogel S, Trapp T, Börger V, Peters C, Lakbir D, Dilloo D, et al. Hepatocyte growth factor-mediated attraction of mesenchymal stem cells for apoptotic neuronal and cardiomyocytic cells. *Cell Mol Life Sci.* 2010;67(2):295-303.
- Ono K, Matsumori A, Shioi T, Furukawa Y, Sasayama S. Enhanced expression of hepatocyte growth factor/c-Met by myocardial ischemia and reperfusion in a rat model. *Circulation*. 1997; 95(11):2552-2558.
- Zhu Y, Hojo Y, Ikeda U, Shimada K. Production of hepatocyte growth factor during acute myocardial infarction. *Heart.* 2000;83(4):450-455.
- Xiao GH, Jeffers M, Bellacosa A, Mitsuuchi Y, Vande Woude GF, Testa JR. Anti-apoptotic signaling by hepatocyte growth factor/Met via the phosphatidylinositol 3-kinase/Akt and mitogenactivated protein kinase pathways. *Proc Natl Acad Sci USA*. 2001;98(1):247-252.
- Aoki M, Morishita R, Taniyama Y, et al. Angiogenesis induced by hepatocyte growth factor in non-infarcted myocardium and infarcted myocardium: up-regulation of essential transcription factor for angiogenesis, ets. *Gene Ther.* 2000;7(5):417-427.
- Okunishi K, Dohi M, Nakagome K, et al. A novel role of hepatocyte growth factor as an immune regulator through suppressing dendritic cell function. J Immunol. 2005;175(7):4745-4753.
- Nakamura T, Mizuno S, Matsumoto K, Sawa Y, Matsuda H, Nakamura T. Myocardial protection from ischemia/reperfusion injury by endogenous and exogenous HGF. J Clin Invest. 2000;106(12): 1511-1519.
- 43. Abbott JD, Huang Y, Liu D, Hickey R, Krause DS, Giordano FJ. Stromal cell-derived factor-1alpha plays a critical role in stem cell recruitment to the heart after myocardial infarction but is not sufficient to induce homing in the absence of injury. *Circulation*. 2004;110(21):3300-3305.
- Mackie AR, Losordo DW. CD34-positive stem cells: in the treatment of heart and vascular disease in human beings. *Tex Heart Inst J.* 2011; 38(5):474-485.
- Hu X, Dai S, Wu WJ, et al. Stromal cell derived factor-1 alpha confers protection against myocardial ischemia/reperfusion injury: role of the cardiac stromal cell derived factor-1 alpha CXCR4 axis. *Circulation*. 2007;116(6):654-663.
- Ziegler M, Elvers M, Baumer Y, et al. The bispecific SDF1-GPVI fusion protein preserves myocardial function after transient ischemia in mice. *Circulation*. 2012;125(5):685-696.
- Geisler T, Fekecs L, Wurster T, et al. Association of platelet-SDF-1 with hemodynamic function and infarct size using cardiac MR in patients with AMI. *Eur J Radiol.* 2012;81(4):e486-e490.

- Stellos K, Bigalke B, Langer H, et al. Expression of stromal-cell-derived factor-1 on circulating platelets is increased in patients with acute coronary syndrome and correlates with the number of CD34+ progenitor cells. *Eur Heart J.* 2009;30(5):584-593.
- Stellos K, Bigalke B, Borst O, et al. Circulating platelet-progenitor cell coaggregate formation is increased in patients with acute coronary syndromes and augments recruitment of CD34+ cells in the ischaemic microcirculation. *Eur Heart J.* 2013;34(32):2548-2556.
- Daub K, Langer H, Seizer P, et al. Platelets induce differentiation of human CD34+ progenitor cells into foam cells and endothelial cells. *FASEB J*. 2006;20(14):2559-2561.
- 51. Elmore S. Apoptosis: a review of programmed cell death. *Toxicol Pathol.* 2007;35(4):495-516.
- Freishtat RJ, Natale J, Benton AS, et al. Sepsis alters the megakaryocyte-platelet transcriptional axis resulting in granzyme B-mediated lymphotoxicity. *Am J Respir Crit Care Med.* 2009; 179(6):467-473.
- Gambim MH, do Carmo AO, Marti L, Veríssimo-Filho S, Lopes LR, Janiszewski M. Plateletderived exosomes induce endothelial cell apoptosis through peroxynitrite generation: experimental evidence for a novel mechanism of septic vascular dysfunction. *Crit Care*. 2007;11(5): R107.
- Janiszewski M, Do Carmo AO, Pedro MA, Silva E, Knobel E, Laurindo FR. Plateletderived exosomes of septic individuals possess proapoptotic NAD(P)H oxidase activity: A novel vascular redox pathway. *Crit Care Med.* 2004; 32(3):818-825.
- Sharron M, Hoptay CE, Wiles AA, et al. Platelets induce apoptosis during sepsis in a contactdependent manner that is inhibited by GPIIb/IIIa blockade. *PLoS ONE*. 2012;7(7):e41549.

- Vasina EM, Cauwenberghs S, Staudt M, Feijge MA, Weber C, Koenen RR, et al. Aging- and activation-induced platelet microparticles suppress apoptosis in monocytic cells and differentially signal to proinflammatory mediator release. *Am J Blood Res.* 2013;3(2):107-123.
- Ashkenazi A, Dixit VM. Death receptors: signaling and modulation. *Science*. 1998;281(5381): 1305-1308.
- Wajant H, Pfizenmaier K, Scheurich P. Tumor necrosis factor signaling. *Cell Death Differ*. 2003; 10(1):45-65.
- Ahmad R, Menezes J, Knafo L, Ahmad A. Activated human platelets express Fas-L and induce apoptosis in Fas-positive tumor cells. *J Leukoc Biol.* 2001;69(1):123-128.
- André P, Nannizzi-Alaimo L, Prasad SK, Phillips DR. Platelet-derived CD40L: the switch-hitting player of cardiovascular disease. *Circulation*. 2002;106(8):896-899.
- Crist SA, Elzey BD, Ludwig AT, Griffith TS, Staack JB, Lentz SR, et al. Expression of TNFrelated approtosis-inducing ligand (TRAIL) in megakaryocytes and platelets. *Exp Hematol.* 2004;32(11):1073-1081.
- Meyer T, Amaya M, Desai H, Robles-Carrillo L, Hatfield M, Francis JL, et al. Human platelets contain and release TWEAK. *Platelets*. 2010; 21(7):571-574.
- Otterdal K, Smith C, Oie E, et al. Platelet-derived LIGHT induces inflammatory responses in endothelial cells and monocytes. *Blood.* 2006; 108(3):928-935.
- Datta SR, Dudek H, Tao X, Masters S, Fu H, Gotoh Y, et al. Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery. *Cell.* 1997;91(2):231-241.
- 65. Hayon Y, Dashevsky O, Shai E, Varon D, Leker RR. Platelet microparticles promote neural stem

cell proliferation, survival and differentiation. *J Mol Neurosci.* 2012;47(3):659-665.

- Mause SF, Ritzel E, Liehn EA, et al. Platelet microparticles enhance the vasoregenerative potential of angiogenic early outgrowth cells after vascular injury. *Circulation.* 2010;122(5):495-506.
- Stellos K, Gawaz M. Platelets and stromal cellderived factor-1 in progenitor cell recruitment. Semin Thromb Hemost. 2007;33(2):159-164.
- Pakala R, Willerson JT, Benedict CR. Mitogenic effect of serotonin on vascular endothelial cells. *Circulation*. 1994;90(4):1919-1926.
- Hisano N, Yatomi Y, Satoh K, Akimoto S, Mitsumata M, Fujino MA, et al. Induction and suppression of endothelial cell apoptosis by sphingolipids: a possible in vitro model for cell-cell interactions between platelets and endothelial cells. *Blood.* 1999;93(12):4293-4299.
- Scaffidi P, Misteli T, Bianchi ME. Release of chromatin protein HMGB1 by necrotic cells triggers inflammation. *Nature*. 2002;418(6894): 191-195.
- Andersson U, Tracey KJ. HMGB1 is a therapeutic target for sterile inflammation and infection. *Annu Rev Immunol.* 2011;29:139-162.
- Tang D, Loze MT, Zeh HJ, Kang R. The redox protein HMGB1 regulates cell death and survival in cancer treatment. *Autophagy*. 2010;6(8): 1181-1183.
- Livesey KM, Kang R, Vernon P, et al. p53/HMGB1 complexes regulate autophagy and apoptosis. *Cancer Res.* 2012;72(8):1996-2005.
- Rouhiainen A, Imai S, Rauvala H, Parkkinen J. Occurrence of amphoterin (HMG1) as an endogenous protein of human platelets that is exported to the cell surface upon platelet activation. *Thromb Haemost.* 2000;84(6): 1087-1094.