

# Allogeneic hematopoietic cell transplantation for XIAP deficiency: an international survey reveals poor outcomes

Rebecca A. Marsh,<sup>1</sup> Kanchan Rao,<sup>2</sup> Prakash Satwani,<sup>3</sup> Kai Lehmsberg,<sup>4</sup> Ingo Müller,<sup>4</sup> Dandan Li,<sup>5</sup> Mi-Ok Kim,<sup>5</sup> Alain Fischer,<sup>6</sup> Sylvain Latour,<sup>7</sup> Petr Sedlacek,<sup>8</sup> Vincent Barlogis,<sup>9</sup> Kazuko Hamamoto,<sup>10</sup> Hirokazu Kanegane,<sup>11</sup> Sam Milanovich,<sup>12</sup> David A. Margolis,<sup>12</sup> David Dimmock,<sup>13</sup> James Casper,<sup>12</sup> Dorothea N. Douglas,<sup>14</sup> Persis J. Amrolia,<sup>2</sup> Paul Veys,<sup>2</sup> Ashish R. Kumar,<sup>1</sup> Michael B. Jordan,<sup>1</sup> Jack J. Bleesing,<sup>1</sup> and Alexandra H. Filipovich<sup>1</sup>

<sup>1</sup>Division of Bone Marrow Transplantation and Immunodeficiency, Cincinnati Children's Hospital Medical Center, Cincinnati, OH; <sup>2</sup>Department of Haematology and Bone Marrow Transplantation, Great Ormond Street Hospital for Sick Children, London, United Kingdom; <sup>3</sup>Division of Pediatric Blood and Marrow Transplantation, Columbia University Medical Center, Morgan Stanley Children's Hospital of New York-Presbyterian, New York, NY; <sup>4</sup>Clinic for Pediatric Hematology and Oncology, University Medical Center Hamburg-Eppendorf, Hamburg, Germany; <sup>5</sup>Division of Biostatistics and Epidemiology, Cincinnati Children's Hospital Medical Center, Cincinnati, OH; <sup>6</sup>Inserm U768, Unité d'Immunologie et d'hématologie, Hôpital Necker, Assistance Publique-Hopitaux de Paris, Université Paris Descartes, Sorbonne Paris Cité, Institut Imagine, Paris, France; <sup>7</sup>Inserm U768, Hôpital Necker, Université Paris Descartes, Sorbonne Paris Cité, Institut Imagine, France; <sup>8</sup>Department of Pediatric Hematology and Oncology, University Hospital Motol, Charles University, Prague, Czech Republic; <sup>9</sup>Hôpital Timone Enfants, Marseille, France; <sup>10</sup>Department of Pediatrics, Hiroshima Red Cross Hospital, Hiroshima, Japan; <sup>11</sup>Department of Pediatrics, Graduate School of Medicine, University of Toyama, Toyama, Japan; <sup>12</sup>Division of Pediatric Hematology, Oncology and Bone Marrow Transplant, Medical College of Wisconsin, Children's Hospital of Wisconsin, Milwaukee, WI; <sup>13</sup>Division of Genetics, Department of Pediatrics, Medical College of Wisconsin, Milwaukee, WI; and <sup>14</sup>University of Arizona College of Medicine, Phoenix Children's Hospital, Phoenix, AZ

## Key Points

- High mortality rates are observed in patients with XIAP deficiency treated with myeloablative conditioning regimens for hematopoietic cell transplantation.

**There have been no studies on patient outcome after allogeneic hematopoietic cell transplantation (HCT) in patients with X-linked inhibitor of apoptosis (XIAP) deficiency. To estimate the success of HCT, we conducted an international survey of transplantation outcomes. Data were reported for 19 patients. Seven patients received busulfan-containing myeloablative conditioning (MAC) regimens. Eleven patients underwent reduced intensity conditioning (RIC) regimens predominantly consisting of alemtuzumab, fludarabine, and melphalan. One patient received an intermediate-intensity regimen. Survival was poor in the MAC group, with only 1 patient surviving (14%). Most deaths were from transplantation-related toxicities, including venoocclusive disease**

**and pulmonary hemorrhage. Of the 11 patients who received RIC, 6 are currently surviving at a median of 570 days after HCT (55%). Preparative regimen and HLH activity affected outcomes, and of RIC patients reported to be in remission from HLH, survival is 86% ( $P = .03$ ). We conclude that MAC regimens should not be used for patients with XIAP deficiency. It is possible that the loss of XIAP and its antiapoptotic functions contributes to the high incidence of toxicities observed with MAC regimens. RIC regimens should be pursued with caution and, if possible, efforts should be made to ensure HLH remission before HCT in these patients. (*Blood*. 2013;121(6):877-883)**

## Introduction

Deficiency of X-linked inhibitor of apoptosis (XIAP) is associated with X-linked lymphoproliferative disease (XLP) and familial hemophagocytic lymphohistiocytosis (FHLH) phenotypes. Traditionally, patients with inherited immune deficiencies that cause HLH have been treated with allogeneic hematopoietic cell transplantation (HCT) because of the life-threatening nature of HLH. There is extensive experience with transplantation in patients with FHLH. Over the past 10 years, survival has generally approximated 60% with myeloablative conditioning (MAC) regimens.<sup>1-7</sup> More recently, improvements have been made with reduced-intensity conditioning (RIC) protocols, and current survival rates are as high as 80%.<sup>8-11</sup> There is less experience with transplantation in patients with XLP because of SLAM-associated protein (SAP) deficiency, but survival is generally accepted to be greater than 70% regardless of the intensity of the conditioning protocol.<sup>12-14</sup>

To date, little has been published concerning the outcomes of HCT for patients with XIAP deficiency. XIAP deficiency was first discovered in 2006,<sup>15</sup> and is associated with XLP, FHLH, and colitis phenotypes.<sup>15-18</sup> Patients with XIAP deficiency are unique compared with patients with the other genetic forms of HLH because, as the name suggests, XIAP is an inhibitor of apoptosis that is widely expressed outside of the immune system.<sup>19</sup> Thymocytes from XIAP-deficient mice have been shown to have normal apoptotic responses to a variety of apoptotic stimuli,<sup>20</sup> but hepatocytes are more sensitive to death induced by treatment with cross-linked Fas ligand.<sup>21</sup> XIAP-deficient mouse embryonic fibroblasts are also more sensitive to death after infection with MHV-68.<sup>22</sup> In addition, there is increasing experience with the use of XIAP inhibitors in conjunction with traditional cancer treatment. In this setting, XIAP inhibitors generally increase the susceptibility

Submitted June 26, 2012; accepted October 11, 2012. Prepublished online as *Blood* First Edition paper, November 6, 2012; DOI 10.1182/blood-2012-06-432500.

The publication costs of this article were defrayed in part by page charge

payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2013 by The American Society of Hematology

**Table 1. Patient characteristics**

Patient no	Age at initial presentation	EBV HLH before HCT	Non-EBV HLH before HCT	HLH not in full remission before HCT	Colitis before HCT	Other	XIAP mutation	Protein expression
1	3 mo	–	+	–	–		1443_1449 delins 24 (P482fsX508)	NE*
2	2 mo	–	+	+	–		1443_1449 delins 24 (P482fsX508)	NE
3	2 mo	–	+	–	–		563 G → A (G188E)	Reduced
4	Asymptomatic (symptomatic brother)	–	–	–	–		563 G → A (G188E)	Reduced
5	15 mo	–	–	–	+	Recurrent enterocutaneous fistulas; multiple episodes of polymicrobial sepsis	608G → A (C203Y)	Reduced
6	9 mo	–	+	–	–		E99KfsX129	Absent
7	9 y	+	–	–	–		497G → T, R166I	NE
8	7 mo	–	+	+	–		1141C → T (R381X)	Reduced
9	Infancy	–	+	–	–		1481 T → A (I494N)	NE
10	4 mo	–	+	+	–		1445 C → G (P482R)	Reduced
11	1 y	+	–	+	–	Repeated infections: pneumonia, otitis media, history of paracentesis, mastoidectomy	1189 delA (I397fsX414)	Absent
12	1 y	+	–	+	–		387_390del (D130fsX140)	Not reported
13	3 mo	–	+	–	–		Gross Deletion Exons 1-5	Absent
14	1 y	–	–	–	+	Recurrent fevers; pneumococcal sepsis.	758 C → G (S253X)	Absent
15	3 y	–	+	–	–	Ventricular septal defect	356_359del (N119fs384)	NE
16	7 y	+	+	–	–		1141C → T (R381X)	Reduced
17	8 y	+	–	+	–		310 C → T (Q104X)	Absent
18	Infancy	–	+	–	–	Liver failure in infancy required liver transplantation; nodular lung disease; positive CMV and fungal elements	Gross deletion exon 6	Truncated (robust detection of a smaller molecular weight protein by Western blot)
19	17 y	+	–	–	–		894_898 del 5 (K299fsX307)	Absent

\*Not examined.

of cancer cells to undergo apoptosis.<sup>23,24</sup> Because of the importance of XIAP in preventing apoptosis, patients with XIAP deficiency may be at increased risk of treatment-related toxicities because of increased sensitivity to chemotherapeutic agents.

To investigate whether deficiency of XIAP adversely affects the survival of patients undergoing allogeneic HCT, we conducted an international survey to collect information regarding the transplantation outcomes of patients confirmed to have XIAP deficiency.

## Methods

### Data collection

Approval for this retrospective study was granted by the Cincinnati Children's Hospital Institutional Review Board. A spreadsheet questionnaire was sent to physicians who provided treatment for patients with XIAP deficiency who underwent allogeneic HCT. Physicians were identified through contact with our center, our review of the literature regarding XIAP deficiency, or a request made to all members of the Histocyte Society.

### Patients

Only patients with a confirmed *XIAP/BIRC4* (baculoviral inhibitor of apoptosis repeat containing protein 4) mutation or with a sibling with a confirmed mutation were included in this study (Table 1), which was

conducted in accordance with the Declaration of Helsinki. Supplemental lymphocyte protein analysis was performed in some patients using either Western blot or intracellular flow cytometric analysis.<sup>15-17,25</sup>

### Transplantation procedures

Patients received transplantation at centers in the United States (n = 12), Europe (n = 6), and Japan (n = 1) between the years 2001-2011. Transplantation procedures were carried out per institutional standard practices. Conditioning regimens and graft characteristics are listed in Table 2. Conditioning regimens were classified as MAC if they contained an alkylating agent (busulfan) or total body irradiation (TBI) at a dose that would not allow autologous BM recovery.<sup>26</sup> Conditioning regimens were classified as RIC if they did not meet the definition of MAC regimen.<sup>26</sup> If there was uncertainty regarding the intensity of the regimen (n = 1, patient 8), it was classified as an intermediate-intensity regimen. Neutrophil engraftment was considered to be the day the neutrophil count reached  $0.5 \times 10^9/L$ . Engraftment studies were done using either XY FISH for sex-mismatched donors or variable number of tandem repeat analysis for same-sex donors. Mixed chimerism was defined as having 5% or more host-derived cells in the whole blood on more than 1 occasion. Acute and chronic GVHD were assessed by standard criteria.<sup>27,28</sup> Patients received GVHD prophylaxis per institutional standard practices. Other routine transplantation care, such as antimicrobial prophylaxis, IV Ig replacement, and fluid and nutrition supplementation when needed, were also provided per institutional standard practices.

**Table 2. Transplantation procedures**

Patient no	Age at HCT, y	Type of conditioning	Conditioning regimen	Graft HLA match*	Graft source	Relationship
1	0.42	MAC	Bu, Mel, ATG	5/6	Cord	Unrelated
2	0.58	MAC	Bu, Cy, ATG, Etop	6/6	Cord	Unrelated
3	1	MAC	Bu, Cy, ATG	7/8	BM	Unrelated
4	4	MAC	Bu, Cy, ATG	10/10	BM	Unrelated
5	5	MAC	Bu, Flu, ATG	6/6	Cord	Unrelated
6	10	MAC	Bu, Cy, ATG	6/6	BM	Unrelated
7	14	MAC	Bu, Cy, ATG, Etop	7/8	PBSCs	Unrelated
8	1	Intermediate	TBI (6 Gy), Flu, Cy, Mel (80 mg/m <sup>2</sup> )	7/8	Cord	Unrelated
9	0.40	RIC	Alem, Flu, Mel	8/8	BM	Unrelated
10	0.98	RIC	Alem, Flu, Mel	9/10	BM	Unrelated
11	2	RIC	Alem, Flu, Mel	9/10	BM	Unrelated
12	3	RIC	Alem, Flu, Mel	9/10	Cord	Unrelated
13	3	RIC	Alem, Flu, Mel	8/8	BM	Unrelated
14	3	RIC	Alem, Flu, Mel	10/10	BM	Unrelated
15	4	RIC	Alem, Flu, Mel	8/8	PBSCs	Maternal
16	7	RIC	Alem, Flu, Treo, Thio	10/10	PBSCs	Unrelated
17	9	RIC	Alem, Flu, Mel	7/8	BM	Unrelated
18	11	RIC	Alem, Flu, Mel	8/8	BM	Unrelated
19	19	RIC	Alem, Flu, Mel	10/10	BM	Sibling

Bu indicates busulfan; Mel, melphalan; ATG, antithymocyte globulin; Cy, cyclophosphamide; Etop, etoposide; Flu, fludarabine; Alem, alemtuzumab; Treo, treosulfan; Thio, thiotepa; and PBSCs, peripheral blood stem cells.

\*Six to 10 alleles (HLA-A, HLA-B, HLA-C, HLA-DRB1, or HLA-DQB1).

### Statistical analysis

Survival was analyzed using Kaplan-Meier curves created with XLSTAT 2011 software (Addinsoft). Comparison of survival curves was done using the log-rank test. For multivariate analysis of survival time and the impact of preparative regimen (MAC vs RIC), donor match, (full match vs mismatch), and HLH activity (remission vs nonremission), Cox proportional hazard regression model analysis was used. The patient who received the intermediate-intensity regimen was excluded from these analyses. Statistical significance was considered as  $P < .05$ .

## Results

### Patients

Nineteen patients with XIAP deficiency underwent allogeneic HCT between 2001 and 2011 at a median age of 3 years (range, 0.4-19). Patient characteristics before HCT and *XIAP/BIRC4* mutations are listed in Table 1. Approximately one-third of patients had developed EBV-related HLH before HCT, and approximately two-thirds of patients had developed non-EBV HLH before HCT. Six of these patients were reported to have either active HLH or HLH in partial remission just before HCT. Two patients with colitis were diagnosed and treated as having Crohn disease before the diagnosis of XIAP deficiency.

### Transplantation procedures

Graft characteristics and conditioning regimens are shown in Table 2. Seven patients received a MAC protocol.<sup>26</sup> Most patients received busulfan, cyclophosphamide, and antithymocyte globulin with or without etoposide ( $n = 5$ ). The remaining 2 patients received busulfan with either fludarabine or melphalan and antithymocyte globulin. Eleven patients received a RIC protocol.<sup>26</sup> Ten RIC patients received alemtuzumab, fludarabine, and melphalan, and 1 patient received alemtuzumab, fludarabine, treosulfan, and thiotepa. The remaining patient (patient 8) received an intermediate protocol consisting of TBI (6 Gy), fludarabine, cyclophosphamide, and melphalan (80 mg/m<sup>2</sup>).

Eleven patients received fully matched related ( $n = 2$ ) or unrelated ( $n = 9$ ) grafts based on typing of 6-10 HLA antigens (HLA-A, HLA-B, HLA-C, HLA-DRB1, and HLA-DQB1). Eight patients received a single allele mismatched graft. The stem cell source was BM in 11 patients, cord blood in 5 patients, and peripheral blood stem cells in 3 patients.

### Engraftment

All patients engrafted with a median of 15 days (range, 8-22) except for patient 11, who died before engraftment on day +13.

### Toxicities

There was a high incidence of conditioning-related toxicities among MAC patients (Table 3). There were 3 cases of hepatic venoocclusive disease (VOD), which contributed to deaths on days +17, +50, and +144 in patients 6, 2, and 1, respectively. Two of these patients also developed pulmonary hemorrhage. One patient (patient 3) developed pulmonary hypertension of uncertain etiology with pulmonary hemorrhage after transplantation and died on day +170. This patient had received MAC after having previously undergone HCT twice with RIC.

There were no cases of hepatic VOD or pulmonary hemorrhage in patients who received RIC. However, 1 patient (patient 11) developed multiorgan failure and cardiac toxicity with asystole and died at day +13. A second patient (patient 15) suffered an unexpected death related to idiopathic pneumonitis and respiratory failure at day +125.

Patient 8, who received the intermediate preparative regimen (consisting of TBI, fludarabine, cyclophosphamide, and melphalan), suffered posttransplantation cytokine storm syndrome with acute respiratory distress syndrome and died on day +22.

### GVHD

Three patients developed acute GVHD of grade 2 or greater (Table 3). One additional patient developed acute GVHD after receiving a donor lymphocyte infusion that was administered as an intervention for declining donor contribution to hematopoiesis.

**Table 3. Toxicities and complications**

Patient no	VOD	Pulmonary hemorrhage	Acute VHD	Pneumonitis or ARDS	Confirmed bacteremia/sepsis	Fungal infection	Viremia with EBV, CMV, adenovirus, or HHV6	BK virus hemorrhagic cystitis
1	+	+	–	NR	NR	NR	NR	NR
2	+	–	–	NR	NR	NR	NR	NR
3	–	+ (shown by autopsy, not clinically)	II	–	+ ( <i>S marcescens</i> )	–	+ (EBV, adenovirus)	–
4	–	+ (related to fungal septic thrombosis of the pulmonary veins and pulmonary artery)	III	–	–	+ (fungal septic thrombosis of the pulmonary veins and pulmonary artery)	+ (EBV, adenovirus)	+
5	–	–	I	–	+ ( <i>K oxytoca</i> , <i>Enterococcus</i> sp, <i>P aeruginosa</i> )	–	+ (CMV, adenovirus, HHV6)	–
6	+	+	–	–	–	–	–	–
7	–	–	III	+	–	–	–	–
8	–	–	–	+	–	–	–	–
9	–	–	–	–	–	–	+ (adenovirus)	–
10	–	–	– (+ after DLI)	–	+ ( <i>K oxytoca</i> , <i>S maltophilia</i> , <i>P aeruginosa</i> )	–	–	–
11	–	–	–	+	–	–	–	–
12	–	–	–	–	–	–	–	+
13	–	–	–	–	–	–	+ (adenovirus)	–
14	–	–	–	–	–	–	+ (EBV, CMV)	–
15	–	–	–	+	–	–	–	–
16	–	–	–	–	–	–	+ (adenovirus)	–
17	–	–	I	–	+ ( <i>S aureus</i> )	–	+ (adenovirus)	–
18	–	–	–	–	+ ( <i>S aureus</i> )	–	+ (CMV)	–
19	–	–	–	–	–	–	–	–

ARDS indicates acute respiratory distress syndrome; NR, not reported; and DLI, donor lymphocyte infusion.

Two patients developed chronic GVHD (limited, n = 1, and extensive, n = 1).

### Infections

Most patients experienced an infectious complication of HCT (Table 3). Common viral complications included EBV viremia (n = 3, all patients received rituximab), CMV viremia (n = 3, all patients received CMV-directed therapy), and adenovirus viremia (n = 7, 4 patients received adenovirus-directed therapy). Other reported viral complications included human herpesvirus 6 (HHV6) viremia and encephalitis (n = 1), varicella zoster (n = 1), and BK virus hemorrhagic cystitis (n = 2).

Reported bacterial infections included pneumonias, bacteremias and episodes of sepsis (n = 5) related to *Serratia marcescens*, *Klebsiella oxytoca*, *Stenotrophomonas maltophilia*, *Enterococcus* sp, *Pseudomonas aeruginosa*, and *Staphylococcus aureus*. One patient developed fatal fungal septic thrombosis of the pulmonary veins and pulmonary artery.

### Donor contribution to hematopoiesis

Six patients were reported to develop mixed donor and recipient chimerism (< 95% donor cells detected in peripheral blood) at a median of 37 days after HCT. All of these patients had received RIC. Patient 12 was reported to lose the graft by 35 days after HCT. For the remaining 5 patients (patients 9, 10, 13, 18, and 19), the lowest observed donor contributions to hematopoiesis ranged from 13.8%-92%. Three patients received a stem cell boost and/or donor lymphocyte infusion(s). At the time of last follow-up at a median of 867 days after HCT (range, 139-1706), all 5 patients possessed greater than 90% donor contribution to hematopoiesis and remained free of disease.

### Survival and outcome

Only 1 of the 7 patients who received MAC is currently surviving, 414 days after HCT (Table 4). The other 6 patients died at a median of 97 days after HCT (range, 17-247) from toxicities and complications including VOD, pulmonary hemorrhage, pulmonary hypertension, GVHD, sepsis, multiorgan failure, and fungal septic thrombosis of pulmonary veins and pulmonary artery with pulmonary hemorrhagic necrosis.

Patient 8, who received an intermediate-conditioning regimen, also died, on day +22, of posttransplantation cytokine storm syndrome with acute respiratory distress syndrome.

Of the patients who received RIC, 6 of 11 are currently alive and well at a median of 570 days after HCT (55%). All but 1 survivor were given a Lansky or Karnofsky score of 100 at the time of last follow-up. Patients 10, 11, 12, 15, and 17 died at a median of 140 days after HCT (range, 13-416). Reported causes of death were heterogeneous and included pneumonitis with respiratory failure, cardiac toxicity with asystole and multiorgan failure, encephalitis, and ongoing CNS HLH (with loss of graft), sepsis, and pneumonia with respiratory failure (Table 4).

The 1-year probabilities of survival for MAC and RIC patients are 14% and 57%, respectively (Figure 1A), with long-term probabilities of survival of 14% and 43%, respectively (Figure 1B).

### Influences on survival

We examined the significance of multiple factors known to influence transplantation outcomes including preparative regimen (MAC vs RIC),<sup>11</sup> donor match,<sup>29</sup> and HLH disease status at the time of transplantation.<sup>2-4</sup> HLH disease status at the time of transplantation was based on the judgment of the treating/contributing physician who reported HLH to be in remission, in partial

**Table 4. Patient outcomes**

Patient no	Follow-up, d	Outcome	Cause of death
1	144	Died	VOD and pulmonary hemorrhage
2	50	Died	VOD and MOF
3	170	Died	Pulmonary hypertension
4	247	Died	Fungal septic thrombosis of pulmonary veins and pulmonary artery with pulmonary hemorrhagic necrosis
5	414	Alive and well; limited skin GVHD	
6	17	Died	Pulmonary hemorrhage, VOD
7	50	Died	GVHD, MOF
8	22	Died	ARDS, posttransplantation cytokine storm syndrome
9	1765	Alive and well	
10	285	Died	Drug-resistant <i>P aeruginosa</i> sepsis
11	13	Died	Cardiac toxicity, MOF, asystole
12	140	Died	Encephalitis, HLH with CNS involvement
13	1057	Alive and well	
14	149	Alive and well	
15	125	Died	Pneumonitis and respiratory failure
16	273	Alive and well	
17	416	Died	Pneumonia and respiratory failure; chronic extensive GVHD
18	867	Alive and well	
19	139	Alive and well	

MOF indicates multiorgan failure; and ARDS, acute respiratory distress syndrome.

remission, or active. The patient who received the intermediate-intensity regimen (patient 8) was excluded from the analysis. Although there are a limited number of patients in our series, it is notable that of the surviving patients ( $n = 7$ ), all were reported to be in remission of HLH at the time of HCT. Of the deceased patients ( $n = 12$ ), half were reported to be in partial remission or have active disease at the time of HCT. It is also notable that of the 7 surviving patients, all but 1 received grafts from HLA-matched donors, whereas of the 12 deceased patients, only 3 received grafts from HLA-matched donors. Multivariate analysis suggested that MAC regimens and HLH that was not in remission conveyed statistically significant negative influences on survival (Figure 1C and Table 5). Match was significant in univariate analysis (data not shown), but was not significant once controlled for conditioning regimen and HLH remission status. Survival for patients receiving RIC who were reported to be in remission from HLH is 86% ( $P = .03$ ; Figure 1C).

Because XIAP functions as an inhibitor of apoptosis and is widely expressed, we also sought to determine whether residual protein expression may offer some protective benefit for survival after allogeneic HCT. Twelve patients were reported to have had analysis of XIAP protein expression. Of 5 patients with no detectable XIAP, 2 are alive and well (40%). Of 7 patients with detectable decreased or truncated protein expression, 3 are alive and well (43%). We conclude that in this limited cohort, the presence of detectable XIAP does not appear to confer a survival advantage.

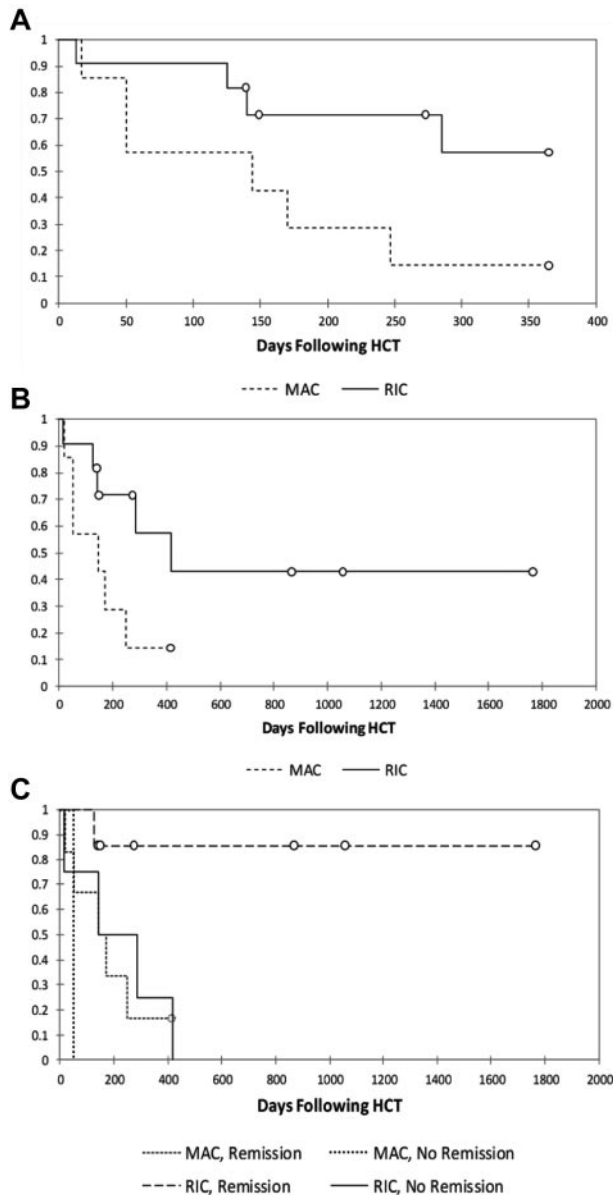
## Discussion

Deficiency of XIAP is a newly recognized disorder, and the results of the present study survey reveal that transplantation outcomes overall appear poor compared with the outcomes typically expected of patients with XLP and FHLH. There was a high incidence of conditioning-related toxicity, which may be related to the lack of ubiquitously expressed XIAP and the resultant loss of its antiapoptotic and other functions. In particular, only 1 patient treated with MAC is currently surviving (14%). This is in sharp contrast to the

typical survival rates in other forms of HLH, which are generally greater than 50%.<sup>1-7,11-13</sup> There was a preponderance of hepatic VOD and pulmonary hemorrhage in MAC patients. Although VOD has been reported in patients with HLH who undergo MAC regimens, it appears that the 50% incidence of VOD in this series is high compared with previous reports of 20%-30%.<sup>3,4</sup> However, because of the small number of patients included in the present study, it is difficult to conclude definitively that XIAP deficiency predisposes patients to an increased risk of liver and pulmonary toxicity. In addition, a high proportion of MAC patients received grafts from HLA-mismatched donors or had HLH that was not in remission at the time of transplantation, which may have contributed to the poor outcomes. Regardless, based on the poor survival outcomes, MAC protocols should be cautioned against and avoided in patients with XIAP deficiency.

With regard to the RIC cohort, the overall survival of just over half of patients appears to be decreased compared with the relatively high survival rates expected for HLH patients undergoing RIC HCT, which are typically greater than 80%.<sup>10,11</sup> However, the causes of death among the patients with XIAP deficiency were heterogeneous and we found no clear evidence to suggest that the deaths were related to deficiency of XIAP. The survival of RIC patients reported to be in remission from HLH was 86%, and the impact of HLH status was significant. This suggests that RIC transplantation outcomes for patients with XIAP deficiency who are in remission from HLH may be equivalent to that of other forms of XLP and FHLH. Infectious complications were common after HCT in both MAC and RIC patients. These complications do not appear to be increased compared with reports of transplantation outcomes for patients with HLH.<sup>9,11</sup>

Given our findings, the question of whether to pursue allogeneic RIC HCT is somewhat difficult to answer and is further complicated by the limited amount of information regarding outcomes of patients with XIAP deficiency not treated with transplantation. In the largest published series to date ( $N = 30$ ), approximately 40% of patients with XIAP deficiency died at a mean age of 16 years predominantly because of HLH, colitis, or complications of allogeneic HCT.<sup>30</sup> Overall, the small numbers of patients make it difficult to draw a firm conclusion regarding recommendations for



**Figure 1. Kaplan-Meier survival analyses.** Shown are analyses of 1-year survival (A), long-term survival (B), and survival stratified by reported HLH status at the time of transplantation (C;  $P = .035$ ) in patients treated with MAC or RIC regimens.

RIC HCT for patients with XIAP deficiency. At this time, based on the available information, it is our opinion that RIC protocols should be pursued with caution in young patients with XIAP deficiency who have a compelling clinical history and for whom a good stem cell donor is available. Preferably, patients should have

## References

- Henter JI, Samuelsson-Horne A, Arico M, et al. Treatment of hemophagocytic lymphohistiocytosis with HLH-94 immunochemotherapy and bone marrow transplantation. *Blood*. 2002;100(7):2367-2373.
- Horne A, Janka G, Maarten Egeler R, et al. Haematopoietic stem cell transplantation in haemophagocytic lymphohistiocytosis. *Br J Haematol*. 2005;129(5):622-630.
- Ouachée-Chardin M, Elie C, de Saint Basile G, et al. Hematopoietic stem cell transplantation in hemophagocytic lymphohistiocytosis: a single-

- center report of 48 patients. *Pediatrics*. 2006;117(4):e743-750.
- Baker KS, Filipovich AH, Gross TG, et al. Unrelated donor hematopoietic cell transplantation for hemophagocytic lymphohistiocytosis. *Bone Marrow Transplant*. 2008;42(3):175-180.
- Cesaro S, Locatelli F, Lanino E, et al. Hematopoietic stem cell transplantation for hemophagocytic lymphohistiocytosis: a retrospective analysis of data from the Italian Association of Pediatric Hematology Oncology (AIEOP). *Haematologica*. 2008;93(11):1694-1701.

- Ohga S, Kudo K, Ishii E, et al. Hematopoietic stem cell transplantation for familial hemophagocytic lymphohistiocytosis and Epstein-Barr virus-associated hemophagocytic lymphohistiocytosis in Japan. *Pediatr Blood Cancer*. 2010;54(2):299-306.
- Yoon HS, Im HJ, Moon HN, et al. The outcome of hematopoietic stem cell transplantation in Korean children with hemophagocytic lymphohistiocytosis. *Pediatr Transplant*. 2010;14(6):735-740.
- Shenoy S, Grossman WJ, DiPersio J, et al. A novel reduced-intensity stem cell transplant

**Table 5. Cox proportional hazard regression model analysis**

Variable	P	HR	HR 95% CI
<b>A</b>			
Conditioning (MAC vs RIC)	.0251	7.524	1.287 44.000
Match (match vs mismatch)	.2744*	0.471	0.122 1.816
HLH activity (not in remission vs remission)	.0806	4.322	0.837 22.330
<b>B</b>			
Conditioning (MAC vs RIC)	.0181	6.348	1.371 29.394
HLH activity (not in remission vs remission)	.0218	5.301	1.275 22.046

In part A of the table, multivariate analysis included preparative regimen, match, and HLH activity; in part B, the effects of preparative regimen and HLH activity were analyzed with removal of the nonsignificant match effect.

HR indicates hazard ratio; and CI, confidence interval.

\*The effect of match was statistically significant in univariate analysis.

no active lymphoproliferative disease or HLH and aggressive efforts should be made to ensure remission of HLH. The outcomes of all patients with XIAP deficiency should be monitored to further support evidence-based decisions regarding optimal treatment strategies.

## Acknowledgments

The authors thank the Histiocyte Society for distributing our request for participation in this study to the members of the Society; Denise Bellman, Laura Hart, Linda Bierman, Angie Bonavita, and Christine Sper; Kejian Zhang and the molecular genetics laboratory at Cincinnati Children's Hospital; the patients and their families; and all of the physicians, nurses, and staff who provided care for patients.

R.A.M. is supported by a grant from the Clinical Immunology Society.

## Authorship

Contribution: R.A.M. and K.R. designed the study, collected and analyzed the patient data, and wrote the manuscript; P.K., K.L., I.M., A.F., S.L., P.S., V.B., K.H., H.K., S.M., D.A.M., D.D., J.C., D.N.D., P.J.A., P.V., A.R.K., M.B.J., and J.J.B. collected the patient data and edited the manuscript; D.L. and M.K. performed the statistical analyses; and A.H.F. designed and oversaw the study and edited the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Rebecca A. Marsh, MD, Division of Bone Marrow Transplantation and Immune Deficiency, Cincinnati Children's Hospital Medical Center, 3333 Burnet Ave, Cincinnati, OH 45229; e-mail: rebecca.marsh@cchmc.org.

- regimen for nonmalignant disorders. *Bone Marrow Transplant*. 2005;35(4):345-352.
9. Cooper N, Rao K, Gilmour K, et al. Stem cell transplantation with reduced-intensity conditioning for hemophagocytic lymphohistiocytosis. *Blood*. 2006;107(3):1233-1236.
  10. Cooper N, Rao K, Goulden N, Webb D, Amrolia P, Veys P. The use of reduced-intensity stem cell transplantation in haemophagocytic lymphohistiocytosis and Langerhans cell histiocytosis. *Bone Marrow Transplant*. 2008;42(suppl 2):S47-50.
  11. Marsh RA, Vaughn G, Kim MO, et al. Reduced-intensity conditioning significantly improves survival of patients with hemophagocytic lymphohistiocytosis undergoing allogeneic hematopoietic cell transplantation. *Blood*. 2010;116(26):5824-5831.
  12. Lankester AC, Visser LF, Hartwig NG, et al. Allogeneic stem cell transplantation in X-linked lymphoproliferative disease: two cases in one family and review of the literature. *Bone Marrow Transplant*. 2005;36(2):99-105.
  13. Booth C, Gilmour KC, Veys P, et al. X-linked lymphoproliferative disease due to SAP/SH2D1A deficiency: a multicenter study on the manifestations, management and outcome of the disease. *Blood*. 2011;117(1):53-62.
  14. Kanegane H, Yang X, Zhao M, et al. Clinical features and outcome of X-linked lymphoproliferative syndrome type 1 (SAP deficiency) in Japan identified by the combination of flow cytometric assay and genetic analysis. *Pediatr Allergy Immunol*. 2012;23(5):488-493.
  15. Rigaud S, Fondaneche MC, Lambert N, et al. XIAP deficiency in humans causes an X-linked lymphoproliferative syndrome. *Nature*. 2006;444(7115):110-114.
  16. Marsh RA, Madden L, Kitchen BJ, et al. XIAP deficiency: a unique primary immunodeficiency best classified as X-linked familial hemophagocytic lymphohistiocytosis and not as X-linked lymphoproliferative disease. *Blood*. 2010;116(7):1079-1082.
  17. Yang X, Kanegane H, Nishida N, et al. Clinical and genetic characteristics of XIAP deficiency in Japan. *J Clin Immunol*. 2012;32(3):411-420.
  18. Worthey EA, Mayer AN, Syverson GD, et al. Making a definitive diagnosis: successful clinical application of whole exome sequencing in a child with intractable inflammatory bowel disease. *Genet Med*. 2011;13(3):255-262.
  19. Duckett CS, Nava VE, Gedrich RW, et al. A conserved family of cellular genes related to the baculovirus iap gene and encoding apoptosis inhibitors. *EMBO J*. 1996;15(11):2685-2694.
  20. Harlin H, Reffey SB, Duckett CS, Lindsten T, Thompson CB. Characterization of XIAP-deficient mice. *Mol Cell Biol*. 2001;21(10):3604-3608.
  21. Jost PJ, Grabow S, Gray D, et al. XIAP discriminates between type I and type II FAS-induced apoptosis. *Nature*. 2009;460(7258):1035-1039.
  22. Rumble JM, Oetjen KA, Stein PL, Schwartzberg PL, Moore BB, Duckett CS. Phenotypic differences between mice deficient in XIAP and SAP, two factors targeted in X-linked lymphoproliferative syndrome (XLP). *Cell Immunol*. 2009;259(1):82-89.
  23. Fakler M, Loeder S, Vogler M, et al. Small molecule XIAP inhibitors cooperate with TRAIL to induce apoptosis in childhood acute leukemia cells and overcome Bcl-2-mediated resistance. *Blood*. 2009;113(8):1710-1722.
  24. Schimmer AD, Estey EH, Borthakur G, et al. Phase I/II trial of AEG35156 X-linked inhibitor of apoptosis protein antisense oligonucleotide combined with idarubicin and cytarabine in patients with relapsed or primary refractory acute myeloid leukemia. *J Clin Oncol*. 2009;27(28):4741-4746.
  25. Marsh RA, Villanueva J, Zhang K, et al. A rapid flow cytometric screening test for X-linked lymphoproliferative disease due to XIAP deficiency. *Cytometry B Clin Cytom*. 2009;76(5):334-344.
  26. Bacigalupo A, Ballen K, Rizzo D, et al. Defining the intensity of conditioning regimens: working definitions. *Biol Blood Marrow Transplant*. 2009;15(12):1628-1633.
  27. Przepiorka D, Weisdorf D, Martin P, et al. 1994 Consensus Conference on Acute GVHD Grading. *Bone Marrow Transplant*. 1995;15(6):825-828.
  28. Filipovich AH. Diagnosis and manifestations of chronic graft-versus-host disease. *Best Pract Res Clin Haematol*. 2008;21(2):251-257.
  29. Shaw BE, Arguello R, Garcia-Sepulveda CA, Madrigal JA. The impact of HLA genotyping on survival following unrelated donor haematopoietic stem cell transplantation. *Br J Haematol*. 2010;150(3):251-258.
  30. Pachlopnik Schmid J, Canioni D, Moshous D, et al. Clinical similarities and differences of patients with X-linked lymphoproliferative syndrome type 1 (XLP-1/SAP deficiency) versus type 2 (XLP-2/XIAP deficiency). *Blood*. 2011;117(5):1522-1529.