

Tim Littlewood

Department of Haematology,
Oxford University Hospitals NHS Trust, Oxford University Hospital,
Oxford, United Kingdom

Tessa L. Holyoake

Paul O'Gorman Leukaemia Research Centre, Institute of Cancer Sciences,
College of Medical, Veterinary and Life Sciences, University of Glasgow,
Glasgow, United Kingdom

Mhairi Copland

Paul O'Gorman Leukaemia Research Centre, Institute of Cancer Sciences,
College of Medical, Veterinary and Life Sciences, University of Glasgow,
Glasgow, United Kingdom

Anthony V. Moorman

Leukaemia Research Cytogenetics Group,
Northern Institute for Cancer Research, Newcastle University,
Newcastle upon Tyne, United Kingdom

Christine J. Harrison

Leukaemia Research Cytogenetics Group,
Northern Institute for Cancer Research, Newcastle University,
Newcastle upon Tyne, United Kingdom

Paresh Vyas

Department of Haematology,
Oxford University Hospitals NHS Trust, Oxford University Hospital and
MRC Molecular Haematology Unit, Weatherall Institute of Molecular Medicine,
University of Oxford,
Oxford, United Kingdom

*T.E., C.J.S., and R.K. contributed equally to this work.

The online version of this article contains a data supplement.

Acknowledgments: P.V. acknowledges funding from the Medical Research Council (MRC) Molecular Hematology Unit, MRC Disease Team Award, the Leukemia Lymphoma Research Specialist Program Grant 08030, Cancer Research UK Program Grant C7893/A12796, and the National Institute for Health Research (NIHR) Oxford Biomedical Research Center based at Oxford University Hospitals Trust, Oxford, United Kingdom. M.C. acknowledges funding from the Scottish Funding Council (Fellowship SCD/04) and Leukemia and Lymphoma Research (grant 11017). C.S., A.V.M. and C.J.H. acknowledge Leukemia Lymphoma Research Specialist Program Grant 11004.

Contribution: T.E., A.P., A.M., T.L., and P.V. collected clinical and laboratory data; C.J.S. performed genetic analysis; R.K. and M.C. performed kinase sensitivity assays and analyzed the data; T.L.H., A.V.M., and C.J.H. analyzed laboratory data; J.S. and A.K.M. performed some of the cytogenetic investigations; T.E. and P.V. wrote the manuscript; and all authors edited the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Paresh Vyas, MRC Molecular Haematology Unit, WIMM, Oxford OX3 9DU, Oxford, United Kingdom; e-mail: paresh.vyas@imm.ox.ac.uk.

References

1. Graux C, Cools J, Melotte C, et al. Fusion of NUP214 to ABL1 on amplified episomes in T-cell acute lymphoblastic leukemia. *Nat Genet*. 2004;36(10):1084-1089.
2. Graux C, Stevens-Kroef M, Lafage M, et al. Heterogeneous patterns of amplification of the NUP214-ABL1 fusion gene in T-cell acute lymphoblastic leukemia. *Leukemia*. 2009;23(1):125-133.
3. Roberts KG, Morin RD, Zhang J, et al. Genetic alterations activating kinase and cytokine receptor signaling in high-risk acute lymphoblastic leukemia. *Cancer Cell*. 2012;22(2):153-166.
4. Mullighan CG, Goorha S, Radtke I, et al. Genome-wide analysis of genetic alterations in acute lymphoblastic leukaemia. *Nature*. 2007;446(7137):758-764.
5. Parker H, An Q, Barber K, et al. The complex genomic profile of ETV6-RUNX1 positive acute lymphoblastic leukemia highlights a recurrent deletion of TBL1XR1. *Genes Chromosomes Cancer*. 2008;47(12):1118-1125.
6. Loudin MG, Wang J, Leung HC, et al. Genomic profiling in Down syndrome acute lymphoblastic leukemia identifies histone gene deletions associated with altered methylation profiles. *Leukemia*. 2011;25(10):1555-1563.
7. De Keersmaecker K, Versele M, Cools J, Superti-Furga G, Hantschel O. Intrinsic differences between the catalytic properties of the oncogenic NUP214-ABL1 and BCR-ABL1 fusion protein kinases. *Leukemia*. 2008;22(12):2208-2216.
8. Deenik W, Beverloo HB, van der Poel-van de Luytgaarde SC, et al. Rapid complete cytogenetic remission after upfront dasatinib monotherapy in a patient with a NUP214-ABL1-positive T-cell acute lymphoblastic leukemia. *Leukemia*. 2009;23(3):627-629.
9. Clarke S, O'Reilly J, Romeo G, Cooney J. NUP214-ABL1 positive T-cell acute lymphoblastic leukemia patient shows an initial favorable response to imatinib therapy post relapse. *Leuk Res*. 2011;35(7):e131-e133.

To the editor:

Role of fecal calprotectin as biomarker of gastrointestinal GVHD after allogeneic stem cell transplantation

We read with interest the article of Rodriguez-Otero et al.¹ The authors studied the ability of fecal calprotectin (FC), α -1 antitrypsin, and elastase to diagnose acute gastrointestinal GVHD (GI-GVHD) after allogeneic stem cell transplantation (SCT). In their experience, FC and α -1 antitrypsin increased in patients with GI-GVHD, but there was no statistic difference compared with control groups. On the other hand, high levels of both markers at the time of diagnosis were predictive of steroid-resistant GVHD. In past years, our group also investigated the role of FC as a noninvasive biomarker of GVHD. We enrolled a cohort of 59 hematologic patients consecutively submitted to allogeneic SCT, and studied the level of FC in patients who developed GI-GVHD, non-GI-GVHD, and in patients with infective colitis. We also included a control group of 9 patients with aspecific colitis after autologous SCT. FC was detected at the onset of symptoms and before starting any therapy. Stool collection was performed by Calprest device and the protein level was measured by ELISA

assay (Calprest test; Eurospital). Data were analyzed using IBM SPSS Statistics 20 Core System and Prism Version 3.0 software (GraphPad). Diagnosis and staging of acute GVHD (aGVHD) and chronic GVHD (cGVHD) was made according to current criteria.^{2,3} FC was higher in patients with acute GI-GVHD (GI-aGVHD) than in non-GI-aGVHD (500 mg/Kg vs 95 mg/Kg; $P = .0003$; Figure 1A), and in stage III-IV GI-aGVHD than in the others; although, no statistic difference was observed in this case.

After treatment, in 2 of 3 responsive patients, FC value decreased to less than 15 mg/Kg. In contrast, FC was lower in patients with infective colitis compared with GI-aGVHD (106 mg/Kg vs 500 mg/Kg; $P = .0039$; Figure 1B). Comparing patients with GI-aGVHD, patients with infective enteritis and patients with both conditions, the median level of FC was 500 mg/Kg, 106 mg/Kg, and 475 mg/Kg, respectively ($P = .0096$; Figure 1C). FC was also lower in the control group of patients submitted to autologous SCT who developed mucositis and

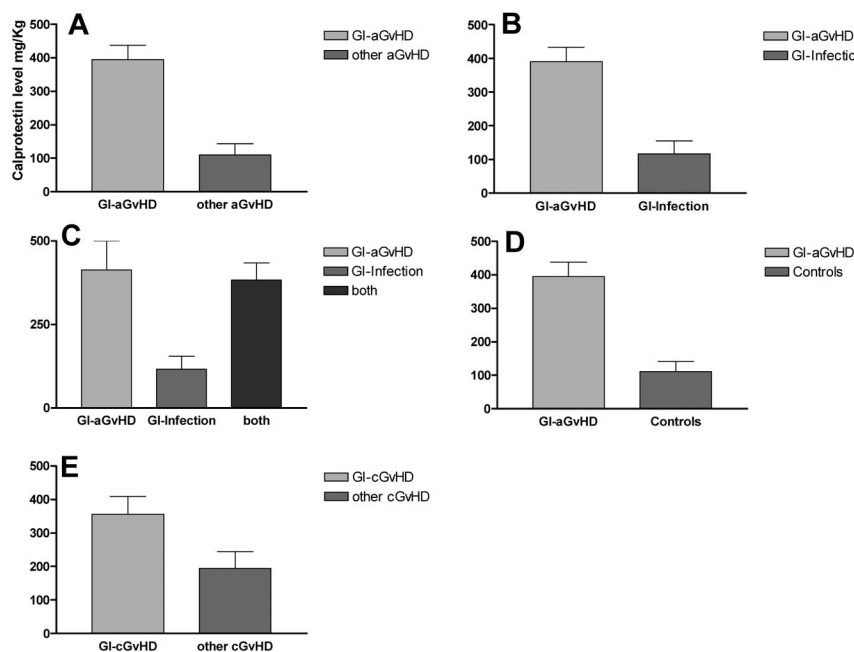


Figure 1. FC levels in different settings. (A) FC in patients with GI-aGVHD and other organ involvement aGVHD. (B) FC in patients with GI-aGVHD and infective enteritis. (C) FC in patients with GI-aGVHD, infective enteritis and concomitant GI-aGVHD, and infective enteritis. (D) FC in patients with GI-aGVHD and patients with diarrhea after autologous SCT. (E) FC in patients with GI-cGVHD and other organ involvement cGVHD.

diarrhea with a FC median level of 92 mg/Kg versus 500 mg/Kg ($P = .0012$; Figure 1D). Furthermore, we analyzed FC level at the onset of cGVHD. Again it was higher in patients with GI involvement than in non-GI-cGVHD (450 mg/Kg vs 94.5 mg/Kg; $P = .0229$; Figure 1E). Although no statistic difference was seen, FC was higher for score-3 GI-cGVHD than in score-2 (475 mg/Kg vs 171.5 mg/Kg, respectively). Using an arbitrary cut-off point value of 160 mg/Kg, sensitivity of the test was 100%, specificity 81.8% with a positive predictive value of 86%, and a negative predictive value of 100%. The area under receiver operating characteristic (ROC) curve for the test was 0.942 (confidence interval: 0.848-1.000). Consistent data are recently reported also by Bastos Oreiro et al.⁴ In conclusion, fecal calprotectin could be considered as a possible sensitive marker of GI-GVHD given its ability to distinguish GI-GVHD manifestation from other causes of diarrhea, such as infective colitis or aspecific enteritis. Moreover, fecal calprotectin was a noninvasive test and samples could be easily collected by patients themselves or by the nursing staff.

***Patrizia Chiusolo**

Hematology Department, Università Cattolica del Sacro Cuore, Rome, Italy

***Elisabetta Metafuni**

Hematology Department, Università Cattolica del Sacro Cuore, Rome, Italy

Sabrina Giammarco

Hematology Department, Università Cattolica del Sacro Cuore, Rome, Italy

Silvia Bellesi

Hematology Department, Università Cattolica del Sacro Cuore, Rome, Italy

Nicola Piccirillo

Hematology Department, Università Cattolica del Sacro Cuore, Rome, Italy

Chiara Fanali

Biochemistry Department, Università Cattolica del Sacro Cuore, Rome, Italy

Massimo Castagnola

Biochemistry Department, Università Cattolica del Sacro Cuore, Rome, Italy

Cecilia Zuppi

Biochemistry Department, Università Cattolica del Sacro Cuore, Rome, Italy

Teresa De Michele

Biochemistry Department, Università Cattolica del Sacro Cuore, Rome, Italy

Giuseppe Leone

Hematology Department, Università Cattolica del Sacro Cuore, Rome, Italy

Simona Sica

Hematology Department, Università Cattolica del Sacro Cuore, Rome, Italy

*P.C. and E.M. contributed equally to this manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Patrizia Chiusolo, MD, Hematology Department, Università Cattolica del Sacro Cuore, Largo Agostino Gemelli 8, 00168 Rome, Italy; e-mail: p.chiusolo@rm.unicatt.it.

References

- Rodriguez-Otero P, Porcher R, de Latour RP, et al. Fecal calprotectin and alpha-1 antitrypsin predict severity and response to corticosteroid in gastrointestinal graft-versus host disease. *Blood*. 2012;119(24):5909-5917.
- Filipovich AH, Weisdorf D, Pavletic S, et al. National Institute of Health consensus development project on criteria for clinical trials in chronic graft-versus-host disease: I. Diagnosis and staging working group report. *Biol Blood Marrow Transplant*. 2005;11(12):945-956.
- Przepiorka D, Weisdorf D, Martin P, et al. 1994 Consensus Conference on acute GVHD grading. *Bone Marrow Transplant*. 1995;15(6):825-828.
- Bastos Oreiro MB, Castilla-Llorente C, de la Guia AL, et al. Fecal calprotectin in allogeneic stem cell transplantation for the diagnosis of acute intestinal graft versus host disease. *Bone Marrow Transplant*. 2012;47(9):1241-1242.