Risk stratification-directed donor lymphocyte infusion could reduce relapse of standard-risk acute leukemia patients after allogeneic hematopoietic stem cell transplantation

*Chen-Hua Yan,¹ *Dai-Hong Liu,¹ Kai-Yan Liu,¹ Lan-Ping Xu,¹ Yan-Rong Liu,¹ Huan Chen,¹ Wei Han,¹ Yu Wang,¹ Ya-Zhen Qin,¹ and Xiao-Jun Huang¹

¹Peking University People's Hospital, Peking University Institute of Hematology, Beijing Key Laboratory of Hematopoietic Stem Cell Transplantation, Beijing, Peoples Republic of China

We studied the impact of risk stratificationdirected interventions for minimal residual disease (MRD) on relapse and disease-free survival (DFS) prospectively in 814 subjects with standard-risk acute leukemia receiving allotransplantation in first or second complete remission. A total of 709 subjects were MRD⁻ after transplantation (Group A); 105 subjects were MRD⁺, 49 received low-dose IL-2 (Group B), and 56 received modified donor lymphocyte infusion (DLI) with or without low-dose IL-2 (Group C). Posttransplantation immune suppression for GVHD was also modified based on MRD state. The cumulative risk of relapse was significantly less and DFS was significantly better in subjects in Group C than in subjects in Group B (P = .001 and P = .002, respectively), but was not different from subjects in Group A (P = .269 and P = .688, respectively). Multivariate analyses confirmed that MRD state and modified DLI were significantly correlated with relapse (P = .000, odds ratio [OR] = 0.255 and P = .000,OR = 0.269)and DFS (P = .001,OR = 0.511 and P = .006. OR = 0.436. respectively). These data suggest that risk stratification-directed interventions with modified DLI in patients with standardrisk acute leukemia who are MRD+ after transplantation may improve transplantation outcomes. (Blood. 2012;119(14): 3256-3262)

Introduction

Allogeneic hematopoietic cell transplantation (HSCT) is an effective therapy for acute leukemia, but relapse remains an important problem.^{1,2} Therapy options for relapse include stopping immune suppression, reinduction of chemotherapy, donor lymphocyte infusion (DLI), and another transplantation used alone or in combination. However, the efficacy of these interventions is limited. One approach to the relapse problem in persons with standard-risk acute leukemia is to intervene before hematologic or pathologic relapse occurs based on posttransplantation analysis or minimal residual disease (MRD) using immune or molecular techniques.

Recent studies have indicated that leukemia-associated aberrant immune phenotypes (LAIPs) identified by multiparameter flow cytometry (FCM) and specific fusion genes detected by real-time quantitative PCR (RQ-PCR) can be used to detect and monitor MRD.^{3,4} Other studies have also confirmed the feasibility of using MRD state as determined by FCM and RO-PCR as prognostic variables in acute leukemia treated with chemotherapy.^{5,6} The use of leukemia-specific fusion genes is also effective for monitoring MRD, but is only useful in approximately 25% of cases. Wilms tumor gene-1 (WTI) is overexpressed in approximately 90% of cases of acute leukemia and is therefore useful in monitoring MRD in most persons with standard-risk acute leukemia. Several studies have suggested that quantification of WT1 by RQ-PCR can be used to monitor MRD after chemotherapy and can predict relapse.^{7,8} Recently, we reported a strong correlation between LAIPs and WT1 and leukemia relapse, disease-free survival (DFS), and survival in patients with acute leukemia receiving allotransplantation.^{9,10}

DLI is an effective posttransplantation therapy for leukemia relapse,^{11,12} but is associated with a substantial risk of GVHD. Recently, we modified the usual DLI procedure by treating patients with G-CSF–mobilized peripheral blood cells, followed by short-term immune suppression after DLI. Our data suggest that modified DLI is associated with less GVHD and that post-DLI immune suppression does not reduce the GVL effects.^{13,14} Our recent data also indicate that low-dose IL-2 given after transplantation reduces relapse risk in persons with high-risk acute lymphoblastic leukemia (ALL) receiving allotransplantation.¹⁵ Based on these observations, we performed a prospective study to investigate the impact of risk stratification–directed interventions on transplantation outcomes in persons with standard-risk acute leukemia receiving allotransplantations based on MRD state.

Methods

Eligibility criteria

Consecutive subjects receiving non-T cell-depleted allotransplantations at Peking University Institute of Hematology from January 1, 2006 to November 30, 2010 were eligible if they met one of these criteria: (1) had standard-risk acute leukemia defined as first or second complete remission without t(9;22), t(15;17), inv(16), t(16;16), or t(8;21) mutations; or (2) had myelodysplastic syndrome–refractory anemia with excess blasts. The study was approved by the ethics committee of Peking University People's

Submitted September 17, 2011; accepted February 10, 2012. Prepublished online as *Blood* First Edition paper, February 14, 2012; DOI 10.1182/blood-2011-09-380386.

*C.-H.Y. and D.-H.L. contributed equally to this work.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2012 by The American Society of Hematology



Figure 1. Intervention strategy. Diagram of intervention strategy based on MRD state (A) and subgroup (B).

Hospital and written informed consent was obtained from all subjects before study entry in accordance with the Declaration of Helsinki.

Transplantations

Transplantations were performed as described previously.^{16,17} Recipients of HLA-identical related transplantations received busulfan (0.8 mg/kg IV every 6 hours for 12 doses) and cyclophosphamide (1.8 g/m²/d for 2 days) or total body irradiation (7.7 Gy) given as 1 fraction followed by cyclophosphamide. Recipients of HLA-haploidentical related transplantation and recipients of HLA-matched transplantation from unrelated donors were conditioned with busulfan, cyclophosphamide, and human antithymocyte globulin 2.5 mg/kg/d IV for 4 days (Sangstat) or total body irradiation, cyclophosphamide, and antithymocyte globulin. All subjects received G-CSF–mobilized BM and blood cells. After transplantation, they received cyclosporine (CSA), mycophenolate mofetil, and short-term methotrexate (MTX).

MRD monitoring and definitions

Data were available on LAIPs at diagnosis in 91.8% (747 of 814) of subjects and on WT1 overexpression in 63.1% (514 of 814) of subjects; 91.8% of subjects had at least 1 measurement, 63.1% of subjects had both. All subjects were monitored for MRD after transplantation using LAIPs and WT1. LAIPs were detected by 4-color FCM. Different Ab combinations were used in B-ALL, T-ALL, and acute myeloid leukemia as described previously.^{10,18} A total of 1 000 000 events were collected for analysis routinely. When cell numbers were limited, a minimal 750 000 events were collected. FCM⁺ was defined as > 0.001% of cells with a LAIP phenotype in ≥ 1 BM samples after transplantation. Sensitivity was 79% and specificity was 85% for persons with ALL.¹⁰ WT1 expression was evaluated by TaqMan-based RQ-PCR technology, as described previously.9 WT1+ was defined as a transcript level > 0.60%. Sensitivity was 96% and specificity was 79% for persons with acute leukemia.9 Samples were analyzed at 1, 2, 3, 4.5, 6, 9, and 12 months after transplantation and at 6-month intervals thereafter. When FCM- or WT1+ was detected, the test was repeated 2 weeks later. Subjects were scored as MRD+ if they had 2 consecutive positive results using FCM or WT1 or were both FCM⁺ and WT1⁺ in a single sample within 1 year after transplantation.

Therapy of MRD⁺ subjects after transplantation

Discontinuing immune suppression. Posttransplantation immune suppression was immediately tapered and then discontinued in subjects who were $MRD^+ \leq 100$ days after transplantation. Subjects who were $MRD^+ > 100$ days after transplantation had immune suppression immediately discontinued.

IL-2 therapy. IL-2 was given at a dose of 1×10^6 U/d subcutaneously for 14 days. One or more subsequent cycles were given after a 14-day interval for up to 12 cycles¹⁵ unless the subject developed GVHD or \geq grade 3 toxicity using NCI Common Terminology Criteria for Adverse Events Version 4.¹⁹

Modified DLI. For modified DLI,^{13,16} G-CSF-mobilized peripheral blood stem cells were given instead of the more common unstimulated donor blood lymphocytes. The median doses of mononuclear cells, CD3⁺ cells, CD4⁺ lymphocytes, and CD8⁺ lymphocytes were 1 (range, 0.75-2.00) × 10⁸/kg, 0.38 (range, $(0.15-0.93) \times 10^{8}$ /kg, 0.19 (range, 0.10-0.54) $\times 10^{8}$ /kg, and 0.14 (range, 0.06-0.43 × 10⁸/kg, respectively. After DLI, subjects received immunosuppressive drugs such as CSA or MTX to prevent GVHD. Subjects receiving DLI from an HLA-identical related donor received GVHD prophylaxis for 2-4 weeks; subjects receiving DLI from an HLA-matched unrelated or HLA-haploidentical donor received GVHD prophylaxis for 4-6 weeks at the discretion of the attending physicians (and usually depending on the patient's GVHD status after DLI). The starting dose of CSA was 2.5 mg/kg/d and the dose was adjusted to maintain a plasma concentration > 100 ng/mL. MTX was given at 10 mg IV on days 1, 4, 8, and weekly thereafter for 2-6 weeks.

Intervention strategy based on MRD state

From January 1, 2006 to December 30, 2006, subjects who were MRD⁺ and without GVHD received low-dose IL-2, whereas subjects with GVHD received GVHD therapy followed by low-dose IL-2. DLI was given if the subject remained MRD⁺ after IL-2 therapy (the intervention strategy is shown in Figure 1A).

From January 1, 2007 to November 30, 2010, DLI or IL-2 was given to MRD⁺ subjects based on whether a DLI donor was available. However, subjects with GVHD first received GVHD therapy, after which time MRD testing was repeated. Those subjects who then became MRD⁻ received low-dose IL-2, whereas those subjects who remained MRD⁺ received DLI or IL-2 based on donor availability. Subjects receiving DLI also received antileukemia chemotherapy 48-72 hours before DLI if they agreed. Antileukemia to myelodysplastic syndrome and MTX 1 g/m² for 7 days in subjects with Acute myeloid leukemia or myelodysplastic syndrome and MTX 1 g/m² for 1 day in subjects with ALL. MRD state was monitored at 1, 2, 3, 4.5, 6, 9, and 12 months after the interventions and at 6-month intervals thereafter.

Definitions and evaluation

Leukemia relapse was scored as BM, extramedullary, or both by common morphological criteria. FCM and molecular data were not used to define relapse. GVHD was scored as acute or chronic based on published criteria.²⁰ Chimerism analyses were done by DNA fingerprinting of short tandem-repeat on blood samples and/or chromosome FISH of BM samples. Evaluations were done at 1, 2, 3, 4.5, 6, 9, and 12 months after transplantation and at 6-month intervals thereafter.

Statistical analyses

Cumulative incidences of relapse, transplant-related mortality (TRM), and GVHD were calculated by accounting for competing risks.²¹ DFS was



Figure 2. Relapse after transplantation. (A) Cumulative incidence of relapse after transplantation between Group A and Groups B and C (n = 814). (B) Cumulative incidence of relapse after transplantation comparing Groups A, B, and C (n = 814). (C) Cumulative incidence of relapse after intervention for MRD between Groups B and C (n = 105). A total of 709 subjects were MRD⁻ after transplantation (Group A); 105 subjects were MRD⁺ after transplantation (Groups B and C); 49 subjects received IL-2 (Group B) and 56 subjects received DLI (Group C).

plotted using the Kaplan-Meier method and was compared using the log-rank test.²² Univariate analyses were performed using the χ^2 test for categorical variables and the Mann-Whitney test for continuous variables. Multivariate analyses were performed using a Cox proportional hazards model. The end point of follow-up for all surviving subjects was May 31, 2011. Unless otherwise specified, all *P* values are 2-sided and *P* < .05 was considered significant. The SPSS 13.0 and R 2.6.1 software packages were used for data analyses.

Results

Subject groups

A total of 709 of 814 consecutive subjects were MRD⁻ at all of the time points tested after transplantation (Group A); 105 subjects were MRD⁺ and in 12 of these, this was based on FCM and *WT1* data in 1 sample. Ten subjects had 2 FCM⁺ results and 83 had 2 *WT1*⁺ results; 49 of these 105 MRD⁺ subjects only received low-dose IL-2 (Group B) and another 56 received DLI with or without low-dose IL-2 (Group C).

From January 1, 2006 to December 30, 2006, 11 subjects received IL-2, 6 of whom finally received DLI because of persistent MRD⁺. From January 1, 2007 to November 30, 2010, 94 subjects received interventions: 41 of 56 subjects without GVHD at the time of MRD⁺ received DLI; the remaining 15 received IL-2 because a DLI donor was unavailable. Thirty-eight additional subjects had GVHD at the time of MRD⁺ after GVHD therapy. Nine of these subjects then received DLI and 7 received IL-2 because of DLI-donor unavailability. The 22 remaining subjects became MRD⁻ after control of GVHD and received IL-2 only (Figure 1B).

Relapse

Cumulative incidence of leukemia relapse at 3 years after transplantation was 22.0% (95% confidence interval [95% CI], 18.4%-25.6%) in the 814 subjects; 18.1% (95% CI, 14.6%-21.6%) in Group A, 64.4% (95% CI, 44.8%-84.0%) in Group B, and 27.8% (95% CI, 12.1%-43.5%) in Group C. Cumulative incidence of relapse in Group A was significantly lower than in Groups B and C combined (P < .000) or Group B alone (P < .000). Cumulative incidence of relapse in Group C was significantly lower than Group B (P = .001) but not significantly different from Group A (P = .269). In addition, there was no significant difference in incidence of relapse after transplantation between the subjects receiving chemotherapy before DLI and subjects not receiving chemotherapy in Group C (P = .822; Figure 2 and Table 1). Disease type (P = .016), remission state (P = .001), MRD state after transplantation (P = .001) and intervention for MRD (P = .001) were significantly correlated with cumulative risk of relapse in univariate analyses. No other variable tested significantly correlated with relapse risk in univariate analysis, including age, patient's sex, cytogenetic subgroup, number of induction chemotherapies, donor type, conditioning regimen, acute and chronic GVHD before MRD, interval from HSCT to MRD, chimerism status and acute and chronic GVHD after intervention of MRD. In multivariate analysis MRD-negativity after transplantation (P = .000, OR = 0.255) and receiving DLI (P = .000, OR = 0.269) were significantly correlated with a lower cumulative risk of leukemia relapse.

TRM

There were no significant differences in the incidence of TRM between Group A as well as Groups B and C combined (P = .080), between Groups B and C (P = .897), between Groups A and C (P = .115) or between Groups A and B (P = .112). In addition, there was a trend that the incidence of TRM after transplantation in the subjects receiving chemotherapy before DLI was higher than that in subjects not receiving chemotherapy in Group C (22.4% vs 8.5%, P = .055; Table 1).

GVHD

Cumulative incidences of acute GVHD of any grade, \geq grade 2 and \geq grade 3 after transplantation were 30.2%, 15.2% and 4.9% in Group A, respectively. Cumulative incidences of chronic and extensive chronic GVHD after transplantation were 38.8% and 32.9% in Group A.

Cumulative incidence of acute GVHD of any grade after interventions was significantly higher in Group C than in Group B (P = .017). There were no significant differences in incidences of \geq grade 3 acute GVHD (P = .471), total chronic GVHD (P = .982) and extensive chronic GVHD (P = .858) after interventions between the Groups (Table 1). In addition, between the subjects receiving chemotherapy before DLI and subjects not receiving chemotherapy in Group C, there were no significant difference in incidence of acute GVHD of any grade (P = .655), \geq grade 3 acute GVHD (P = .752), total chronic GVHD (P = .934) and extensive chronic GVHD (P = .283).

OS

The median overall survival (OS) was 44 months (95% CI, 42-46 months). OS in Group A was significantly longer than that

Table 1. Subject characteristics and outcomes (N = 814)

		MRD ⁺	P †	MRD ⁺			DLI		
Variable	MRD-			IL-2	DLI	P ‡	Chemo- therapy	No chemo- therapy	P§
No. of subjects	709	105		49	56		32	24	
Subject age, y (range)	29 (2-59)	23 (4-59)	.012	25 (4-59)	22 (6-55)	.882	20 (6-54)	24.5 (12-55)	.379
Subject sex, n (%)	. ,	. ,	.556			.181		. ,	.938
Male	438 (61.8)	68 (64.8)		35 (71.4)	33 (58.9)		19 (59.4)	14 (58.3)	
Female	271 (38.2)	37 (35.2)		14 (28.6)	23 (41.1)		13 (40.6)	10 (41.7)	
Disease type, n (%)	ζ, γ	, , ,	.170	, ,	, ,	.950	. ,	. ,	.940
AML	378 (53.3)	61 (58.1)		29 (59.2)	32 (57.1)		18 (56.3)	14 (58.3)	
ALL	247 (34.8)	38 (36.2)		17 (34.7)	21 (37.5)		12 (37.5)	9 (37.5)	
MDS-RAEB	84 (11.8)	6 (5.7)		3 (6.1)	3 (5.4)		2 (6.3)	1 (4.2)	
Remission state, n (%)	, , , , , , , , , , , , , , , , , , ,	, ,	.103	. ,	. ,	.037		, ,	.679
CR1	559 (88.6)	85 (81.0)		42 (85.7)	43 (76.8)		22 (73.3)	18 (78.3)	
CR2	72 (11.4)	20 (19.0)		7 (14.3)	13 (23.2)		8 (26.7)	5 (21.7)	
Cvtogenetic subGroup, n (%)*	~ /	- (/	.103	· · · /	- (-)	.615	- (-)	- ()	.626
Intermediate	466 (65.7)	62 (59.0)		30 (61.2)	32 (57.1)		22 (68.8)	16 (66.7)	
Poor	63 (8.9)	14 (13.3)		5 (10.2)	9 (16.1)		4 (12.5)	5 (20.8)	
Not available	180 (25.4)	29 (27.6)		14 (28.6)	15 (26.8)		8 (18.8)	3 (12.5)	
No. of induction chemotherapies. n (%)			.095			.989	- ()	- ()	.892
1	433 (68.6)	66 (62.9)		31 (63.3)	35 (62.5)		20 (66.7)	14 (60.9)	
2	149 (23.6)	22 (20.9)		10 (20.4)	12 (21.4)		6 (20.0)	5 (21.7)	
- >2	49 (7.8)	17 (16.2)		8 (16.3)	9 (16.1)		4 (13.3)	4 (17.4)	
Type of donor, n (%)			080	- (,	- ()	002	. ()	. ()	214
HI A-identical related	215 (30.3)	40 (38 1)		14 (28.6)	26 (46 4)		13 (40.6)	13 (54 2)	
HLA-haploidentical related	438 (61.8)	53 (50.5)		24 (49.0)	29 (51.8)		19 (59.4)	10 (41.7)	
Unrelated	56 (7.9)	12 (11 4)		11 (22 4)	1 (1 8)		0 (0 0)	1 (4 2)	
Conditioning regimen, n (%)	00 (710)	.= ()	003	()	. (376	0 (010)	. (=)	797
TBI-based	2 (0,3)	4 (3.8)		1 (2 0)	1 (1.8)		1 (3 1)	2 (8.3)	
BU-based	707 (99 7)	101 (96.2)		48 (98 0)	55 (98.2)		31 (96.9)	22 (91 7)	
Acute GvHD pre-MBD p (%)				15 (30.6)	18 (32 1)	762	11 (34.3)	7 (29 1)	370
> Grade 2				8 (16.3)	8 (14.3)	708	6 (18 8)	2 (8.3)	269
> Grade 3				0 (0 0)	0 (0 0)	989	0 (0 0)	0 (0 0)	1 000
Chronic GvHD pre-MBD n (%)				13 (26 5)	12 (21.4)	326	7 (21.9)	5 (20.8)	788
Extensive GvHD pre-MBD				4 (8 2)	5 (8 9)	768	3 (9.3)	2 (8.3)	.700
Chimerism status pre-MBD n (%)				. (0.2)	0 (010)	856	0 (010)	2 (0.0)	986
Full dopor				46 (93 9)	54 (96.4)		31 (96 9)	23 (95.8)	
Mixed				3 (6 1)	2 (3 6)		1 (3 1)	1 (4 2)	
Full recipient				0 (0 0)	0 (0 0)		0 (0 0)	0 (0 0)	
Interval from HSCT to MBD_d (range)				140 (30-365)	135 (30-435)	455	132 (30-435)	135 (30-365)	980
Chemotherapy before intervention n (%)				0 (0 0)	32 (57 1)	.400	102 (00 400)	100 (00 000)	.000
Discontinuing immunosuppressions before				26 (53 1)	28 (50 0)	754	16 (50.0)	12 (50.0)	989
intervention n (%)				20 (00.1)	20 (00.0)		10 (00.0)	12 (00.0)	.000
Acute GvHD after intervention, n (%)				10.2	30.8	017	33.3	29.0	655
> Grade 2				82	27.9	017	30.3	26.2	701
> Grade 3				4.1	84	471	9.8	7.3	752
Chronic GvHD after intervention n (%)				37.3	42.9	982	45.5	39.4	934
Extensive chronic GvHD				30.5	34.2	858	39.4	30.1	283
Incidence of TBM at 3 v after HSCT n (%)	19.7	12 7	080	11.4	14.4	897	22.4	85	.200
Actual relanse rate in (%)	103 (14 5)	37 (35.2)	.000	25 (51 0)	12 (21 4)	.007	5 (15 6)	7 (29 2)	222
Incidence of relapse at 3 y after HSCT,	18.1	46.0	.000	64.4	27.8	.001	26.4	32.7	.822
OS at 3 v after HSCT_n (%)	66.0	44 0	005	28.1	58.3	022	60.5	57.8	496
Median OS after HSCT mo (95% CI)	44.39	33.17	.005	18.97	38.53	.022	40.32	38.53	.496
	(33.87-54.90)	(25.23-41.11)		(3.13-34.80)	(33.78-50.86)	.022	(33.78-50.86)	(33.32-54.25)	
DES at 3 y atter HSUI, n	01.0	40.3	.041	24.1	0.50	.002	51.2	ο/.ö	.327
Median DFS after HSC1, mo (95% CI)	42.40	28.00	.041	13.33	38.53	.002	40.52	38.53	.327
	(33.85-54.90)	(19.12 - 36.88)		(8.78-17.88)	(32.00-59.92)		(32.0-49.04)	(32.40-54.05)	

AML indicates acute myelogenous leukemia; MDS-RAEB, myelodysplastic syndrome-refractory anemia with excess blasts; CR, complete remission; BU, busulfan; and TBI, total body irradiation.

*Cytogenetic subGroups were according to the published data.23-25

†Comparison was performed between Group A and Groups B and C.

‡Comparison was performed between Groups B and C.

\$Comparison was performed between the subjects receiving chemotherapy prior to DLI and subjects not receiving chemotherapy in Group C.

in Groups B and C combined (P = .005) or Group B alone B (P = .022) and was not significantly different from Group (P = .027). OS in Group C was significantly longer than in Group A (P = .695). In addition, there was no significant difference in OS



Figure 3. OS after transplantation. (A) OS after transplantation between Group A and Groups B and C (n = 814). (B) OS after transplantation comparing Groups A, B, and C (n = 814). (C) OS after intervention for MRD between Groups B and C (n = 105). A total of 709 subjects were MRD⁻ after transplantation (Group A); 105 subjects were MRD⁺ after transplantation (Group B) and C); 49 subjects received IL-2 (Group B) and 56 subjects received DLI (Group C).

between the subjects receiving chemotherapy before DLI and subjects not receiving chemotherapy in Group C (P = .496; Figure 3 and Table 1).

Disease type (P = .025), remission state (P = .002), MRD state after transplantation (P = .003), and intervention for MRD (P = .001) were significantly correlated with OS in univariate analyses. No other variable tested was correlated significantly with OS in univariate analyses, including age, sex, cytogenetic subgroup, number of induction chemotherapies, donor type, conditioning regimen, acute and chronic GVHD pre-MRD, interval from HSCT to MRD, chimerism status, and acute and chronic GVHD after treatment of MRD. In multivariate analyses, MRD-negativity after transplantation (P = .007, OR = 0.644) and receiving DLI (P = .002, OR = 0.553) were correlated significantly with longer OS.

DFS

The median DFS was 42 months (95% CI, 40-44 months). DFS in Group A was significantly longer than that in Groups B and C combined (P = .041) or Group B alone (P = .001). DFS in Group C was significantly longer than in Group B (P = .002) and was not significantly different from Group A (P = .688). In addition, there was no significant difference in DFS between the subjects receiving chemotherapy before DLI and subjects not receiving chemotherapy in Group C (P = .327; Figure 4 and Table 1).

Disease type (P = .017), remission state (P = .002), MRD state after transplantation (P = .003), and intervention for MRD (P = .005) were correlated significantly with DFS in univariate analyses. In multivariate analyses, MRD-negativity after transplantation (P = .001, OR = 0.511) and receiving DLI (P = .006, OR = 0.436) were correlated significantly with longer DFS.

Discussion

The incidence of relapse in subjects who were MRD⁻ after transplantation was lower than in subjects who were MRD⁺ after transplantation (Groups B and C combined) and was lower than in the subset of patients who received only low-dose IL-2 (Group B). However, subjects in Group C who were MRD⁺ after transplantation and received modified DLI with or without IL-2 had a leukemia relapse rate similar to subjects in Group A. These data suggest that the intervention of DLI with or without IL-2 in MRD⁺ subjects after transplantation reduced relapse risk.

Similar results were seen with a DFS end point: comparable results with Groups A and C, both of which were significantly better than Group B. These data suggest that giving DLI with or without IL-2 to subjects who are MRD⁺ after transplantation results in no decrement in survival from causes other than relapse. Clearly, modified DLI with or without IL-2 produced better results than IL-2 only. Subjects receiving DLI with or without IL-2 had lower relapse rates (29.8% vs 68.2%, P = .001) and better DFS (52.5% vs 20.8%, P = .002).

In the present study, the incidence of relapse was 22.0% in the total population and 18.1% in subjects who were MRD⁻ after transplantation. Recent studies, however, have reported the cumulative relapse risk to be approximately 30%-40%.²⁶⁻³¹ This may be partly because the application of risk stratification–directed interventions for persons who were MRD⁺ reduced relapse after transplantation. These results also confirmed that risk stratification by MRD state after transplantation is reliable and is the foundation of risk stratification–directed interventions. However, it is notable that data from Group A subjects indicate that MRD negativity after transplantation does not guarantee continued complete remission.



Figure 4. DFS after transplantation. (A) DFS after transplantation between Group A and Groups B and C (n = 814). (B) DFS after transplantation comparing Groups A, B, and C (n = 814). (C) DFS after intervention for MRD between Groups B and C (n = 105). A total of 709 subjects were MRD⁻ after transplantation (Group A); 105 subjects were MRD⁺ after transplantation (Group B and C); 49 subjects received IL-2 (Group B) and 56 subjects received DLI (Group C).

The imperfect sensitivity was possibly because the MRD-detection platform itself was imperfect. Although the sensitivities for FCM⁺ in ALL subjects¹⁰ and *WT1*⁺ for acute leukemia subjects⁹ were more than 75% in previous studies, our definition of MRD⁺ in this study was stricter than in most other reports to avoid the excessive TRM related to the interventions for MRD⁺ state after transplantation. This certainly resulted in an imperfect sensitivity of MRD testing but increasing specificity. In the future, MRD markers used in risk stratification should be analyzed further to increase the sensitivity but to not reduce specificity. Nevertheless, in the present study, persons with standard-risk acute leukemia still benefited from risk stratification and risk stratification–directed interventions.

Our study was not randomized, a factor that mandates a replication of our results, ideally in a randomized clinical trial. We tried to compensate for the lack of randomization by considering variables know to be correlated with transplantation outcomes in this setting. Most were comparable between the cohort except for remission state before transplantation (P = .037), donor types (P = .002), and antileukemia chemotherapy pre-intervention (P = .001). In subjects receiving DLI with or without IL-2, more were in second complete remission before transplantation than those receiving only IL-2 (23.2% vs 14.3%, P = .037). However, the impact of this imbalance should be to increase rather than to decrease relapse risk. In addition, more subjects in the DLI cohort received antileukemia chemotherapy pre-intervention than in the IL-2–only cohort (57.1% vs none, P = .001). Although this may have potentially biased the results in favor of the patients receiving DLI, our data showed that there were no significant differences in the incidence of relapse, OS, or DFS in the subjects receiving chemotherapy before DLI and subjects not receiving chemotherapy in Group C (P > .05). In this study, the subjects in Group C received chemotherapy based on their choice and were not randomized. Furthermore, because the interval from chemotherapy to DLI was only 48-72 hours, it is difficult to evaluate whether the efficacy observed in subjects in Group C was due to chemotherapy or to DLI. However, in the present study, chemotherapy with DLI or DLI alone were effective methods of preventing relapse and improving survival in persons with standard-risk acute leukemia. In the future, a randomized study should be designed to compare the effect of chemotherapy plus DLI or DLI or chemotherapy alone on relapse and survival. In the present study, it is difficult to separate the role of discontinuation of immunosuppression from interventions (IL-2/ DLI), because approximately 50% of subjects receiving interventions had already discontinued immunosuppression.

Although results in subjects in Group B were inferior to those in Group C, our data do not rule out a benefit for intervention with IL-2 only in subjects MRD^+ after transplantation. Our previous study suggested a 75% relapse rate in subjects who were MRD^+

References

- Mortimer J, Blinder MA, Schulman S, et al. Relapse of acute leukemia after marrow transplantation: natural history and results of subsequent therapy. J Clin Oncol. 1989;7(1):50-57.
- Frassoni F, Barrett AJ, Grañena A, et al. Relapse after allogeneic bone marrow transplantation for acute leukaemia: a survey by the E.B.M.T. of 117 cases. *Br J Haematol.* 1988;70(3):317-320.
- Kern W, Danhauser-Riedl S, Ratei R, et al. Detection of minimal residual disease in unselected patients with acute myeloid leukemia using multiparameter flow cytometry for definition of leukemia-associated immunophenotypes and determination of their frequencies in normal bone marrow. *Haematologica*. 2003;88(6):646-653.
- Schnittger S, Weisser M, Schoch C, Hiddemann W, Haferlach T, Kern W. New score predicting for prognosis in PML-RARA+, AML1-ETO+, or CBFBMYH11+ acute myeloid leukemia based on quantification of fusion transcripts. *Blood*. 2003; 102(8):2746-2755.
- San Miguel JF, Vidriales MB, Lopez-Berges C, et al. Early immunophenotypical evaluation of minimal residual disease in acute myeloid leukemia identifies different patient risk groups and may contribute to postinduction treatment stratification. *Blood.* 2001;98(6):1746-1751.
- Kern W, Haferlach C, Haferlach T, Schnittger S. Monitoring of minimal residual disease in acute myeloid leukemia. *Cancer.* 2008;112(1):4-16.

after transplantation but not receiving any interventions.⁹ These results suggest that the GVL effects of IL-2 were probably inferior to DLI, but that treatment with IL-2 before MRD⁺ may be useful in patients with standard-risk acute leukemia.

GVHD is an important risk of DLI. However, we found no significant differences in subjects receiving IL-2 or DLI (P = .471) in the incidence of \geq grade 3 acute GVHD or TRM (P = .897). Our previous study suggested that modified DLI has the same antileukemia efficacy as conventional DLI but less acute GVHD and better survival. These observations may result from the different composition of the infused cells and short-term use of immunosuppressive drugs.^{13,14} Data from a randomized trial are needed to confirm our observation. Huang et al reported that the use of G-CSF may decrease T-cell responses indirectly through selective increase of plasmacytoid dendritic cell and monocytes and down-regulation of the costimulatory signal of CD28/B7.³²⁻³⁴ Morris et al confirmed that the use of G-CSF during blood-cell mobilization augments natural killer/T-cell-dependent CD8⁺ cytotoxicity, purportedly separating GVHD and GVL.³⁴

In conclusion, our study is the first to suggest that risk stratification-directed modified DLI may reduce relapse and improve survival of subjects with standard-risk acute leukemia after HSCT.

Acknowledgments

The authors thank Prof Robert Peter Gale (Imperial College, London, United Kingdom, and Celgene Corporation, Summit, NJ), for revising the manuscript.

This study was supported by National Natural Science Foundation of China (grant number 30971292), the HI-Tech Research Development Program of China 863 (2010-2014), and the Leading Program of Clinical Faculty accredited by the Ministry of Health of China (2010-2012).

Authorship

Contribution: C.-H.Y. and D.-H.L. analyzed the data and wrote the manuscript; Y.-R.L. and Y.-Z.Q. detected *WT1* and LAIPs and analyzed the data; X.-J.H. designed the research; and all authors provided patient data.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Xiao-Jun Huang, Peking University People's Hospital, Peking University Institute of Hematology, Beijing 100044, China; e-mail: xjhrm@medmail.com.cn.

- Weisser M, Kern W, Rauhut S, et al. Prognostic impact of RT-PCR-based quantification of WT1 gene expression during MRD monitoring of acute myeloid leukemia. *Leukemia*. 2005;19(8):1416-1423.
- Lapillonne H, Renneville A, Auvrignon A, et al. High WT1 expression after induction therapy predicts high risk of relapse and death in pediatric acute myeloid leukemia. *J Clin Oncol.* 2006; 24(10):1507-1515.
- Zhao XS, Jin S, Zhu HH, et al. Wilms' tumor gene 1 expression: an independent acute leukemia prognostic indicator following allogeneic hematopoietic SCT [published online ahead of print June 6, 2011]. Bone Marrow Transplant. doi:10.1038/bmt.2011.121.

- Zhao XS, Liu YR, Zhu HH, et al. Monitoring MRD with flow cytometry: an effective method to predict relapse for ALL patients after allogeneic hematopoietic stem cell transplantation. Ann Hematol. 2012;91(2):183-192.
- Collins Jr RH, Shpilberg O, Drobyski WR, et al. Donor leukocyte infusions in 140 patients with relapsed malignancy after allogeneic bone marrow transplantation. J Clin Oncol. 1997;15(2): 433-444.
- Dazzi F, Szydlo RM, Cross NCP, et al. Durability of responses following donor lymphocyte infusions for patients who relapse after allogeneic stem cell transplantation for chronic myeloid leukemia. *Blood.* 2000;96(8):2712-2716.
- Huang XJ, Wang Y, Liu DH, et al. Administration of short-term immunosuppressive agents after DLI reduces the incidence of DLI associated acute GVHD without influencing the GVL effect. Bone Marrow Transplant. 2009;44(9):309-316.
- Huang XJ, Guo NL, Ren HY, et al. The comparison of GVL effects between the patients receiving donor peripheral blood stem cells and donor lymphocytes after allogeneic bone marrow transplantation. J Peking University (health science). 2003; 35(1):103-107.
- Liu KY, Chen YH, Liu DH, Xu LP, Huang XJ. A pilot study of low-dose recombinant interleukin-2 for acute lymphoblastic malignancy after unmanipulated allogeneic blood and marrow transplantation. *Bone Marrow Transplant.* 2008; 42(8):535-539.
- Huang XJ, Wang Y, Liu DH, et al. Modified donor lymphocyte infusion (DLI) for the prophylaxis of leukemia relapse after hematopoietic stem cell transplantation in patients with advanced leukemia – feasibility and safety study. J Clin Immunol. 2008;28(4):390-397.
- Huang XJ, Liu DH, Liu KY, et al. Haploidentical hematopoietic stem cell transplantation without in vitro T cell depletion for the treatment of hematological malignancies. *Bone Marrow Transplant*. 2006;38(4):291-297.
- 18. Wang YZ, Liu YR, Zhu HH, et al. Prognostic sig-

nificance of minimal residual disease detected by multiparameter flow cytometry in acute myeloid leukemia. *Zhongguo Shi Yan Xue Ye Xue Za Zhi.* 2009;17(3):551-556.

- Cancer Therapy Evaluation Program. NCI Common Terminology Criteria for Adverse Events (CT-CAE) v4.0. http://www.ctep.cancer.gov. Published May 28, 2009.
- Thomas ED, Storb R, Clift RA, et al. Bone marrow transplantation. N Engl J Med. 1975;292(17): 895.
- Scrucca L, Santucci A, Aversa F. Regression modeling of competing risk using R: an in depth guide for clinicians. *Bone Marrow Transplant*. 2010;45(9):1388-1395.
- Efron B. Logistic regression, survival analysis and the Kaplan-Meier curve. J Am Stat Assoc. 1988; 83(214):414-425.
- Slovak ML, Kopecky KJ, Cassileth PA, et al. Karyotypic analysis predicts outcome of preremission and postremission therapy in adult acute myeloid leukemia: a Southwest Oncology Group/Eastern Cooperative Oncology Group study. *Blood.* 2000;96(13):4075-4083.
- Moorman AV, Harrison CJ, Buck GAN, et al. Karyotype is an independent prognostic factor in acute lymphoblastic leukaemia: analysis of cytogenetic data from patients treated on the MRC UKALL XII/ECOG 2993 trial. *Blood.* 2007;109(8): 3189-3197.
- Greenberg P, Cox C, LeBeau MM, Fenaux P, et al. International scoring systems for evaluating prognosis in myelodysplastic syndromes. *Blood*. 1997;89(6):2079-2088.
- Mohty M, Labopin M, Volin L, et al. Reducedintensity versus conventional myeloablative conditioning allogeneic stem cell transplantation for patients with acute lymphoblastic leukemia: a retrospective study from the European Group for Blood and Marrow Transplantation. *Blood*. 2010; 116(22):4439-4443.
- Kröger N, Brand R, van Biezen A, et al. Risk factors for therapy-related myelodysplastic syndrome and acute myeloid leukemia treated with

allogeneic stem cell transplantation. *Haematologica*. 2009;94(4):542-549.

- Klingebiel T, Cornish J, Labopin M, et al. Results and factors influencing outcome after fully haploidentical hematopoietic stem cell transplantation in children with very high-risk acute lymphoblastic leukemia: impact of center size: an analysis on behalf of the Acute Leukemia and Pediatric Disease Working Parties of the European Blood and Marrow Transplant group. *Blood*. 2010;115(17):3437-3446.
- Arellano ML, Langston A, Winton E, Flowers CR, Waller EK. Treatment of relapsed acute leukemia after allogeneic transplantation: a single center experience. *Biol Blood Marrow Transplant*. 2007; 13(1):116-123.
- Garderet L, Labopin M, Gorin NC, et al. Patients with acute lymphoblastic leukaemia allografted with a matched unrelated donor may have a lower survival with a peripheral blood stem cell graft compared to bone marrow. *Bone Marrow Transplant*. 2003;31(1):23-29.
- de Witte T, Hermans J, Vossen J, et al. Haematopoietic stem cell transplantation for patients with myelo-dysplastic syndromes and secondary acute myeloid leukaemias: a report on behalf of the Chronic Leukaemia Working Party of the European Group for Blood and Marrow Transplantation (EBMT). Br J Haematol. 2000;110(3):620-632.
- Jun HX, Jun CY, Yu ZX. In vivo induction of T-cell hyporesponsiveness and alteration of immunological cells of bone marrow grafts using granulocyte colony-stimulating factor. *Haematologica*. 2004;89:1517-1524.
- Huang XJ, Chang YJ, Zhao XY. Maintaining hyporesponsiveness and polarization potential of T cells after in vitro mixture of G-CSF mobilized peripheral blood grafts and G-CSF primed bone marrow grafts in different proportions. *Transpl Immunol.* 2007;17:193-197.
- Morris ES, Macdonald KP, Rowe V, et al. NKT cell-dependent leukemia eradication following stem cell mobilization with potent G-CSF analogs. J Clin Invest. 2005;115(11):3093-3103.