

microtubule-mediated movement of lytic granules to the immunological synapse. *Nat Immunol.* 2003;4(11):1111-1120.

7. Swank RT, Novak EK, McGarry MP, Rusiniak ME, Feng L. Mouse models of Hermansky Pudlak syndrome: a review. *Pigment Cell Res.* 1998;11(2):60-80.
8. Falcón-Pérez JM, Dell'Angelica EC. The pallidin (Pldn) gene and the role of SNARE proteins in melanosome biogenesis. *Pigment Cell Res.* 2002;15(2):82-86.
9. Ribeiro-Gomes FL, Moniz-de-Souza MC, Alexandre-Moreira MS, et al. Neutrophils activate macrophages for intracellular killing of *Leishmania major* through recruitment of TLR4 by neutrophil elastase. *J Immunol.* 2007;179(6):3988-3994.

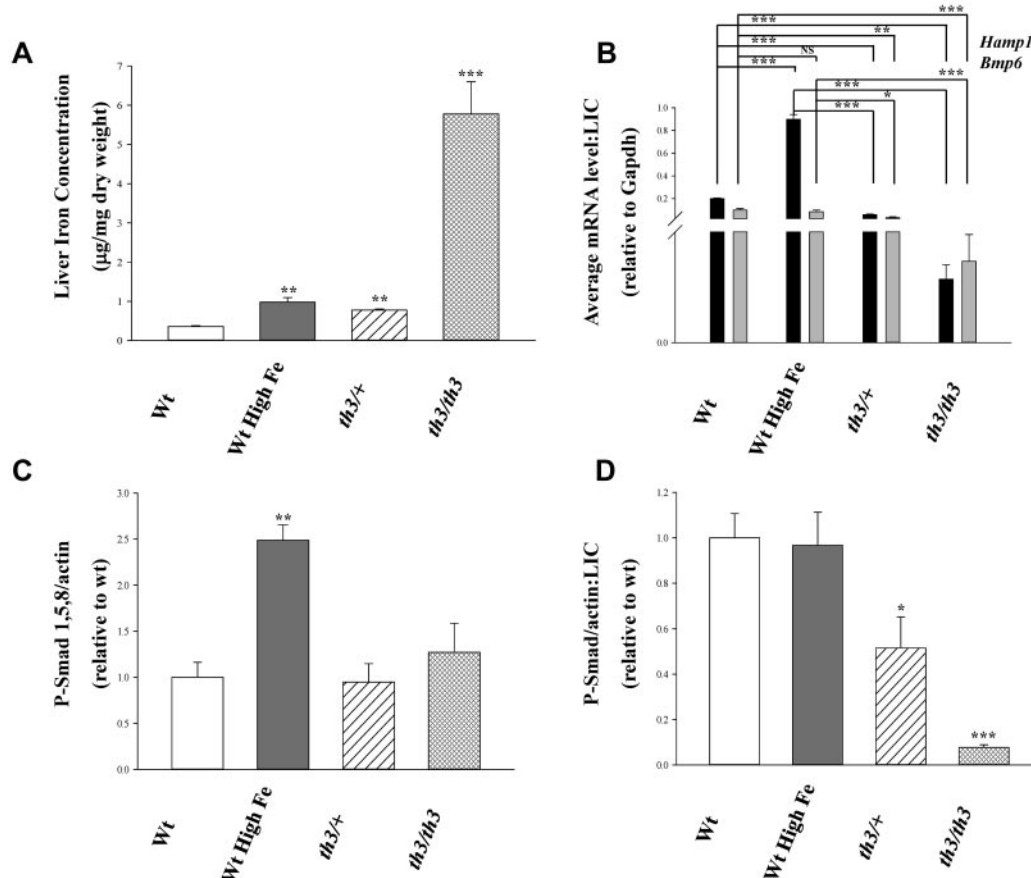
10. Huang L, Kuo YM, Gitschier J. The pallid gene encodes a novel, syntaxin 13-interacting protein involved in platelet storage pool deficiency. *Nat Genet.* 1999;23(3):329-332.
11. Bell CJ, Dinwiddie DL, Miller NA, et al. Carrier testing for severe childhood recessive diseases by next generation sequencing. *Sci Transl Med.* 2011;3(65):65ra4.
12. Moretta A, Bottino C, Vitale M, et al. Activating receptors and coreceptors involved in human natural killer cell-mediated cytotoxicity. *Annu Rev Immunol.* 2001;19:197-223.
13. Cullinane AR, Curry JA, Carmona-Rivera C, et al. A BLOC-1 mutation screen reveals that PLDN is mutated in Hermansky-Pudlak Syndrome type 9. *Am J Hum Genet.* 2011;88(6):778-787.

## To the editor:

### Decreased hepcidin expression in murine $\beta$ -thalassemia is associated with suppression of Bmp/Smad signaling

$\beta$ -thalassemia is a genetic disorder of hemoglobin production characterized by ineffective erythropoiesis and anemia.<sup>1</sup> Iron overload, a major source of morbidity, results from inappropriately low expression of the gene encoding hepcidin (*Hamp1*).<sup>1</sup> *Hamp1* controls plasma iron concentration by facilitating the degradation of the

iron efflux protein ferroportin.<sup>2</sup> In healthy individuals, hepcidin is transcriptionally responsive to iron via bone morphogenetic protein (Bmp)/Smad pathway-mediated phosphorylation of Smad 1,5,8.<sup>3</sup> Phosphorylated Smad 1,5,8 is an essential component of the transcriptional complex that induces *Hamp1* expression in response to iron.<sup>4</sup> We investigated the



**Figure 1. Bmp/Smad signaling is hyporesponsive to iron in  $\beta$ -thalassemic mice.** (A) Liver iron concentrations ( $\mu\text{g}/\text{mg}$  dry weight) from wt mice fed a standard diet (200 ppm iron), wt mice fed a high iron diet (standard chow supplemented with 2.5% carbonyl iron), *th3/+*, and *th3/th3* mice were determined by atomic absorption and are expressed as mean  $\pm$  SEM. (B) Average fold difference in the transcription levels of the Bmp/Smad pathway member *Bmp6* and target gene *Hamp1* were normalized to LIC. Data were normalized to mouse *Gapdh* and are presented as the mean  $\pm$  SEM ( $n \geq 3$ ). All averaged values are the product of duplicate determinations. (C) Chemiluminescence signals from Western blots of liver lysates reacted with antibodies to phosphorylated Smad 1,5,8 and to  $\beta$ -actin were quantified and data expressed as the mean  $\pm$  SEM of their ratios. (D) Phosphorylated Smad 1,5,8 levels normalized to  $\beta$ -actin are expressed relative to LIC. Data are presented as mean  $\pm$  SEM. Statistical significance was determined using the Student *t* test. \* $P \leq .05$ , \*\* $P \leq .01$ , \*\*\* $P \leq .001$ . Significance with respect to wt or iron-loaded wt values are indicated by the brackets.

relationship between iron status, Bmp/Smad signaling, and *Hamp1* expression in mouse models of  $\beta$ -thalassemia intermedia (*th3/+*) and major (*th3/th3*).<sup>5</sup>

Wild-type (wt) C57BL/6 mice were placed on a high iron diet to match the elevated liver iron concentrations (LICs) observed in *th3/+* mice (Figure 1A). The severity of iron overload in *th3/th3* was even higher, by 5-fold, relative to iron-loaded wt mice (Figure 1A). Dietary iron loading of wt mice resulted in the expected increase in *Hamp1* and *Bmp6* expression (measured by quantitative RT-PCR). However, despite the similarly elevated LICs in *th3/+* mice, neither *Bmp6* nor *Hamp1* expression was significantly and proportionally increased. Moreover, the highly iron loaded *th3/th3* mice demonstrated a decrease in expression of both *Bmp6* and *Hamp1*. Thus, relative to hepatic iron load, expression of these genes is significantly decreased in both  $\beta$ -thalassemia models (Figure 1B). Furthermore, hepatic phosphorylated Smad 1,5,8 protein concentration is significantly increased only in liver samples from wt mice fed a high-iron diet relative to controls (Figure 1C). When normalized to LIC, our data reveal significantly decreased phosphorylated Smad 1,5,8 in *th3/+* and *th3/th3* samples compared with wt and wt fed a high-iron diet (Figure 1D). These results present strong evidence of a blunted relationship between LIC and *Bmp6* mRNA expression in  $\beta$ -thalassemic mice.

No prior studies have examined *Bmp6* expression or Smad signaling in models of  $\beta$ -thalassemia. Suppressed *Hamp1* transcription relative to the degree of iron overload in  $\beta$ -thalassemic mice has been postulated to occur as a consequence of expanded or ineffective erythropoiesis and effects of certain erythroid factors (eg, GDF15 or TWSG1) on hepatocellular hepcidin expression.<sup>6-8</sup> Our results suggest that the effects of ineffective erythropoiesis on *Hamp1* expression include a suppression of *Bmp6* mRNA expression relative to LIC, with consequent decreased Smad signaling. In contrast, studies in mice examining the acute effects of erythropoietin administration on *Bmp6* and *Hamp1* demonstrated decreased *Hamp1* expression but no effect on *Bmp6* expression.<sup>9</sup> This observation supports the proposal that different erythroid signals regulate hepcidin in normal and ineffective erythropoiesis.<sup>8</sup> Whether or not as a consequence of an erythroid factor, our findings demonstrate that in  $\beta$ -thalassemic mice the normal relationship between iron status and liver *Bmp6* mRNA expression is blunted. This dysregulation results in low *Hamp1* levels relative to iron stores and excess iron absorption relative to tissue demands. Continued exploration of the mechanisms underlying blunted Bmp/Smad signaling in  $\beta$ -thalassemia will enhance understanding of the iron overload that is central to the pathophysiology of this disease.

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## References

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1. Ginzburg Y, Rivella S. Beta-thalassemia: a model for elucidating the dynamic regulation of ineffective erythropoiesis and iron metabolism. *Blood*. 2011; 118(16):4321-4330.
2. Nemeth E, Tuttle MS, Powelson J, et al. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science*. 2004; 306(5704):2090-2093.
3. Babitt JL, Huang FW, Wrighting DM, et al. Bone morphogenetic protein signaling by hemojuvelin regulates hepcidin expression. *Nat Genet*. 2006;38(5): 531-539.
4. Kautz L, Meynard D, Monnier A, et al. Iron regulates phosphorylation of Smad1/5/8 and gene expression of Bmp6, Smad7, Id1, and Atoh8 in the mouse liver. *Blood*. 2008;112(4):1503-1509.
5. Rivella S, May C, Chadburn A, et al. A novel murine model of Cooley anemia and its rescue by lentiviral-mediated human beta-globin gene transfer. *Blood*. 2003;101(8):2932-2939.
6. Pak M, Lopez MA, Gabayan V, et al. Suppression of hepcidin during anemia requires erythropoietic activity. *Blood*. 2006;108(12):3730-3735.
7. Gardenghi S, Marongiu MF, Ramos P, et al. Ineffective erythropoiesis in beta-thalassemia is characterized by increased iron absorption mediated by down-regulation of hepcidin and up-regulation of ferroportin. *Blood*. 2007;109(11): 5027-5035.
8. Tanno T, Noel P, and Miller JL. Growth differentiation factor 15 in erythroid health and disease. *Curr Opin Hematol*. 2010;17(3):184-190.
9. Krijt J, Jonášová A, Neuwirtová R, Necas E. Effect of erythropoietin on hepcidin expression in hemojuvelin-mutant mice. *Blood Cells Mol Dis*. 2010;44(4): 257-261.