

# Prognostic significance of additional cytogenetic aberrations in 733 de novo pediatric 11q23/*MLL*-rearranged AML patients: results of an international study

Eva A. Coenen,<sup>1</sup> \*Susana C. Raimondi,<sup>2,3</sup> \*Jochen Harbott,<sup>4</sup> Martin Zimmermann,<sup>5</sup> Todd A. Alonzo,<sup>3</sup> Anne Auvrignon,<sup>6</sup> H. Berna Beverloo,<sup>7,8</sup> Myron Chang,<sup>9</sup> Ursula Creutzig,<sup>10</sup> Michael N. Dworzak,<sup>11</sup> Erik Forestier,<sup>12</sup> Brenda Gibson,<sup>13</sup> Henrik Hasle,<sup>14</sup> Christine J. Harrison,<sup>15</sup> Nyla A. Heerema,<sup>3,16</sup> Gertjan J. L. Kaspers,<sup>17-19</sup> Anna Leszl,<sup>20</sup> Nathalia Litvinko,<sup>21</sup> Luca Lo Nigro,<sup>22</sup> Akira Morimoto,<sup>23,24</sup> Christine Perot,<sup>6</sup> Dirk Reinhardt,<sup>5</sup> Jeffrey E. Rubnitz,<sup>2</sup> Franklin O. Smith,<sup>3,25</sup> Jan Stary,<sup>26</sup> Irina Stasevich,<sup>21</sup> Sabine Strehl,<sup>11</sup> Takashi Taga,<sup>23,27</sup> Daisuke Tomizawa,<sup>23,28</sup> David Webb,<sup>18,29</sup> Zuzana Zemanova,<sup>30</sup> Rob Pieters,<sup>1</sup> †C. Michel Zwaan,<sup>1,17</sup> and †Marry M. van den Heuvel-Eibrink<sup>1,17</sup>

<sup>1</sup>Department of Pediatric Oncology/Hematology, Erasmus MC–Sophia Children's Hospital, Rotterdam, The Netherlands; <sup>2</sup>St Jude Children's Research Hospital, Memphis, TN; <sup>3</sup>Children's Oncology Group, Arcadia, CA; <sup>4</sup>Acute Myeloid Leukemia-Berlin-Frankfurt-Münster Study Group, Department of Pediatric Hematology and Oncology, Justus-Liebig-University, Giessen, Germany; <sup>5</sup>Acute Myeloid Leukemia-Berlin-Frankfurt-Münster Study Group, Pediatric Hematology/Oncology, Medical School Hannover, Hannover, Germany; <sup>6</sup>French Leucémie Aigue Myeloïde Enfant, Hôpital Trousseau, Paris, France; <sup>7</sup>Dutch Childhood Oncology Group, Dutch Working Group on Hemato-Oncologic Genome Diagnostics, The Hague, The Netherlands; <sup>8</sup>Department of Clinical Genetics, Erasmus MC, Rotterdam, The Netherlands; <sup>9</sup>Children's Oncology Group, Data Center, Gainesville, FL; <sup>10</sup>Acute Myeloid Leukemia-Berlin-Frankfurt-Münster Study Group, Pediatric Hematology/Oncology, University Hospital, Münster, Germany; <sup>11</sup>Children's Cancer Research Institute, Vienna, Austria; <sup>12</sup>Nordic Society for Pediatric Hematology and Oncology, Department of Clinical Science, Pediatrics, Umeå University, Umeå, Sweden; <sup>13</sup>Department of Pediatric Oncology/Hematology, Royal Hospital for Sick Children, Glasgow, United Kingdom; <sup>14</sup>Nordic Society for Pediatric Hematology and Oncology, Department of Pediatrics, Aarhus University Hospital Skejby, Aarhus, Denmark; <sup>15</sup>Northern Institute for Cancer Research, Newcastle University, Newcastle upon Tyne, United Kingdom; <sup>16</sup>Department of Pathology, The Ohio State University, Columbus, OH; <sup>17</sup>Dutch Childhood Oncology Group, The Hague, The Netherlands; <sup>18</sup>Acute Myeloid Leukemia Committee International-Berlin-Frankfurt-Münster Study Group; <sup>19</sup>Department of Pediatric Oncology/Hematology, VU University Medical Center, Amsterdam, The Netherlands; <sup>20</sup>Italian Association of Pediatric Hematology Oncology, Clinica Pediatrica, Università Padova, Padova, Italy; <sup>21</sup>Research Center for Pediatric Oncology and Hematology, Minsk, Belarus; <sup>22</sup>Italian Association of Pediatric Hematology Oncology, Clinica Pediatrica, Università Catania, Catania, Italy; <sup>23</sup>Japanese Pediatric Leukemia/Lymphoma Study Group, Nagoya, Japan; <sup>24</sup>Department of Pediatrics, Jichi Medical University School of Medicine, Tochigi, Japan; <sup>25</sup>Hematology/Oncology and Pediatrics, Cincinnati Children's Hospital Medical Center and the University of Cincinnati College of Medicine, Cincinnati, OH; <sup>26</sup>Czech Pediatric Hematology/Oncology, University Hospital Motol and 2nd Medical School, Charles University, Prague, Czech Republic; <sup>27</sup>Department of Pediatrics, Shiga University of Medical Science, Shiga, Japan; <sup>28</sup>Department of Pediatrics and Developmental Biology, Tokyo Medical and Dental University, Tokyo, Japan; <sup>29</sup>Great Ormond Street Hospital for Children, London, United Kingdom; and <sup>30</sup>Center of Oncocytogenetics, General University Hospital and First Faculty of Medicine, Charles University, Prague, Czech Republic

We previously demonstrated that outcome of pediatric 11q23/*MLL*-rearranged AML depends on the translocation partner (TP). In this multicenter international study on 733 children with 11q23/*MLL*-rearranged AML, we further analyzed which additional cytogenetic aberrations (ACA) had prognostic significance. ACAs occurred in 344 (47%) of 733 and were associated with unfavorable outcome (5-year overall survival [OS] 47% vs 62%,  $P < .001$ ). Trisomy 8, the most frequent specific ACA ( $n = 130/344$ , 38%), indepen-

dently predicted favorable outcome within the ACAs group (OS 61% vs 39%,  $P = .003$ ; Cox model for OS hazard ratio (HR) 0.54,  $P = .03$ ), on the basis of reduced relapse rate (26% vs 49%,  $P < .001$ ). Trisomy 19 ( $n = 37/344$ , 11%) independently predicted poor prognosis in ACAs cases, which was partly caused by refractory disease (remission rate 74% vs 89%,  $P = .04$ ; OS 24% vs 50%,  $P < .001$ ; HR 1.77,  $P = .01$ ). Structural ACAs had independent adverse prognostic value for event-free survival (HR 1.36,  $P = .01$ ).

Complex karyotype, defined as  $\geq 3$  abnormalities, was present in 26% ( $n = 192/733$ ) and showed worse outcome than those without complex karyotype (OS 45% vs 59%,  $P = .003$ ) in univariate analysis only. In conclusion, like TP, specific ACAs have independent prognostic significance in pediatric 11q23/*MLL*-rearranged AML, and the mechanism underlying these prognostic differences should be studied. (*Blood*. 2011;117(26):7102-7111)

## Introduction

Pediatric acute myeloid leukemia (AML) is a clinically and genetically heterogeneous disease. In addition to the patient's initial response to treatment, its prognosis is largely determined by the presence of cytogenetic abnormalities and genetic lesions.<sup>1-6</sup> Several recurrent cytogenetic abnormalities, such as 11q23/*MLL*-rearrangements, predict outcome in myeloid neoplasms and acute

leukemia.<sup>7</sup> So far,  $> 60$  different translocation partners (TPs) have been identified, and new partners are still being reported to add to the diversity of *MLL*-rearranged leukemia.<sup>8,9</sup> The authors of a recent international study<sup>10</sup> highlighted the heterogeneity of 11q23/*MLL*-rearranged pediatric AML by demonstrating that outcome is dependent on TPs. This study also revealed that additional

Submitted December 30, 2010; accepted April 13, 2011. Prepublished online as *Blood* First Edition paper, May 6, 2011; DOI 10.1182/blood-2010-12-328302.

\*S.C.R. and J.H. contributed equally to this article.

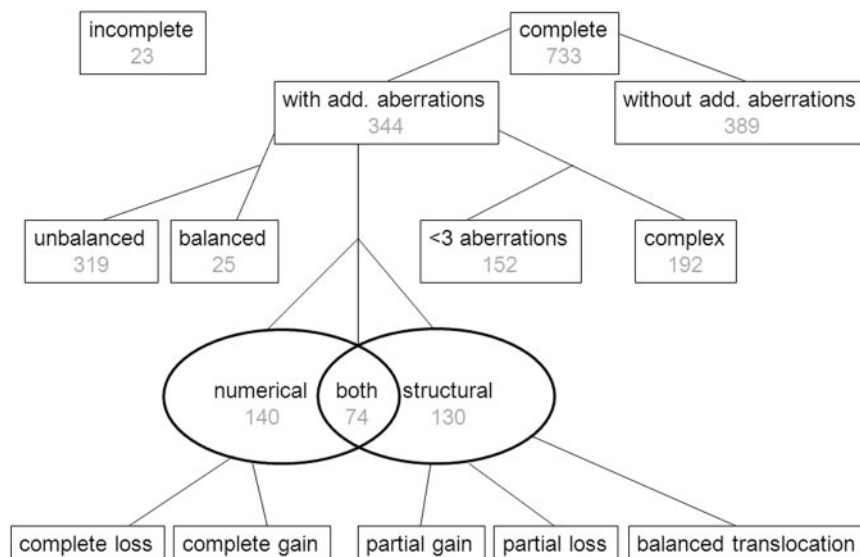
†C.M.Z. and M.M.v.d.H.-E. contributed equally to this article.

The online version of this article contains a data supplement.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2011 by The American Society of Hematology

**Figure 1. Flow chart showing the presence and type of ACAs in 756 pediatric patients with 11q23/*MLL*-rearranged AML.** Complete karyotypes were not available for 23 patients, and they were therefore excluded from analyses. The presence or absence of ACAs was determined for 733 patients for whom complete karyotypes were available. In the cohort having ACAs balanced karyotype was coded for 25 patients; the remaining had an unbalanced karyotype. The types of aberrations were coded as numerical, structural, or both, and the number of aberrations was also coded. Losses and gains are further coded in other figures.



cytogenetic aberrations (ACAs) were an independent adverse prognostic factor,<sup>10</sup> but so far, it is unknown which additional aberration(s) determine this unfavorable outcome signature.

The authors of a recent large study in an adult AML cohort<sup>11</sup> showed that additional cytogenetic abnormalities in t(9;11)(p22;q23) AML did not affect outcome. However, the Berlin-Frankfurt-Münster group showed that children with t(9;11)(p22;q23) with additional aberrations had lower rates of overall survival (OS) than those with other subgroups of AML.<sup>6</sup>

To date, no large studies have been undertaken to study the prognostic relevance of specific ACAs in pediatric *MLL*-rearranged AML. In this multicenter international study, we retrospectively analyzed data from a large cohort (n = 733) to determine which ACAs contribute to the prognostic effect in pediatric *MLL*-rearranged AML.

## Patients and methods

### Patients

Patients' data collected in the retrospective international study by Balgobind et al<sup>10</sup> were included in this study. In summary, data from 756 patients with 11q23/*MLL*-rearranged pediatric AML were collected from 11 collaborative study groups—the Berlin-Frankfurt-Münster Study Group (Germany and Austria); the Japanese Pediatric Leukemia/Lymphoma Study Group (Japan); the Leucémies Aiguës Myéloblastiques de l'Enfant Cooperative Group (France); the Czech Pediatric Hematology Working Group (Czech Republic); the St Jude Children's Research Hospital (United States); the Associazione Italiana Ematologia Oncologia Pediatrica (Italy); Research Center for Pediatric Oncology and Hematology (Belarus); the Children's Oncology Group (United States); the Nordic Society for Pediatric Hematology and Oncology (Denmark, Finland, Iceland, Norway, and Sweden); the Dutch Children's Oncology Group (The Netherlands); and 2 centers of the Medical Research Council (United Kingdom). Patients were treated by national/collaborative group AML trials.<sup>12-22</sup> The treatment protocols were approved according to local law and guidelines and by the institutional review boards of each participating center, with informed consent obtained from the patients' parents or legal guardians in accordance with the Declaration of Helsinki.

Inclusion criteria for the current analyses were diagnosis between January 1, 1993, and January 1, 2005; younger than 18 years of age at diagnosis; and involvement of 11q23 or *MLL* as determined by G-, Q-, or

R-banded karyotyping; FISH; or RT-PCR. Exclusion criteria were secondary AML after congenital BM failure disorders, aplastic anemia, previous chemotherapy or radiotherapy for other diseases, and previous myelodysplastic syndrome (MDS). Patients with Down syndrome were included if they met the other inclusion criteria. All clinical data obtained at initial diagnosis, data on treatment (therapy protocol, including HSCT), and all events during follow-up were checked for consistency and completeness.<sup>10</sup>

### Cytogenetic analysis

All karyotypes were centrally reviewed by 2 cytogeneticists (J.H., S.C.R.) and assigned to 11q23/*MLL*-rearranged groups on the basis of TP.<sup>10</sup> All karyotypes were designated according to the International System for Human Cytogenetic Nomenclature 2005.<sup>23</sup>

To analyze ACAs, data from all patients with incomplete karyotypes were excluded. For all cases included in the analysis, the number of aberrations was counted. Each aberration separated from the rest of the karyotype by a comma was counted as one abnormality (regardless of its complexity), every aberration was counted only once (if present in multiple clones), and constitutional aberrations were excluded. Triploidy and tetraploidy were counted as 1 aberration (1 event). In this cohort of 11q23/*MLL*-rearranged cases, ACAs cases were defined as having 2 or more aberrations, including the 11q23/*MLL*-rearrangement (n = 344). All cases with 3 or more aberrations were considered having a complex karyotype, consistent with previously used definitions.<sup>24,25</sup> Numerical aberrations were defined as loss or gain of a full chromosome. Balanced translocations were defined as translocations in which no material seemed to be gained or lost as determined by conventional karyotyping. Structural aberrations were defined as aberrations resulting from breakpoints within a chromosome. In all unbalanced translocations we described which material was lost and gained and also whether 11q23 was involved. The presence of a balanced overall karyotype was defined as a karyotype with 2 complete copies of all autosomes and complete copies of sex chromosomes without any additional material (2n). Definitions used for cytogenetic classification are summarized in supplemental Table 1 (available on the *Blood* Web site; see the Supplemental Materials link at the top of the online article).

### Statistical analyses

Complete remission (CR) was defined as < 5% blasts in the BM, with regeneration of trilineage hematopoiesis plus absence of extramedullary disease.<sup>26</sup> Early death was defined as any death within the first 6 weeks of treatment. Treatment of patients who did not obtain CR within the specified time in the protocol was considered a failure on day 0. OS was measured from the date of diagnosis to the date of last follow-up or death from any

**Table 1. Distribution of ACAs by translocation partner and clinically relevant parameters**

	n	ACAs, n (%)	ACAs type		
			Numerical, n (%)	Structural, n (%)	Both, n (%)
TP group					
9p22	316	148 (47)	84 (57)*	40 (27)*	24 (16)*
10p12	96	48 (50)	13 (27)*	26 (54)*	9 (19)*
6q27	35	17 (49)	8 (47)*	7 (41)*	2 (12)*
19p13	30	10 (33)	6 (60)*	1 (10)*	3 (30)*
19p13.1	34	13 (38)	5 (38)*	4 (31)*	4 (31)*
19p13.3	25	13 (52)	5 (38)*	4 (31)*	4 (31)*
1q21	24	6 (25)	2 (33)*	3 (50)*	1 (17)*
4q21	13	8 (62)	2 (25)*	4 (50)*	2 (25)*
10p11.2	12	7 (58)	0*	5 (71)*	2 (29)*
17q21	12	3 (25)	1 (33)*	1 (33)*	1 (33)*
other	136	71 (52)	14 (20)*	35 (49)*	22 (31)*
	733				
Sex					
Male	358	171 (48)	78 (46)	63 (37)	30 (18)
Female	375	173 (46)	62 (36)	67 (39)	44 (25)
	733				
Age, y					
< 2	344	143 (42)	45 (31)†	68 (48)†	30 (21)†
2-9	219	115 (53)	57 (50)†	35 (30)†	23 (20)†
≥ 10	170	86 (51)	38 (44)†	27 (31)†	21 (24)†
	733				
WBC, ×10 <sup>9</sup> /L					
< 20	339	175 (52)	84 (49)†	51 (29)†	40 (23)†
20-99	203	87 (43)	28 (32)†	40 (46)†	19 (22)†
≥ 100	171	73 (43)	25 (34)†	35 (48)†	13 (18)†
	713				
FAB					
M0	23	12 (52)	6 (50)	3 (25)	3 (25)
M1	39	20 (51)	9 (45)	7 (35)	4 (20)
M2	32	12 (38)	7 (58)	4 (33)	1 (8)
M4	134	49 (37)	21 (43)	21 (43)	7 (14)
M5	446	217 (49)	88 (41)	83 (38)	46 (21)
M7	19	15 (79)	7 (47)	2 (13)	6 (4)
n.d.	7	5 (71)	0	2 (40)	3 (60)
	700				
Overall		344 (47)	140 (41)	130 (38)	74 (22)

ACAs (%) indicates number of cases with additional aberrations and percentage within this group; Numerical (%), number of cases with only numerical additional aberrations and percentage of specific group (row); Structural (%), number of cases with only structural additional aberrations and percentage of specific group (row); Both (%), number of cases with both numerical and structural additional aberrations and percentage of specific group (row); and TP group, site of translocation on partner chromosome

ACA indicates additional cytogenetic aberrations; dx, diagnosis; FAB, French American British morphology classification subtype; n.d., not determined; TP, translocation partner; and WBC, white blood cell count.

\*Values significantly different at the  $P < .01$  level ( $\chi^2$ ).

†Values significantly different at the  $P < .05$  level ( $\chi^2$ ).

cause. Event-free survival (EFS) was calculated from the date of diagnosis to the first event or to the date of last follow-up. Events included nonremittance, relapse, secondary malignancy, or death from any cause. Cumulative incidence of relapse (CIR) was calculated from the date of CR to the first relapse. Refractory disease was included in the EFS and CIR analyses by arbitrarily setting the event date on day 0. For OS, EFS, and CIR analyses, patients who did not experience an event were censored at the time of last follow-up.

The Kaplan-Meier method was used to estimate the 5-year probabilities of OS and EFS, and survival estimates were compared by the log-rank test. The Gray test for competing risks was used for CIR analysis. Multivariate analyses were performed with the Cox proportional hazards model. Continuous variables known to be of prognostic value in AML were categorized according to cutoff points (eg,  $> 2$  or 10 years of age, white blood cell [WBC] count  $< 20 \times 10^9/L$  or  $> 100 \times 10^9/L$ ). The  $\chi^2$  or Fisher exact test was used to compare differences in proportions of variables among groups; the Mann-Whitney  $U$  test was used for continuous variables. All  $P$  values are descriptive and explorative and were considered

significant if  $\leq .05$ . All statistical data were analyzed by the use of SAS-PC, Version 9.1 (SAS Institute Inc).

## Results

### Distribution of ACAs

Of the 756 patients, 733 (97%) had complete karyotypes, and their data were included in the study (see flowchart in Figure 1). There were no significant differences in the patients included ( $n = 733$ ) and not included ( $n = 23$ ) in this study with respect to sex, age, WBC count, and TP group (data not shown). ACAs were found in 344 (47%) of 733 cases (Figure 1). The number of additional aberrations ranged from 0 to 15 (mean, 1.2 additional aberrations; supplemental Figure 1).

There were 3 or more aberrations (including the 11q23/*MLL*-rearrangement) in 192 of 733 (26%) cases, which were therefore

**Table 2. Number of aberrations by 11q23 translocation partner and clinically relevant parameters**

	Number of aberrations							All
	0	1	2	3	4	5	> 5	
TP group								
9p22		168 (44)*	75 (49)*	33 (41)*	19 (43)*	7 (28)*	14 (33)*	316 (43)
10p12	1 (14)*	47 (12)*	19 (13)*	12 (15)*	7 (16)*	5 (20)*	5 (12)*	96 (13)
6q27	1 (14)*	17 (4)*	7 (5)*	1 (1)*			9 (21)*	35 (5)
19p13		20 (5)*	3 (2)*	4 (5)*	1 (2)*	1 (4)*	1 (2)*	30 (4)
19p13.1	1 (14)*	20 (5)*	7 (5)*	2 (3)*	1 (2)*	3 (12)*		34 (5)
19p13.3		12 (3)*	7 (5)*		3 (7)*	2 (8)*	1 (2)*	25 (3)
1q21		18 (5)*	3 (2)*	1 (1)*	1 (2)*		1 (2)*	24 (3)
4q21		5 (1)*	2 (1)*	4 (5)*	1 (2)*	1 (4)*		13 (2)
10p11.2		5 (1)*	2 (1)*	3 (4)*		2 (8)*		12 (2)
17q21		9 (2)*	1 (1)*	1 (1)*			1 (2)*	12 (2)
Other	4 (57)*	61 (16)*	26 (17)*	19 (24)*	11 (25)*	4 (16)*	11 (26)*	136 (19)
								733
Sex								
Male		187 (49)†	89 (59)†	35 (44)†	13 (30)†	11 (44)†	23 (53)†	358 (49)
Female	7 (100)†	195 (51)†	63 (41)†	45 (56)†	31 (70)†	14 (56)†	20 (47)†	375 (51)
								733
Age, y								
< 2	4 (57)*	197 (52)*	61 (40)*	39 (49)*	12 (27)*	16 (64)*	15 (35)*	344 (47)
2-9		104 (27)*	49 (32)*	29 (36)*	22 (50)*	5 (20)*	10 (23)*	219 (30)
≥ 10	3 (43)*	81 (21)*	42 (28)*	12 (15)*	10 (23)*	4 (16)*	18 (42)*	170 (23)
								733
WBC, ×10 <sup>9</sup> /L								
< 20	5 (71)	159 (42)	76 (50)	39 (49)	23 (52)	16 (64)	21 (49)	339 (46)
20-99	1 (14)	115 (30)	38 (25)	20 (25)	14 (32)	5 (20)	10 (23)	203 (28)
≥ 100	1 (14)	97 (25)	34 (22)	19 (24)	7 (16)	2 (8)	11 (26)	171 (23)
								713 (97)
FAB								
M0		11 (3)*	4 (3)*	2 (3)*	3 (7)*	1 (4)*	2 (5)*	23 (3)
M1		19 (5)*	12 (8)*	4 (5)*	1 (2)*		3 (7)*	39 (5)
M2	1 (14)*	19 (5)*	7 (5)*	2 (3)*	3 (7)*			32 (4)
M4	2 (29)*	83 (22)*	25 (16)*	12 (15)*	5 (11)*	2 (6)*	5 (12)*	134 (18)
M5	4 (57)*	225 (59)*	97 (64)*	54 (68)*	24 (55)*	17 (68)*	25 (58)*	446 (61)
M7		4 (1)*	3 (2)*	2 (3)*	4 (9)*		6 (14)*	19 (3)
n.d.		2 (1)*	1 (1)*	1 (1)*	1 (2)*	1 (4)*	1 (2)*	7 (1)
								700 (95)
Overall	7 (1)	382 (52)	152 (21)	80 (11)	44 (6)	25 (3)	43 (6)	733

The number of aberrations indicates total number of aberrations in the karyotype, including 11q23/MLL-rearrangement, percentages per group shown in parentheses (per column).

dx indicates diagnosis; FAB, French American British morphology classification subtype; n.d., not determined; TP, translocation partner; and WBC, white blood cell count.

\*Significantly different at the  $P < .01$  level ( $\chi^2$ ).

†Significantly different at the  $P < .05$  level ( $\chi^2$ ).

defined as complex karyotypes. Of the 344 cases with ACAs, 140 (41%) had numerical ACAs only, 130 (38%) had structural ACAs only, and 74 (22%) had both numerical and structural ACAs (Figure 1). There were 25 (7%) cases of ACA that had only balanced structural abnormalities in their karyotypes (Figure 1).

#### Distribution of ACAs in clinically relevant groups

Tables 1 and 2 show the distribution of ACAs by TP group and clinically relevant parameters (sex, age, WBC count, and FAB [ie, French-American-British] subtype). TP groups 9p22 and 19p13 were characterized by a relatively high frequency of numerical ACAs, whereas groups 10p12, 10p11.2, and 4q21 showed greater prevalence of structural ACAs ( $P < .001$ ; Table 1). Also, there were significant differences in the number of aberrations among TP groups: the 6q27 group had a relatively high number of ACAs ( $P = .002$ ), whereas groups 9p22, 19p13, and 1q21 had a lower number of ACAs (Table 2).

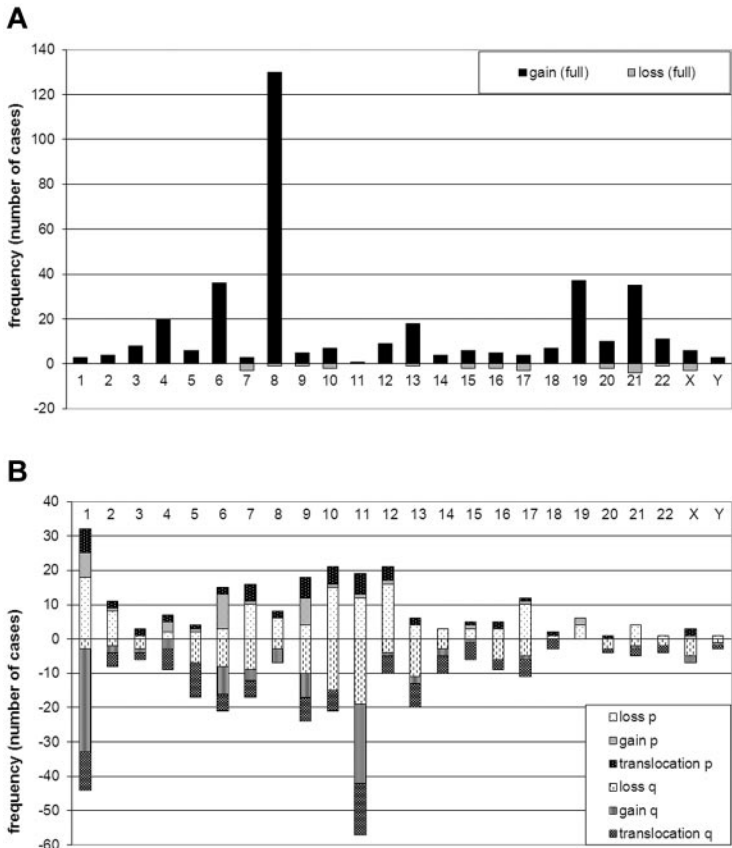
ACAs were less likely to occur in young children (< 2 years of age) than in children 2-9 years of age or 10 years or older (42% vs 53% vs 51%,  $P = .02$ ; Table 1). However, structural ACAs were more frequent

in children < 2 years of age than in children 2-9 years of age or 10-18 years of age (48% vs 30% vs 31%,  $P < .01$ ; Table 1). There was a greater prevalence of highly complex karyotypes (> 5 aberrations) in children 10-18 years of age than those younger than 2 years or 2-9 years of age (11% vs 4% vs 5%,  $P = .02$ , Table 2).

Although the number of patients with FAB M7 was small, ACAs were more likely to occur in patients with AML FAB M7 compared with those with other FAB types (79% vs 46%,  $P = .008$ ), whereas patients with AML FAB M2 and M4 had the lowest occurrence of ACAs (Table 1). Also, patients with AML FAB M7 seem to have a higher number of aberrations than those with other FAB morphologies ( $P = .003$ ; Table 2).

#### Specific recurrent aberrations

Trisomy 8 was the most frequently occurring numerical abnormality (130/733, 18% of all cases and 38% of ACA cases, Figure 2A). In addition, trisomy 4, 6, 13, 19, and 21 were recurrent ACAs (at least 15 cases each). Two cases with Down syndrome were included in this study. However, because constitutional aberrations



**Figure 2. Frequency (number of cases) of numerical and structural ACAs.** (A) Numerical ACAs. Gains are shown on the positive y-axis, and losses are shown on the negative y-axis. Chromosomes are on the x-axis. (B) Structural ACAs. The short arms (p) of the chromosomes are shown on the positive y-axis and the long arms (q) on the negative y-axis. Lightest shades are used for losses, medium-shaded colors are used for gains, and the darkest-shaded colors for breakpoints of balanced translocations. Chromosomes are on the x-axis. Balanced 11q23 translocations are not included in the figure.

were not included in the additional aberrations, they were not included in the trisomy 21 group. Only 11 patients had losses of full chromosomes, collectively accounting for 25 monosomies (Figure 2A).

Figure 2B shows the collective analysis of structural ACAs per chromosome arm but does not include breakpoints involved in balanced 11q23/*MLL*-translocations. However, the figure includes unbalanced 11q23/*MLL*-translocations in which chromosomal

**Table 3. Univariate survival analysis of the complete cohort (n = 733)**

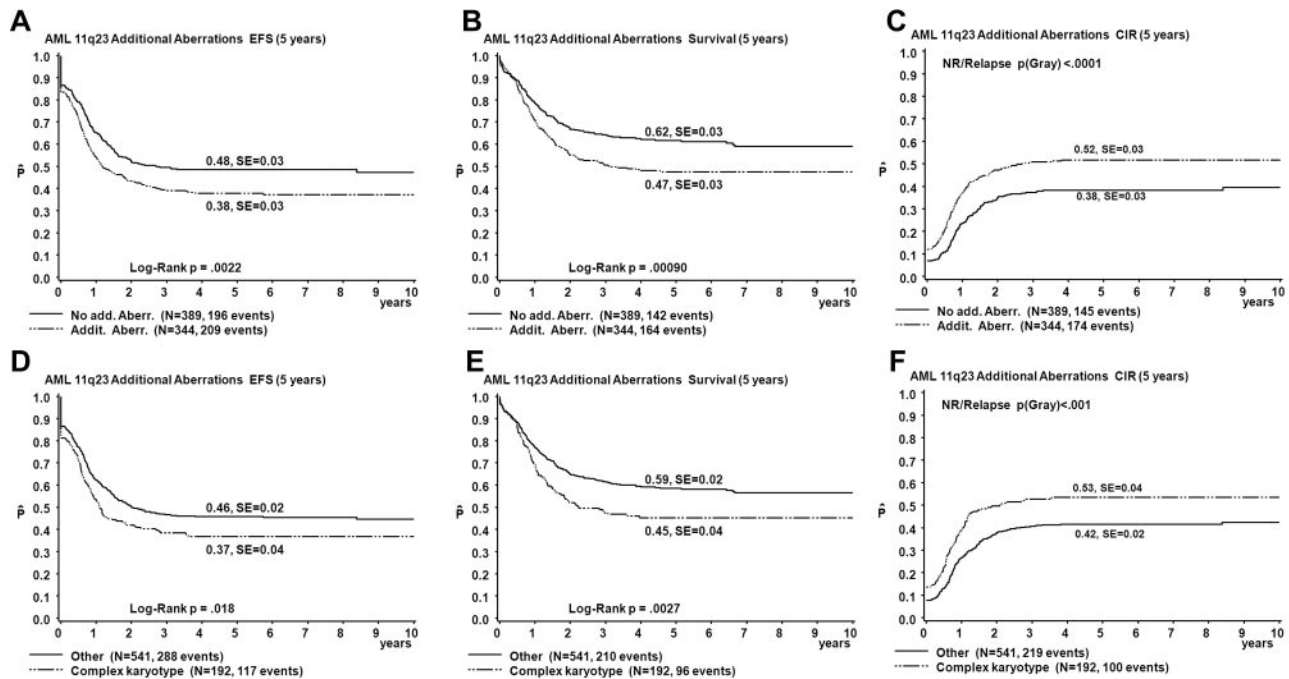
	n	EFS	Complete cohort			
			P (log-rank)	OS	P (log-rank)	CIR
<b>Additional aberrations</b>			.002†		< .001†	< .001†
Absent	389	0.48		0.62		0.38
Present	344	0.38		0.47		0.52
<b>No. of aberrations</b>			< .001†		< .001†	.001†
2	152	0.39		0.50		0.50
3	80	0.45		0.53		0.48
≥ 3	192	0.37	.018*	0.45	.003†	0.53
4	44	0.40		0.50		0.53
5	25	0.36		0.43		0.60
> 5	43	0.18	< .001†	0.25		0.61
<b>Type</b>			.001†		.003†	< .001†
Numerical	140	0.47		0.56		0.41
Structural	130	0.32		0.43		0.59
Both	74	0.31		0.40		0.59
<b>Trisomy</b>						
4	20	0.43	.72	0.52	.87	0.52
6	36	0.35	.43	0.35	.029*	0.54
8	130	0.53	< .001†	0.61	.003†	0.35
13	18	0.49	.52	0.64	.41	0.40
19	37	0.17	.003†	0.24	< .001†	0.54
21	35	0.19	.007†	0.28	.015*	0.69

CIR, indicates 5-year cumulative incidence of relapse; EFS, 5-year event-free survival estimates; n, number of patients; OS, 5-year overall survival estimate; P (Gray), P value from the Gray test; and P (log-rank), P value from log-rank test.

\*Significant at P < .05 level.

†Significant at P < .01 level.





**Figure 3.** Survival curves obtained from univariate analysis comparing patients with ACAs to patients without ACAs and comparing patients with complex karyotype with all patients with < 3 aberrations. (A-C) Patients with ACAs are compared to patients without ACAs. (D-F) Patients with complex karyotype are compared to patients with < 3 aberrations. EFS (A,D), OS (Survival; B,E), and CIR (C,F).

material was lost or gained. Chromosomes 1 and 11 were most frequently involved in structural ACAs. Analysis of specific breakpoints showed that 11q23 was the only breakpoint found more than 10 times (data not shown).

#### Univariate analysis of the prognostic impact of ACAs on survival

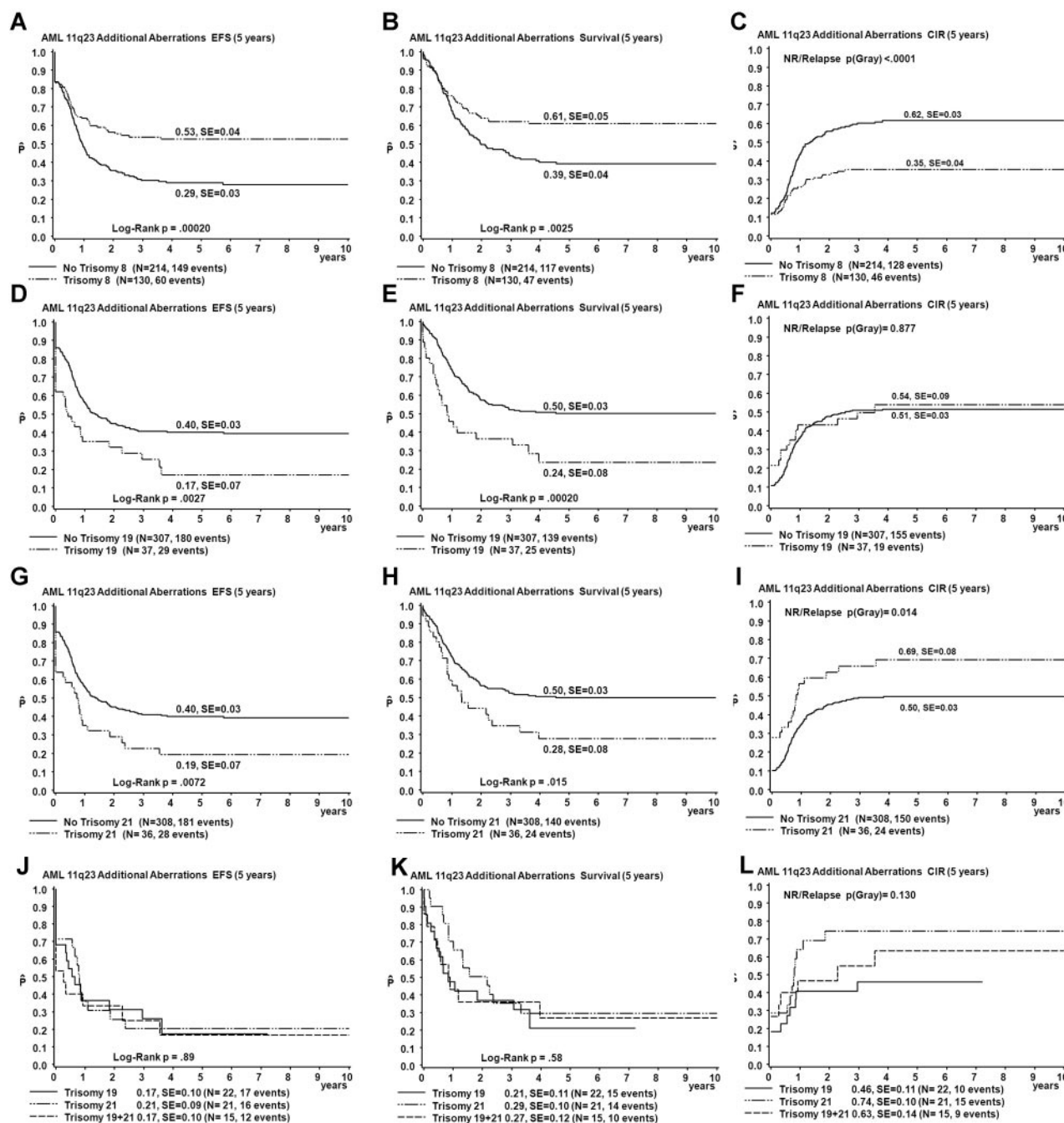
Table 3 summarizes results of the univariate analysis of survival parameters. The EFS and OS estimates of patients with ACAs were significantly lower than those without ACAs (EFS 38% vs 48%,  $P = .002$ ; OS 47% vs 62%,  $P < .001$ ; Figure 3). CIR estimates of patients with ACAs were significantly greater than for those without ACAs (52% vs 38%,  $P < .001$ ; Figure 3). Patients with complex karyotypes had significantly worse outcomes than those without complex karyotypes (EFS 37% vs 46%,  $P = .02$ ; OS 45% vs 59%,  $P = .003$ ; CIR 53% vs 42%,  $P < .001$ ; Figure 3).

The presence of trisomy 8 ( $n = 130$ ) was a favorable prognostic factor (EFS 53% vs 29% for patients without trisomy 8,  $P < .001$ ; OS 61% vs 39% for patients without trisomy 8,  $P = .003$ ; CIR 35% vs 62% for patients without trisomy 8,  $P < .001$ ; Figure 4). Survival differences are mainly explained by reduced relapse rate in trisomy 8 patients (relapse rate 26% vs 49% for patients without trisomy 8,  $P < .001$ ; Figure 4). The presence of trisomy 19 ( $n = 37$ ) and trisomy 21 ( $n = 36$ ) was an unfavorable prognostic factor (EFS 17% vs 40% for patients without trisomy 19,  $P = .003$ ; OS 24% vs 50% for patients without trisomy 19,  $P < .001$ ; CIR 54% vs 51% for patients without trisomy 19,  $P = .88$ ; and EFS 19% vs 40% for patients without trisomy 21,  $P = .007$ ; OS 28% vs 50% for patients without trisomy 21,  $P = .02$ ; CIR 69% vs 50% for patients without trisomy 21,  $P = .01$ ; Figure 4). Both trisomies 19 and 21 were present in 15 patients. Survival curves for patients with either trisomy 19 or 21 were not different from those for patients with both trisomies 19 and 21 (Figure 4). Combined trisomy 19 and trisomy 8 was present in 23 patients. These patients

showed a survival curve intermediate to that of trisomy 8 and trisomy 19 cases (EFS 30%, data not shown). The survival disadvantage of patients with trisomy 19 seems to be determined by refractory disease (probability of CR 74% for patients with trisomy 19 vs 89% for patients with other ACAs, as calculated over the fraction of patients who survive beyond the first 6 weeks after diagnosis,  $P = .04$ ) rather than relapse. In addition, patients with trisomy 19 had a significantly greater incidence of early death (16% vs 3.3% in other ACA cases,  $P = .004$ ), which could not be explained by adverse clinical prognostic factors such as greater WBC or age. Structural aberrations were diverse and randomly distributed among TP groups and survival analysis of patients with specific breakpoints was not feasible because none of the breakpoints was involved > 10 times.

#### Multivariate analyses of the prognostic impact of ACAs on survival

Table 4 summarizes results of the multivariate survival analysis. Cox proportional hazards model for EFS, OS, and relapse incidence of the full cohort ( $n = 733$ ) showed that trisomy 8 and trisomy 19 were independent prognostic factors at  $P < .05$  for EFS (hazard ratio [HR] 0.57,  $P = .02$ ; and HR 1.77,  $P = .01$ ) and OS (HR 0.54,  $P = .03$ ; and HR 2.11,  $P = .002$ ; Table 4). Structural aberrations as a general finding predicted EFS (HR 1.39,  $P = .01$ ; Table 4). The TPs identified by Balgobind et al<sup>10</sup> (10p12, 6q27, 1q21, and 10p11.2) remained significant independent prognostic factors in these models. Trisomy 8, 19, and 21 were not significant factors in the model for the prediction of relapse incidence. Complexity of the karyotype, tested by different cutoff values ( $\geq 2$  aberrations,  $\geq 3$  aberrations, and > 5 aberrations), was not a significant factor for outcome in all models and was therefore excluded from the final model. A separate analysis of t(9;11)(p22;q23) cases showed that they did not differ considerably from the complete cohort (supplemental Figure 2 and supplemental Table 2).



**Figure 4.** Comparison of survival curves obtained from univariate analysis for patients with trisomy 8, trisomy 19, and those with trisomy 21 and defined by strata of occurrence of trisomy 19 and trisomy 21. For curves A-I, patients with a specific trisomy are compared with patients with other ACAs. Patients with trisomy 8 are shown in panels A-C, patients with trisomy 19 in panels D-F, and patients with trisomy 21 in panels G-I. The strata of occurrence of trisomy 19 and trisomy 21 are shown in panels J-L. EFS (A,D,G,J), OS (Survival; B,E,H,K), and CIR (C,F,I,L).

## Discussion

The heterogeneity of pediatric AML is mainly determined by specific karyotypes and molecular aberrations, which have become important prognosticators.<sup>1,3-6,8,11,27-33</sup> In addition, within distinct groups such as 11q23/*MLL*-rearranged AML, we have reported that additional cytogenetic aberrations are of prognostic relevance.<sup>10</sup> In the present exploratory study, we identified trisomy 8, trisomy 19, and trisomy 21 to be recurrent ACAs of prognostic significance in pediatric 11q23/*MLL*-rearranged AML. Multivariate analysis

showed that only trisomy 8 and trisomy 19 as additional aberrations were of independent prognostic value. Notably, the adverse outcome for 11q23/*MLL*-rearranged AML patients harboring trisomy 19 was because of refractory disease and early death rather than an increased rate of relapse. Complex karyotype was a frequent finding (26%) and a negative prognostic factor in univariate analysis only.

Trisomy 19 in AML is an aberration that is rarely found as the sole aberration.<sup>34</sup> In infants with AML it is associated with t(7;12)(q36;p13) and t(7;12)(q32;p13).<sup>35</sup> In most of such cases it can seem to be the sole aberration because of the cryptic t(7;12).<sup>35</sup>

**Table 4. Multivariate survival analysis of the complete cohort by use of the Cox proportional hazards model**

	Cox proportional hazards model								
	EFS			OS			Relapse incidence		
	HR	CI	P	HR	CI	P	HR	CI	P
<b>TP</b>									
9p22	1	reference		1	reference		1	reference	
other	1.15	(0.87-1.51)	.328	1.13	(0.82-1.57)	.461	1.17	(0.92-1.47)	.195
10p12	1.36	(1.01-1.83)	.042*	1.62	(1.16-2.27)	.005†	1.76	(1.36-2.29)	.000†
6q27	2.29	(1.54-3.39)	.000†	2.72	(1.77-4.19)	.000†	2.79	(1.80-4.33)	.000†
19p13	1.06	(0.62-1.80)	.832	1.44	(0.82-2.51)	.204	0.88	(0.57-1.37)	.579
19p13.1	1.11	(0.69-1.79)	.667	0.97	(0.53-1.77)	.931	1.04	(0.71-1.53)	.841
19p13.3	1.06	(0.60-1.88)	.832	1.64	(0.90-3.00)	.105	1.18	(0.71-1.94)	.522
1q21	0.12	(0.03-0.49)	.003†	0.00			0.68	(0.44-1.05)	.080
4q21	1.46	(0.74-2.88)	.276	2.04	(1.02-4.09)	.043*	1.84	(0.99-3.43)	.054
10p11.2	2.12	(1.10-4.06)	.024*	2.56	(1.24-5.32)	.011*	1.37	(0.67-2.78)	.384
17q21	1.14	(0.53-2.43)	.743	1.15	(0.47-2.82)	.763	1.28	(0.68-2.42)	.446
<b>Trisomy</b>									
No trisomy	1	reference		1	reference		1	reference	
8	0.57	(0.36-0.92)	.022*	0.54	(0.32-0.94)	.028*	0.79	(0.56-1.12)	.188
19	1.77	(1.13-2.78)	.012*	2.11	(1.31-3.42)	.002†	1.15	(0.68-1.94)	.596
21	1.35	(0.85-2.13)	.198	1.25	(0.76-2.03)	.377	0.98	(0.60-1.60)	.926
<b>Type</b>									
No ACAs	1	reference		1	reference		1	reference	
numerical	1.16	(0.83-1.63)	.376	1.17	(0.84-1.62)	.353	1.09	(0.81-1.47)	.588
structural	1.39	(1.07-1.80)	.013*	1.27	(0.98-1.63)	.068	1.13	(0.90-1.43)	.288

Results are of 3 independent analyses.

ACA indicates additional cytogenetic aberrations; CI, 95% confidence interval; EFS, event-free survival; HR, hazard ratio; and OS, overall survival.

\*Significant at  $P < .05$  level.

†Significant at  $P < .01$  level.

Trisomy 19 has been described as an additional aberration with adverse prognostic significance in adult AML.<sup>11</sup> It has been postulated that a gene dosage effect of the DNA methyltransferase 1 located on 19p13.2 contributes to the hypermethylation found in patients with MDS and thereby to prognosis.<sup>36</sup> Future studies may reveal whether this mechanism also contributes to aberrant methylation found in pediatric 11q23/*MLL*-rearranged AML.<sup>37</sup>

In our study, trisomy 8 was found to be an independent favorable prognostic factor. Kok et al<sup>38</sup> identified a gene expression signature with high *HOXA* gene expression in adult AML patients with AML with trisomy 8 as the sole abnormality, which clustered together with patients with *MLL*-rearranged AML. This finding may suggest similarities in the biology of these diseases. In contrast, in pediatric MDS, trisomy 8 is recognized as a positive prognostic factor, possibly because of differences in apoptosis regulation between cells with trisomy 8 and cells with other abnormalities.<sup>39,40</sup> To date, it is not clear how trisomy 8 influences the biology of *MLL*-rearranged AML.

Interestingly, in our study, although 26% of all cases of 11q23/*MLL*-rearranged had complex karyotypes, this ACA was not an independent prognostic factor. Although the use of definitions on complex karyotypes is not uniform, the occurrence of complex karyotypes in pediatric AML cohorts has been reported to range from 7% to 15%.<sup>2,6,14,41</sup> A Cancer and Leukemia Group B study on adult de novo AML showed that patients with increased number of aberrations had significantly worse outcome than those with normal karyotypes.<sup>42</sup> Recently, Göhring et al<sup>43</sup> used a new definition of "structural complex karyotype," defined as a karyotype with  $\geq 3$  chromosomal aberrations including at least one structural aberration. This specific karyotype independently predicted very poor survival in a cohort of 192 children with advanced MDS.<sup>43</sup>

Although all the cases of complex karyotype in our study fit their definition, we did not find the presence of such karyotype to be associated with the poor prognosis that was reported in pediatric advanced MDS.<sup>43</sup> Only some studies have specifically shown a correlation between complexity of the karyotype and outcome in pediatric AML.<sup>2,6,14,33,44</sup> EFS rates for patients with complex karyotype have ranged from 29% to 42% in these studies, which is comparable with the EFS obtained in our study. Alternatively, a strong negative association between monosomal karyotype, defined as a karyotype with at least 2 monosomies or 1 monosomy combined with at least 1 structural aberration, and outcome was described in adult AML.<sup>45</sup> This monosomal karyotype was only present in 1.5% ( $n = 11$ ) of our cases and therefore it was not possible to evaluate the predictive value in our pediatric 11q23/*MLL*-rearranged AML cohort.

Although we have added additional prognostic factors in our study, the multivariate models still point out that previously determined risk factors (among which the TPs) retain their independent prognostic significance irrespective of ACA status.

A limitation of our study is the variety of treatment regimens, although all protocols had a similar backbone, including intensive chemotherapy with cytarabine/anthracycline. Unfortunately, numbers were too small to do specific analyses for different protocols, or to draw any meaningful conclusion regarding provided treatment and outcome.

In separate analysis of t(9;11)(p22;q23) cases, we confirmed most of the findings from the complete cohort, regarding frequent recurrent aberrations and predictive factors. In addition, FAB M5 morphology was still recognized as independent favorable prognostic factor in this group of patients.



In conclusion, in this exploratory study we have identified specific ACAs as novel independent prognostic variables in pediatric 11q23/*MLL*-rearranged AML, which can be identified by conventional karyotyping. Future studies should be aimed to test the associations found in this study in different patient cohorts. Our findings may also guide further studies that unravel the biologic differences that determine outcome differences in 11q23/*MLL*-rearranged AML as well as future treatment stratification.

## Acknowledgments

We thank Brian V. Balgobind for data collection and Vani Shanker from the Department for Scientific Editing of the St Jude Children's Research Hospital for her input.

This work was funded by the Rotterdam Oncology Research Foundation KOCR (E.A.C.), the Parents' Foundation Giessen (J.H.), the Swedish Childhood Cancer Foundation (E.S.F.), and by a grant for Clinical Cancer Research from the Ministry of Health, Labor and Welfare, Japan (A.M., T.T., D.T.).

## References

- Balgobind BV, Zwaan CM, Reinhardt D, et al. High BCR expression in pediatric *MLL*-rearranged AML is associated with favorable outcome. *Leukemia*. 2010;24(12):2048-2055.
- Harrison CJ, Hills RK, Moorman AV, et al. Cytogenetics of childhood acute myeloid leukemia: United Kingdom Medical Research Council Treatment trials AML 10 and 12. *J Clin Oncol*. 2010;28(16):2674-2681.
- Hollink IH, van den Heuvel-Eibrink MM, Zimmermann M, et al. Clinical relevance of Wilms tumor 1 gene mutations in childhood acute myeloid leukemia. *Blood*. 2009;113(23):5951-5960.
- Hollink IH, Zwaan CM, Zimmermann M, et al. Favorable prognostic impact of NPM1 gene mutations in childhood acute myeloid leukemia, with emphasis on cytogenetically normal AML. *Leukemia*. 2009;23(2):262-270.
- Kuipers JE, Coenen EA, Balgobind BV, et al. High IGSF4 expression in pediatric M5 acute myeloid leukemia with t(9;11)(p22;q23). *Blood*. 2011;117(3):928-935.
- von Neuhoff C, Reinhardt D, Sander A, et al. Prognostic impact of specific chromosomal aberrations in a large group of pediatric patients with acute myeloid leukemia treated uniformly according to trial AML-BFM 98. *J Clin Oncol*. 2010;28(16):2682-2689.
- Vardiman JW, Thiele J, Arber DA, et al. The 2008 revision of the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia: rationale and important changes. *Blood*. 2009;114(5):937-951.
- Balgobind BV, Zwaan CM, Meyer C, et al. NRIP3: a novel translocation partner of *MLL* detected in a pediatric acute myeloid leukemia with complex chromosome 11 rearrangements. *Haematologica*. 2009;94(7):1033.
- Coenen EA, Zwaan CM, Meyer C, et al. KIAA1524: A novel *MLL* translocation partner in acute myeloid leukemia. *Leuk Res*. 2011;35(1):133-135.
- Balgobind BV, Raimondi SC, Harbott J, et al. Novel prognostic subgroups in childhood 11q23/*MLL*-rearranged acute myeloid leukemia: results of an international retrospective study. *Blood*. 2009;114(12):2489-2496.
- Grimwade D, Hills RK, Moorman AV, et al. Refinement of cytogenetic classification in acute myeloid leukemia: determination of prognostic significance of rare recurring chromosomal abnormalities among 5876 younger adult patients treated in the United Kingdom Medical Research Council trials. *Blood*. 2010;116(3):354-365.
- Creutzig U, Zimmermann M, Lehnbecher T, et al. Less toxicity by optimizing chemotherapy, but not by addition of granulocyte colony-stimulating factor in children and adolescents with acute myeloid leukemia: results of AML-BFM 98. *J Clin Oncol*. 2006;24(27):4499-4506.
- Creutzig U, Zimmermann M, Ritter J, et al. Treatment strategies and long-term results in paediatric patients treated in four consecutive AML-BFM trials. *Leukemia*. 2005;19(12):2030-2042.
- Gibson BE, Wheatley K, Hann IM, et al. Treatment strategy and long-term results in paediatric patients treated in consecutive UK AML trials. *Leukemia*. 2005;19(12):2130-2138.
- Katano N, Tsurusawa M, Hirota T, et al. Treatment outcome and prognostic factors in childhood acute myeloblastic leukemia: a report from the Japanese Children's Cancer and Leukemia Study Group (CCLSG). *Int J Hematol*. 1997;66(1):103-110.
- Lange BJ, Smith FO, Feusner J, et al. Outcomes in CCG-2961, a children's oncology group phase 3 trial for untreated pediatric acute myeloid leukemia: a report from the children's oncology group. *Blood*. 2008;111(3):1044-1053.
- Lie SO, Abrahamsson J, Clausen N, et al. Treatment stratification based on initial in vivo response in acute myeloid leukaemia in children without Down's syndrome: results of NOPHO-AML trials. *Br J Haematol*. 2003;122(2):217-225.
- Perel Y, Auvrignon A, Leblanc T, et al. Treatment of childhood acute myeloblastic leukemia: dose intensification improves outcome and maintenance therapy is of no benefit—multicenter studies of the French LAME (Leucemie Aigue Myeloblastique Enfant) Cooperative Group. *Leukemia*. 2005;19(12):2082-2089.
- Pession A, Rondelli R, Basso G, et al. Treatment and long-term results in children with acute myeloid leukaemia treated according to the AIEOP AML protocols. *Leukemia*. 2005;19(12):2043-2053.
- Ravindranath Y, Chang M, Steuber CP, et al. Pediatric Oncology Group (POG) studies of acute myeloid leukemia (AML): a review of four consecutive childhood AML trials conducted between 1981 and 2000. *Leukemia*. 2005;19(12):2101-2116.
- Ribeiro RC, Razzouk BI, Pounds S, Hijjiya N, Pui CH, Rubnitz JE. Successive clinical trials for childhood acute myeloid leukemia at St Jude Children's Research Hospital, from 1980 to 2000. *Leukemia*. 2005;19(12):2125-2129.
- Smith FO, Alonzo TA, Gerbing RB, Woods WG, Arceci RJ. Long-term results of children with acute myeloid leukemia: a report of three consecutive Phase III trials by the Children's Cancer Group: CCG 251, CCG 213 and CCG 2891. *Leukemia*. 2005;19(12):2054-2062.
- Shaffer LG, Tommerup N, eds. *ISCN 2005: An International System for Human Cytogenetic Nomenclature*. Basel: S. Karger; 2005.
- Betts DR, Ammann RA, Hirt A, et al. The prognostic significance of cytogenetic aberrations in childhood acute myeloid leukaemia. A study of the Swiss Paediatric Oncology Group (SPOG). *Eur J Haematol*. 2007;78(6):468-476.
- Schoch C, Haeflrich T, Haase D, et al. Patients with de novo acute myeloid leukaemia and complex karyotype aberrations show a poor prognosis despite intensive treatment: a study of 90 patients. *Br J Haematol*. 2001;112(1):118-126.
- Creutzig U, Kaspers GJ. Revised recommendations of the International Working Group for diagnosis, standardization of response criteria, treatment outcomes, and reporting standards for therapeutic trials in acute myeloid leukemia. *J Clin Oncol*. 2004;22(16):3432-3433.
- Balgobind BV, Hollink IH, Reinhardt D, et al. Low frequency of *MLL*-partial tandem duplications in paediatric acute myeloid leukaemia using MLPA as a novel DNA screenings technique. *Eur J Cancer*. 2010;46(10):1892-1899.
- Balgobind BV, Lugthart S, Hollink IH, et al. EVI1 overexpression in distinct subtypes of pediatric acute myeloid leukemia. *Leukemia*. 2010;24(5):942-949.
- Balgobind BV, Van den Heuvel-Eibrink MM, De Menezes RX, et al. Evaluation of gene expression signatures predictive of cytogenetic and molecular subtypes of pediatric acute myeloid leukemia. *Haematologica*. 2011;96(2):221-230.
- Balgobind BV, Van Vlierberghe P, van den Ouweland AM, et al. Leukemia-associated NF1 inactivation in patients with pediatric T-ALL and AML lacking evidence for neurofibromatosis. *Blood*. 2008;111(8):4322-4328.
- Hollink IH, van den Heuvel-Eibrink MM, Zimmermann M, et al. No prognostic impact of the WT1 gene single nucleotide polymorphism rs16754 in pediatric acute myeloid leukemia.

## Authorship

Contribution: E.A.C., S.C.R., J.H., R.P., C.M.Z., and M.M.v.d.H.-E. conveyed and planned the study, analyzed the data, and wrote the paper; M.Z. performed the statistical analyses and wrote the paper; and T.A.A., A.A., H.B.B., M.C., U.C., M.N.D., E.F., B.G., H.H., C.J.H., N.A.H., G.J.L.K., A.L., N.L., L.L.N., A.M., C.P., D.R., J.E.R., F.O.S., J.S., I.S., S.S., T.T., D.T., D.W., and Z.Z. participated in data collection and in critical review and final approval of the paper.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Marry M. van den Heuvel-Eibrink, MD, PhD, Associate Professor in Pediatric Oncology/Hematology, Erasmus MC/Sophia Children's Hospital, Department of Pediatric Oncology/Hematology, Rm Sp2568, Dr. Molewaterplein 60, PO Box 2060, 3000 CB Rotterdam, The Netherlands; e-mail: m.vandenheuvel@erasmusmc.nl.

- J Clin Oncol*. 2010;28(28):e523-526; author reply e527-e528.
32. Hollink IH, van den Heuvel-Eibrink MM, Zwaan CM. CEBPA resembles Roman god Janus. *Blood*. 2009;113(26):6501-6502.
  33. Raimondi SC, Chang MN, Ravindranath Y, et al. Chromosomal abnormalities in 478 children with acute myeloid leukemia: clinical characteristics and treatment outcome in a cooperative pediatric oncology group study-POG 8821. *Blood*. 1999;94(11):3707-3716.
  34. National Cancer Institute. Mitelman Database of Chromosome Aberrations and Gene Fusions in Cancer. 2010. <http://cgap.nci.nih.gov/Chromosomes/Mitelman>. Accessed November 4, 2010.
  35. Slater RM, von Drunen E, Kroes WG, et al. t(7;12)(q36;p13) and t(7;12)(q32;p13)—translocations involving ETV6 in children 18 months of age or younger with myeloid disorders. *Leukemia*. 2001;15(6):915-920.
  36. Länger F, Dingemann J, Kreipe H, Lehmann U. Up-regulation of DNA methyltransferases DNMT1, 3A, and 3B in myelodysplastic syndrome. *Leuk Res*. 2005;29(3):325-329.
  37. Alvarez S, Suela J, Valencia A, et al. DNA methylation profiles and their relationship with cytogenetic status in adult acute myeloid leukemia. *PLoS ONE*. 2010;5(8):e12197.
  38. Kok CH, Brown AL, Ekert PG, D'Andrea RJ. Gene expression analysis reveals HOX gene upregulation in trisomy 8 AML. *Leukemia*. 2010;24(6):1239-1243.
  39. Sloand EM, Kim S, Fuhrer M, et al. Fas-mediated apoptosis is important in regulating cell replication and death in trisomy 8 hematopoietic cells but not in cells with other cytogenetic abnormalities. *Blood*. 2002;100(13):4427-4432.
  40. Sloand EM, Pfannes L, Chen G, et al. CD34 cells from patients with trisomy 8 myelodysplastic syndrome (MDS) express early apoptotic markers but avoid programmed cell death by up-regulation of antiapoptotic proteins. *Blood*. 2007;109(6):2399-2405.
  41. Entz-Werle N, Suci S, van der Werff ten Bosch J, et al. Results of 58872 and 58921 trials in acute myeloblastic leukemia and relative value of chemotherapy vs allogeneic bone marrow transplantation in first complete remission: the EORTC Children Leukemia Group report. *Leukemia*. 2005;19(12):2072-2081.
  42. Byrd JC, Mrozek K, Dodge RK, et al. Pretreatment cytogenetic abnormalities are predictive of induction success, cumulative incidence of relapse, and overall survival in adult patients with de novo acute myeloid leukemia: results from Cancer and Leukemia Group B (CALGB 8461). *Blood*. 2002;100(13):4325-4336.
  43. Göhring G, Michalova K, Beverloo HB, et al. Complex karyotype newly defined: the strongest prognostic factor in advanced childhood myelodysplastic syndrome. *Blood*. 2010;116(19):3766-3769.
  44. Stark B, Jeison M, Gabay LG, et al. Classical and molecular cytogenetic abnormalities and outcome of childhood acute myeloid leukaemia: report from a referral centre in Israel. *Br J Haematol*. 2004;126(3):320-337.
  45. Breems DA, Van Putten WL, De Greef GE, et al. Monosomal karyotype in acute myeloid leukemia: a better indicator of poor prognosis than a complex karyotype. *J Clin Oncol*. 2008;26(29):4791-4797.