provide novel therapeutic targets for CAD, hemophilia, and/or thrombosis. High levels of plasma FVIII (> 150%) are a major risk factor for CAD and both arterial and venous thrombosis in humans. Up-regulation of LDLR protein expression may be of therapeutic interest for patients who have elevated plasma FVIII levels. Enhancement of LDLR protein expression is achieved by treatment with 3hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (also called statins). Statins are widely recognized in the treatment of hypercholesterolemia in humans. It would be appealing to study whether statins have the potential to lower elevated levels of plasma FVIII in humans, with the ultimate goal of reducing the risk of atherosclerotic and thrombotic events.

Conflict-of-interest disclosure: The author declares no competing financial interests.

REFERENCES

1. Martinelli N, Girelli D, Lunghi B, et al. Polymorphisms at LDLR locus may be associated with coronary artery disease through modulation of coagulation factor VIII activity and independently from lipid profile. *Blood.* 2010;116(25): 5688-5697.

 Brown MS, Goldstein JL. A receptor-mediated pathway for cholesterol homeostasis. *Science*. 1986;232(4746): 34-47.

3. Bovenschen N, Mertens K, Hu L, Havekes LM, van Vlijmen BJ. LDL receptor cooperates with LDL receptor-related protein in regulating plasma levels of coagulation factor VIII in vivo. *Blood.* 2005;106(3):906-912.

 Willnow TE, Nykjaer A, Herz J. Lipoprotein receptors: new roles for ancient proteins. *Nat Cell Biol.* 1999;1(6): E157-E162.

 Bovenschen N, Herz J, Grimbergen JM, et al. Elevated plasma factor VIII in a mouse model of low-density lipoprotein receptor-related protein deficiency. *Blood*. 2003; 101(10):3933-3939.

6. Lenting PJ, Neels JG, van den Berg BM, et al. The light chain of factor VIII comprises a binding site for low density lipoprotein receptor-related protein. *J Biol Chem.* 1999; 274(34):23734-23739.

7. Ananyeva NM, Makogonenko YM, Kouiavskaia DV, et al. The binding sites for the very low density lipoprotein receptor and low-density lipoprotein receptor-related protein are shared within coagulation factor VIII. *Blood Coagul Fibrinolysis*. 2008;19(2):166–177.

8. Ananyeva NM, Makogonenko YM, Sarafanov AG, et al. Interaction of coagulation factor VIII with members of the low-density lipoprotein receptor family follows common mechanism and involves consensus residues within the A2 binding site 484-509. *Blood Coagul Fibrinolysis*. 2008;19(6): 543-555.

 Marchetti G, Lunghi B, Legnani C, et al. Contribution of low density lipoprotein receptor-related protein genotypes to coagulation factor VIII levels in thrombotic women. *Haematologica*. 2006;91(9): 1261-1263.

10. Vormittag R, Bencur P, Ay C, et al. Low-density lipoprotein receptor-related protein 1 polymorphism 663 C > T affects clotting factor VIII activity and increases the risk of venous thromboembolism. *J Thromb Haemost*. 2007;5(3):497-502.

• • • TRANSPLANTATION

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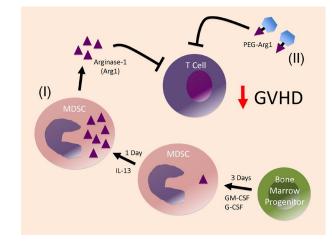
Exploiting arginase to prevent GVHD

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In this issue of *Blood*, Highfill and colleagues introduce a novel procedure to generate arginase-producing MDSCs that can prevent GVHD while maintaining antileukemic responses. An alternative non-cell–based approach using pegylated arginase-1 is also introduced.

llogeneic hematopoietic stem cell transplantation (HSCT) can be an effective treatment for certain hematologic malignancies due to the graft-versus-tumor (GVT) response mediated by donor T cells. However, this procedure has significant hurdles including graft-versus-host-disease (GVHD), infectious complications, and leukemia resistance/ relapse. GVHD is a complex disease that occurs after HSCT when donor T cells recognize foreign histocompatibility antigens on recipient tissues. The resulting donor T-cell reactivity can facilitate the destruction of several host tissues including skin, lung, liver, gut, and those of hematopoietic origin.1 Preserving GVT responses while eliminating GVHD has been a long-standing goal of both basic and clinical researchers.

Myeloid-derived suppressor cells (MDSCs) comprise a heterogeneous population of cells known to suppress T-cell activation and the functions of other immune cells.² These cells arise from bone marrow progenitors and form distinct lineages of cells based on the combination of factors that influence their growth, including vascular endothelial growth factor (VEGF), granulocyte-macrophage colony stimulating factor (GM-CSF), granulocytecolony stimulating factor (G-CSF), and other immunomodulatory cytokines.3 Murine MDSCs are CD11b+ and Gr-1+ (Ly6C/ Ly6G) and are further delineated based on the selective expression of Ly6G (granulocytic MDSCs) or Ly6C (monocytic MDSCs).4 MDSCs suppress T cells through a variety of mechanisms including the production of arginase-1, an enzyme that depletes arginine from the local microenvironment. In this issue, Highfill and colleagues report a new method for generating MDSCs from nonseparated bone marrow cells.5 A single infusion of these MDSCs at the time of transplantation suppressed the proliferation of donor T cells, decreased expression of the CD32 chain, and reduced production of interferon- γ . Importantly, these effects resulted in decreased GVHD-related mortality without eliminating the GVT effect. These MDSCs express arginase-1, and the



Arginase-1-producing MDSC derived from bone marrow progenitors prevent GVHD. Nonseparated bone marrow cells are cultured with GM-CSF and G-CSF for 3 days to generate MDSC and then an additional day with IL-13 to produce high levels of arginase-1 in the MDSC (I). These MDSC then suppress alloreactive T cells primarily through arginase-1 production. As an alternative pathway, pegylated-arginase-1 (PEG-Arg1) can be used to inhibit T-cell alloreactivity (II).

investigators clearly demonstrate that arginase-1 is primarily responsible for their ability to inhibit alloresponses. Importantly, these MDSCs migrated to tissues where donor T-cell priming occurs.

Recently, Zhou and colleagues also reported on the generation of MDSCs from stem cells.6 Those MDSCs were also able to inhibit GVHD, but it is worth highlighting several key differences between their report and the current study. First, while Zhou et al demonstrated that MDSCs could be generated from bone marrow progenitors, their GVHD experiments focused on the use of MDSCs derived from embryonic stem cells. The second major difference between these studies is the culture conditions used to generate MDSCs. The MDSCs generated by Zhou et al were derived from purified progenitor cell populations; they required 8-17 days of culture in rather complex mixtures of 7 different cytokines. In contrast, MDSCs generated in the current study involved the culture of nonseparated bone marrow cells for only 4 days in the presence of 3 cytokines: GM-CSF, G-CSF, and interleukin-13 (IL-13). The presence of IL-13 (during the last day of culture) is the most likely factor responsible for the high levels of arginase-1 (please refer to "(I)" in the figure), because IL-13 has been shown to induce production of arginase-1 by macrophages.7 A third major difference between these studies is the mechanism of inhibition: Zhou et al identified inducible NO synthase, IL-10, and regulatory T-cell induction as the primary suppressive mediators, while the current study by Highfill et al found that

arginase-1 was primarily responsible for the suppressive activity. Although the different suppressive mechanisms observed in these 2 studies may be due to the different culture conditions used to generate the MDSCs, the progenitor cell sources could partly account for the differences. Finally, the current study involved only a single infusion of MDSCs (at the time of transplantation), while Zhou and colleagues administered a total of 3 MDSC infusions in their GVHD studies.

As an interesting alternative to the use of arginase-1+ MDSC cellular therapy, Highfill and colleagues also demonstrate the ability of pegylated-arginase-1 (PEG-arg1) to inhibit GVHD. PEG-arg1 was tested based on the reported ability of this compound to target malignant cells dependent upon arginine for their proliferation.8,9 Remarkably, the GVHD inhibitory effects mediated by PEG-arg1 were quite similar to those induced by the arginase-1+ MDSCs. Although the ability of PEG-arg1 to inhibit GVHD needs to be investigated further, based on the challenges associated with getting cell-based therapies in the clinic, administration of PEG-arg1 to inhibit GVHD may prove to be a more readily translatable approach (refer to "(II)" in the figure).

In conclusion, the report by Highfill et al provides a novel process for generating highly suppressive MDSCs that are dependent upon arginase-1 activity for their suppressive function. These cells inhibit GVHD while maintaining beneficial GVT effects. In the future, it will be interesting to determine whether additional MDSC infusions can be more efficacious, and a more detailed study of the impact on GVT reactivity is warranted because it is possible that arginine starvation could negatively impact the tumor-reactive donor T cells. It is also conceivable that GVT reactivity would be more negatively impacted by systemic PEG-arg1 administration than infusion of arginase-1⁺ MDSCs, which may primarily act in localized tissue microenvironments.

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REFERENCES

1. Welniak LA, Blazar BR, Murphy WJ. Immunobiology of allogeneic hematopoietic stem cell transplantation. *Annu Rev Immunol.* 2007;25:139-170.

 Ostrand-Rosenberg S, Sinha P. Myeloid-derived suppressor cells: linking inflammation and cancer. *J Immunol.* 2009;182(8):4499-4506.

3. Nagaraj S, Gabrilovich DI. Myeloid-derived suppressor cells. *Adv Exp Med Biol.* 2007;601:213–223.

 Youn JI, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumor-bearing mice. *J Immunol.* 2008;181(8):5791-5802.

5. Highfill SL, Rodriguez PC, Zhou Q, et al. Bone marrow myeloid-derived suppressor cells (MDSCs) inhibit graft-versus-host disease (GVHD) via an arginase-1–dependent mechanism that is upregulated by IL-13. *Blood.* 2010; 116(25):5738-5747.

6. Zhou Z, French DL, Ma G, et al. Development and function of myeloid-derived suppressor cells generated from mouse embryonic and hematopoietic stem cells. *Stem Cells.* 28(3):620-632.

7. Rodriguez PC, Quiceno DG, Zabaleta J, et al. Arginase I production in the tumor microenvironment by mature myeloid cells inhibits T-cell receptor expression and antigenspecific T-cell responses. *Cancer Res.* 2004;64(16):5839-5849.

8. Cheng PN, Lam TL, Lam WM, et al. Pegylated recombinant human arginase (rhArg-peg5,000mw) inhibits the in vitro and in vivo proliferation of human hepatocellular carcinoma through arginine depletion. *Cancer Res.* 2007; 67(1):309–317.

 Hernandez CP, Morrow K, Lopez-Barcons LA, et al. Pegylated arginase I: a potential therapeutic approach in T-ALL. *Blood*. 2010;115(25):5214–5221.