

*Conflict-of-interest disclosure: The authors declare no competing financial interests.* ■

**REFERENCES**

1. Mercher T, Cornejo MG, Sears C, et al. Notch signaling specifies megakaryocyte development from hematopoietic stem cells. *Cell Stem Cell*. 2008;3(3):314-326.  
 2. Poirault-Chassac S, Six E, Catelain C, et al. Notch/Delta4 signaling inhibits human megakaryocytic terminal differentiation. *Blood*. 2010;116(25):5670-5678.

3. Radtke F, Fasnacht N, Macdonald HR. Notch signaling in the immune system. *Immunity*. 2010;32(1):14-27.  
 4. Ishiko E, Matsumura I, Ezoe S, et al. Notch signals inhibit the development of erythroid/megakaryocytic cells by suppressing GATA-1 activity through the induction of HES1. *J Biol Chem*. 2005;280(6):4929-4939.  
 5. Mercher T, Raffel GD, Moore SA, et al. The OTT-MAL fusion oncogene activates RBPJ-mediated transcription and induces acute megakaryoblastic leukemia in a knockin mouse model. *J Clin Invest*. 2009;119(4):852-864.

● ● ● **THROMBOSIS & HEMOSTASIS**

Comment on Martinelli et al, page 5688

# LDL receptor polymorphisms revisited

Niels Bovenschen UNIVERSITY MEDICAL CENTER UTRECHT

In this issue of *Blood*, Martinelli and colleagues describe that certain single-nucleotide polymorphisms in the *LDL receptor* gene independently associate with high plasma levels of coagulation factor VIII in patients with coronary artery disease.<sup>1</sup>

In 1985, Drs M. S. Brown and J. L. Goldstein received the Nobel Prize for their discoveries concerning the regulation of cholesterol homeostasis. They identified the low-density lipoprotein receptor (LDLR) as a pivotal player in cholesterol metabolism through binding to apolipoproteins E and B-100.<sup>2</sup> The importance of the LDLR is demonstrated by the fact that genetic defects within the *LDLR* gene are the underlying cause of familial hypercholesterolemia.<sup>2</sup> These patients display elevated plasma LDL choles-

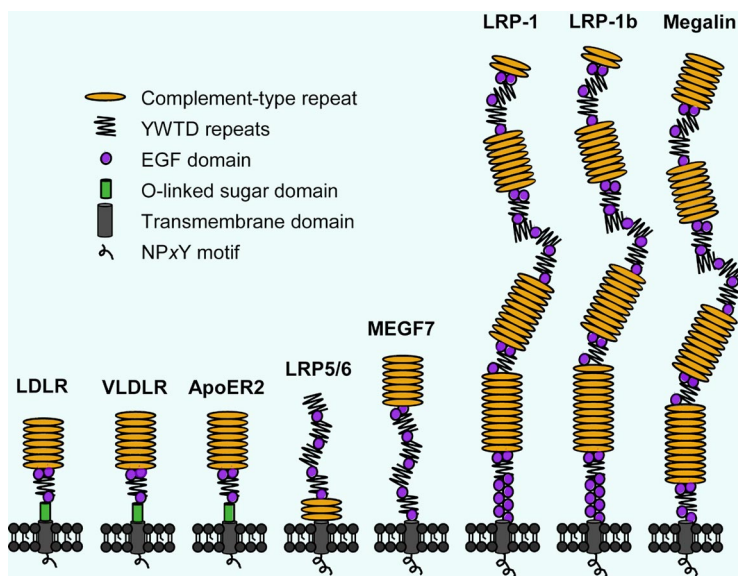
terol concentrations and have a concomitant increased risk for atherosclerosis and coronary artery disease (CAD). Cholesterol-containing apolipoproteins were the only identified LDLR ligands for many years. In 2005, we demonstrated that LDLR also binds to coagulation factor VIII (FVIII) in vitro and contributes to the catabolism of FVIII in mice.<sup>3</sup> In this issue of *Blood*, Martinelli and colleagues build further on these studies by convincingly showing for the first time an independent association between certain LDLR single-nucleotide

polymorphisms (SNPs) and increased plasma levels of FVIII in human subjects with CAD.<sup>1</sup> Their interesting findings support the hypothesis that, in addition to cholesterol metabolism, LDLR also plays a critical role in regulating plasma FVIII levels in humans.

In their study, Martinelli et al measure FVIII coagulant activity in human subjects with or without CAD and confirm that high FVIII levels are an independent risk factor for CAD.<sup>1</sup> Interestingly, high levels of FVIII in plasma positively associate with the occurrence of LDLR T-allele SNPs rs688 and rs2228671.<sup>1</sup> Whereas the LDLR rs688 polymorphism is an independent predictor of plasma FVIII and increased risk of CAD, it does not significantly associate with an altered lipid profile in plasma.<sup>1</sup> This indicates that this particular LDLR polymorphism is associated with cardiovascular disease independent of plasma lipids in these patients, probably through a FVIII-related mechanism. The molecular mechanism by which these LDLR SNPs may regulate plasma FVIII remains an intriguing question that deserves further study. Both SNPs do not result in an amino acid change, but rs688 has been postulated to increase transmembrane-less soluble LDLR in plasma that could bind to FVIII and may interfere with its uptake by LDLR on cells.

LDLR is a member of the *LDLR* gene family that includes at least 8 homologous receptors in mammals, which play partially overlapping roles in many aspects of cell physiology (see figure).<sup>4</sup> Our previous mouse studies have shown that LDLR cooperates with LDLR-related protein-1 (LRP-1) in regulating plasma FVIII levels in vivo.<sup>3,5</sup> In in vitro-binding assays, FVIII directly binds to 4 members of the LDLR family, including LDLR, LRP-1, megalin (LRP-2), and very-low-density lipoprotein receptor (VLDLR; see figure).<sup>3,5-8</sup> Interestingly, 2 polymorphisms in the *LRP-1* gene have previously been proposed as independent predictors of plasma FVIII activity that increase the risk of venous thrombotic events.<sup>9,10</sup> Although Martinelli and colleagues are unable to link plasma FVIII with 1 known LRP-1 polymorphism (-25C/G) in their study group,<sup>1</sup> it would be exciting to further investigate SNPs in LDLR family genes in general as determinants of FVIII levels in plasma, either alone or in combination.

Understanding the catabolism of FVIII in vivo is of particular interest because it could



Structural representation of mammalian LDLR family members. The LDLR family consists of at least 8 homologous transmembrane receptors in mammals, which are composed of the same protein domains with similar topological organizations. Coagulation factor VIII directly binds to LDLR, LRP-1, VLDLR, and megalin (LRP-2) via their clusters of complement-type repeats.<sup>3,5-8</sup> ApoER2 indicates apolipoprotein E receptor 2; and MEGF7, multiple EGF repeat-containing protein 7.

Downloaded from <http://ashpublications.net/blood/article-pdf/116/25/5439/1461521/h805110005439.pdf> by guest on 02 June 2024

provide novel therapeutic targets for CAD, hemophilia, and/or thrombosis. High levels of plasma FVIII (> 150%) are a major risk factor for CAD and both arterial and venous thrombosis in humans. Up-regulation of LDLR protein expression may be of therapeutic interest for patients who have elevated plasma FVIII levels. Enhancement of LDLR protein expression is achieved by treatment with 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (also called statins). Statins are widely recognized in the treatment of hypercholesterolemia in humans. It would be appealing to study whether statins have the potential to lower elevated levels of plasma FVIII in humans, with the ultimate goal of reducing the risk of atherosclerotic and thrombotic events.

*Conflict-of-interest disclosure: The author declares no competing financial interests.* ■

## REFERENCES

- Martinelli N, Girelli D, Lunghi B, et al. Polymorphisms at LDLR locus may be associated with coronary artery disease through modulation of coagulation factor VIII activity and independently from lipid profile. *Blood*. 2010;116(25):5688-5697.
- Brown MS, Goldstein JL. A receptor-mediated pathway for cholesterol homeostasis. *Science*. 1986;232(4746):34-47.
- Bovenschen N, Mertens K, Hu L, Havekes LM, van Vlijmen BJ. LDL receptor cooperates with LDL receptor-related protein in regulating plasma levels of coagulation factor VIII in vivo. *Blood*. 2005;106(3):906-912.
- Willnow TE, Nykjaer A, Herz J. Lipoprotein receptors: new roles for ancient proteins. *Nat Cell Biol*. 1999;1(6):E157-E162.
- Bovenschen N, Herz J, Grimbergen JM, et al. Elevated plasma factor VIII in a mouse model of low-density lipoprotein receptor-related protein deficiency. *Blood*. 2003;101(10):3933-3939.
- Lenting PJ, Neels JG, van den Berg BM, et al. The light chain of factor VIII comprises a binding site for low density lipoprotein receptor-related protein. *J Biol Chem*. 1999;274(34):23734-23739.
- Ananyeva NM, Makogonenko YM, Kouivaskaia DV, et al. The binding sites for the very low density lipoprotein receptor and low-density lipoprotein receptor-related protein are shared within coagulation factor VIII. *Blood Coagul Fibrinolysis*. 2008;19(2):166-177.
- Ananyeva NM, Makogonenko YM, Sarafanov AG, et al. Interaction of coagulation factor VIII with members of the low-density lipoprotein receptor family follows common mechanism and involves consensus residues within the A2 binding site 484-509. *Blood Coagul Fibrinolysis*. 2008;19(6):543-555.
- Marchetti G, Lunghi B, Legnani C, et al. Contribution of low density lipoprotein receptor-related protein genotypes to coagulation factor VIII levels in thrombotic women. *Haematologica*. 2006;91(9):1261-1263.
- Vormittag R, Bencur P, Ay C, et al. Low-density lipoprotein receptor-related protein 1 polymorphism 663 C > T affects clotting factor VIII activity and increases the risk of venous thromboembolism. *J Thromb Haemost*. 2007;5(3):497-502.

## ● ● ● TRANSPLANTATION

Comment on Highfill et al, page 5738

# Exploiting arginase to prevent GVHD

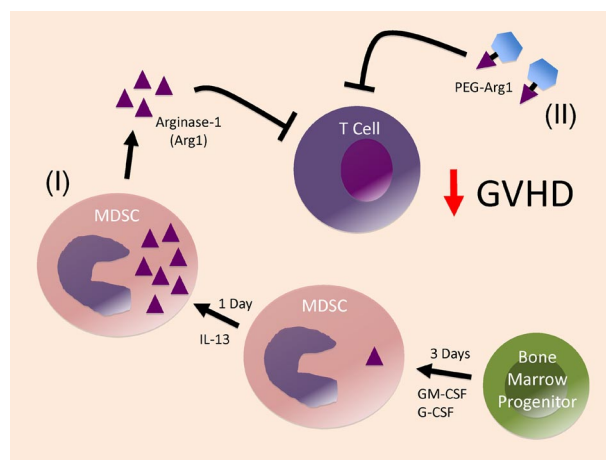
William Hallett and Bryon Johnson MEDICAL COLLEGE OF WISCONSIN

In this issue of *Blood*, Highfill and colleagues introduce a novel procedure to generate arginase-producing MDSCs that can prevent GVHD while maintaining antileukemic responses. An alternative non-cell-based approach using pegylated arginase-1 is also introduced.

**A**llogeneic hematopoietic stem cell transplantation (HSCT) can be an effective treatment for certain hematologic malignancies due to the graft-versus-tumor (GVT) response mediated by donor T cells. However, this procedure has significant hurdles including graft-versus-host-disease (GVHD), infectious complications, and leukemia resistance/relapse. GVHD is a complex disease that occurs after HSCT when donor T cells recognize foreign histocompatibility antigens on recipient tissues. The resulting donor T-cell reactivity can facilitate the destruction of several host tissues including skin, lung, liver, gut, and those of hematopoietic origin.<sup>1</sup> Preserving GVT responses while eliminating GVHD has been a long-standing goal of both basic and clinical researchers.

Myeloid-derived suppressor cells (MDSCs) comprise a heterogeneous population of cells known to suppress T-cell activation and the functions of other immune cells.<sup>2</sup> These cells arise from bone marrow progenitors and form distinct lineages of cells based on the combina-

tion of factors that influence their growth, including vascular endothelial growth factor (VEGF), granulocyte-macrophage colony stimulating factor (GM-CSF), granulocyte-colony stimulating factor (G-CSF), and other immunomodulatory cytokines.<sup>3</sup> Murine MDSCs are CD11b<sup>+</sup> and Gr-1<sup>+</sup> (Ly6C/Ly6G) and are further delineated based on the selective expression of Ly6G (granulocytic MDSCs) or Ly6C (monocytic MDSCs).<sup>4</sup> MDSCs suppress T cells through a variety of mechanisms including the production of arginase-1, an enzyme that depletes arginine from the local microenvironment. In this issue, Highfill and colleagues report a new method for generating MDSCs from nonseparated bone marrow cells.<sup>5</sup> A single infusion of these MDSCs at the time of transplantation suppressed the proliferation of donor T cells, decreased expression of the CD3ζ chain, and reduced production of interferon-γ. Importantly, these effects resulted in decreased GVHD-related mortality without eliminating the GVT effect. These MDSCs express arginase-1, and the



**Arginase-1-producing MDSC derived from bone marrow progenitors prevent GVHD. Nonseparated bone marrow cells are cultured with GM-CSF and G-CSF for 3 days to generate MDSC and then an additional day with IL-13 to produce high levels of arginase-1 in the MDSC (I). These MDSC then suppress alloreactive T cells primarily through arginase-1 production. As an alternative pathway, pegylated-arginase-1 (PEG-Arg1) can be used to inhibit T-cell alloreactivity (II).**