

of the impact of different treatment arms on MRD kinetics. The observation time is still fairly short. I expect that these issues will be the object of future updates of the study. Nevertheless, the importance of results so far achieved by MRD analysis is clear, as documented by the inclusion of MRD as secondary endpoint in most new-generation MCL trials, such as the recently launched IIL-MCL-0208 trial from Intergruppo Italiano Linfomi/European MCL Network (Clinic Gov EudraCT Number 2009-012807-25). In the near future, prospective studies in MCL will probably also address the use of MRD as a surrogate marker, suitable for risk-adapted treatment.

As MRD becomes increasingly important in the clinical management of MCL patients, the issue of standardization and adoption of consensus definitions is becoming critical. During the Second International Symposium on MRD assessment held in Kiel, Germany, in September 2008 (proceedings are currently in the publication process), expert consensus has been established on a number of technical requirements and definitions suitable for clinical use. Moreover, several MRD laboratories have started to perform quality-control rounds in the context of the European Study Group for MRD detection in acute lymphoblastic leukemia and non-Hodgkin lymphoma (now called Euro-MRD). The ultimate aim of such an effort will be the full standardization of PCR techniques in mature lymphoid disorders as already achieved in the acute lymphoblastic leukemia field.⁹

Conflict-of-interest disclosure: M.L. received research funding from Roche, Celgene, and Am-

gen and honoraria from Roche, Amgen, and Bayer. ■

REFERENCES

1. Pott C, Hoster E, Delfau-Larue M-H, et al. Molecular remission is an independent predictor of clinical outcome in patients with mantle cell lymphoma after combined immunotherapy: a European MCL intergroup study. *Blood*. 2010;115(16):3215-3223.
2. Gribben JG, Freedman AS, Neuberg D, et al. Immunologic purging of marrow assessed by PCR before autologous bone marrow transplantation for B-cell lymphoma. *N Engl J Med*. 1991;325(22):1525-1533.
3. Ladetto M, Pagliano G, Ferrero S, et al. Major tumor shrinking and persistent molecular remissions after consolidation with bortezomib, thalidomide, and dexamethasone in patients with autografted myeloma. *J Clin Oncol*. 2010 Mar 22. [Epub ahead of print]
4. Ladetto M, De Marco F, Benedetti F, et al. Prospective, multicenter randomized GITMO/IIL trial comparing intensive (R-HDS) versus conventional (CHOP-R) chemotherapy in high-risk follicular lymphoma at diagnosis: the superior disease control of R-HDS does not translate into an overall survival advantage. *Blood*. 2008;111(8):4004-4013.
5. Pott C, Schrader C, Gesk S, et al. Quantitative assessment of molecular remission after high-dose therapy with autologous stem cell transplantation predicts long-term remission in mantle cell lymphoma. *Blood*. 2006;107(6):2271-2278.
6. Corradini P, Ladetto M, Zallio F, et al. Long-term follow-up of indolent lymphoma patients treated with high-dose sequential chemotherapy and autografting: evidence that durable molecular and clinical remission frequently can be attained only in follicular subtypes. *J Clin Oncol*. 2004;22(8):1460-1468.
7. Magni M, Di Nicola M, Devizzi L et al. Successful in vivo purging of CD34-containing peripheral blood harvests in mantle cell and indolent lymphoma: evidence for a role of both chemotherapy and rituximab infusion. *Blood*. 2000; 96(3):864-869.
8. Andersen NS, Pedersen LB, Laurell A, et al. Pre-emptive treatment with rituximab of molecular relapse after autologous stem cell transplantation in mantle cell lymphoma. *J Clin Oncol*. 2009;27(26):4365-4370.
9. van der Velden VH, Cazzaniga G, Schrauder A, et al. Analysis of minimal residual disease by Ig/TCR gene rearrangements: guidelines for interpretation of real-time quantitative PCR data. *Leukemia*. 2007;21(4):604-611.

to be elucidated. In this issue of *Blood*, Kase et al demonstrate a role for α B-crystallin and VEGF in the intraocular neovascularization process.¹

The α crystallins represent half of the lens protein content, are expressed in the retinal tissues, and protect cells from thermal and metabolic stress. The α crystallins comprise 2 family members, α A and α B. Both are small heat shock proteins, although only α B-crystallin is stress-inducible. α B-crystallin/HSPB5 functions as a chaperone protein for partially unfolded proteins and regulates cytoskeletal integrity. Under physiologic conditions, α B-crystallin is a cytoplasmic protein, colocalizes with vimentin, and can associate with the Golgi apparatus. During cell stress and pathologic conditions, α B-crystallin protects against apoptosis and prevents protein aggregation. Accordingly, α crystallins are responsible for lens transparency and play an important function preventing protein aggregation that would lead to opacity of the lens.³ Correspondingly, α B-crystallin-deficient mice display severe lens degeneration after chemical hypoxia due to excessive protein aggregation.⁴ Further, under hypoxia, α B-crystallin is phosphorylated at serine 59, resulting in enhanced chaperone activity.⁵ In previous studies, the authors and others demonstrated that α B-crystallin protects retinal pigment epithelium (RPE) cells from apoptosis induced by oxidative stress.^{6,7} Thus, α B-crystallin is important in retinal stress responses and the inhibition of RPE apoptosis. Levels of α B-crystallin increase in the eye during retinal degeneration and cataract formation; however, the chaperone activity of α B-crystallin is reduced with aging. This decrease in function may result in the accumulation of misfolded proteins and, thus, leave the retina unable to cope with various stresses, such as hypoxia, oxidative stress, and injury.

Neovascularization is one response of retinal vessels to hypoxic stress. Several recent studies have demonstrated roles for α B-crystallin in angiogenesis. α B-crystallin displays increased expression and phosphorylation in endothelial cells during tube formation in vitro.⁸ Tumors undergo neovascularization as they grow and the inner mass becomes hypoxic. Tumor vasculature in α B-crystallin-deficient mice displays high levels of apoptosis and decreased vessel formation.⁸ Thus, there is a precedence

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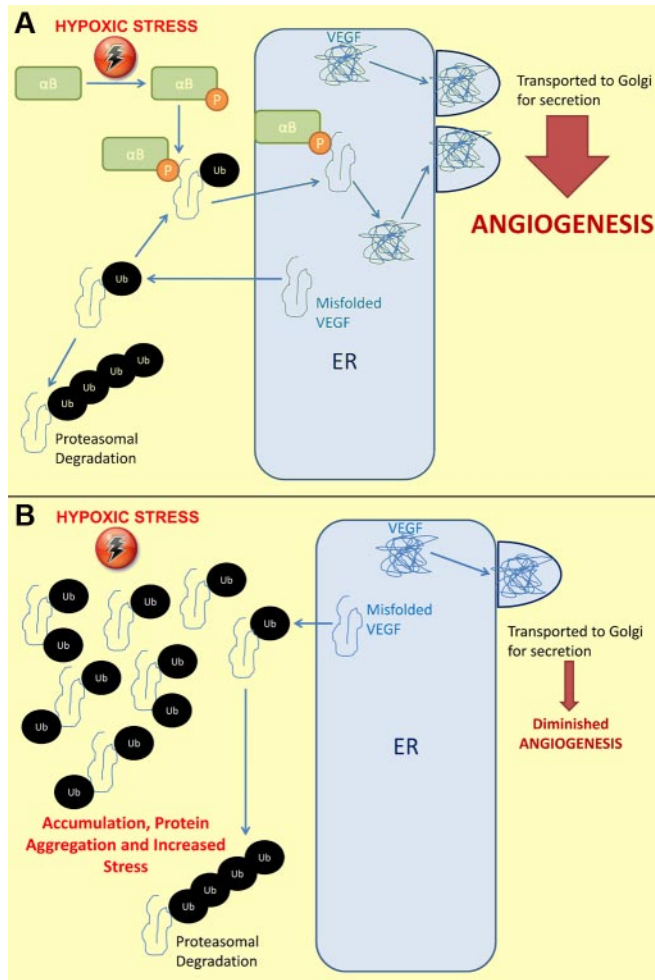
α B-crystallin: a novel VEGF chaperone

Bethany A. Kerr and Tatiana V. Byzova CLEVELAND CLINIC FOUNDATION

In this issue of *Blood*, Kase and colleagues demonstrate that α B-crystallin controls stress-induced intraocular neovascularization via regulation of VEGF secretion.¹

Intraocular neovascularization caused by retinopathy is the primary cause of blindness. Retinopathies, such as retinopathy of prematurity (ROP), diabetic retinopathy (DR), or age-related macular degeneration

(AMD), typically occur from an initial stress or injury inducing a hypoxic response with resultant neovascularization.² The mechanisms controlling neovascularization and stress responses in these retinopathies remain



Hypoxic stress results in increased expression of vascular endothelial growth factor (VEGF), a known stimulator of angiogenesis. Increases in expression result in both properly folded and misfolded VEGF proteins. Misfolded VEGF is exported from the endoplasmic reticulum (ER) into the cytoplasm and ubiquitinated. (A) Hypoxic stress also stimulates phosphorylation of αB -crystallin (αB). Phosphorylated αB -crystallin binds misfolded, monoubiquitinated VEGF and returns it to the ER, where it is folded correctly and transported to the Golgi apparatus for secretion. Thus, the secretion of VEGF is up-regulated, angiogenesis occurs, and hypoxic stress may be reduced. (B) Properly folded VEGF is still transported to the Golgi and secreted. In the absence of αB -crystallin, misfolded, monoubiquitinated VEGF accumulates in the cytoplasm. Some of this will become polyubiquitinated and degrade. The rest of the misfolded VEGF can aggregate, leading to increased stress on the cell. Because misfolded VEGF cannot be transported back to the ER to be refolded, decreased secretion of VEGF occurs and hypoxic stress continues.

of αB -crystallin stimulating neovascularization in hypoxic tissues. In this issue of *Blood*, Kase et al study the role of αB -crystallin in retinal neovascularization using oxygen-induced retinopathy (OIR) and laser-induced choroidal neovascularization (CNV).¹ In CNV, a laser is used to rupture the Bruch membrane, resulting in hypoxic stress and choroidal neovascularization within 7 to 12 days. In OIR, mice are exposed to a high oxygen (75%) environment for the first 7 to 10 days of life during which the retinal vasculature completes development. Five days after a return to normoxia (21%), neovascularization develops due to changes in oxygen tension, resulting in oxidative stress.⁹ In agree-

ment with previous studies on αB -crystallin in neovascularization, OIR- and CNV-injured retinas from αB -crystallin-null mice demonstrate increased endothelial apoptosis and diminished intraocular angiogenesis.¹ Thus, αB -crystallin supports neovascularization in stressed retinas.

αB -crystallin interacts with several important human growth factors, including vascular endothelial growth factor (VEGF). It is interesting to note that the interaction of αB -crystallin with VEGF occurs in the same region αB -crystallin uses to recognize and bind misfolded proteins.¹⁰ This implies that αB -crystallin may chaperone and stabilize misfolded VEGF. Interestingly, VEGF stimulated αB -crystallin

phosphorylation and function in microcapillary endothelial cells during angiogenesis.⁸ Thus, a positive feedback loop may exist between the 2 proteins. In the retina, VEGF is secreted by a majority of the retinal cell types, is important in retinal development, and is expressed at high levels in retinopathies.² In the current study, Kase et al demonstrate that VEGF protein expression decreases in αB -crystallin-deficient mice compared with wild-type mice after injury. The decreased VEGF protein levels in αB -null mice are caused not by changes in mRNA expression, but by altered protein secretion and ubiquitination (see figure). Interestingly, VEGF is polyubiquitinated equally in control and αB -crystallin knockdown RPE cells. However, mono-tetra-ubiquitinated VEGF increases in αB -crystallin-deficient RPE cells. Additionally, OIR and CNV injury increase the levels of phosphorylated αB -crystallin in retinal tissues,¹ which has been associated with increased αB -crystallin chaperone activity.⁵ αB -crystallin binds and colocalizes with VEGF in the endoplasmic reticulum (ER), which may stimulate VEGF secretion and subsequent angiogenesis. Thus, in the absence of αB -crystallin chaperone function, VEGF accumulates and degrades, resulting in decreased secretion. Inhibition of the 26S proteasome in αB -crystallin-deficient mice partially rescues VEGF secretion resulting in increased retinal angiogenesis. Thus, Kase et al demonstrate the novel finding that phosphorylated αB -crystallin chaperones VEGF protein to the ER in hypoxic cells, resulting in proper VEGF folding and secretion. In the absence of αB -crystallin, misfolded, monoubiquitinated VEGF may be exported to the cytoplasm becoming ubiquitinated and degraded (see figure).¹

This novel regulation of VEGF folding and secretion by the chaperone αB -crystallin represents an important control mechanism in the development of retinopathies. Although VEGF is important in physiologic retinal maintenance, it is up-regulated in pathologic retinopathies. In this regard, the stabilization of VEGF by phosphorylated αB -crystallin may contribute to the development of retinopathies. Currently, VEGF antagonists are a main treatment for retinopathies, although VEGF inhibition may lead to atrophy of the retina, particularly in AMD.² Further, systemic inhibition of VEGF may result in various adverse side effects. Thus, phosphorylated αB -crystallin may represent an alternative therapeutic target for controlling intraocular neovascularization. This new mechanism of

VEGF regulation may also function in other cases of hypoxia- or trauma-induced neovascularization, including ischemic pathologies, tumor progression, and wound healing. It remains to be seen whether α B-crystallin is involved in normal homeostasis of endothelium, known to be dependent on VEGF.¹¹ However, because α B-crystallin-null mice display no major problems with vasculature during development or endothelial dysfunction, it appears that α B-crystallin regulates VEGF secretion primarily during postnatal neoangiogenesis. Therefore, manipulation of α B-crystallin phosphorylation and function could be used to inhibit VEGF secretion in pathologic neovascularization or to stimulate VEGF secretion in ischemic tissues. The role of chaperoning of VEGF by α B-crystallin in other tissues and pa-

thologies remains to be revealed, but will lead to exciting advances in the field of angiogenesis.

Conflict-of-interest disclosure: The authors declare no competing financial interests. ■

REFERENCES

1. Kase S, He S, Sonoda S, et al. AlphaB-crystallin regulation of angiogenesis by modulation of VEGF. *Blood*. 2010;115(16):3398-3406.
2. Penn JS, Madan A, Caldwell RB, Bartoli M, Caldwell RW, Hartnett ME. Vascular endothelial growth factor in eye disease. *Prog Retin Eye Res*. 2008;27(4):331-371.
3. Andley UP, Song Z, Wawrousek EF, Brady JP, Bassnett S, Fleming TP. Lens epithelial cells derived from α B-crystallin knockout mice demonstrate hyperproliferation and genomic instability. *FASEB J*. 2001;15(1):221-229.
4. Yaung J, Kannan R, Wawrousek EF, Spee C, Sreekumar PG, Hinton DR. Exacerbation of retinal degeneration in the absence of alpha crystallins in an in vivo model of chemically induced hypoxia. *Exp Eye Res*. 2008;86(2):355-365.
5. Ito H, Okamoto K, Nakayama H, Isobe T, Kato K.

Phosphorylation of alphaB-crystallin in response to various types of stress. *J Biol Chem*. 1997;272(47):29934-29941.

6. Yaung J, Jin M, Barron E, et al. α -Crystallin distribution in retinal pigment epithelium and effect of gene knockouts on sensitivity to oxidative stress. *Mol Vis*. 2007;13:566-577.
7. Alge CS, Priglinger SG, Neubauer AS, et al. Retinal pigment epithelium is protected against apoptosis by α B-crystallin. *Invest Ophthalmol Vis Sci*. 2002;43(11):3575-3582.
8. Dimberg A, Rylova S, Dieterich LC, et al. α B-crystallin promotes tumor angiogenesis by increasing vascular survival during tube morphogenesis. *Blood*. 2008;111(4):2015-2023.
9. Montezuma SR, Vavvas D, Miller JW. Review of the ocular angiogenesis animal models. *Semin Ophthalmol*. 2009;24(2):52-61.
10. Ghosh JG, Shenoy AK Jr, Clark JJ. Interactions between important regulatory proteins and human α B-crystallin. *Biochemistry*. 2007;46(21):6308-6317.
11. Lee S, Chen TT, Barber CL, et al. Autocrine VEGF signaling is required for vascular homeostasis. *Cell*. 2007;130(4):691-703.