Dasatinib treatment of chronic-phase chronic myeloid leukemia: analysis of responses according to preexisting *BCR-ABL* mutations

Martin C. Müller,¹ Jorge E. Cortes,² Dong-Wook Kim,³ Brian J. Druker,⁴ Philipp Erben,¹ Ricardo Pasquini,⁵ Susan Branford,⁶ Timothy P. Hughes,⁶ Jerald P. Radich,⁷ Lynn Ploughman,⁸ Jaydip Mukhopadhyay,⁸ and Andreas Hochhaus^{1,9}

¹III Medizinische Klinik, Universitätsmedizin Mannheim, Universität Heidelberg, Mannheim, Germany; ²Leukemia Department, M. D. Anderson Cancer Center, Houston, TX; ³Seoul St Mary's Hospital, The Catholic University of Korea, Seoul, South Korea; ⁴Oregon Health & Science University Cancer Institute, Portland; ⁵Universidade Federal do Paranà, Curitiba, Brazil; ⁶Institute of Medical and Veterinary Science, Adelaide, Australia; ⁷Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, WA; ⁸Bristol-Myers Squibb, Wallingford, CT; and ⁹Department Hematology/Oncology, Universitätsklinikum Jena, Jena, Germany

Dasatinib is a BCR-ABL inhibitor with 325fold higher potency than imatinib against unmutated BCR-ABL in vitro. Imatinib failure is commonly caused by *BCR-ABL* mutations. Here, dasatinib efficacy was analyzed in patients recruited to phase 2/3 trials with chronic-phase chronic myeloid leukemia with or without *BCR-ABL* mutations after prior imatinib. Among 1043 patients, 39% had a preexisting *BCR-ABL* mutation, including 48% of 805 patients with imatinib resistance or suboptimal response. Sixty-three different *BCR-ABL* mutations affecting 49 amino acids were detected at baseline, with G250, M351, M244, and F359 most frequently affected. After 2 years of followup, dasatinib treatment of imatinib-resistant patients with or without a mutation resulted in notable response rates (complete cytogenetic response: 43% vs 47%) and durable progression-free survival (70% vs 80%). High response rates were achieved with different mutations except T315I, including highly imatinib-resistant mutations in the P-loop region. Impaired responses were observed with some mutations with a dasatinib median inhibitory concentration (IC_{50}) greater than 3nM; among patients with mutations with lower or unknown IC_{50} , efficacy was comparable with those with no mutation. Overall, dasatinib has durable efficacy in patients with or without *BCR-ABL* mutations. All trials were registered at http://www.clinicaltrials.gov as NCT00123474, NCT00101660, and NCT00103844. (Blood. 2009;114:4944-4953)

Introduction

In chronic myeloid leukemia (CML), proliferation is driven by the constitutive tyrosine kinase activity of BCR-ABL, a chimeric oncoprotein encoded after the fusion of the *BCR* gene from chromosome 22 and *ABL* kinase gene from chromosome 9.¹ Most patients ($\sim 80\%$) are diagnosed with CML during the initial chronic phase (CP).² Current therapeutic approaches for CML-CP aim to decrease leukemic cell numbers and prevent disease progression by inhibiting BCR-ABL kinase activity.

Dasatinib (SPRYCEL; Bristol-Myers Squibb) is a potent BCR-ABL inhibitor and is 325-fold more potent than imatinib (Glivec/Gleevec; Novartis) in vitro against unmutated BCR-ABL.³ Across a series of international phase 2/3 trials with more than 2 years of follow-up, dasatinib has demonstrated durable responses in patients with CML after resistance, suboptimal response, or intolerance to imatinib.⁴⁻⁸ After a phase 3 dose optimization trial showing equivalent efficacy and significantly fewer key side effects compared with alternative dosing schedules, dasatinib 100 mg once daily is now the approved dose in patients with CML-CP.⁸

BCR-ABL mutations are a common cause of inadequate response or relapse during imatinib treatment. In a recent study, a *BCR-ABL* mutation was detected in 4 of 34 patients (13%) with CML-CP who experienced a suboptimal response to first-line imatinib, and 18 of 72 (25%) who met criteria for imatinib failure.⁹ Among patients with

CML-CP who developed imatinib resistance after prior interferon-alpha treatment, a *BCR-ABL* mutation was reported in 31% to 42% of patients.^{10,11} In studies of patients with imatinib-resistant advanced-phase CML or Philadelphia chromosome–positive acute lymphoblastic leukemia, a *BCR-ABL* mutation was detected in 30% to 83% of patients.¹⁰⁻¹² Although more than 90 different *BCR-ABL* mutations have been reported affecting more than 55 amino acids, 15 amino acid substitutions account for more than 85% of reported mutations, and 7 amino acids (ABL G250, Y253, E255, T315, M351, F359, H396) are responsible for two-thirds.^{13,14}

In clinical trials, dasatinib treatment responses have been similar in patients with or without *BCR-ABL* mutations, although a detailed analysis has not been performed.⁴⁻⁸ The objective of the current report was to analyze in detail the efficacy of dasatinib in patients with CML-CP recruited to phase 2/3 trials according to *BCR-ABL* mutation status.

Methods

Patients and treatment

Patient data were analyzed from 3 dasatinib trials in CML-CP: CA180-013 (START-C), a phase 2 trial of dasatinib 70 mg twice daily in 387 patients

Submitted April 2, 2009; accepted August 29, 2009. Prepublished online as *Blood* First Edition paper, September 24, 2009; DOI 10.1182/blood-2009-04-214221.

The online version of this article contains a data supplement.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2009 by The American Society of Hematology

An Inside *Blood* analysis of this article appears at the front of this issue.

Table 1. Baseline characteristics and dosing for patients treated in dasatinib trials in chronic-phase CML who had a mutational assessment performed at baseline

	Total, n = 1043	Imatinib resistant, n = 805	Imatinib intolerant, n = 238
Male, no. (%)	516 (49)	429 (53)	87 (37)
Median age, y (range)	56 (18-85)	55 (18-85)	57 (19-84)
Median duration of CML, mo (range)	58 (1-251)	68 (3-251)	24 (1-183)
Highest imatinib dose, no. (%)			
Less than 400 mg/day	1 (< 1)	1 (< 1)	0 (0)
400 to 600 mg/day	643 (62)	422 (52)	221 (93)
More than 600 mg/day	398 (38)	382 (48)	16 (7)
Prior imatinib treatment, %			
Less than 1 y	213 (20)	72 (9)	141 (59)
1 to 3 y	356 (34)	291 (36)	65 (27)
More than 3 y	473 (45)	442 (55)	31 (13)
Other prior therapy, no. (%)			
Interferon-a	614 (59)	508 (63)	106 (45)
Stem cell transplantation	72 (7)	62 (8)	10 (4)
Response achieved prior to imatinib			
failure, no. (%)			
CHR	882 (85)	709 (88)	173 (73)
MCyR	387 (37)	289 (36)	98 (41)
CCyR	192 (18)	135 (17)	57 (24)
Baseline BCR-ABL mutation, no. (%)			
Yes	402 (39)	384 (48)	18 (8)
No	641 (61)	421 (52)	220 (92)
Dasatinib dose received, no. (%)			
100 mg QD	147 (14)	112 (14)	35 (15)
70 mg BID	608 (58)	475 (59)	133 (56)
140 mg QD	139 (13)	105 (13)	34 (14)
50 mg BID	149 (14)	113 (14)	36 (15)

CML indicates chronic myeloid leukemia; CHR, complete hematologic response; MCyR, major cytogenetic response; CCyR, complete cytogenetic response; BID, twice daily; and QD, once daily.

with resistance (n = 288) or intolerance (n = 99) to imatinib^{4,15}; CA180-017 (START-R), a phase 2 randomized trial of dasatinib 70 mg twice daily versus imatinib 800 mg/day in patients with resistance to imatinib 400 to 600 mg/day (n = 150), including 101 patients who were randomized to dasatinib7; and CA180-034, a phase 3 dose-optimization trial in patients with resistance, suboptimal response, or intolerance to imatinib (N = 670), in which patients were randomized using a 2×2 factorial design to receive 1 of 4 dosing schedules: 100 mg once daily (n = 167), 50 mg twice daily (n = 168), 140 mg once daily (n = 167), or 70 mg twice daily (n = 168).⁸ In all 3 studies, dasatinib dose escalation (to a maximum of 180 mg once daily or 90 mg twice daily) was permitted for inadequate response, and dose interruption or reduction (to a minimum of 80 mg once daily or 40 mg twice daily) was permitted for adverse events. Trials were conducted in accordance with the Declaration of Helsinki and were approved by the responsible Institutional Review Board or Ethics Committee of all participating centers. Written informed consent was obtained from all patients.

Eligibility criteria differed between trials for patients with a *BCR-ABL* mutation. In Study 013, patients resistant to imatinib at a dose of 600 mg/day or less were eligible if they had any of the following *BCR-ABL* mutations associated with high-level imatinib resistance: L248V, G250E, Q252H/R, Y253F/H, E255K/V, T315I, F317L, or H396P/R; no specific mutation status was required for patients resistant to imatinib doses higher than 600 mg/day. In Study 017, patients known to have a *BCR-ABL* mutation associated with high-level imatinib resistance (as listed for Study 013) were excluded from enrollment; however, mutation screening prior to enrollment was not obligatory. In Study 034, patients who had any *BCR-ABL* mutation were eligible, including those who had achieved a major cytogenetic response (MCyR) with imatinib at a dose of 400 mg/day or higher.

Assessments

Patient samples were assayed at 1 of 4 laboratories: the University of Heidelberg; Institute of Medical and Veterinary Science; Fred Hutchinson

Cancer Research Center (Studies 013 and 017); or ACGT Inc (Study 034, coordinated by Bristol-Myers Squibb). Peripheral blood cell mRNA was collected at baseline and analyzed for point mutations in the *BCR-ABL* kinase domain by reverse transcription–polymerase chain reaction and direct sequencing, with or without screening by denaturing high-performance liquid chromatography, according to the standard practices of the laboratories. All mutation data in the current analysis were based on conventional direct sequencing. For analyses of individual *BCR-ABL* mutations, patients who had more than 1 mutation were included in separate analyses for each mutation that was detected. Known single nucleotide polymorphisms within the *BCR-ABL* kinase domain¹⁶ were not considered to represent mutations and patients with only polymorphisms were included in analyses of patients with no mutation.

Standard efficacy assessments, including complete hematologic response (CHR), MCyR, and complete cytogenetic response (CCyR), were used to classify treatment responses.^{7,15} All cytogenetic responses were determined using conventional assessment of at least 20 metaphases. A major molecular response (MMR) was defined as a reduction of *BCR-ABL* transcripts to 0.1% or less according to the international scale.¹⁷

Analyses

Time to response was analyzed using a Kaplan-Meier technique in which all nonresponders were censored at the same maximum assessment time. The resulting curve represents the response rate over the observed period and reaches a maximal value corresponding to the overall observed rate. Standard Kaplan-Meier analysis was used to estimate duration of response, progression-free survival (PFS), transformation-free survival (TFS), and overall survival (OS), and 95% confidence intervals (CIs) were calculated. Progression was defined as any of the following events: confirmed accelerated- or blast-phase (AP/BP) disease, loss of a previous CHR or MCyR, increase in Philadelphia chromosome–positive metaphases of 30% or more (Study 034 only), increase in white blood cell count (doubling from lowest value to more than 20×10^9 /L [20 000/mm³] or an increase of more

Table 2. Frequencies of individual baseline *BCR-ABL* mutations and polymorphisms within the total analysis population

Mutation	
H201L	1
Y232S	1
M237V	1
1242T	1
M244V	46
L248V	15
del248-274	3
G250E	60
G250V	1
Q252H	6
Y253F	3
Y253H*	23
F255K	16
E255V	11
E2580	1
L 273M	3
D0760	3
D278G	8
E2/9K	8
E281XT	1
V289I	1
E292V	1
L298V	1
V299L	1
F311I	4
F311L	2
T315I	21
F317L	14
Y342H	1
M351T	54
E355G	19
F359C	5
F359I	12
F359V	27
D363Y	1
L364I	1
A365V	1
A366G	1
V379I	2
1 384M	2
L 387M	5
M388I	3
W308E	4
13930	1
H396P	4
H396R	33
A397P	2
S417Y	1
1418S	1
l418V	2
S438C	3
P441L	1
E450A	2
E450G	2
E450K	2
E450V	1
E453K	4
E453V	1
E459G	1
E459K	12
M4721	12
P480	9
F486S	10
DE04D	13
	1
00145	1

Table 2. Frequencies of individual baseline *BCR-ABL* mutations and polymorphisms within the total analysis population (continued)

	No
Polymorphism	
T240T	1
K247R	6
F311V	1
E499E	19

Excluding known polymorphisms, a total of 480 *BCR-ABL* mutations, comprising 63 different types of mutation, were detected in 402 patients.

*Includes 1 patient whose mutation was incorrectly listed in the Study 013 database as $\ensuremath{\mathsf{Y253K}}$.

[†]X indicates a stop codon.¹⁶

No

than 50 \times 10⁹/L [50 000/mm³] on 2 occasions, performed at least 2 weeks apart), or death at any time regardless of cause^{8,15}; occasional cases where disease progression was reported by the investigator but the progression event was not provided were also included for progression analyses. Loss of CHR was defined as no longer meeting all of the criteria for CHR for a consecutive 2-week period. The range of events considered in the PFS analysis was therefore similar to analyses of event-free survival presented elsewhere.18 Lack of response was not defined as a progression event, and progression in nonresponding patients was defined using the same events as in responding patients. For TFS analyses, transformation was defined as confirmed AP or BP disease or death at any time regardless of cause, and patients with other progression events were not considered in the analysis. Duration of response was measured from the time of response until progression event. For PFS/TFS/duration of response analysis, patients who neither progressed/lost response nor died were censored at last assessment prior to loss to follow-up. Patients were not followed for PFS/TFS/duration of response analysis after study drug discontinuation for protocol-stipulated reasons that included the following: withdrawal of informed consent; any adverse event, laboratory abnormality, or intercurrent illness that, in the investigator's opinion, indicated that continued treatment was not in the patient's best interest; pregnancy; and stem cell transplantation (Study 034). For OS analysis, living patients or patients lost to follow-up were censored on last known alive date. In Study 034, patients were followed for survival status regardless of whether they discontinued study drug. In START-R, OS was not assessed because of the crossover design of the study.

For analysis by median inhibitory concentration to dasatinib, cellular IC50 values for individual mutations were derived from the reports by O'Hare et al3 and Redaelli et al.19 Mutations were grouped according to whether the highest dasatinib IC50 value reported was greater than 3nM, or 3nM or less. Because the IC₅₀ value for T315I is substantially higher than all other mutations, patients with T315I were not included in the IC₅₀-based analysis. Using these criteria, mutations with a dasatinib IC₅₀ greater than 3nM (intermediate IC₅₀) were L248V, G250E, Q252H, E255K/V, V299L, F317L, L384M, and F486S; mutations with a dasatinib IC50 of 3nM or less (low IC₅₀) were M244V, Y253F/H, D276G, E279K, F311L, M351T, F359V, V379I, L387M, and H396P/R; all other mutations were considered to have an unknown IC50. For analysis of responses according to IC50 in patients who had more than 1 baseline mutation and the different mutations belonged to separate IC50 groupings, patients were classified according to highest known IC₅₀, that is, patients with a concurrent mutation with IC₅₀ greater than 3nM were excluded from the group with IC₅₀ of 3nM or less, and patients with a mutation with known IC_{50} (as listed in the current paragraph) were excluded from the unknown IC₅₀ group.

Results

Patient characteristics

A total of 1158 patients with CML-CP after imatinib failure were randomized to (Studies 017 and 034) or treated with (Study 013) dasatinib. Of these, 1043 patients had a mutational assessment performed at baseline and were eligible for analysis, including

Table 3. Cumulative response rates with dasatinib treatment according to BCR-ABL mutation status

	n		n (%)	or n/N (%)	
		CHR	MCyR	CCyR	MMR*
All eligible patients	1043	955 (92)	639 (61)	533 (51)	399/992(40)
Any BCR-ABL mutation	402	362 (90)	225 (56)	175 (44)	125/383(33)
No BCR-ABL mutation	641	593 (93)	414 (65)	358 (56)	274/609(45)
Patients with resistance or suboptimal	805	735 (91)	454 (56)	360 (45)	260/771(34)
response to imatinib					
Any BCR-ABL mutation	384	345 (90)	211 (55)	164 (43)	115/367(31)
No BCR-ABL mutation	421	390 (93)	243 (58)	196 (47)	145/404(36)
Patients with intolerance to imatinib	238	220 (92)	185 (78)	173 (73)	139/221(63)
Any BCR-ABL mutation	18	17 (94)	14 (78)	11 (61)	10/16(63)
No BCR-ABL mutation	220	203 (92)	171 (78)	162 (74)	129/205(63)
Patients with any BCR-ABL mutation					
Grouping by dasatinib IC ₅₀					
IC ₅₀ greater than 3nM (excluding T315I)†	121	110 (91)	56 (46)	39 (32)	26/114(23)
IC ₅₀ 3nM or less‡	176	167 (95)	107 (61)	93 (53)	65/171(38)
IC ₅₀ unknown§	84	79 (94)	60 (71)	43 (51)	34/78(44)
Grouping by dose schedule					
100 mg QD	49	44 (90)	27 (55)	20 (41)	17/47(36)
70 mg BID	240	219 (91)	140 (58)	115 (48)	79/231(34)
140 mg QD	50	42 (84)	28 (56)	17 (34)	11/46(24)
50 mg BID	63	57 (90)	30 (48)	23 (37)	18/59(31)
Patients with 2 or more concurrent BCR-ABL					
mutations at baseline	64	58 (91)	33 (52)	23 (36)	16/61(26)
2 mutations	50	47 (94)	29 (58)	21 (42)	15/48(31)
3 mutations	14	11 (79)	4 (29)	2 (14)	1/13 (8)

CHR indicates complete hematologic response; MCyR, major cytogenetic response; CCyR, complete cytogenetic response; MMR, major molecular response; IC₅₀, median inhibition concentration; QD, once daily; and BID, twice daily.

*MMR rates are calculated in patients who had BCR-ABL transcript levels assessed.

†L248V, G250E, Q252H, E255K/V, V299L, F317L, L384M, and F486S.

‡M244V, Y253F/H, D276G, E279K, F311L, M351T, F359V, V379I, L387M, and H396P/R. Patients were excluded if they had a concurrent mutation with a dasatinib IC₅₀ higher than 3 nM.

§Patients were excluded if they had concurrent mutations with a known dasatinib IC₅₀ (as listed in † and ‡).

805 with imatinib resistance and 238 with imatinib intolerance. Patient characteristics were representative of the broader patient population with imatinib-resistant or -intolerant CML-CP (Table 1). Patients had a median duration of disease prior to dasatinib treatment of 58 months (68 vs 24 months in resistant/suboptimal vs intolerant population). Pretreatment included imatinib at a dose greater than 600 mg/day in 38% and prior interferon- α in 59% of patients. In all 3 studies, minimum follow-up was 24 months from last patient first visit to database lock. Median follow-up in each study (with range) was 25 months (0.1-32.4) for Study 013, 26 months (0.9-32.7) for Study 017, and 26 months (0.1-34.5) for Study 034.



Figure 1. Cumulative response rates in patients with or without a *BCR-ABL* mutation who received dasatinib treatment at any dose or 100 mg once daily (QD) after resistance or suboptimal response to imatinib. CHR indicates complete hematologic response; MCyR, major cytogenetic response; CCyR, complete cytogenetic response; and MMR, major molecular response.



Figure 2. Time to first major cytogenetic response (MCyR). (A) Patients with or without a *BCR-ABL* mutation who received dasatinib treatment at any dose or 100 mg once daily (QD) after resistance or suboptimal response to imatinib. (B) Patients within the total analysis population with commonly mutated amino acids ($n \ge 20$).



Figure 3. Kaplan-Meier analyses of patients with or without a *BCR-ABL* mutation who received dasatinib treatment at any dose or 100 mg once daily (QD) after resistance or suboptimal response to imatinib. (A) Duration of complete cytogenetic response (CCyR). (B) Progression-free survival (PFS). (C) Overall survival (OS).

Baseline BCR-ABL mutation characteristics

Excluding known single nucleotide polymorphisms, a baseline mutation was detected in 402 patients (39%), comprising 384 patients with imatinib resistance/suboptimal response (48% of resistant patients) and 18 patients with imatinib intolerance (8% of intolerant patients). In the 3 clinical studies included in the analysis, a baseline BCR-ABL mutation was detected in 40% of patients from Study 013, 44% of patients from Study 017, and 36% of patients from Study 034. A total of 63 different BCR-ABL mutations were detected affecting 49 amino acids (Table 2). Amino acids most frequently mutated were G250 (n = 61), M351 (n = 54), M244 (n = 46), F359 (n = 42), H396 (n = 37), Y253 (n = 26), and E255 (n = 25). A broad range of mutations were represented in each of the 3 clinical studies, with some sampling differences observed in the frequencies of individual mutations (supplemental Table 1. available on the *Blood* website: see the Supplemental Materials link at the top of the online article). Overall, a T315I mutation was detected in 21 patients (5% of patients with a mutation), which represents a lower frequency than would be expected in the clinical population; this likely reflects a lack of

recruitment by investigators based on the known unresponsiveness of T315I to dasatinib. More than 1 concurrent *BCR-ABL* mutation was detected in 64 patients (2 mutations in 50 patients and 3 mutations in 14 patients). Known single nucleotide polymorphisms detected comprised T240T (n = 1), K247R (n = 6), F311V (n = 1), and E499E (n = 19).

Efficacy according to BCR-ABL mutation status

Patients with or without a baseline BCR-ABL mutation achieved notable rates of response with dasatinib treatment (Table 3; Figure 1). Among patients with imatinib resistance or suboptimal response, cytogenetic response rates in patients with or without a BCR-ABL mutation were MCyR in 55% and 58%, respectively, and CCyR in 43% and 47%, respectively. Rates were similar after 1 year of follow-up (MCyR: 53% and 55%, respectively; CCyR: 40% and 41%, respectively), indicating that most responses were achieved within the first year of treatment. Among patients who achieved a MCyR, median times to response for those with or without a mutation, respectively, were 3.0 (95% CI: 2.8-3.1) versus 2.9 (95% CI: 2.8-3.0) months (Figure 2A), and 91% versus 92% had maintained their MCyR at 12 months. Among patients who achieved a CCyR, median times to CCyR in patients with or without a mutation, respectively, were 5.4 (95% CI: 3.1-5.6) versus 3.1 (95% CI: 2.9-5.5) months, and CCyRs were maintained at 12 months by 95% versus 95% of patients (Figure 3A). Among the total group of patients with imatinib resistance or suboptimal response, rates at 24 months in patients with or without a BCR-ABL mutation, respectively, were 70% versus 80% for PFS, 86% versus 90% for TFS, and 88% versus 92% for OS (Figure 3B-C).

Among patients who were treated with dasatinib 100 mg once daily, time to response, response rates, duration of response, PFS, TFS, and OS in patients with or without *BCR-ABL* mutations were similar to those observed in the total group (Figures 1-3).

Sixty-four patients had 2 or more *BCR-ABL* mutations at baseline, including 5 patients who had T315I plus another mutation (Table 3). Excluding patients with T315I, response rates in patients with 2 or more mutations (CHR in 97%, MCyR in 54%, CCyR in 39%, and MMR in 29%) appeared comparable with the total group of patients with any mutation at baseline. PFS rates at 12 and 24 months in patients with 2 or more mutations were 77% and 55%, respectively (81% and 57%, respectively, excluding patients with T315I).

Efficacy according to individual BCR-ABL mutations

High response rates were observed in patients with the majority of different *BCR-ABL* mutation types, including highly imatinibresistant mutations in L248, Y253, E255, F359, and H396, in addition to other common mutations in G250 and M351 (Figure 4). MCyRs were rapidly achieved in patients with the most commonly mutated amino acids, except T315I (Figure 2B). During dasatinib treatment, patients with an F317L mutation had a high rate of CHR (13/14, 93%), but low rates of MCyR (2/14, 14%) and CCyR (1/14, 7%). Among responding patients with F317L, 5 of 13 had a CHR at baseline (data not available for 4 patients) and 1 patient had a MCyR at baseline (data not available for 1 patient). For the 2 patients with a baseline F317L mutation who had a MCyR recorded on study, durations of MCyR during dasatinib treatment were 85 and 507 days (the latter patient maintained a CCyR for 416 days).

As expected, few responses were observed in the 21 patients who had a baseline T315I mutation: CHR in 6 patients, MCyR in 2 patients (maintained for 381 and 485 days), and no CCyRs.

Figure 4. Best response achieved in each patient and overall rates of hematologic or cytogenetic response after dasatinib treatment of patients with different *BCR-ABL* mutations. Individual mutations occurring in nanomolar), as reported by O'Hare et al³ and Redaelli et al.¹⁹ Other indicates total of all patients with other mutations that are not listed.



Among responding patients with T315I, 4 of 6 had a CHR at baseline (data not available for 1 patient) and 1 patient had a MCyR at baseline (data not available for 1 patient). Of the 21 patients with T315I at baseline, 16 had T315I alone and responses on study were CHR in 5 and MCyR in 1 patient. Of the 5 patients with a T315I mutation who had a different concurrent mutation, 1 patient had a CHR and 1 had a MCyR on study.

Analysis according to dasatinib IC₅₀

To investigate the efficacy of dasatinib against mutations associated with an increased level of in vitro insensitivity, patients with mutations were grouped according to the dasatinib IC₅₀ value for mutations reported by O'Hare et al³ and Redaelli et al.¹⁹ Consistent with previous suggestions,²⁰ an IC₅₀ value greater than 3nM was used to classify mutations as having an intermediate in vitro sensitivity to dasatinib. Other subgroups comprised patients with mutations with low IC₅₀ values (\leq 3nM; ie, high sensitivity to dasatinib) or unknown IC₅₀. Because the clinical resistance and high IC₅₀ of the T315I mutation is well established, patients with this mutation were excluded from the IC₅₀-based analysis.

In 121 patients who had a *BCR-ABL* mutation with an intermediate IC₅₀ (> 3nM), response rates were MCyR in 46%, CCyR in 32%, and MMR in 23% (Table 3). In patients who had a mutation with a low IC₅₀ (n = 176) or unknown IC₅₀ (n = 84), respectively, response rates were MCyR in 61% and 71%, CCyR in 53% and 51%, and MMR in 38% and 44%. Among responding patients in each subgroup, median time to MCyR was similar (2.8-3.0 months; Figure 5A), and MCyR was maintained at 12 months by 87% of responders in the intermediate IC₅₀ subgroup and 93% in both the low and unknown IC₅₀ subgroups. At 24 months, PFS rates were 67% to 80%, TFS rates were 85% to 90%, and OS rates were 89% to 91% (Figure 5B-C).

Mutational assessment at the end of dasatinib treatment

Of the 1043 patients analyzed, 536 (51%) progressed or discontinued dasatinib therapy, and 174 (17%) had a mutational assessment

near to or after the time of progression/discontinuation. Among these 174 patients, 54 new mutations were detected in 47 patients (7 patients developed more than 1 new mutation), including 19 patients who developed a new mutation with a dasatinib IC50 greater than 3nM. The most commonly detected new mutations were T315I, V299L, and F317L (Figure 6). Among patients with or without a new mutation, respectively, 70% versus 37% had a preexisting mutation after prior imatinib treatment and 62% versus 32% had received a prior imatinib dose greater than 600 mg/day; other baseline characteristics were similar (Table 4). Response rates to dasatinib in the 174 patients among those who did eventually develop a new mutation compared with those who did not, respectively, were CHR in 85% versus 79%, MCyR in 34% versus 20%, and CCyR in 23% versus 13%. Among the 47 patients who had developed a new mutation, 24 had experienced a progression event, including 4 who progressed to AP or BP CML at the time of the assessment.

Among the 174 patients who had a mutational assessment close to or after progression/discontinuation, 45 patients had a baseline *BCR-ABL* mutation that persisted until the end of dasatinib treatment (16 with dasatinib $IC_{50} > 3nM$), including 11 patients who both retained a mutation and developed a new mutation. In addition, 42 patients had lost a *BCR-ABL* mutation (19 with dasatinib $IC_{50} > 3nM$), including 24 patients who both lost a mutation and gained a new mutation.

Discussion

This report of more than 1000 patients, representing one of the largest analyses of *BCR-ABL* mutations to date, demonstrates that dasatinib is associated with favorable response rates in patients with or without *BCR-ABL* mutations after prior imatinib treatment, and that responses were durable throughout more than 2 years of follow-up. Among patients from the subgroup with resistance/



Figure 5. Analysis of responses and outcomes according to dasatinib IC_{50} , based on cellular IC_{50} values reported by O'Hare et al³ and Redaelli et al.¹⁹ (A) Time to major cytogenetic response (MCyR). (B) Progression-free survival (PFS). (C) Transformation-free survival (TFS).

suboptimal response to prior imatinib, low rates of progression events (PFS 70% to 80%) and high rates of survival (88% to 92%) were observed in patients with or without a baseline mutation. Across various measures, the efficacy achieved with dasatinib 100 mg once daily, which is associated with optimal tolerability, was similar to other doses including the previously approved 70-mg twice-daily dose.

Notable rates of hematologic, cytogenetic, and molecular responses were observed across almost all individual mutation types. As observed previously, patients with T315I responded poorly to dasatinib. Among patients who had other baseline mutations with a dasatinib IC_{50} greater than 3nM, that is, an intermediate IC_{50} value, response rates appeared to be slightly lower than those observed in patients with mutations having a low dasatinib IC_{50} (\leq 3nM), or unknown IC_{50} . When mutations in the intermediate IC_{50} subgroup were examined individually, a favorable response rate was achieved with several mutations, including patients with highly imatinib-resistant E255K/V mutations (CCyR in 38%/36%, maintained for at least 450 days in 7 of these 9 patients), and also patients with L248V (CCyR in 40%) or G250E (CCyR in 33%). CCyR rates appeared to be lower in patients with



Figure 6. New *BCR-ABL* mutations detected at the time of progression or dasatinib discontinuation (n = 174). Other progression event indicates loss of complete hematologic response, loss of major cytogenetic response, or increasing white cell count. The progression event was not specified by the investigator for 3 patients who developed M244V, T315I, or V299L. AP indicates accelerated phase; BP, blast phase.

F317L (1/14, 7%), Q252H (1/6, 17%), L384M (0/2, 0%), or V299L (0/1, 0%), although because of the small numbers of patients in some cases, caution is advised when interpreting these response rates. These data indicate that the IC₅₀ cutoff of 3nM, as derived from in vitro studies, is only partially predictive of a decreased level of clinical responses to dasatinib, and several mutations with an IC₅₀ greater than 3nM are associated with favorable responses to dasatinib. In groups of patients with mutations having a low or unknown dasatinib IC50, response rates were comparable with those observed in patients with no mutations. Newly emerging mutations associated with clinical resistance to dasatinib have been reported in amino acids T315, F317, and V299, which all have a low in vitro sensitivity to dasatinib.21-24 Overall, these data indicate that dasatinib has a similar potential for efficacy in the majority of patients with imatinib resistance (with or without a BCR-ABL mutation). Alternative treatment options should be considered for patients with baseline mutations that are clearly associated with resistance to dasatinib or lower levels of response, that is, T315I, F317L, and V299L. In the current analysis, these mutations represent less than 5% of imatinib-resistant patients, although the overall incidence may be higher in the clinical population based on the likely underrepresentation of T315I.

In addition to dasatinib, nilotinib (Tasigna; Novartis) is an alternative second-generation BCR-ABL inhibitor that is approved for the treatment of patients with CML-CP with resistance or intolerance to imatinib. Dasatinib and nilotinib have distinct effects on different types of BCR-ABL mutants, although T315I mutants are highly resistant to both agents (and imatinib). In patients who have developed nilotinib resistance, frequently detected mutations include T315I, some P-loop mutations (Y253H and E255K/V), and F359C/V.^{23,25} Unlike dasatinib, F317L has not been commonly associated with nilotinib resistance. In a recent analysis of nilotinib clinical study data in CML-CP, the CCyR rate after 12 months of nilotinib treatment in patients with a mutation in amino acids Y253H, E255K/V, or F359C/V was 0%.25 In the current analysis, dasatinib treatment was associated with favorable CCyR rates in patients with these mutations (61% for Y253H, 38%/36% for E255K/V, and 60%/52% for F359C/V). Response rates to nilotinib were not reported for other individual mutations, preventing further comparisons.

A recent manuscript has reported an IC_{50} -based mutational analysis of patients who were treated at a single center (M. D. Anderson Cancer Center) with either dasatinib or nilotinib after prior imatinib failure.²⁶ Within the study group, 30 patients had CML-CP and a *BCR-ABL* mutation; approximately one-half were

Table 4. Baseline characteristics and	treatment responses in	n patients who had	l a mutational assessm	ent close to or follo	wing progression
or discontinuation of dasatinib therap	y (n = 174) who did or	r did not develop a	new BCR-ABL mutatic	n	

	With new mutation, $n = 47$	Without new mutation, n = 127
Male, no. (%)	26 (55)	57 (45)
Median age, y (range)	60 (32-82)	57 (20-85)
Median duration of CML, mo (range)	72 (4-246)	61 (4-163)
Highest imatinib dose, no. (%)		
Less than 400 mg/d	0 (0)	1 (1)
400 to 600 mg/d	18 (38)	85 (67)
More than 600 mg/d	29 (62)	41 (32)
Prior imatinib treatment, %		
Less than 1 y	5 (11)	21 (17)
1 to 3 y	16 (34)	51 (40)
More than 3 y	26 (55)	55 (43)
Other prior therapy, no. (%)		
Interferon-a	25 (53)	79 (62)
Stem cell transplantation	2 (4)	7 (6)
Response achieved prior to imatinib failure, no. (%)		
CHR	34 (72)	103 (81)
MCyR	11 (23)	32 (25)
CCyR	8 (17)	12 (9)
MMR	4 (9)	8 (6)
Baseline BCR-ABL mutation, no. (%)		
Yes	33 (70)	47 (37)
No	14 (30)	80 (63)
Response achieved on dasatinib therapy, no. (%)		
CHR	40 (85)	100 (79)
MCyR	16 (34)	26 (20)
CCyR	11 (23)	16 (13)

CML indicates chronic myeloid leukemia; CHR, complete hematologic response; MCyR, major cytogenetic response; CCyR, complete cytogenetic response; and MMR, major molecular response.

treated with dasatinib. Compared with the subgroup of patients with CML-CP whose mutations had low IC_{50} values for dasatinib or nilotinib (n = 15), patients with mutations with intermediate IC_{50} values (n = 8) had a substantially lower rate of PFS at 24 months (22% vs 78%) and decreased survival (70% vs 100%). In the current analysis of dasatinib-treated patients only, this was not observed. Reasons for the divergent findings include the possible selection bias due to the small number of patients with CML-CP included in the M. D. Anderson analysis and the effects of nilotinib treatment.

In previous reports, detection of a BCR-ABL mutation has been associated with disease relapse or progression during imatinib treatment.12,27-30 The current analysis suggests that, with the exception of a small number of well-characterized mutations, dasatinib treatment can overcome this negative prognosis. This is consistent with the conclusions of other analyses where patients with mutations received a second-line TKI after imatinib resistance.^{11,31,32} Dasatinib has several attributes that may explain its ability to overcome the majority of imatinib-resistant mutations. BCR-ABL mutations arising during imatinib treatment mostly cluster into the imatinib binding site and other regions in ABL involved in conformational changes required for imatinib binding.^{1,33} Compared with imatinib, dasatinib has a distinct chemistry and a different (although overlapping) binding site within ABL, requiring fewer critical binding residues. In particular, interaction with the P-loop region appears less important for dasatinib than imatinib.³⁴ In addition, in vitro modeling indicates that dasatinib has less stringent conformational requirements for ABL binding and predicts that dasatinib can bind both the catalytically active and inactive conformations of ABL, whereas imatinib can bind only the inactive conformation.³⁴ Furthermore, dasatinib has more than 300-fold higher potency than imatinib against unmutated BCR-ABL, and this increased potency is maintained against most mutated BCR-ABL proteins tested.³

In addition to patients with a BCR-ABL mutation at baseline, response rates and durability of responses were also notable in imatinib-resistant patients without a mutation. This suggests the high potency and distinct chemistry of dasatinib compared with imatinib can overcome other mechanisms of imatinib resistance, such as BCR-ABL overexpression. In addition, low activity of the human OCT-1 influx protein is associated with lower clinical response rates to imatinib,35 and recent data have shown that unlike imatinib, dasatinib influx into cells is not dependent on OCT-1.36 Furthermore, dasatinib inhibits the activity of SRC-family kinases, which have been associated with imatinib resistance.37,38 It should also be noted that a lack of normal hematopoietic reserve in patients with prolonged disease can result in failure to achieve a cytogenetic response, and that this might explain the lack of response observed in some patients without a mutation or with a dasatinib-sensitive mutation.

Overall, the results of this analysis demonstrate that dasatinib is an effective treatment for the majority of patients with CML-CP who have developed an imatinib-resistant *BCR-ABL* mutation and is associated with durable responses and favorable long-term outcomes. In vitro sensitivity of different *BCR-ABL* mutants only partially predicts clinical outcome. Patients who fail imatinib therapy should undergo mutational analysis to facilitate rational selection of second-line therapy.

Acknowledgments

Funding for clinical trials and statistical analysis was provided by Bristol-Myers Squibb (BMS). Professional medical writing assistance was provided by Jeremy Gardner of StemScientific, funded by Bristol-Myers Squibb. A.H. was supported by the German José-Carreras Foundation (DJCLS H 03/01). Clinical trial numbers: CA180-034 (NCT00123474), CA180-013 (NCT00101660), CA180-017 (NCT00103844).

Authorship

Contribution: M.C.M. designed and performed research, collected, analyzed, and interpreted data, and participated in writing the manuscript; J.E.C. collected and interpreted data, performed research, and participated in writing the manuscript; D.-W.K. and B.J.D. collected data; P.E. performed research, collected, analyzed, and interpreted data, and participated in writing the manuscript; R.P. collected data; S.B. and T.P.H. performed research and interpreted data; J.P.R. performed research, interpreted data, and

participated in writing the manuscript; L.P. and J.M. collected data, performed statistical analysis, and interpreted data; and A.H. designed and performed research, collected, analyzed, and interpreted data, and participated in writing the manuscript.

Conflict-of-interest disclosure: J.E.C. and A.H. have received research funding from BMS, Novartis, and Wyeth. D.-W.K. has received research funding from BMS, Novartis, Wyeth, and Merck, and has acted in a consultant/speaker role for Novartis and Wyeth. Oregon Health & Science University and B.J.D. have a financial interest in MolecularMD; B.J.D. receives clinical trial funding from Novartis and BMS. S.B. has received research funding and honoraria from BMS and Novartis. T.P.H. has received research funding and has acted in a consultant/speaker role for BMS and Novartis. J.P.R. has acted in a consultant role for BMS and Novartis and has received research funding and honoraria from BMS and Novartis. L.P. and J.M. are employees of BMS. The remaining authors declare no competing financial interests.

Correspondence: Andreas Hochhaus, Abt Hämatologie und Internistische Onkologie, Universitätsklinikum Jena, Erlanger Allee 101, 07740 Jena, Germany; e-mail: andreas.hochhaus@ med.uni-jena.de.

References

- Sherbenou DW, Druker BJ. Applying the discovery of the Philadelphia chromosome. J Clin Invest. 2007;117(8):2067-2074.
- Cortes J. Natural history and staging of chronic myelogenous leukemia. *Hematol Oncol Clin North Am.* 2004;18(3):569-584.
- O'Hare T, Walters DK, Stoffregen EP, et al. In vitro activity of Bcr-Abi inhibitors AMN107 and BMS-354825 against clinically relevant imatinibresistant Abi kinase domain mutants. *Cancer Res.* 2005;65(11):4500-4505.
- Hochhaus A, Baccarani M, Deininger M, et al. Dasatinib induces durable cytogenetic responses in patients with chronic myelogenous leukemia in chronic phase with resistance or intolerance to imatinib. Leukemia. 2008;22(6):1200-1206.
- Guilhot F, Apperley J, Kim DW, et al. Dasatinib induces significant hematologic and cytogenetic responses in patients with imatinib-resistant or -intolerant chronic myeloid leukemia in accelerated phase. *Blood.* 2007;109(10):4143-4150.
- Cortes J, Kim DW, Raffoux E, et al. Efficacy and safety of dasatinib in imatinib-resistant or -intolerant patients with chronic myeloid leukemia in blast phase. *Leukemia*. 2008;22(12):2176-2183.
- Kantarjian H, Pasquini R, Hamerschlak N, et al. Dasatinib or high-dose imatinib for chronic-phase chronic myeloid leukemia after failure of first-line imatinib: a randomized phase 2 trial. *Blood.* 2007; 109(12):5143-5150.
- Shah NP, Kantarjian HM, Kim DW, et al. Intermittent target inhibition with dasatinib 100 mg once daily preserves efficacy and improves tolerability in imatinib-resistant and -intolerant chronic-phase chronic myeloid leukemia. J Clin Oncol. 2008; 26(19):3204-3212.
- Soverini S, Gnani A, Colarossi S, et al. Abl kinase domain mutations are infrequent in early-chronic phase chronic myeloid leukemia patients resistant to imatinib [abstract]. *Haematologica*. 2008; 93(suppl 1):43. Abstract 0107.
- Soverini S, Colarossi S, Gnani A, et al. Contribution of ABL kinase domain mutations to imatinib resistance in different subsets of Philadelphiapositive patients: by the GIMEMA Working Party on Chronic Myeloid Leukemia. *Clin Cancer Res.* 2006;12(24):7374-7379.
- 11. Jabbour E, Kantarjian H, Jones D, et al. Frequency and clinical significance of BCR-ABL mu-

tations in patients with chronic myeloid leukemia treated with imatinib mesylate. *Leukemia*. 2006; 20(10):1767-1773.

- Lahaye T, Riehm B, Berger U, et al. Response and resistance in 300 patients with BCR-ABLpositive leukemias treated with imatinib in a single center: a 4.5-year follow-up. *Cancer.* 2005; 103(8):1659-1669.
- Branford S. Chronic myeloid leukemia: molecular monitoring in clinical practice. *Hematology Am* Soc Hematol Educ Program. 2007:376-383.
- Apperley JF. Part I: mechanisms of resistance to imatinib in chronic myeloid leukaemia. *Lancet* Oncol. 2007;8(11):1018-1029.
- Hochhaus A, Kantarjian HM, Baccarani M, et al. Dasatinib induces notable hematologic and cytogenetic responses in chronic-phase chronic myeloid leukemia after failure of imatinib therapy. *Blood.* 2007;109(6):2303-2309.
- Ernst T, Hoffmann J, Erben P, et al. ABL single nucleotide polymorphisms may masquerade as BCR-ABL mutations associated with resistance to tyrosine kinase inhibitors in patients with chronic myeloid leukemia. *Haematologica*. 2008;93(9): 1389-1393.
- Hughes T, Deininger M, Hochhaus A, et al. Monitoring CML patients responding to treatment with tyrosine kinase inhibitors: review and recommendations for harmonizing current methodology for detecting BCR-ABL transcripts and kinase domain mutations and for expressing results. *Blood.* 2006;108(1):28-37.
- Druker BJ, Guilhot F, O'Brien SG, et al. Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. *N Engl J Med.* 2006;355(23): 2408-2417.
- Redaelli S, Piazza R, Rostagno R, et al. Activity of bosutinib, dasatinib, and nilotinib against 18 imatinib-resistant BCR/ABL mutants. *J Clin Oncol.* 2009;27(3):469-471.
- O'Hare T, Eide CA, Deininger MW. Bcr-Abl kinase domain mutations, drug resistance and the road to a cure of chronic myeloid leukemia. *Blood*. 2007;110(7):2242-2249.
- Soverini S, Colarossi S, Gnani A, et al. Resistance to dasatinib in Philadelphia-positive leukemia patients and the presence or the selection of mutations at residues 315 and 317 in the BCR-ABL kinase domain. *Haematologica*. 2007;92(3): 401-404.

- Downloaded from http://ashpublications.net/blood/article-pdf/114/24/4944/1320265/zh804909004944.pdf by guest on 08 June 2022
- Khorashad JS, Milojkovic D, Mehta P, et al. In vivo kinetics of kinase domain mutations in CML patients treated with dasatinib after failing imatinib. *Blood.* 2008;111(4):2378-2381.
- Cortes J, Jabbour E, Kantarjian H, et al. Dynamics of BCR-ABL kinase domain mutations in chronic myeloid leukemia after sequential treatment with multiple tyrosine kinase inhibitors. *Blood*. 2007;110(12):4005-4011.
- Shah NP, Skaggs BJ, Branford S, et al. Sequential ABL kinase inhibitor therapy selects for compound drug-resistant BCR-ABL mutations with altered oncogenic potency. *J Clin Invest.* 2007; 117(9):2562-2569.
- Hughes P, Saglio G, Branford S, et al. Impact of baseline BCR-ABL mutations on response to nilotinib in chronic myeloid leukemia patients in chronic phase (CML-CP). J Clin Oncol. 2009; 27(25):4204-4210.
- Jabbour E, Jones D, Kantarjian HM, et al. Longterm outcome of patients with chronic myeloid leukemia treated with second generation tyrosine kinase inhibitors after imatinib failure is predicted by the in vitro sensitivity of BCR-ABL kinase domain mutations. *Blood.* 2009;114(10):2037-2043.
- Ernst T, Erben P, Müller MC, et al. Dynamics of BCR-ABL mutated clones prior to hematologic or cytogenetic resistance to imatinib. *Haematologica*. 2008;93(2):186-192.
- Branford S, Rudzki Z, Walsh S, et al. Detection of BCR-ABL mutations in patients with CML treated with imatinib is virtually always accompanied by clinical resistance, and mutations in the ATP phosphate-binding loop (P-loop) are associated with a poor prognosis. *Blood*. 2003;102(1):276-283.
- Khorashad JS, de LH, Apperley JF, et al. Finding of kinase domain mutations in patients with chronic phase chronic myeloid leukemia responding to imatinib may identify those at high risk of disease progression. *J Clin Oncol.* 2008;26(29): 4806-4813.
- Hochhaus A, Kreil S, Corbin AS, et al. Molecular and chromosomal mechanisms of resistance to imatinib (STI571) therapy. *Leukemia*. 2002; 16(11):2190-2196.
- Jabbour E, Kantarjian H, Jones D, et al. Characteristics and outcomes of patients with chronic myeloid leukemia and T315I mutation following

failure of imatinib mesylate therapy. *Blood.* 2008; 112(1):53-55.

- Nicolini FE, Corm S, Le QH, Roche-Lestienne C, Preudhomme C. The prognosis impact of BCR-ABL P-loop mutations: worse or not worse? *Leukemia*. 2007;21(2):193-194.
- Deininger M, Buchdunger E, Druker BJ. The development of imatinib as a therapeutic agent for chronic myeloid leukemia. *Blood.* 2005;105(7): 2640-2653.
- 34. Tokarski JS, Newitt JA, Chang CY, et al. The structure of dasatinib (BMS-354825) bound to

activated ABL kinase domain elucidates its inhibitory activity against imatinib-resistant ABL mutants. *Cancer Res.* 2006;66(11):5790-5797.

- White DL, Saunders VA, Dang P, et al. Most CML patients who have a suboptimal response to imatinib have low OCT-1 activity: higher doses of imatinib may overcome the negative impact of low OCT-1 activity. *Blood*. 2007;110(12):4064-4072.
- 36. Giannoudis A, Davies A, Lucas CM, et al. Effective dasatinib uptake may occur without human organic cation transporter 1 (hOCT1): implications for the treatment of imatinib-resistant

chronic myeloid leukemia. *Blood.* 2008;112(8): 3348-3354.

- Donato NJ, Wu JY, Stapley J, et al. BCR-ABL independence and LYN kinase overexpression in chronic myelogenous leukemia cells selected for resistance to STI571. *Blood*. 2003;101(2):690-698.
- Wu J, Meng F, Kong LY, et al. Association between imatinib-resistant BCR-ABL mutationnegative leukemia and persistent activation of LYN kinase. *J Natl Cancer Inst.* 2008;100(13): 926-939.