

Cytohesin-1 controls the activation of RhoA and modulates integrin-dependent adhesion and migration of dendritic cells

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Adhesion and motility of mammalian leukocytes are essential requirements for innate and adaptive immune defense mechanisms. We show here that the guanine nucleotide exchange factor cytohesin-1, which had previously been demonstrated to be an important component of beta-2 integrin activation in lymphocytes, regulates the activation of the small GTPase RhoA in primary dendritic

cells (DCs). Cytohesin-1 and RhoA are both required for the induction of chemokine-dependent conformational changes of the integrin beta-2 subunit of DCs during adhesion under physiological flow conditions. Furthermore, use of RNAi in murine bone marrow DCs (BM-DCs) revealed that interference with cytohesin-1 signaling impairs migration of wild-type dendritic cells in complex 3D environments and in

vivo. This phenotype was not observed in the complete absence of integrins. We thus demonstrate an essential role of cytohesin-1/RhoA during amoeboid migration in the presence of integrins and further suggest that DCs without integrins switch to a different migration mode. (Blood. 2009;113:5801-5810)

Introduction

Integrin-mediated adhesion is an essential function of most metazoan cells, and plays a key role in, for example, organ development, the maintenance of tissue architecture, blood clotting, and many aspects of leukocyte biology. The mammalian integrin family consists of 24 heterodimeric cell surface receptors that bear distinct functional binding specificities for extracellular matrix proteins and plasma membrane counterreceptors.¹ Immune cells have highly specific requirements for timing and location of cell-cell and -matrix contacts, which need to be finely tuned to variable conditions of homeostasis and inflammation.

An important hallmark of integrin function in the immune system is the rapid adhesion of leukocytes to activated endothelia, a process that is essential for immune cell egress from the vasculature into either lymphoid organs or into infected tissues. To perform such functions, integrins on leukocytes have to be activated by a process that has been termed “inside-out” signaling. It is now generally believed that integrins on circulating, resting leukocytes are conformationally restricted, so that they normally cannot bind to their protein ligands, such as ICAM-1 or VCAM-1. However, if a leukocyte becomes tethered to an endothelial cell, for example, by selectin-mediated adhesive “rolling,” immune cell chemokine receptors such as CCR7 or CXCR4 may be activated by chemokines present on the activated vessel wall.^{2,3} This, in turn, results in intracellular, heterotrimeric G-protein-coupled signal transduction, which enables very fast conformational changes and ligand binding of cell surface integrins, resulting in a robust firm adhesion, the so-called integrin-mediated leukocyte “arrest.”³ Shear forces imposed by the blood flow apparently play an important role in the

regulation of this process.⁴ The mode of activation of immune cell integrins is incompletely understood. Several intracellular proteins that directly or indirectly interact with the cytoplasmic domains of the leukocyte-specific beta-2 integrins, such as LFA-1, have been implicated in this function. These include the actin- and integrin-binding protein talin, which is important for the general coupling of integrin-mediated cell adhesion to the actin cytoskeleton,⁵⁻⁷ the GTPase Rap1⁷⁻⁹ and its effector proteins RAPL and RIAM,⁹⁻¹¹ the ADAP/Vav/SLP76 complex in T cells,¹²⁻¹⁴ the Rho GTPases RhoA¹⁵ and Cdc42,¹⁶ and, recently, the so-called kindlin proteins.^{17,18}

Our laboratory has identified the guanine nucleotide exchange factor (GEF) for ADP-ribosylation factor (ARF) GTPases, cytohesin-1, as an LFA-1 interacting protein. Cytohesin-1 binds specifically to the cytoplasmic portion of the integrin beta-2 subunit CD18.¹⁹ In Jurkat T cells, overexpression of cytohesin-1 results in up-regulation of LFA-1 adhesion to ICAM-1, whereas the expression of the isolated PH domain of cytohesin-1, which is required for inositol-phospholipid-mediated membrane recruitment of the protein, abrogates LFA-1 activation.^{20,21} Cytohesin-1 was also found to be an important regulator of beta-2 integrin-mediated recruitment of monocytic cells to human endothelial cells under physiological flow conditions.²² The precise mechanism of integrin activation by cytohesin-1 is not understood but it involves both direct interactions, possibly resulting in triggering conformational switches, and signal transduction through its GEF domain, which activates ARF GTPases, a function that is important for cell spreading.²³

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Another important requirement for immune cell functionality is their ability to rapidly move through complex environments in the interstitium. So-called dendritic cells (DCs) of myeloid origin become activated by pathogen encounter in infected tissues through Toll-like receptor and inflammatory cytokine signaling. During this process, DCs “mature,” that is, they undergo massive changes in morphology that enable their rapid motility. At the same time, DCs up-regulate surface proteins that enable their trafficking to lymphoid organs, for example, chemokine receptors such as CCR7, as well as costimulatory receptors and MHC molecules that are required for proper T-cell activation by DCs in secondary lymphoid organs. The importance of integrins in coupling ECM recognition to cytoskeletal rearrangements in interstitial immune cell migration has been subject to debate. Several years ago, seminal work by Friedl et al²⁴ suggested that integrins might not be required for interstitial motility of T cells. Recently, an elegant study published by Lämmermann et al,²⁵ which used dendritic cells and neutrophil granulocytes from mice that lack all integrins on the leukocyte surface, showed that integrin-deficient immune cells apparently migrate normally *in vitro* and *in vivo*. However, rapid interstitial migration of these cells was shown to be dependent on rapid forward flow of actin at the cell front, and on their capacity to deform organelles such as the nucleus to “squeeze” through narrow passages in collagen/matrix webs. This latter ability was dependent on intracellular signaling pathways controlled by the GTPase RhoA.²⁵

We show here that the ARF-GEF and beta-2 integrin binding protein cytohesin-1 serves a novel, dual role in mediating dendritic cell adhesion and migration, and the common denominator of both functions is the positioning of the protein upstream of RhoA. The expression of cytohesin-1 protein has been efficiently silenced by RNAi in mature dendritic cells of both human and murine origin. Interestingly, mature DCs are capable of activating their beta-2 integrins by chemokine signaling under physiological flow conditions, a function that is linked to PI3-kinase and cytohesin-1-mediated inside-out signal transduction. Furthermore, cytohesin-1 is shown for the first time to control DC migration in 3D collagen gels, and to regulate mature DC migration *in vivo* by functioning upstream of RhoA activation. Strikingly, cytohesin-1 is apparently not required for 3D migration of integrin knockout DCs (quadruple ablation of the integrin alpha-v, beta-1, beta-2, and beta-7 genes, respectively²⁵). These findings suggest that immune cell migration may also be subclassified into distinct modes, and that the integrin-dependent mode selectively requires the function of cytohesin-1 upstream of RhoA.

Methods

Generation of human monocyte-derived DCs

See supplemental methods (available on the *Blood* website; see the Supplemental Materials link at the top of the online article). Animal care and experiments were done in compliance with institutional guidelines and the German law for Welfare of Laboratory Animals. Isolation of primary cells was approved by the University of Bonn ethics committee.

Generation of murine bone marrow-derived DCs

See supplemental methods.

Generation of integrin-deficient bone marrow-derived DCs

Generation of integrin alpha-v-, beta-1-, beta-2-, and beta-7-deficient bone marrow-derived DCs (BM-DCs) has been described by Lämmermann et al.²⁵

Antibodies

The following monoclonal antibodies were used against the human antigens: HLA-DR, CD80, CD86 (all from BD Pharmingen, San Diego, CA), anti-CCR7 and anti-CXCR4 (both from R&D Systems, Minneapolis, MN), anti-CD18 MHM23 (Dako, Carpinteria, CA). The 327C^{26,27} specific for activated human CD18 epitopes was a gift from D. Staunton (Seattle, WA). E. Kremmer (Munich, Germany) generated a rat monoclonal antibody (7H2) against human cytohesin-1 as previously described²³ and 2D7, which also detects cytohesin. These monoclonal antibodies were against the following mouse antigens: MHCII, CD80 (clone 16-10A1), CD86 (clone B7-2; all from eBiosciences, San Diego, CA). These monoclonal antibodies were against both human and mouse antigens: anti-RhoA (Santa Cruz Biotechnology, Santa Cruz, CA) and anti-Rac (Chemicon, Temecula, CA). For flow cytometry analysis, the following isotype-matched antibody controls were used: rat IgG1 PE (BD Pharmingen) and rat IgG1 FITC (clone eBRG1; eBiosciences).

Oligonucleotide sequences and plasmids

See supplemental methods.

siRNA transfections

Oligonucleotides (5 μg siRNA against the target sequence of cytohesin-1; 10 μg against the target sequence of RhoA or Rac1, respectively) were transferred to a 4-mm cuvette (Bridge, Providence, RI) and incubated for 3 minutes with 4×10^6 immature DCs (imDCs) in 100 μL Optimem (Invitrogen, Frederick, MD) before electroporation in a Gene Pulser X cell + CE module (Bio-Rad, Hercules, CA). Pulse conditions were square wave, 1000 V, 2 pulses, and 0.5-ms pulse length. Forty-eight hours after electroporation, imDCs were stimulated to mature (Document S1). Seventy-two to 96 hours after electroporation, RNAi efficiency was tested using a standard Western blot protocol and silenced mature DCs (mDCs) were used for functional assays.

Transfection of monocyte-derived DCs with plasmid DNA

See supplemental methods.

Confocal laser scanning microscopy

See supplemental methods.

Static adhesion assays

See supplemental methods.

Static adhesion assay with GFP-transfected monocyte-derived DCs

See supplemental methods.

Laminar flow assays

See supplemental methods.

BM-DC/T-cell adhesion

See supplemental methods.

Transwell migration assay

See supplemental methods.

Migration assay with GFP-transfected monocyte-derived dendritic cells

See supplemental methods.

In vivo migration assays

See supplemental methods.

3D collagen gel chemotaxis assay

3D chemotaxis assays were processed as described by Friedl et al²⁴ with the following modifications. Collagen I (Pure Col; Inamed, Fremont, CA) was mixed with 0.7% sodium bicarbonate (Invitrogen) and 1 × MEM (Sigma-Aldrich, St Louis, MO) on ice. BM-DCs in VLE-RPMI/0.5% FCS were carefully mixed with the collagen solution at a 2:1 ratio, resulting in gels with a collagen concentration of 1.6 mg/mL and a final cell concentration of 1.5 × 10⁶ cells/mL gel. Collagen–BM-DC mixtures were placed in custom-made chemotaxis chambers and incubated at 37°C/5% CO₂ for 90 minutes. After collagen polymerization, 600 ng/mL CCL19 (R&D Systems) in VLE-RPMI/0.5% FCS was added on top of the gel. Time-lapse series of motile BM-DCs in 3D collagen gel were recorded using a fully automated inverted TE Eclipse microscope (Nikon, Tokyo, Japan), equipped with a 10 × phase-contrast objective (Nikon), motorized xyz-stage (Märzhäuser, Wetzlar, Germany), a climate chamber (37°C/5% CO₂ humidity), and a CCD-1300 camera (Vosskühler, Osnabrück, Germany). For every single experiment 60 individual BM-DCs were tracked over 3 hours (5 minutes/frame) using the manual tracking plug-in of ImageJ (National Institutes of Health [NIH], Bethesda, MD). Subsequently, cell directionality, forward migration index, and velocity were calculated and visualized as plots and videos by analyzing the acquired data with the Chemotaxis and Migration tool plug-in (Ibidi, Martinsried, Germany) for ImageJ.

GTPase activation assays

See supplemental methods.

Statistical analysis

Statistical significance between data groups was determined by analysis of variance (ANOVA) with the InStat software (GraphPad, San Diego, CA) and considered to be significantly different at *P* values less than .05. The *P* values are depicted as asterisks in the figures. *** indicates *P* value less than .001; **, *P* value less than .01; and *, *P* value less than .05.

Results

Efficient RNAi of cytohesin-1 does not affect maturation of human or murine dendritic cells

The mRNA of cytohesin-1 is up-regulated in human monocyte-derived dendritic cells (Mo-DCs) upon maturation induced by lipopolysaccharide (LPS) and TNF- α .²⁶ We performed Western blot analysis to establish the up-regulation on the protein level (Figure 1A). On day 7 after isolation of monocytes, expression of cytohesin-1 was detectable in immature dendritic cells by the specific anti-cytohesin-1 antibody 7H2. Incubation of the cells with LPS and TNF- α yielded a strong increase of cytohesin-1 expression, which reaches peak levels at day 9. The maturation of DCs was determined by controlling the expression of the characteristic markers HLA-DR, CD80, and CD86 by fluorescence-activated cell sorting (FACS) analysis.

RNA interference can be used to specifically inhibit gene expression and to analyze gene function.²⁷ Using electroporation, we silenced cytohesin-1 protein expression during maturation of DCs. Compared with Mo-DCs that were transfected with a control siRNA, RNAi of cytohesin-1 dramatically reduced specific protein expression, as detected by Western blot analysis (Figure 1B). In contrast to human Mo-DCs, cytohesin-1 is constitutively expressed in murine BM-DCs (not shown). Nonetheless, RNAi-mediated silencing of cytohesin-1 yielded a similar reduction of specific protein expression in this cell type (Figure 1C).

To test whether RNAi of cytohesin-1 interferes with maturation of DCs, we measured the cell surface expression of maturation markers with the help of flow cytometry (FACS). Figure S9A

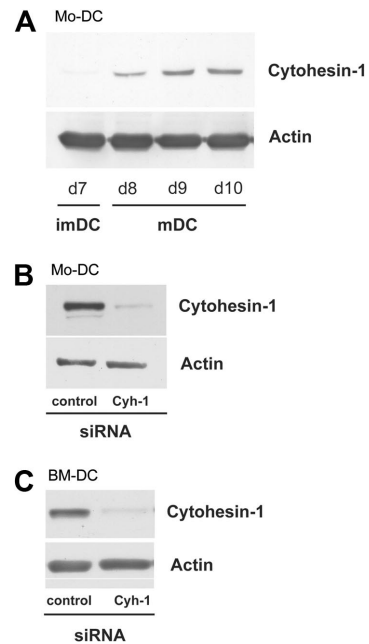


Figure 1. Cytohesin-1 is up-regulated in human Mo-DCs during maturation, and is efficiently silenced by RNAi in both human and murine DCs. Western blot analysis shows that cytohesin-1 expression is up-regulated in human monocyte-derived dendritic cells (Mo-DCs) upon maturation induced by LPS (1 μ g/mL)/TNF α (50 ng/mL; panel A lanes d8, d9, d10). Expression of cytohesin-1 is barely detectable in immature dendritic cells (imDCs), but maturation results in a strong increase of cytohesin-1 expression, reaching maximum levels at day 9 (A). Using square wave electroporation, cytohesin-1 expression was efficiently silenced in mature DCs. Western blot analysis shows that RNAi of cytohesin-1 reduces specific protein expression 96 hours after electroporation in human Mo-DCs (B) or murine bone marrow-derived DCs (BM-DCs, C).

shows that maturation of Mo-DCs is not altered after transfection with a cytohesin-1–specific siRNA, compared with control cells. Furthermore, we assessed potential effects of cytohesin-1 knockdown on the expression of CCR7 and CXCR4 on mature Mo-DCs. CCR7 and CXCR4 are receptors for the chemokines CCL19 and CXCL12, respectively, which are important mediators of mDC migration in vivo. As shown in Figure S9A, RNAi of cytohesin-1 interfere neither with the expression of CCR7, nor with that of CXCR4. Hence, the reduction of cytohesin-1 protein expression does not affect the cellular levels of a variety of important surface proteins of mature Mo-DCs. Consistently, RNAi of cytohesin-1 does not significantly affect maturation marker expression of murine BM-DCs, compared with control samples (Figure S9B).

Cytohesin-1 mediates chemokine-induced activation of CD18 in mature Mo-DCs

We have previously shown that overexpressed cytohesin-1 colocalizes with CD18 in Jurkat cells and activates LFA-1 adhesion to ICAM-1.²³ Here, we demonstrate that endogenous cytohesin-1 colocalizes with the beta-2 integrin subunit CD18 in mature Mo-DCs upon chemokine stimulation (Figure 2A). Immunofluorescence tracings reveal that endogenous cytohesin-1 is predominantly located in the cytoplasm in Mo-DCs. However, stimulation of mature Mo-DCs with the chemokine CCL19 induces translocation of cytohesin-1 to the plasma membrane (Figure 2A middle panel) where it remains colocalized with CD18 (after antibody-mediated clustering of the integrin (Figure 2A bottom panel). To elucidate the functional significance of this finding, we analyzed

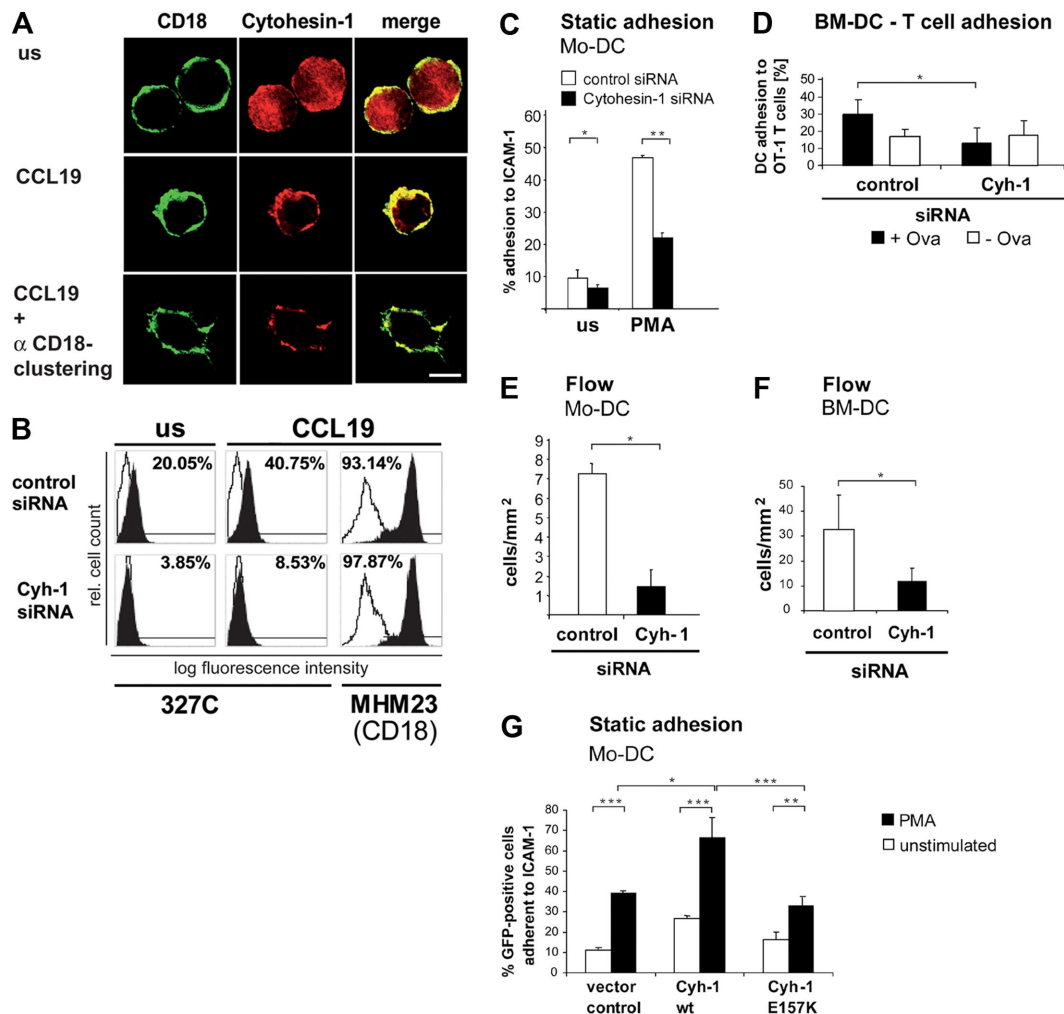


Figure 2. Cytohesin-1 (red fluorescence) colocalizes with CD18 (green fluorescence) and mediates both CD18 activation and adhesion. In contrast to unstimulated mature Mo-DCs where cytohesin-1 (detected by mAb 7H2) localizes to the cytoplasm (A top panel), stimulation with 200 ng/mL CCL19 for 10 minutes induces translocation of cytohesin-1 to the plasma membrane (A middle panel) where it colocalizes with beta-2 integrins, clustered by an anti-CD18 antibody (A bottom panel; bar represents 5 μ m). All shown fluorescence staining images are confocal images, focused on the medial z-position of the respective cells, that is, the cytoplasm and the plasma membrane (A). The different fluorochromes of secondary antibodies in the costaining of cytohesin-1 and CD18 were acquired sequentially by the use of a confocal laser scanning microscope (A). Flow cytometric analysis shows that CCL19 (200 ng/mL for 10 minutes) strongly increases the expression of the high-affinity state (detected by mAb 327C) of LFA-1 (B). RNAi of cytohesin-1 reduces expression of the activation epitope, compared with control cells (B). In contrast, a constitutive CD18 epitope (detected by mAb MHM23) is not altered after RNAi of cytohesin-1 (B). The empty profiles in the flow cytometry histograms indicate the background cell stainings by isotype controls (B). RNAi of cytohesin-1 strongly reduces static adhesion of Mo-DCs to ICAM-1-Fc compared with control cells (C). Cytohesin-1 siRNAs selectively down-regulate antigen-specific BM-DC adhesion to OT-1 T cells (D). Under flow conditions, RNAi of cytohesin-1 reduces dynamic adhesion of Mo-DCs to human umbilical vein endothelial cells (HUVECs; E) or of BM-DCs to brain endothelioma cells (B end5; F). Overexpression of wild-type cytohesin-1 but not of the GEF-deficient E157K mutant increases static adhesion of GFP-cotransfected Mo-DCs to ICAM-1-Fc (G). In the static adhesion assays, Mo-DCs were stimulated with 50 ng/mL PMA for 60 minutes at 37°C (C,G). Error bars indicate \pm SD (C-G). *** P < .001, ** P < .01, * P < .05. Each experiment was repeated at least 3 times independently. Each single experiment was performed in duplicate.

the affinity state of beta-2 integrins on mature Mo-DCs after stimulation with CCL19. Chemokines induce a rapid affinity increase of LFA-1, which is sufficient to cause an arrest of circulating lymphocytes on endothelial cells.²⁸ Integrin activation is accompanied by conformational changes. The antibody 327C may be used to report CD18 epitopes that are expressed only on activated beta-2 integrins because 327C detects the high-affinity state of LFA-1.^{29,30} Using this antibody, we performed FACS staining of unstimulated and CCL19-stimulated mature Mo-DCs (Figure 2B). Unstimulated Mo-DCs showed partial activation of CD18, the level of which differed between cell donors (data not shown). However, CCL19 strongly increased the expression of the 327C epitope in Mo-DCs, whereas RNAi of cytohesin-1 completely abrogates expression of this epitope (Figure 2B). To test whether RNAi of cytohesin-1 reduces surface expression of CD18, we also analyzed steady-state beta-2 integrin expression on Mo-

DCs, using antibody MHM23, which recognizes a constitutive CD18 epitope. CD18 expression is not altered after RNAi of cytohesin-1, compared with control cells (Figure 2B). Therefore, cytohesin-1 is an important positive regulator of CD18 affinity in mature Mo-DCs and mediates chemokine-induced inside-out activation of beta-2 integrins on this cell type.

Cytohesin-1 regulates adhesion of human and murine dendritic cells

We then assessed static cell adhesion to ICAM-1 using cytohesin-1 knockdown Mo-DCs and control cells. Our results clearly establish cytohesin-1 as an endogenous regulator of beta-2 integrin-mediated adhesion, because RNAi of cytohesin-1 markedly reduced static adhesion of unstimulated and PMA-stimulated Mo-DCs to ICAM-1 (Figure 2C). To test whether cytohesin-1 regulates

DC adhesion in more physiological settings, we analyzed antigen-specific adhesion of BM-DCs to transgenic OT-1 T cells, which specifically recognize an epitope of the ovalbumin protein. Figure 2D shows that ovalbumin-loaded BM-DCs specifically adhered to immobilized OT-1 T cells. Importantly, this antigen-specific cell-cell interaction is abrogated if cytohesin-1 expression is selectively silenced in the dendritic cell, suggesting that beta-2 integrin activation on DCs contributes to DC/T-cell interactions.

Furthermore, we analyzed the interactions of Mo-DCs and BM-DCs with endothelial cells under physiological flow conditions. To this end, we first tested dynamic adhesion of mature dendritic cells to human umbilical vein endothelial cells (HUVECs) with the help of time-lapse video imaging using a flow chamber. HUVECs were stimulated to express chemokines and adhesion molecules by incubation with TNF- α overnight. In addition, CXCL12 was added, which binds to the surface of endothelial cells. Mature DCs were then injected into the chamber at a steady flow rate of 0.2 mL/minute (1 dyne/cm²) and subsequent DC interactions with HUVECs were evaluated in 6 different optical fields within 3 minutes. Adhesion to unstimulated HUVEC monolayers under flow was not observed (not shown). RNAi of cytohesin-1 clearly reduced adhesion of mDCs to HUVECs under flow compared with control cells (Figure 2E). The importance of cytohesin-1 for the activation of DC integrins under flow conditions was also confirmed for the murine system, by assessing BM-DC adhesion to TNF- α -activated murine brain endothelium cells (Figure 2F). All these data are in accordance with our earlier observation, which revealed that cytohesin-1 overexpression enhanced LFA-1-mediated arrest of leukocytes on activated endothelium under flow.²² Finally, we assessed whether the GEF function of cytohesin-1 is important for regulating adhesive events of DCs. To this end, we used the novel microporation method “Transfection of monocyte-derived DCs with plasmid DNA” to overexpress wild-type cytohesin-1 or the GEF-deficient mutant E157K²³ together with a cotransfected plasmid that directs expression of the green fluorescent protein. Figure 2G shows that overexpression of wild-type cytohesin-1, but not of the E157K mutant, strongly enhances GFP-positive Mo-DC adhesion to ICAM-1.

Taken together, we show strong and coherent evidence that cytohesin-1—which requires an active GEF function—is an important regulator of integrin-mediated adhesion of dendritic cells in several systems.

DC migration strongly depends on cytohesin-1 function in vitro and in vivo

An important hallmark of mature DCs is their ability to migrate rapidly and directionally in chemokine gradients. To assess the chemotaxis of Mo-DCs, we first performed so-called transwell migration assays. To this end, transwell polycarbonate filters were either used uncoated, or were coated with the integrin ligands ICAM-1-Fc, fibronectin, or collagen I, respectively. RNAi of cytohesin-1 strongly reduced CCL19- or CXCL12- (not shown) stimulated migration of Mo-DCs in all conditions tested (Figure 3A-D). Overexpression analysis revealed that the GEF function of the protein is also required for this function (Figure 3E). Thus, a GEF-competent cytohesin-1 strongly regulates migration of Mo-DCs in vitro.

We investigated whether cytohesin-1 is important for migration of mature DCs in vivo. Using RNAi we reduced the expression of cytohesin-1 in mature, murine bone marrow-derived DCs (BM-DCs) to approximately 10% of endogenous levels (Figure 1C).

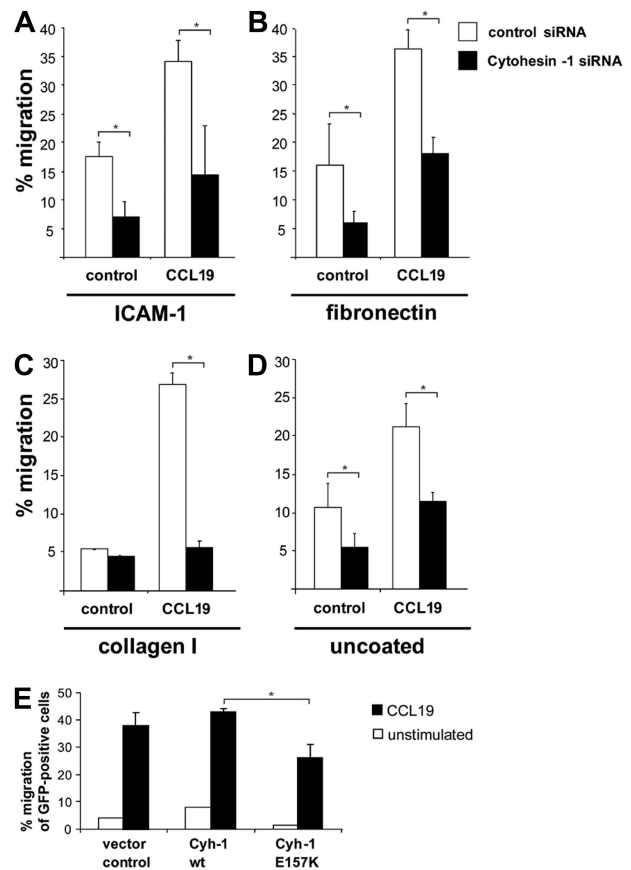


Figure 3. RNAi of cytohesin-1 reduces chemotaxis of mature Mo-DCs. Transwell migration assays show that CCL19 (200 ng/mL)-induced chemotaxis is overall strongly reduced in cytohesin-1 knockdown mature Mo-DCs, using integrin ligands ICAM-1-Fc (~10 μ g/mL; A), fibronectin (50 μ g/mL; B), collagen I (30 μ g/mL; C), or uncoated filters (D). Overexpression of wild-type cytohesin-1 but not of the GEF-deficient E157K mutant increases chemotaxis of GFP-cotransfected Mo-DCs (uncoated filters; E). Error bars indicate \pm SD. * P < .05. Each experiment was repeated at least 3 times independently. Each single experiment was performed in duplicate.

Control cells and cytohesin-1 knockdown cells were differentially labeled with either CFSE or TAMRA dyes, and equal numbers of these cell groups were subsequently injected into the hind footpad of wild-type C57/BL6 mice.³¹ To exclude a perturbing effect of the dyes themselves on in vivo migration of DCs, the labeling of the respective populations was interchanged between experiments (Figure 4B,C). Twenty-four to 30 hours after injection, the draining (popliteal) lymph nodes (LNs) were extracted from these animals; single-cell suspensions were prepared and analyzed by FACS. Figure 4A shows that cytohesin-1 knockdown strongly reduced the migration of mature DCs to the draining lymph node in vivo. The numbers of BM-DCs that reached the LN were decreased by approximately 50% compared with control BM-DCs (set to 100% migration). Therefore, cytohesin-1 protein expression in DCs appears highly important for efficient migration of such cells from the tissue periphery to secondary lymphoid organs.

Cytohesin-1 selectively regulates 3D migration of integrin-bearing DCs

Using live cell imaging microscopy, we further studied the bulk migratory behavior of mature BM-DCs. Migration within tissues requires coordinated interactions with constituents of the extracellular matrix.³² With the help of 3D collagen type I gels as a structural scaffold, we modeled such complex conditions and

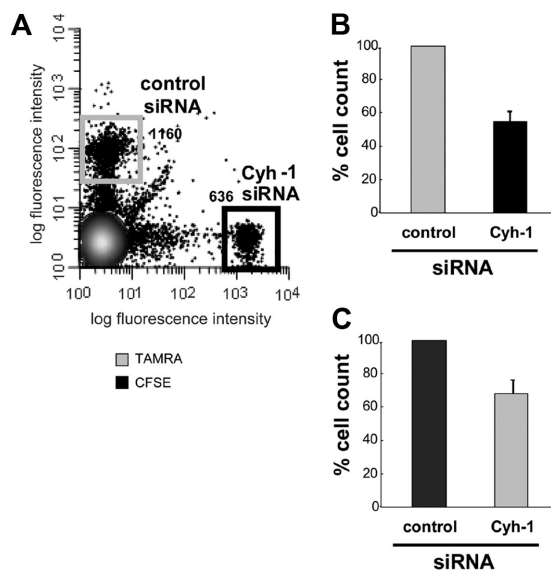


Figure 4. RNAi of cytohesin-1 strongly reduces in vivo migration of mature BM-DCs. Mature BM-DCs after transfection with control siRNA or cytohesin-1 siRNA (for silencing efficiency, see Western blot analysis of cytohesin-1 expression after RNAi of cytohesin-1, panel 1C) were labeled with 10 μ M TAMRA or 1 μ M CFSE, respectively. Cytohesin-1 knockdown DCs (2.5×10^5) and control DCs (2.5×10^5) were injected into the hind footpad of wild-type C57/BL6 mice. Flow cytometric analysis of cell suspensions from the extracted draining (popliteal) lymph nodes 24 to 30 hours after injection shows that in vivo migration of cytohesin-1 knockdown BM-DCs is strongly reduced, compared with control cells (A,B). To exclude fluorochrome effects on DC migration, the staining was switched between the experiments (C). Control BM-DCs are set to 100% migration (B,C). Error bars indicate \pm SD. Each experiment was repeated 3 times independently.

analyzed the migration of BM-DCs in these settings. Stimulation with CCL19 induced vigorous and directional migration of control DCs (Figure 5A,B and corresponding Videos S1-S4). In contrast, RNAi of cytohesin-1 strongly impaired CCL19-stimulated migration of BM-DCs within collagen (Figure 5C and corresponding Videos S5,S6). Using the ImageJ plug-in for manual tracking, we detected that average migratory speed (velocity), forward migration index, and directionality of cytohesin-1 knockdown DCs were significantly reduced compared with control cells (Figure 5E,F). Because Lämmermann et al had shown that the inhibition of the RhoA/myosin II pathway by blebbistatin resulted in the abrogation of DC contractility in 3D environments,²⁵ we opted to use RNAi of RhoA in our assays. Strikingly, knockdown of RhoA yielded a highly similar phenotype as that of cytohesin-1 (Figure 5D-F and corresponding Videos S7,S8).

Lämmermann et al²⁵ had shown that integrins are not required for BM-DC migration in 3D environments and in vivo. Using quadruple knockout BM-DCs, which do not express integrins on their surfaces,²⁵ we therefore assessed whether cytohesin-1 was also required for integrin-independent migration of BM-DCs in 3D collagen. Figure 5 shows that 3D migration of wild-type cells was comparable with integrin-deficient BM-DCs, as expected (Figure 5I,K). Most interestingly, however, we found that RNAi of cytohesin-1, which was similarly efficient in both cell types (Figure 5O), selectively affected 3D migration of integrin-bearing wild-type cells (Figure 5J,M,N). These findings suggest an important limiting function of cytohesin-1 in 3D leukocyte migration of integrin-positive DCs.

Cytohesin-1 controls activation of RhoA in several cell types

The apparent involvement of cytohesin-1 in beta-2 integrin activation and in integrin-independent interstitial migration of dendritic

cells prompted us to investigate a possible and as-yet-unappreciated upstream role of cytohesin-1 in RhoA activation of dendritic cells. Giagulli et al¹⁵ had shown that RhoA is an important regulator of chemokine-induced T-cell adhesion to activated endothelium in vitro and in vivo, possibly by inducing or by stabilizing the high-affinity state of LFA-1. Rho GTPases are furthermore well known as key regulators of cell migration. In controlling diverse signal transduction pathways that link cell surface receptors to a variety of intracellular responses, they influence actin filament and microtubule dynamics, cell polarity, vesicular trafficking, and gene expression.³³ Furthermore, Rho GTPases are essential in maintaining immune responses as they regulate lymphocyte and dendritic cell function and migration.³⁴ Rho GTPases cycle between an active GTP-bound and an inactive GDP-bound state. The active GTP-loaded enzyme selectively interacts with downstream effector molecules. Activation of the GTPases was analyzed with the help of enzyme linked immunosorbent assays (ELISAs), using immobilized Rac or RhoA effector proteins PAK-1 and rhotekin, respectively. We observed that RNAi of cytohesin-1 reduces GTP loading of RhoA, whereas the activity of Rac is enhanced by cytohesin-1 knockdown in Mo-DCs (Figure 6A). Furthermore, a consistent reduction of RhoA activity is observed upon cytohesin-1 or RhoA protein knockdown (Figure 6B) in BM-DCs.

Significantly, the control of RhoA activity by cytohesin-1 is not restricted to immune cells. Overexpression of wild-type cytohesin-1 in HeLa cells results in an increase of RhoA activity, compared with the control sample that had been transfected with the vector. The GEF function of cytohesin-1 is required for RhoA activation through this pathway, because overexpression of the cytohesin-1 E157K (GEF minus) mutant²³ abrogates this effect (Figure 6C). A role of cytohesin-1 function upstream of RhoA in this cell type was also confirmed by the alternative method of effector protein pull-down and subsequent Western blot analysis (Figure 6D). Consistently, Rac activity was significantly enhanced by cytohesin-1 RNAi in HeLa cells as well (Figure 6D).

To corroborate a full coherence of the functional activities of RhoA and cytohesin-1 in all test systems used, we confirmed that RNAi of RhoA also reduces a CCL19-induced activation epitope expression of beta-2 integrins in Mo-DCs (Figure 6E). Furthermore, RhoA was found to be required for BM-DC adhesion to endothelioma cells under physiological flow conditions (Figure 6F). Finally, overexpression of RhoA enhances adhesion of Mo-DCs to ICAM-1-Fc, whereas the expression of the GTP-loading-deficient mutant T19N abrogates this effect (Figure 6G).

We had previously shown that the functional activity of cytohesin-1 in human lymphocytes is regulated by PI3-kinase-dependent membrane recruitment of the protein.²⁰ Consistently, we found that CCL19-dependent membrane recruitment of cytohesin-1, or LFA-1 activation epitope expression in Mo-DCs (Figure S10A,B), can be blocked using the PI3 kinase inhibitor LY-294002. In accordance with these data, inhibition of PI3-kinase in mature BM-DCs yielded a reduced RhoA activity (Figure S10C).

Taken together, we conclude that cytohesin-1 acts upstream of RhoA in directing integrin-dependent dendritic cell adhesion, their migration in 3D collagen gels, and likely their migratory behavior in inflamed tissues.

Actin polymerization and the generation of membrane protrusions are not affected by cytohesin-1 in migrating DCs

Cell migration highly depends on actin polymerization leading to membrane protrusions such as lamellipodia and filopodia, and is

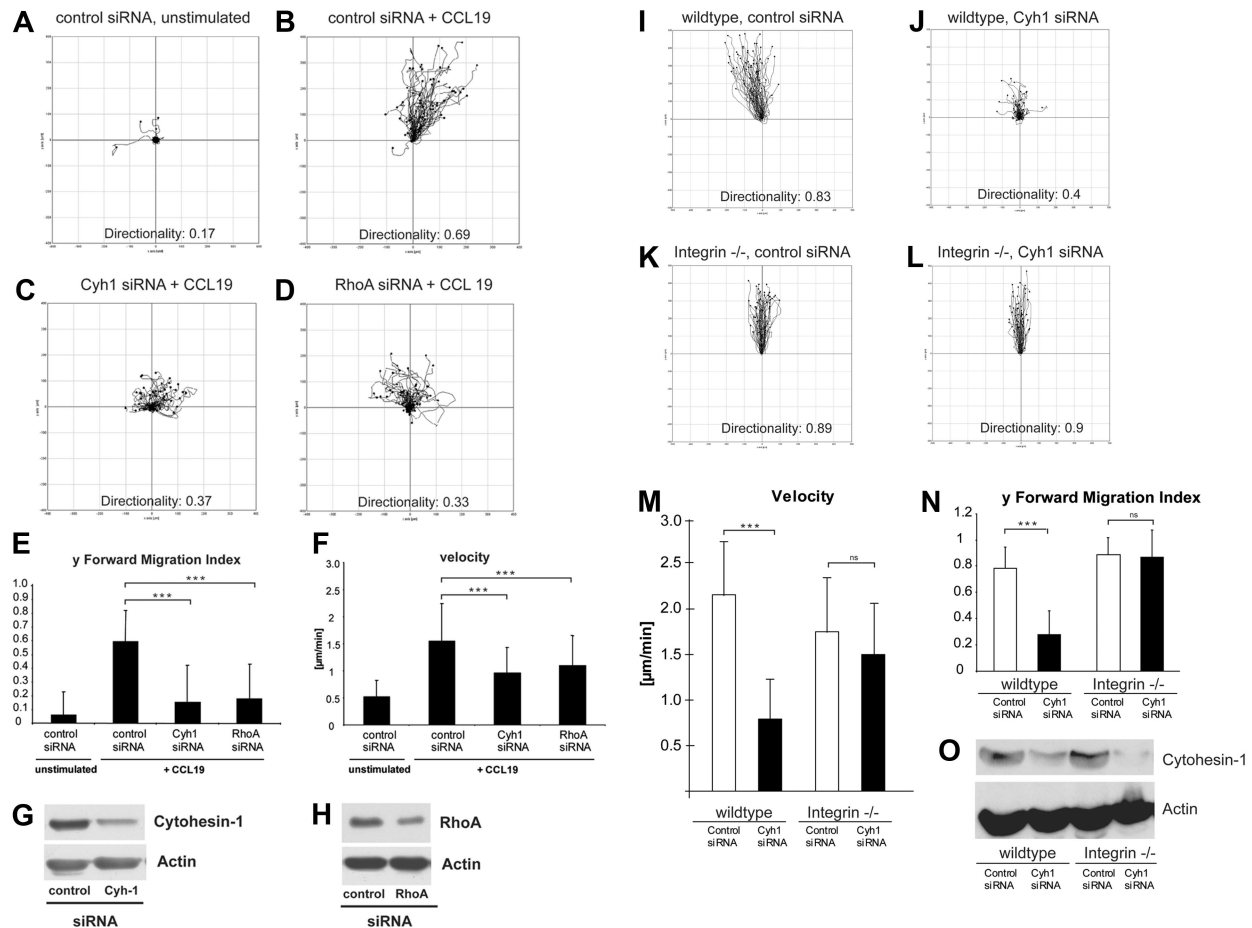


Figure 5. RNAi of cytohesin-1 strongly reduces chemotaxis of mature integrin(+/+) but not of integrin(-/-) BM-DCs in 3-dimensional collagen. Using 3D collagen type I gels, we analyzed the role of cytohesin-1 in BM-DC complex matrix migration. Quantification of time series after live cell imaging shows that directionality, y-forward migration index, and velocity of cytohesin-1 knockdown cells toward the chemokine CCL19 are strongly reduced (C,E,F). RNAi of RhoA results in a highly similar phenotype (D-F). Respective silencing efficiencies are shown by Western blot analysis (G,H). Quantification of 3D migration of integrin-deficient BM-DCs shows that directionality, y-forward migration index, and velocity of cytohesin-1 knockdown cells toward the chemokine CCL19 are strongly reduced in wild-type, that is, integrin(+/+) BM-DCs (J,M,N) but not in integrin(-/-) BM-DCs (L-N). Efficiency of cytohesin-1 RNAi was comparable in both cell types, as shown by Western blot analysis (O). For every 3D migration experiment, the tracks of 60 individual cells were monitored over a period of 3 hours by capturing digital images every 5 minutes. Cell directionality, forward migration index, and velocity were calculated and visualized as plots (A-D, I-L) and animated plots (Videos S1-S8) by analyzing the acquired data with the Chemotaxis and Migration tool plug-in (Ibidi) for ImageJ. Error bars indicate \pm SD. *** $P < .001$; ns indicates not significant. The data are representative for 3 independent experiments.

regulated by tight coordination of Rho GTPase activity.^{35,36} Because cytohesin-1 knockdown strongly impairs DC migration and activation of RhoA, we further analyzed whether the actin network in migrating DCs is affected by cytohesin-1. To this end, DCs were fixed by formaldehyde within 3D collagen gels and stained with Cy3-conjugated phalloidin. Mature DCs transfected with control siRNA show F-actin at the cell cortex and within membrane protrusions (Figure 7A). Mature DCs lack stress fibers, which is known to be characteristic for leukocytes.³⁷ The same, unaltered phenotype can be detected when cytohesin-1 expression in DCs is repressed by RNAi (Figure 7A). Thus, cytohesin-1 does apparently not affect polymerization and arrangement of actin fibers in membrane protrusions. Similarly, RNAi of RhoA does not influence the integrity of the peripheral actin network in mature DCs (Figure 7B), whereas, in contrast, RNAi of Rac1 completely inhibits generation of actin-based membrane protrusions (Figure 7C). These results are in good accordance with the recently published paper by Lämmermann et al,²⁵ who showed that RhoA does not regulate the front of migrating DCs, but that the small GTPase instead controls contractility of the cell body, which is important for flexible cell shape changes during DC migration in tissue environments.

Discussion

In this study, we have described an important role of the guanine nucleotide exchange factor cytohesin-1 in the regulation of cellular processes, as well as signaling pathways, which affect the regulation of adhesion and migration of human and murine dendritic cells. Overexpression analyses had implicated cytohesin-1 early on in the activation of the beta-2 integrin LFA-1 in T cells.¹⁹ We show here that cytohesin-1 expression in dendritic cells, which were derived from primary monocytes, is required for beta-2 integrin adhesion under static and under dynamic flow conditions, as well as for chemokine-dependent conformational activation of this adhesion receptor. Furthermore, we discovered an entirely novel function of cytohesin-1 in controlling interstitial migration of integrin-bearing DCs, but not of integrin-depleted DCs. The signaling pathway discriminating between the 2 conditions has been identified as well: cytohesin-1 acts upstream of the GTPase RhoA, and this novel signaling axis is required for integrin-mediated functions in the regulation of immune cell adhesion and migration.

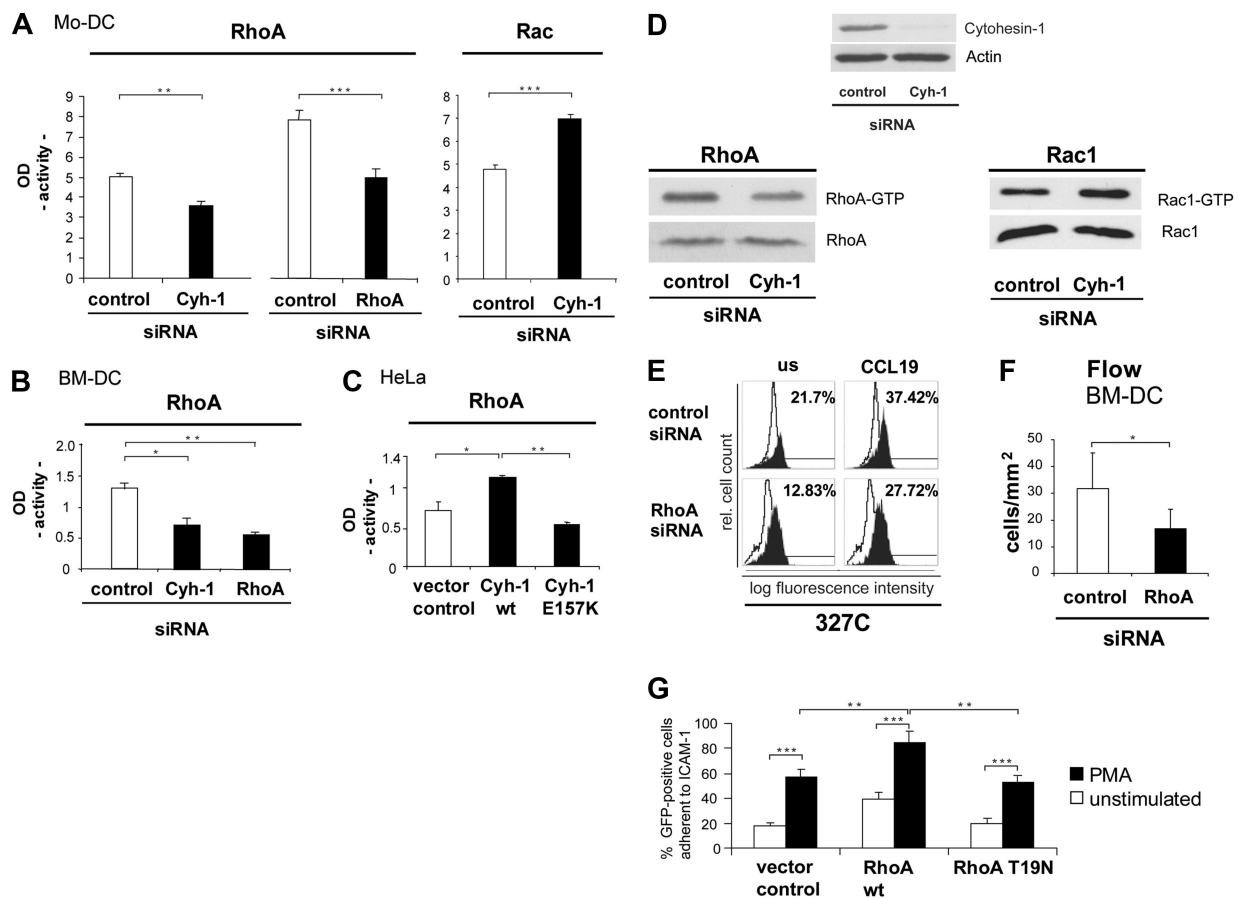


Figure 6. Cytohesin-1 controls the activity of RhoA in several cell types. RhoA is an important regulator of beta-2 integrin activation epitope expression and adhesion of DCs. Using a commercial enzyme linked assay (ELISA), we could show that in Mo-DCs the activity (GTP loading) of the small GTPase RhoA is strongly reduced in cytohesin-1 knockdown cells, compared with control cells. As a positive control for the assay, RhoA activity of RhoA knockdown Mo-DCs was analyzed, which also shows the expected reduction of RhoA activity. The same system was used to measure Rac activity, where RNAi of cytohesin-1 results in an increase of GTP loading (A). Consistently, RhoA activity of RhoA knockdown BM-DCs was also found reduced (B). Overexpression of wild-type cytohesin-1 in EGF-stimulated (50 nM, 1 hour) HeLa cells results in an increase of RhoA activity, compared with the vector control. In contrast, overexpression of the cytohesin-1 E157K mutant results in reduced RhoA activity (C). Cytohesin-1 knockdown down-regulates RhoA activity, but up-regulates GTP loading of Rac1 in EGF-stimulated HeLa (50 nM; 1 hour) cells, as detected by effector protein pull-down and Western blot analysis (panel D bottom panels). Knockdown of RhoA specifically reduces expression of the CD18 327C epitope in Mo-DCs (E) or adhesion under flow of BM-DCs to brain endothelioma (B end5) cells (F). Overexpression of wild-type RhoA but not of the GTP-loading mutant T19N increases static adhesion of GFP-cotransfected Mo-DCs to ICAM-1-Fc (G). In the static adhesion assay, Mo-DCs were stimulated with 50 ng/mL PMA for 60 minutes at 37°C (G). Error bars indicate \pm SD. *** $P < .001$, ** $P < .01$, * $P < .05$. Each experiment was repeated at least 3 times independently. Each single experiment was performed in duplicate.

Beta-2 integrin activation by chemokines is a complex process that involves the regulation of conformational equilibria of these proteins.²⁸ At least 3 conformations have been reported: resting (none or very little ligand binding), and intermediate and high affinity for ligand. Dendritic cells had been reported earlier to be much more restricted in beta-2 integrin activation than other leukocytes under certain conditions, but these studies had not used chemokine-dependent triggering or physiological flow conditions to test for integrin activation of DCs. Woolf et al³⁸ had shown that immobilized chemokines and flow forces are important requirements for the activation of circulating as well as migrating leukocytes, even if these cells migrate in lymph nodes. Our novel data reveal that integrins of mature DCs are capable of adhering to activated endothelia under physiological flow conditions in a cytohesin-1-dependent fashion. Apparently, beta-2 integrins of mature human dendritic cells respond normally to such stimuli, which is in full accordance with the aforementioned study.

The precise sequence of events of chemokine-mediated integrin activation during physiological responses is not known, but forces imposed by the blood flow apparently play an important role as well as protein binding events at the cytoplasmic tails of integrins. Cytohesin-1 had been shown to bind to the membrane-proximal

region of the beta-2 integrin cytoplasmic tail, and this is required for adhesion and activation epitope expression. Importantly, the ARF-GEF function is not required for integrin activation, but was found to be important for adhesion and spreading.²³ RhoA is a small GTPase that links integrin signaling to actin cytoskeletal rearrangements in many cell types. By use of blocking peptides, Giagulli et al¹⁵ had found that RhoA is required for leukocyte beta-2 integrin activation in endothelial arrest. These findings were recently confirmed and extended by Pasvolsky et al,³⁹ who showed that RhoA blocking peptides regulate intermediate affinity epitope expression of LFA-1 in T cells in flow. We now show that cytohesin-1 regulates the GTP loading of RhoA in leukocytes and we consistently report that both cytohesin-1 and RhoA mediate chemokine-induced activation epitope expression of CD18 in dendritic cells (Figure 2, Figure 6E, and data not shown). This places cytohesin-1 upstream of RhoA in beta-2 integrin activation. It is important to note here that this does not rule at all against an additional direct role of cytohesin-1/CD18 interactions in LFA-1 activation.²³ Taken together, our novel data reveal a signaling link between cytohesin-1 and RhoA in beta-2 integrin activation and adhesion of dendritic cells, which likely reflects a dual role of cytohesin-1 in integrin binding and in downstream signaling to RhoA.

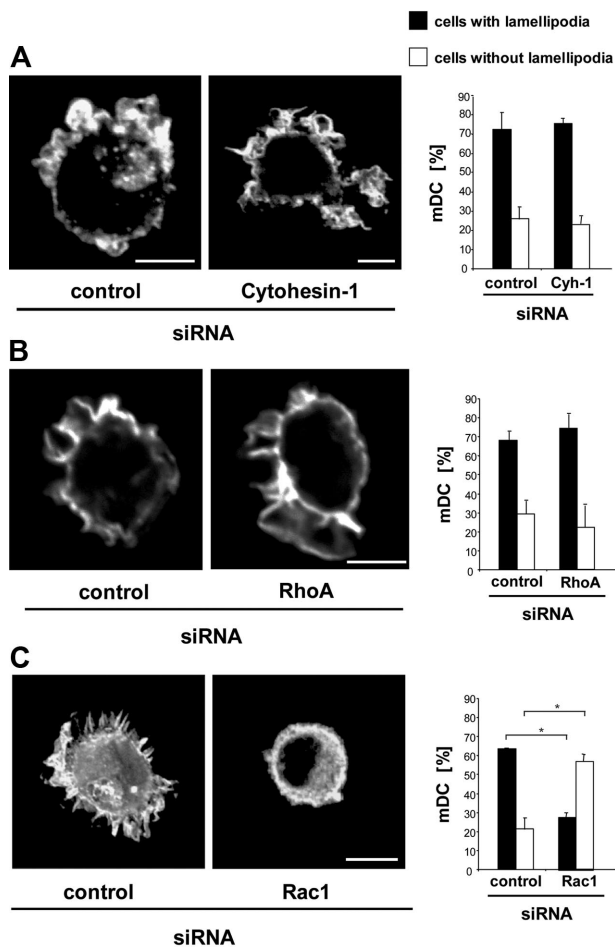


Figure 7. Cytohesin-1 is not involved in the formation of membrane protrusions. Microscopic visualization of F-actin by the use of Cy3-phalloidin shows that the formation of membrane protrusions in mature Mo-DCs is not influenced by RNAi of cytohesin-1, compared with control cells (A). RNAi of cytohesin-1, compared with control cells (A). RNAi of RhoA also shows the same unaltered phenotype (B), whereas RNAi of Rac1 results in strong deficiencies in the formation of membrane protrusions (C). Quantification of membrane protrusions shows that after RNAi of cytohesin-1 respective RhoA 70% of all mature Mo-DCs have lamellipodia and do not differ from control cells (A,B). In contrast, RNAi of Rac1 reduces formation of lamellipodia approximately 50% (C). Bars represent 5 μ m. In each individual experiment, 200 mature DCs were counted. Each single experiment was performed in duplicate. Error bars indicate \pm SD. * $P < .05$. Each experiment was repeated at least 3 times independently.

We show further and for the first time that cytohesin-1 is required for migration of BM-DCs to lymph nodes *in vivo* and for directional migration of BM-DCs in 3D collagen gels *in vitro*. RhoA had been implicated in both functions. In fact, RNAi of RhoA or cytohesin-1 resulted in a highly similar phenotype in interstitial migration of DCs in our assay systems. Specific treatment of BM-DCs with RhoA or cytohesin-1 siRNA yielded a strong abrogation of chemokine-gradient direction sensing and migratory persistence (track length). Most strikingly, however, our analyses show that 3D migration of leukocytes in the presence or absence of integrins is phenotypically similar but not identical. Whereas integrin-positive cells specifically require the cytohesin-1/RhoA signaling axis, integrin-null DCs require RhoA,²⁵ but not cytohesin-1 (shown in this study) for 3D motility. It thus becomes apparent that leukocyte migration in 3D environments may also be subclassified into different types or “modes,” reminiscent of what has been described for certain tumor cells.^{40,41} The fact that cytohesin-1 dependency discriminates between leukocytes migrating in the absence and presence of integrins raises the question how

the Rho pathway is regulated when both integrins and cytohesin-1 are absent and why this alternative pathway is not used when only cytohesin-1 is missing? One likely possibility is that in the absence of integrins, DCs use another migratory mode that does not depend on cytohesin as a Rho activator. This would imply that DCs can use phenotypically identical but molecularly distinct migratory strategies and suggest that there is more plasticity within the amoeboid migration mode than expected. In these experiments, we have used cells that lack all surface integrins to elucidate a principal role of cytohesin-1 in 3D immune cell migration. Consistent with all of our earlier findings, we furthermore show a strong functional association of this protein with beta-2 integrins. However, these data do not exclude a possible cooperation of cytohesin-1 with other integrins in these functions.

How are cytohesin-1 functions linked to RhoA signaling? Cytohesin-1 is a guanine nucleotide exchange factor for ARF-GTPases, and GDP/GTP exchange activities for other GTPases have never been reported. It is important to note here that a large body of work implicates cytohesin-2 (or ARNO) and -3 (Grp1) in ARF6 signaling upstream of Rac-1.⁴² Cytohesin-1 has never been implicated in direct activation of the endosomal and cytoskeletal signaling GTPase ARF6. In fact, data from our laboratory rule against such a link in T cells.⁴³ RhoA and Rac activity are tightly coordinated in cell migration and are often controlled in a reciprocal fashion.⁴⁴⁻⁴⁶ Interestingly, we observed not only a down-regulation of RhoA activity by cytohesin-1 knockdown but an up-regulation of Rac activity as well. This indicates that the control of Rho GTPase activities by cytohesins in immune and nonimmune cells might be aspects of a common theme, involving specific roles of cytohesin-1 in the RhoA pathway and of cytohesin-2/3 in Rac signaling.

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Authorship

Contribution: T.Q., B.T., C.S., J.G., N.C., K.D., and T.L. carried out the experiments; R.F., R.A., C.W., and M.S. contributed to research design and data analysis; J.G. and L.F. established study protocols; and W.K. designed the study, wrote the paper, and acquired research funding.

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