

**Biosynthesis of the P<sub>1</sub>, P<sup>k</sup>, P, and LKE antigens.** GLU indicates glucose; and CER, ceramide. Professional illustration by Marie Dauenheimer.

p phenotype), and in HeLa and Jurkat cells modified to alter levels of P<sup>k</sup> expression.<sup>5</sup>

The results were significant. P<sup>k</sup> cells were protected from R5 and X4 HIV-1 infection and had increased CD4, coreceptor, and P<sup>k</sup> expression, while PBMCs with the p phenotype had heightened susceptibility to R5 and X4 HIV-1 infection. Further supporting a role for P<sup>k</sup> was that P<sup>k</sup>-liposome fusion in Jurkat cells decreased susceptibility to HIV and cells transduced with P<sup>k</sup>-synthase (augmenting P<sup>k</sup> expression) showed decreased HIV-1 infect-

ability, while P<sup>k</sup> depletion or P<sup>k</sup>-synthase gene silencing promoted HIV infection.

Mechanistically, it may prove to be that the P<sup>k</sup> effect is due to altered lipid raft formation and/or HIV receptor or coreceptor localization, function, or accessibility. Alternatively or perhaps additionally, P<sup>k</sup> may be critical to viral internalization.

That P<sup>k</sup> and indeed other blood groups have a more defined role in host-microbial infection is an important new finding. Other examples include Fy<sup>a</sup> as the binding epitope

for plasmodium<sup>6</sup> and the P antigen as a receptor for the B19 parvovirus.<sup>7</sup> With this discovery, P<sup>k</sup> adds to our list of genetic polymorphisms that individually and/or in concert contribute to HIV resistance/susceptibility and potentially generates a site for possible future therapeutic use.<sup>8</sup>

*Conflict-of-interest disclosure:* The author declares no competing financial interests. ■

## REFERENCES

- Westhoff CM, Reid ME. ABO and Related Antigens and Antibodies. In: CD Hillyer, LE Silberstein, PM Ness, KC Anderson, JD Roback, eds. *Blood Banking and Transfusion Medicine*. 2nd Ed. Philadelphia, PA: Churchill Livingstone; 2006:p76.
- Lund N, Branch DR, Sakac D, et al. Lack of susceptibility of cells from patients with Fabry disease to productive infection with R5 human immunodeficiency virus. *AIDS*. 2005;19:1543-1546.
- Lund N, Branch DR, Mylvaganam MC, et al. A novel soluble mimic of the glycolipid, globotriaosyl ceramide inhibits HIV infection. *AIDS*. 2006;20:333-343.
- Fantini J, Tamalet C, Hammache D, et al. HIV-1-induced perturbations of glycosphingolipid metabolism are cell-specific and can be detected at early stages of HIV-1 infection. *J Acquir Immune Defic Syndr Hum Retrovirol*. 1998;19:221-229.
- Lund N, Olsson ML, Ramkumar S, et al. The human P<sup>k</sup> histo-blood group antigen provides protection against HIV-1 infection. *Blood*. 2009;113:4980-4991.
- Miller LH, Mason SJ, Clyde DF, et al. The resistance factor to Plasmodium vivax in blacks. The Duffy-blood-group genotype, FyFy. *N Engl J Med*. 1976;295:302-304.
- Brown KE, Anderson SM, Young NS. Erythrocyte P antigen: cellular receptor for B19 parvovirus. *Science*. 1993;262:114-117.
- Piacentini L, Biasin M, Fenizia C, et al. Genetic correlates of protection against HIV infection: the ally within. *J Intern Med*. 2009;265:110-124.

## ● ● ● VASCULAR BIOLOGY

Comment on Bieker et al, page 5019

# Flipping the wound that doesn't heal: the upside of coagulation in cancer

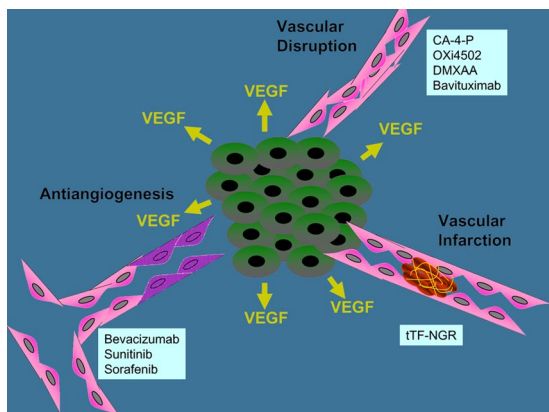
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Two mechanisms for targeting tumor vasculature are well recognized: antiangiogenesis and vascular disruption. In this issue of *Blood*, Bieker and colleagues take therapeutic advantage of coagulation and add a third mechanism, vascular infarction, to this group by demonstrating the effect through first-in-man administration of tTF-NGR.

**W**ith the approval of several antiangiogenic agents including bevacizumab, sunitinib, and sorafenib, the usefulness of targeting tumor vasculature with inhibitors has been clinically established. Antiangiogenic agents may be described as drugs that block angiogenic signaling either by neutralizing secreted proangiogenic factors or by inhibiting

intracellular signaling by their receptors resulting in regression of neovasculature in the tumor.<sup>1</sup> A second approach to targeting the tumor vasculature therapeutically is vasculature disrupting agents. Vasculature disrupting agents are designed to produce rapid vascular collapse or shutdown in tumors.<sup>2,3</sup> The resulting ischemia results in tumor cell killing,

which causes extensive tumor necrosis. Although no vascular disrupting agents are approved as yet, several including combretastatin derivatives (CA4P and OXi4503), the dimethylxanthone DMXAA, and the antiphosphatidylserine bavituximab are progressing through clinical trial. In preclinical in vivo models, decreased tumor blood flow can



**Three mechanisms for therapeutically targeting tumor vasculature, antiangiogenesis, vascular disruption, and vascular infarction, and examples of agents.**

be detected in a few minutes after administration of these agents, and nearly complete vascular shutdown can occur in less than 1 hour.

While the recognition that tumor growth is angiogenesis-dependent and that angiogenesis could be an important target for cancer therapeutics is attributed to Folkman in 1971, and the recognition that tumors can be described as wounds that do not heal is attributed to Dvorak in 1986, the observation that tumors are hypercoagulable occurred much earlier, being attributed to Trousseau in 1865. The association of thrombosis in the extremities (thrombophlebitis) with a diagnosis of pancreatic or gastric cancer has historically been termed Trousseau syndrome. Growing tumors often develop a fibrin “capsule.” The fibrin clots attract host cells including platelets, monocytes/macrophages, and infiltrating lymphocytes, leading to continued matrix formation and formation of new blood vessels. These events mimic the early stages of wound

healing which, in malignant diseases, becomes a continuous non-resolving process. Many malignant tissues express tissue factor, a pro-coagulant and key activator of the coagulation cascade. Disseminated intravascular coagulation is the most frequent coagulopathy associated with cancer and results from generalized activation of coagulation pathways. Cancer is the third most common cause of disseminated intravascular coagulation.<sup>4</sup>

Modern technologies have been applied to understanding the connection between cancer and

wound healing. In 2004, Chang et al developed a wound-response gene signature including genes involved in matrix remodeling, cell motility, and angiogenesis.<sup>5</sup> Expression of the wound-response gene signature predicted poor overall survival and increased risk of metastasis in a subset of patients with breast, lung, and gastric cancers. Later, the wound-response gene signature’s prognostic value was validated in a data set of nearly 300 breast cancer patients.

In the current issue, Bieker et al take advantage of the coagulation cascade and offer a third approach to therapeutically targeting the tumor vascular supply by selectively focusing thrombosis in the tumor blood vessels.<sup>6</sup> These investigators fused the extracellular domain of (truncated) tissue factor (tTF) with the peptide GNGRAHA, which was identified through phage display to bind to aminopeptidase N (CD13) and integrin  $\alpha v \beta 3$  (CD51/CD61) which are cell surface protein up-

regulated on tumor vascular endothelial cells. The fusion protein therapeutic—designated tTF-NGR produces thrombosis in the tumor vasculature, resulting in infarction of tumor vessels.<sup>6</sup> Previous studies have tried other tumor-targeting carrier molecules, such as the ED-B domain of fibronectin, PSA, VCAM-1 and integrins, to deliver tissue factor to tumor vasculature.<sup>7</sup> Bieker et al found that the NGR peptide resulted in rapid homing to tumor vessel and rapid thrombosis. Studies in 3 human tumor xenograft tumors confirmed the efficacy of tTF-NGR. Furthermore, clinical first-in-man treatment of 5 patients with low doses of systemically administered tTF-NGR showed decreased tumor perfusion by MRI.

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## REFERENCES

- Jubb AM, Oates AJ, Holden S, Koeppen H. Predicting benefit from anti-angiogenic agents in malignancy. *Nat Rev Cancer*. 2006;6:626-635.
- Tozer GM, Kanthou C, Baguley BC. Disrupting tumor blood vessels. *Nat Rev Cancer*. 2005;5:423-435.
- Ran S, He J, Huang X, Soares M, Scothorn D, Thorpe PE. Antitumor effects of a monoclonal antibody that binds anionic phospholipids on the surface of tumor blood vessels in mice. *Clin Cancer Res*. 2005;11:1551-1562.
- Belt RJ, Leite C, Haas CD, Stephens RL. Incidence of hemorrhagic complications in patients with cancer. *JAMA*. 1978;239:2571-2574.
- Chang HY, Nuyten DSA, Sneddon JB, et al. Robustness, scalability, and integration of a wound-response gene expression signature in predicting breast cancer survival. *Proc Natl Acad Sci U S A*. 2005;102:3738-3743.
- Bieker R, Kessler T, Schwöppe C, et al. Infarction of tumor vessels by NGR-peptide-directed targeting of tissue factor: experimental results and first-in-man experience. *Blood*. 2009;113:5019-5027.
- Kessler T, Bieker R, Padró T, et al. Inhibition of tumor growth by RGD peptide-directed delivery of truncated tissue factor to the tumor vasculature. *Clin Cancer Res*. 2005;11:6317-6324.