with at least folic acid, riboflavin, pyridoxin, and cyanocobalamin in proper dosing.

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Conflict-of-interest disclosure: The author declares no competing financial interests.

#### References

- den Heijer M, Willems HP, Blom HJ, et al. Homocysteine lowering by B vitamins and the secondary prevention of deep vein thrombosis and pulmonary embolism: a randomized, placebo-controlled, double-blind trial. Blood. 2007;109: 139-144.
- Scott JM. Genetic diversity and disease: opportunities and challenge. Proc Natl Acad Sci U S A. 2001;98:14754-14756.

## Response:

- Yamada K, Chen Z, Rozen R, Matthews RG. Effects of common polymorphisms on the properties of recombinant human methylenetetrahydrofolate reductase. Proc Natl Acad Sci U S A. 2001;98:14853-14858.
- Ames BN, Elson-Schwab I, Silver EA. High-dose vitamin therapy stimulates variant enzymes with decreased coenzyme binding affinity (increased K(m)): relevance to genetic disease and polymorphisms. Am J Clin Nutr. 2002;75:616-658.
- Sibani S, Leclerc D, Weisberg IS, et al. Characterization of mutations in severe methylenetetrahydrofolate reductase deficiency reveals an FAD-responsive mutation. Hum Mutat. 2003;21:509-520.
- Rozen R. Genetic modulation of homocysteinemia. Semin Thromb Hemost. 2000;26:255-261.
- McKinley MC, McNulty H, McPartlin J, Strain JJ, Scott JM. Effect of riboflavin supplementation on plasma homocysteine in elderly people with low riboflavin status. Eur J Clin Nutr. 2002;56:850-856.
- Rozen,R. Methylenetetrahydrofolate reductase: a link between folate and riboflavin? Am J Clin Nutr. 2002;76:301-302.
- McNulty H, Dowey LE, RC, Strain JJ, et al. Riboflavin lowers homocysteine in individuals homozygous for the MTHFR 677C → T polymorphism. Circulation. 2006;113:74-80.

### Homocysteine lowering and recurrent venous thrombosis: the VITRO trial

We welcome the opportunity to respond to the comments of Tsamaloukas and the comments of Marcucci and colleagues. The primary aim of our study was to investigate the effect of vitamin supplementation on the incidence of recurrent venous thrombosis. Because factor V Leiden and prothrombin G20210A are not important risk factors for recurrent events, we did not control for these genetic factors.<sup>1</sup> The same holds true for the *MTHFR* C677T polymorphism. which is not thought to influence recurrence risk independent of homocysteine levels. Moreover, in a randomized trial one assumes that confounding by extraneous risk factors is controlled by the randomization procedure. Nonetheless, we have measured these 3 polymorphisms<sup>2</sup>; their distribution is listed in Table1. Adjustment for these polymorphisms did not change the hazard ratios for treatment effect substantially.

The prevalence of hormonal therapy use at time of the first event in hyperhomocysteinemic patients was 14% in the vitamin group and 15% in the placebo group; in the normohomocysteinemic group, it was 19% in the vitamin group and 12% in the placebo group. Adjustment for hormonal use did not change the hazard ratio for treatment effect substantially. We have no data on renal function (creatinine concentration), but there is no reason to assume any bias because of the randomized nature of the study.

We agree that the homocysteine levels in Table 1 of our paper might be somewhat puzzling. The explanation for this is that the distinction between the hyperhomocysteinemic and normohomocysteinemic groups was made on the basis of homocysteine measurements in acidic citrate tubes as stated in "Patients, materials, and methods." If people met the other inclusion criteria, they were randomized to vitamin and placebo. Just before entering the study, another blood sample was taken in EDTA, which was used for the baseline value in the table. Because of variation in homocysteine levels, some patients in the hyperhomocysteinemic group had homocysteine levels lower than the cutoff level, and some patients in the normohomocysteinemic group had values higher than the cutoff level. In general, there is a clear contrast in homocysteine level between the hyper- and normohomocysteinemic groups (15.5 µM vs 9 µM). As for a trial in intermediate hyperhomocysteinemia, when we stratified patients in quartiles based on the homocysteine levels at baseline (just before they start using the study medication), there was no stronger effect in the highest quartile (with homocysteine levels between 14.7 and 108 µM).

Regarding the study medication, we intended to achieve a maximal homocysteine-lowering effect. The choice for the combination of folate (5 mg), vitamin  $B_{12}$  (0.4 mg), and vitamin  $B_6$ 

Table 1. Distribution of	polymor	phisms in	MTHFR. F	FII G20210A.	and factor V	Leiden in the	VITRO study
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Polymorphism	Hyperhomocysteinen	nic group; n = 360	Normohomocysteinemic group; n = 341		
	Multivitamin; n = 177	Placebo; n = 183	Multivitamin; n = 176	Placebo; n = 165	
MTHFR					
cc, no. (%)	68 (40)	80 (47)	92 (52)	80 (48)	
ct, no. (%)	74 (44)	70 (41)	75 (43)	65 (39)	
tt, no. (%)	28 (16)	22 (13)	9 (5)	20 (12)	
Total no.	170	172	176	165	
FII					
gg, no. (%)	161 (96)	160 (95)	166 (96)	155 (95)	
ga, no. (%)	7 (4)	9 (5)	7 (4)	9 (5)	
Total no.	168	169	173	164	
Factor V Leiden					
gg, no. (%)	128 (81)	139 (87)	144 (84)	135 (84)	
ga, no. (%)	30 (19)	19 (12)	28 (16)	23 (14)	
aa, no. (%)	0 (0)	2 (1)	0 (0)	3 (2)	
Total no.	158	160	172	161	

(50 mg) was based on an earlier study<sup>3</sup> in which this regimen showed the strongest effect on homocysteine levels; this dose is relatively high compared with several other homocysteinelowering trials. We are aware that vitamin  $B_2$  is a coenzyme of *MTHFR*, and that riboflavin supplementation lowers homocysteine levels in vitamin  $B_2$ -deficient patients.<sup>4</sup> However, no data are available on the additional effect of riboflavin next to the combination of folate, vitamin  $B_{12}$ , and vitamin  $B_6$ . Only 1 study shows a small additional effect of riboflavin next to folate alone.<sup>5</sup>

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Conflict-of-interest disclosure: The authors declare no competing financial interests.

# To the editor:

## References

- Christiansen SC, Cannegieter SC, Koster T, Vandenbroucke JP, Rosendaal FR. Thrombophilia, clinical factors, and recurrent venous thrombotic events. JAMA. 2005;293:2352-2361.
- Keijzer MB, Borm GF, Blom HJ, Bos GM, Rosendaal FR, den Heijer M. No interaction between factor V Leiden and hyperhomocysteinemia or MTHFR 677TT genotype in venous thrombosis: results of a meta-analysis of published studies and a large case-only study. Thromb Haemost. 2007;97:32-37.
- den Heijer M, Brouwer IA, Bos GM, et al. Vitamin supplementation reduces blood homocysteine levels: a controlled trial in patients with venous thrombosis and healthy volunteers. Arterioscler Thromb Vasc Biol. 1998;18:356-361.
- 4. McNulty H, Dowey le RC, Strain JJ, et al. Riboflavin lowers homocysteine in individuals homozygous for the MTHFR 677C  $\rightarrow$  T polymorphism. Circulation. 2006;113:74-80.
- Chuang CZ, Boyles A, Legardeur B, Su J, Japa S, Lopez-S A. Effects of riboflavin and folic acid supplementation on plasma homocysteine levels in healthy subjects. Am J Med Sci. 2006;331:65-71.

## RAG1 and BRCA2 polymorphisms in non-Hodgkin lymphoma

A recent *Blood* paper by Hill et al<sup>1</sup> suggested a link between the risk of developing non-Hodgkin lymphoma (NHL) and polymorphic variations in several genes involved in DNA strand break induction and DNA repair pathways. We examined 2 of these polymorphisms where Hill et al reported statistically significant associations, in the Recombination-activating 1 (*RAG1*) gene (K820R; odds ratio [OR] = 2.7 [AA vs GG], 95% confidence interval [CI], 1.4-5.0) and the *Breast cancer 2 (BRCA2)* gene (N372H; OR = 1.5 [AA vs CC], 95% CI, 1.0-2.1), in a United Kingdom case-control study of lymphoma.

Local ethical approval was obtained for this research. Informed consent was obtained in accordance with the Declaration of Helsinki.

A total of 741 white patients with NHL and 806 white control participants randomly selected from population registers were recruited as part of a lymphoma case-control study conducted in parts of north and southwest England.<sup>2</sup> Our study participants consented to give a blood sample, and represented 67% of patients

with incident NHL aged 16 to 69 years in the study area and 66% of the control participants who were contacted successfully. We genotyped the participants' DNA while blinded to case-control status using TaqMan SNP genotyping assays (Applied Biosystems, Warrington, United Kingdom) and ABI PRISM 7000 software (Applied Biosystems). To assess the reliability of the Taqman assays, 20 samples were randomly selected for direct DNA sequencing: concordance with TaqMan was 100% for both polymorphisms. Among our control population, RAG1 K820R and BRCA2 N372H were in Hardy-Weinberg equilibrium, and the distributions of variants were consistent with those reported by Hill et al and the cancer genome anatomy project SNP500 cancer database.<sup>3</sup> Unmatched statistical analyses to estimate ORs and 95% CIs were conducted using unconditional logistic regression adjusting for age and sex,<sup>4</sup> and the likelihood ratio test was used to test for interactions.

Table 1. Number of patients and control participation	ts, adjusted ORs, and 95% CIs for BRCA2 N372H	and RAG1 K820R by immunophenotype
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		NHL		B-cell lymphoma		T-cell lymphoma	
Genes and variants	Controls, no. (%)	Patients, no. (%)	OR (95% CI)	Patients, no. (%)	OR (95% CI)	Patients, no. (%)	OR (95% CI)
Total	806 (100)	741 (100)	NA	665 (100)	NA	59 (100)	NA
BRCA2 N372H, A > C*							
AA	387 (51.1)	375 (55.5)	1 (reference)	340 (56.4)	1 (reference)	28 (48.3)	1 (reference)
AC	307 (40.6)	253 (37.4)	0.85 (0.68-1.05)	224 (37.1)	0.83 (0.66-1.03)	22 (37.9)	1.03 (0.58-1.85)
CC	63 (8.3)	48 (7.1)	0.79 (0.53-1.18)	39 (6.5)	0.71 (0.46-1.09)	8 (13.8)	1.61 (0.70-3.75)
AC + CC	370 (48.9)	301 (44.5)	0.84 (0.68-1.03)	263 (43.6)	0.81 (0.65-1.00)	30 (51.7)	1.14 (0.67-1.96)
RAG1 K820R, A > G*							
AA	622 (81.4)	557 (81.2)	1 (reference)	496 (81.3)	1 (reference)	46 (78.0)	1 (reference)
AG	136 (17.8)	124 (18.1)	1.01 (0.77-1.33)	109 (17.9)	1.00 (0.76-1.32)	13 (22.0)	1.38 (0.72-2.65)
GG	6 (0.8)	5 (0.7)	0.94 (0.29-3.10)	5 (0.8)	1.07 (0.32-3.52)	0 (0.0)	0 (0-8.84)†
AG + GG	142 (18.6)	129 (18.8)	1.01 (0.78-1.33)	114 (18.7)	1.00 (0.76-1.32)	13 (22.0)	1.31 (0.69-2.51)

B-cell lymphoma includes International Classification of Diseases for Oncology version 3 codes 9673/3, 9679/3, 9680/3, 9684/3, 9689/3, 9699/3, 9691/3, 9695/3, 9698/3, and 9699/3; T-cell lymphoma includes 9700/3, 9701/3, 9702/3, 9705/3, 9709/3, 9709/3, 9714/3, 9716/3, 9717/3, 9719/3, and 9827/3.

Immunophenotype was not known for 17 patients. ORs and 95% CIs were calculated using unconditional logistic regression adjusting for age and sex.

NA indicates not applicable.

\*BRCA2 N372H and RAG1 K820R were in Hardy-Weinberg equilibrium among controls ( $\chi^2 = 0.04$ , P = 0.85;  $\chi^2 = 0.23$ , P = 0.63, respectively). Samples did not amplify for 65 patients and 49 control participants for BRCA2 N372H, and 55 patients and 42 control participants for RAG1 K820R.

†OR and 95% CI were calculated using exact methods.