

Brief report

Acute myeloid leukemia bearing cytoplasmic nucleophosmin (NPMc⁺ AML) shows a distinct gene expression profile characterized by up-regulation of genes involved in stem-cell maintenance

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Approximately one third of acute myeloid leukemias (AMLs) are characterized by aberrant cytoplasmic localization of nucleophosmin (NPMc⁺ AML), consequent to mutations in the NPM putative nucleolar localization signal. These events are mutually exclusive with the major AML-associated chromosomal rearrangements, and are frequently associated with normal karyotype, *FLT3* mutations, and multilineage involvement. We report the

gene expression profiles of 78 de novo AMLs (72 with normal karyotype; 6 without major chromosomal abnormalities) that were characterized for the subcellular localization and mutation status of NPM. Unsupervised clustering clearly separated NPMc⁺ from NPMc⁻ AMLs, regardless of the presence of *FLT3* mutations or non-major chromosomal rearrangements, supporting the concept that NPMc⁺ AML represents a distinct entity.

The molecular signature of NPMc⁺ AML includes up-regulation of several genes putatively involved in the maintenance of a stem-cell phenotype, suggesting that NPMc⁺ AML may derive from a multipotent hematopoietic progenitor. (*Blood*. 2005;106:899-902)

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Introduction

The most frequent chromosomal rearrangements in acute myeloid leukemias (AMLs) are t(8;21), t(15;17), inv(16), and t(9;11), which, with their variants, account for approximately 40% of cases.¹ The resulting fusion genes encode for oncogenic proteins capable of initiating leukemia in mice.² Many other chromosomal abnormalities have been described, representing, however, less than 10% of AMLs. The remaining 50% of cases carry a normal karyotype or, less frequently, random chromosomal aberrations, and the underlying genetic lesion is unknown.³

A recent survey of nucleophosmin (NPM) subcellular localization in a large series of de novo AMLs revealed aberrant cytoplasmic NPM localization (NPMc⁺) in about 35% of cases.⁴ Analysis of the *NPM* gene identified mutations within the putative NPM nucleolar localization signal in all NPMc⁺ AMLs.⁴ These mutations cause cytoplasmic localization of the abnormal protein⁴ and, being mutually exclusive with recurrent AML-associated chromosomal abnormalities, are likely to play a key role in leukemogenesis. NPMc⁺ AML encompasses a wide French-American-British (FAB) morphologic spectrum, frequently displays multilineage involvement, and usually shows normal karyotype and *FLT3* mutations. We studied the global expression profiles

of de novo AMLs without major chromosomal translocations and here demonstrate that NPMc⁺ AMLs display a specific gene expression profile dominated by a stem-cell molecular signature.

Study design

Tumor samples

We studied 78 patients with de novo AMLs (age, 15-60 years, other than M3) from the GIMEMA (Gruppo Italiano Malattie Ematologiche Maligne dell'Adulto) LAM 99P and GIMEMA/EORTC (European Organization for Research on Treatment of Cancer) AML12 trials, showing greater than 70% bone marrow infiltration by leukemic cells, previously characterized for subcellular NPM localization, karyotype, reverse transcription-polymerase chain reaction (RT-PCR) for major fusion transcripts, *MLL* status, *FLT3* mutations.⁴ NPM subcellular localization was detected in bone marrow paraffin sections⁴ using specific anti-NPM monoclonal antibodies^{5,6} and the alkaline phosphatase anti-alkaline phosphatase technique.⁷

NPM mutations were previously reported in 24 of 78 of cases.⁴ Additional mutational analysis of *NPM* transcript was performed by PCR amplification of cDNA with the following primers: 5'-region, Fw1,

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A complete list of the members of the GIMEMA (Gruppo Italiano Malattie Ematologiche Maligne dell'Adulto) Acute Leukemia Working Party appears

in the "Appendix."

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The online version of the article contains a data supplement.

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Table 1. Main features of the AML cases used for analysis of gene expression profiles

Patient no.	NPM loc	NPM gene	FAB	FLT3	Karyotype	CD34
Training set						
1	NPM c ⁺	Mut	M1	ITD	NK	Neg
2	NPM c ⁺	Mut	M1	ITD	NK	Neg
3	NPM c ⁺	Mut	M1	ITD	NK	Neg
4	NPM c ⁺	Mut	M1	ITD	NK	Neg
5	NPM c ⁺	Mut	M1	ITD	NK	Neg
6	NPM c ⁺	Mut	M1	ITD	NK	Neg
7	NPM c ⁺	Mut	M1	WT	NK	Neg
8	NPM c ⁺	Mut	M1	WT	NK	Neg
9	NPM c ⁺	Mut	M1	WT	NK	Neg
10	NPM c ⁺	Mut	M2	ITD	NK	Pos
11	NPM c ⁺	Mut	M2	ITD	NK	Neg
12	NPM c ⁺	Mut	M2	ITD	NK	Neg
13	NPM c ⁺	Mut	M2	ITD	NK	Neg
14	NPM c ⁺	Mut	M2	ITD	NK	Neg
15	NPM c ⁺	Mut	M2	ITD	NK	Neg
16	NPM c ⁺	Mut	M2	WT	NK	Neg
17	NPM c ⁺	Mut	M2	WT	NK	Neg
18	NPM c ⁺	Mut	M2	WT	NK	Neg
19	NPM c ⁺	Mut	M2	WT	NK	Pos
20	NPM c ⁺	Mut	M4	ITD	NK	Neg
21	NPM c ⁺	Mut	M4	ITD	NK	Neg
22	NPM c ⁺	Mut	M4	WT	NK	Neg
23	NPM c ⁺	Mut	M4	WT	NK	Neg
24	NPM c ⁺	Mut	M5a	WT	NK	Neg
25	NPM c ⁺	Mut	M5a	WT	NK	Neg
26	NPM c ⁺	Mut	M5a	WT	NK	Neg
27	NPM c ⁺	Mut	M5b	D835	NK	Neg
28	NPM c ⁺	Mut	M5b	D835	NK	Neg
29	NPM c ⁺	Mut	M5b	WT	NK	Neg
30	NPM c ⁻	WT	M0	ITD	NK	Pos
31	NPM c ⁻	WT	M0	WT	NK	Pos
32	NPM c ⁻	WT	M1	WT	NK	Pos
33	NPM c ⁻	WT	M1	WT	NK	Pos
34	NPM c ⁻	WT	M1	WT	NK	Pos
35	NPM c ⁻	WT	M2	WT	NK	Pos
36	NPM c ⁻	WT	M4	WT	NK	Pos
37	NPM c ⁻	WT	M4	WT	NK	Pos
38	NPM c ⁻	WT	M5a	ITD	NK	Neg
39	NPM c ⁻	WT	M5a	ITD	NK	Neg
Test set						
40	NPM c ⁺	Mut	M1	ITD	NK	Neg
41	NPM c ⁺	Mut	M1	WT	NK	Neg
42	NPM c ⁺	Mut	M2	D835	NK	Neg
43	NPM c ⁺	Mut	M2	ITD	NK	Pos
44	NPM c ⁺	Mut	M2	WT	NK	Neg
45	NPM c ⁺	Mut	M2	WT	NK	Neg
46	NPM c ⁺	Mut	M4	D835	NK	Neg
47	NPM c ⁺	Mut	M4	ITD	NK	Neg
48	NPM c ⁺	Mut	M4	ITD	NK	Neg
49	NPM c ⁺	Mut	M4	ITD	NK	Neg
50	NPM c ⁺	Mut	M4	ITD/D835	NK	ND
51	NPM c ⁺	Mut	M4	WT	NK	Neg
52	NPM c ⁺	Mut	M5	ITD	NK	Neg
53	NPM c ⁺	Mut	M5a	ITD	NK	Neg
54	NPM c ⁺	Mut	M5a	ITD	NK	Neg
55	NPM c ⁺	Mut	M5a	WT	NK	Neg
56	NPM c ⁺	Mut	M5b	ITD	NK	Neg
57	NPM c ⁺	Mut	M5b	ITD	NK	Neg
58	NPM c ⁺	Mut	M5b	WT	NK	Neg
59	NPM c ⁺	Mut	M5b	WT	NK	Neg
60	NPM c ⁺	Mut	M5b	WT	NK	Neg
61	NPM c ⁺	Mut	M5b	WT	NK	Neg
62	NPM c ⁺	Mut	M0	WT	del(11)	Pos
63	NPM c ⁺	Mut	M1	WT	add(1)	Neg

Table 1. Main features of the AML cases used for analysis of gene expression profiles (continued)

Patient no.	NPM loc	NPM gene	FAB	FLT3	Karyotype	CD34
64	NPM c ⁺	Mut	M4	ITD	inv(3)	Neg
65	NPM c ⁺	Mut	M4	WT	tri(8)	Pos
66	NPM c ⁺	Mut	M5b	ITD	del(9)	Neg
67	NPM c ⁺	Mut	M5b	ITD/D835	tri(8)	Neg
68	NPM c ⁺	Mut	M0	WT	NK	Pos
69	NPM c ⁻	WT	M1	ITD	NK	Neg
70	NPM c ⁻	WT	M1	WT	NK	Pos
71	NPM c ⁻	WT	M2	ITD	NK	Pos
72	NPM c ⁻	WT	M2	WT	NK	Pos
73	NPM c ⁻	WT	M2	WT	NK	Pos
74	NPM c ⁻	WT	M4	ITD	NK	Pos
75	NPM c ⁻	WT	M4	ITD	NK	Pos
76	NPM c ⁻	WT	M4	ITD	NK	Pos
77	NPM c ⁻	WT	M4	WT	NK	Pos
78	NPM c ⁻	WT	M4e	WT	NK	Pos

NPM loc indicates NPM localization (c⁺, cytoplasmic and nuclear; c⁻, nuclear); NPM gene, status of the NPM gene (Mut, mutated; WT, wild type); FAB, M0 to M5 subtypes; FLT3, status of the FLT3 gene (ITD, internal tandem duplication; D835 mutation; WT, wild type); NK, normal karyotype; CD34, immunohistochemical expression of CD34 (Pos, 20% positive blasts; Neg, < 20% positive blasts; ND, no data available).

5'-GGTTGTTCTCTGGAGCAGCGTTC-3', and Rev1, 5'-GGAGTATCTCG-TATAGATTCTTCAC-3'; 3'-region, Fw2, 5'-GGAGGAGGATGTGAA-ACTCTTAAG-3', and Rev2, 5'-ACTGCCAGATATCAACTGTTACAG-3'.

Microarray analysis and real-time RT-PCR

Microarray and RT-PCR methods and data analysis are described in detail in Document S1 (available at the *Blood* website; see the Supplemental Materials link at the top of the online article). Briefly, Affymetrix HG-U133A chips were hybridized with labeled targets obtained from 2 to 5 µg total RNA as described.⁸ Data were analyzed with MASv5 (Affymetrix, Santa Clara, CA), and further elaborated with GeneSpring 6.1 (Silicon Genetics, Redwood City, CA) or Significance Analysis of Microarrays (SAM).⁹ RT-PCR was performed using TaqMan GeneExpression Assays (<http://myscience.appliedbiosystems.com>).

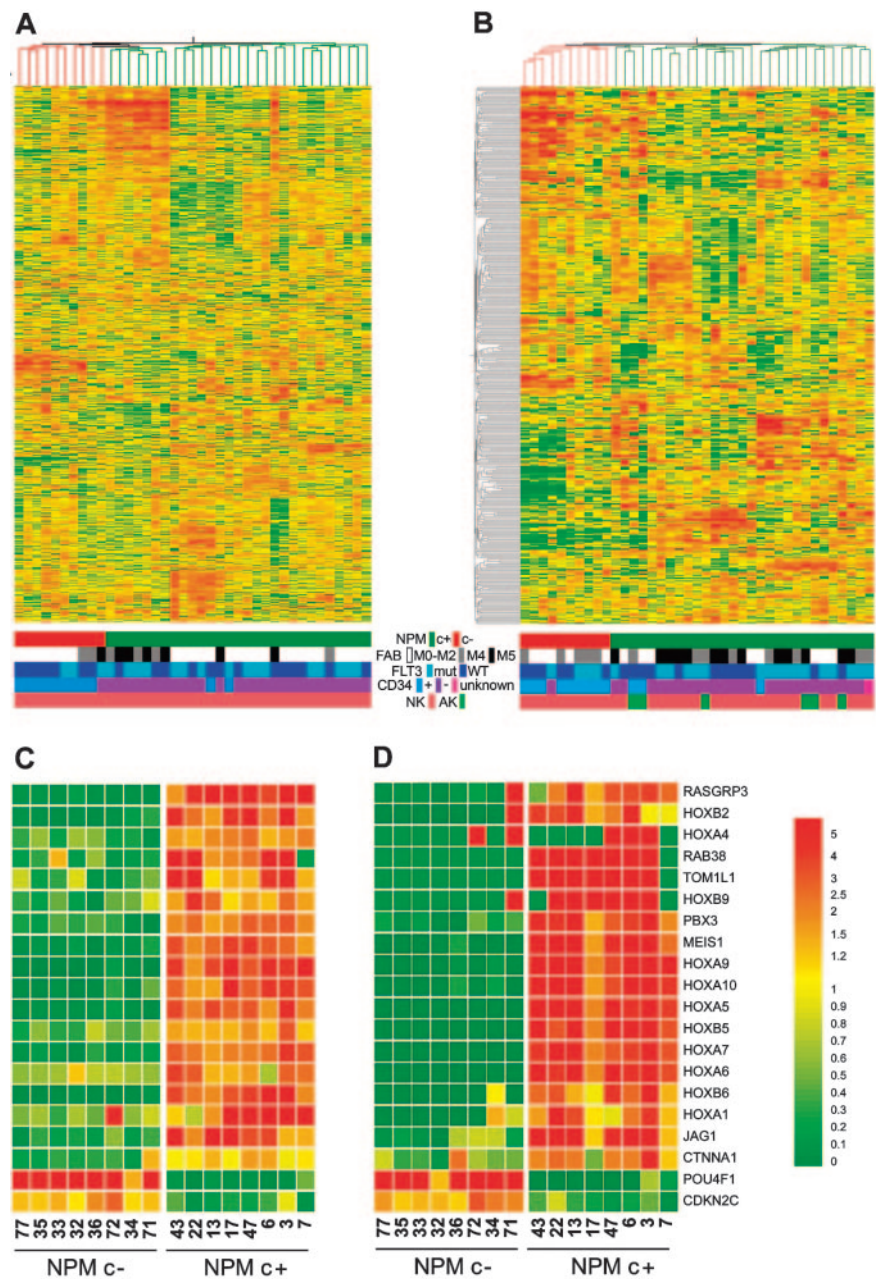
Results and discussion

We studied 78 patients with de novo AMLs (58 NPMc⁺ and 20 NPMc⁻), negative for AML-associated chromosomal translocations at cytogenetic and/or molecular level (Table S2). NPMc⁺ cases were representative of the 3 major genetic features of this novel AML entity⁴: prevalence of normal karyotype (52 of 58), frequent occurrence of FLT3 mutations (32 of 58), and presence in all cases of NPM mutations (Table 1). As reported,⁴ the 20 NPMc⁻ AMLs with normal karyotype included in this study had a lower frequency of FLT3 mutations (8 of 20), and none harbored NPM mutations. Complete description of cytogenetic, molecular, and FAB characteristics is shown in Supplemental Table S2.

To investigate whether NPMc⁺ AMLs are associated with a specific pattern of gene expression, we divided the 78 AML samples into 2 groups and analyzed them on Affymetrix HG-U133A chips. The first group (training set) was used to identify genes that function as putative predictors of NPM status, whereas the second group (test set) was used to assess the validity of the identified predictors. Details of microarray methods, data analysis, and statistical tests are described in Supplemental Document S1.

The training set included 39 AMLs with normal karyotype, differing for NPM and FLT3 status, and FAB subtype (Table 1). Unsupervised

Figure 1. Gene-expression profiles of NPMc⁺ and NPMc⁻ AML. (A) Unsupervised hierarchical clustering of the training set. The dendrogram at the top was obtained using a list of 7197 selected genes (see Document S1, available on the *Blood* website; see the Supplemental Materials link at the top of the online article). The strongest parameter in determining AML clustering is NPM localization. Each of the 39 columns represents an AML sample, and each of the 7197 rows represents a gene (probe set). Genes were clustered according to Pearson correlation (the structure of the gene tree is not shown). (B) Hierarchical clustering of the test set using 369 probe sets obtained from an analysis of variance of the training set (see Document S1). The predictor genes efficiently discriminate AML cases according to NPM localization. Each of the 39 columns represents an AML sample, and each of the 369 rows represents a gene (probe set). Genes were clustered according to Pearson correlation, and the structure of the gene tree is shown. Color scheme used to identify sample characteristics is shown between panels A and B. NK indicates normal karyotype; AK, abnormal karyotype. (C-D) Affymetrix (C) and reverse transcription–quantitative polymerase chain reaction (RT-qPCR) (D) analysis of expression levels of 20 genes (with the highest scores among the 369 predictors) evaluated in 16 patients (8 NPMc⁺ and 8 NPMc⁻, identified by numbers). (D) Relative expression levels are calculated as deviation from the median, and expression values for each gene in each sample are calculated as $2^{-\Delta\text{CT}}$ (ΔCT = difference between the mean threshold cycle for each specific gene and for the 18S control ribosomal RNA gene). In the vast majority of cases, RT-qPCR reflects the results expected from microarray analysis. Homeobox gene expression levels appear to be particularly elevated in NPMc⁺ AML. The color bar (D, right) represents the color scheme applied to all parts of the figure.



hierarchical clustering showed that, strikingly, the strongest clustering parameter was NPM status (Figure 1A). FAB subtype determined partial subclustering, (Figure 1A) with 1 of the 2 major branches of NPMc⁺ samples containing more M4 and M5 cases, whereas *FLT3* mutations did not show specific subclustering. To identify genes that best discriminate the NPMc⁺ from NPMc⁻ AMLs, we performed a supervised analysis of variance (ANOVA) and identified 369 probe sets (Table S3), corresponding to 330 nonredundant genes. An independent method to identify discriminating genes (Significance Analysis of Microarrays⁹) generated largely overlapping results (Table S4).

We used the 369 putative NPMc⁺ predictors to study the test set of AMLs, which included 29 NPMc⁺ and 10 NPMc⁻ AML (Table 1), and, like the training set, was heterogeneous for FAB subtype and *FLT3* mutations. This group of samples, however, also included 6 of 29 NPMc⁺ cases with rare chromosomal abnormalities (add(1), del(11), inv(3), del(9), and trisomy 8 in 2 cases).

Hierarchical clustering efficiently segregated patients with NPMc⁻ from patients with NPMc⁺, suggesting that these 369 genes reliably recapitulate the gene expression profile of NPMc⁺ AMLs, even in cases with rare chromosomal abnormalities (Figure 1B).

Levels of NPM1 mRNA do not differ in the 2 groups of samples (not shown), indicating that NPM1 mutations are not accompanied by quantitative differences in NPM1 expression. CD34 and CD133/prominin-1, which are rarely expressed in NPMc⁺ AML⁴ (Supplemental Table S2), are strongly repressed in NPMc⁺ cases and appear among the predictors (Supplemental Table S3), supporting the reliability of our results. For further validation, we analyzed by real-time RT-PCR the expression levels of other 20 predictor genes, chosen for function (relevance in hematopoiesis and/or cell differentiation) and predictive strength. Figure 1D shows analysis of 16 AMLs (8 NPMc⁺ and 8 NPMc⁻). Comparison of RT-PCR and microarray data resulted in a large overlap (Figure 1C).

Previous studies demonstrated that AMLs with recurrent chromosomal rearrangements show distinct gene-expression signatures, while AMLs with normal karyotype segregate within 2 or more clusters,¹⁰⁻¹³ none of which carries a unifying genetic lesion. Our analysis identifies, within AMLs with normal karyotype, a distinct subgroup unambiguously characterized by cytoplasmic dislocation of NPM and *NPM* gene mutations. Notably, the NPMc⁺ cluster also contained samples with rare chromosomal abnormalities, reinforcing the concept that NPMc⁺ AML represents a distinct subgroup regardless of the karyotype. Non-major chromosomal rearrangements rarely accompanying *NPM* mutations are, therefore, likely to be secondary events.⁴

A striking feature of NPMc⁺ gene-expression signature is the activation of numerous members of the homeodomain-containing family of transcription factors, including *HOX* and *TALE* genes, some of which are oncogenes in myeloid leukemias and implicated in hematopoietic development.¹⁴ Several *HOX* genes are highly expressed in hematopoietic stem cells (HSCs), and their expression decreases with differentiation.¹⁵ Their concerted overexpression in NPMc⁺ AML blasts may, therefore, contribute to the maintenance of a stem-cell phenotype. Notably, NPMc⁺ AMLs also display induction of the Notch1-ligand *JAG1* and repression of *CDKN2C/p18-INK4C*, which are associated to expansion of the HSC pool.^{16,17} Repression of the HSC-associated genes *CD34* and *CD133/PROM1* is not necessarily in contrast with this view, since a HSC subpopulation negative for both lineage- and HSC-associated markers has been identified.¹⁸ Consistent with the view that NPMc⁺ AMLs might derive from a HSC, they show a wide FAB morphologic spectrum and frequent multilineage involvement.⁴

A similar *HOX* signature was previously reported in an uncharacterized subset of AML with normal karyotype.¹¹ We demonstrate

that this is a specific feature of NPMc⁺ AMLs. Acute leukemias carrying *MLL* rearrangements also display induction of *HOX* genes,¹⁹ possibly due to direct binding of mixed lineage leukemia (*MLL*) fusion proteins to *HOX* promoters.²⁰ A similar mechanism is not likely for NPMc⁺ AML: NPM is a nuclear chaperone²¹ that regulates diverse processes (such as assembly and transport of preribosomal particles, and centrosome duplication^{22,23}); interacts with tumor suppressors proteins p53, p19, and retinoblastoma pRB; and is crucial for p53 stabilization after stress.²⁴⁻²⁷ Homeobox activation in NPMc⁺ AML might, therefore, reflect the molecular status of the leukemic target cell rather than represent a direct consequence of NPM mutations.

Appendix

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